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A New Class of Modular P,N-Ligand Library for Asymmetric Pd-Catalyzed Allylic Substitution Reactions: A Study of the Key Pd $-\pi$ -Allyl Intermediates

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Abstract: A new class of modular P,Nligand library has been synthesized and screened in the Pd-catalyzed allylic substitution reactions of several substrate types. These series of ligands can be prepared efficiently from easily accessible hydroxyl–oxazole/thiazole derivatives. Their modular nature enables the bridge length, the substituents at the heterocyclic ring and in the alkyl backbone chain, the configuration of the ligand backbone, and the substituents/configurations in the biaryl phosphite moiety to be easily and systematically varied. By carefully selecting the ligand components, therefore, high regio- and enantioselectivities (*ee*

Keywords: allylic substitution • asymmetric catalysis • ligand design • N,P ligands • palladium values up to 96%) and good activities are achieved in a broad range of mono-, di-, and trisubstituted linear hindered and unhindered substrates and cyclic substrates. The NMR spectroscopic and DFT studies on the Pd– π -allyl intermediates provide a deeper understanding of the effect of ligand parameters on the origin of enantioselectivity.

Introduction

The development of methods for enantioselective carbon– carbon and carbon–heteroatom bond formation is one of the key issues in organic synthesis. A versatile method for forming these bonds is palladium-catalyzed asymmetric allylic substitution.^[1]

Most of the successful ligands reported to date for this process have been designed using three main strategies. The first one, developed by Hayashi and co-workers, was the use of a secondary interaction of the nucleophile with a side chain of the ligand to direct the approach of the nucleophile to one of the allylic terminal carbon atoms.^[2] The second one, developed by Trost and co-workers, was to increase the

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bite angle of the ligand to create a chiral cavity in which the allyl system is embedded. This idea opened up the successful application of ligands with large bite angles for the allylic substitution of sterically undemanding substrates.^[1,3] The third strategy, developed by groups led by Helmchen, Pfaltz, and Williams, was the use of heterodonor ligands that result in an electronic discrimination of the two allylic terminal carbon atoms due to the different *trans* influences of the donor groups.^[1,4] This made it possible to successfully use a wide range of heterodonor ligands (mainly P,N ligands) in allylic substitution reactions.^[1]

Nowadays, many chiral ligands (mainly P and N ligands), which possess either C_2 or C_1 symmetry, have been developed, and they provide high enantiomeric excesses for several types of disubstituted substrates.^[1] Nevertheless, in general, there is still a problem of substrate specificity (for example, ee values are high in disubstituted linear hindered substrates and low in unhindered substrates, and vice versa) and reaction rates. Other types of substrates still require much attention. For example, for monosubstituted substrates, more active and more regio- and enantioselective Pd catalysts are needed.^[1] Another challenging class of substrates is that of trisubstituted substrates. Although a few good enantioselective Pd-catalytic systems have been reported, their activities are still very low.^[1] More research is needed on the development of new ligands that can overcome these limitations.



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In this context, we recently demonstrated that the presence of biaryl phosphite moieties in ligand design is highly advantageous because 1) substrate specificity decreases because the chiral pocket created (i.e., the chiral cavity in which the allyl is embedded) is flexible enough to enable the perfect coordination of hindered and unhindered substrates,^[5] 2) reaction rates increase thanks to the larger π -acceptor ability of these moieties,^[6] and 3) regioselectivity towards the desired branched isomer in monosubstituted linear substrates increases, thanks to the π -acceptor ability of the phosphite moiety that enhances the S_N1 character of the nucleophilic attack.^[7]

Due to our interest in discovering faster and more versatile Pd-catalytic systems, we decided to go one step further in the design of a new ligand library for this process. Therefore, we developed a ligand library, the design of which in-

corporates the advantages of heterodonor and biaryl phosphite ligands and also allows extra control of the flexibility of the chiral pocket by changing the size of the chelate ring. To do this, we synthesized and screened a library of 56 potential new phosphite-oxazole/ thiazole ligands.^[8] The highly modular construction of these ligands enabled a systematic study of the effect of bridge length (ligands L1 and L6), the substituent at the heterocyclic ring (ligands L1-L4) and in the alkyl backbone chain (ligands L6 and L7), the configuration of the ligand backbone (ligands L1 vs. L5 and L7 vs. L8), and the substituents and configurations in the biaryl phosphite moiety (**a**–**g**). By carefully selecting these elements, we achieved high selectivities (regioand enantioselectivities) and activities in a wide range of mono-, di-, and trisubstituted substrates. In this paper we also discuss the synthesis and characterization of the Pd– π allyl intermediates to provide greater insight into the origin of enantioselectivity in these catalytic systems.

Results and Discussion

Synthesis of the ligand library: Scheme 1 illustrates the sequence of ligand synthesis. Ligands L1–L8a–g were synthesized very efficiently from the corresponding easily accessible ketone–oxazole or thiazole–ester derivatives (2–5 and 12, Scheme 1). Compounds 2–5 and 12 are easily made in



Scheme 1. Synthesis of a new phosphite–nitrogen ligand library **L1–L8a–g**. a), b) see ref. [9a] and the Experimental Section; c), d) see ref. [9b]; e) CH₃MgCl/THF/CeCl₃ (yield: 71 %); f) ClP(OR)₂; (OR)₂=**a–g**/pyridine/toluene (yields: 42–76 %).



two steps from the corresponding dimedone 1 and ketoester 11, respectively (see the Experimental Section).^[9] Ketoneoxazoles 2-5 were then reduced using (R)-Me-CBS or $NaBH_4$ (CBS = oxazaborolidine; Scheme 1, step b). Enantioselective reduction of 2 using (R)-Me-CBS followed by single recrystallization afforded hydroxyl-oxazole 6 in >99% ee.^[9a] The same procedure applied to ketones 3 and 4 afforded hydroxyl-oxazoles 8 and 9 but with ee values of <80%. Thus, further enantiomer resolution was achieved using preparative chiral HPLC. For compounds 7 and 10, the corresponding ketone-oxazoles 2 and 5 were reduced using NaBH₄ followed by enantiomer resolution using preparative HPLC. Reduction of 12 using LiAlH₄ (Scheme 1, step d)^[9b] or MeMgCl (Scheme 1, step e) gave good yields of the corresponding racemic alcohols 13 and 14, respectively. This was followed by the resolution of racemates into their enantiomers by preparative chiral HPLC.^[9]

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The last step of the ligand synthesis is common to all of them (Scheme 1, step f). Therefore, treatment of the corresponding hydroxyl-oxazole (6-10) or hydroxyl-thiazole (13 and 14) with 1 equiv of the corresponding in situ formed biaryl phosphorochloridite $(CIP(OR)_2; (OR)_2 = a-g)$ in the presence of pyridine, in a parallel manner, provided easy access to the desired ligands L1-L8a-g (see the Experimental Section for details). All of the ligands were stable during purification on neutral alumina under an atmosphere of argon and isolated in moderate-to-good yields as white solids. They were stable at room temperature and very stable to hydrolysis. The elemental analyses were in agreement with the assigned structure. The ¹H, ¹³C, and ³¹P NMR spectra were as expected for these C_1 ligands. One singlet for each compound was observed in the ³¹P NMR spectrum (see the Experimental Section). Rapid ring inversions (atropoisomerization) in the biphenylphosphorus moieties (a-c) occurred on the NMR spectroscopic timescale because the expected diastereoisomers were not detected by low-temperature phosphorus NMR spectroscopy.^[10]

Allylic substitution of symmetrical 1,3-disubstituted allylic substrates: In this section, we report the use of the chiral phosphite-nitrogen ligand library (L1–L8a–g) in the Pd-catalyzed allylic substitution of linear disubstituted substrates with different steric properties [Eq. (1a)]: *rac*-1,3-diphenyl-3-acetoxyprop-1-ene (S1) (widely used as a model substrate), *rac*-(*E*)-ethyl-2,5-dimethyl-3-hex-4-enylcarbonate (S2), and *rac*-1,3-dimethyl-3-acetoxyprop-1-ene (S3); and cyclic substrates [Eq. (1b)]: *rac*-3-acetoxycyclohexene (S4) (widely used as a model substrate), *rac*-3-acetoxycycloheptene (S5), and *rac*-3-acetoxycyclopentene (S6). Two nucleophiles were tested. In all cases, the catalysts were generated in situ from π -allyl-palladium chloride dimer [{PdCl(η^3 -C₃H₅)}₂] and the corresponding ligand.^[1]



Allylic substitution of S1 using dimethyl malonate and benzylamine as nucleophiles: In the first set of experiments, we used the palladium-catalyzed asymmetric substitution reactions of S1 [Eq. (1 a); R=Ph, X=OAc], with dimethyl malonate and benzylamine as nucleophiles, to study the potential of the phosphite–nitrogen ligand library L1-L8a-g. Compound S1 was chosen as a substrate because the reaction was performed with a wide range of ligands, which enabled the efficiency of the various ligand systems to be compared directly.^[1]

First, we studied the effect of the reaction conditions by conducting a series of experiments with two ligands (L1a and L6a) using different solvents (tetrahydrofuran, toluene, and dichloromethane) and ligand-to-palladium ratios (L/Pd=0.75, L/Pd=1, and L/Pd=2). We found that the efficiency of the process was strongly dependent on the nature of the solvent and the ligand-to-palladium ratio (Table 1).

Table 1. Selected results for the Pd-catalyzed allylic alkylation of **S10** by using ligand library **L1–L8a–g**. Effects of the solvent and the ligand-to-palladium ratio.^[a]

Entry	Ligand	Solvent	L/Pd	Conv [%] (t [min]) ^[b]	ee [%] ^[c]
1	L1a	CH_2Cl_2	1	100 (30)	82 (<i>S</i>)
2	L1 a	THF	1	89 (60)	73 (S)
3	L1 a	toluene	1	100 (1440)	92 (S)
4	L6 a	CH_2Cl_2	1	100 (30)	21 (S)
5	L6 a	THF	1	85 (180)	18 (S)
6	L6 a	toluene	1	64 (600)	34 (S)
7	L1 a	CH_2Cl_2	0.75	100 (60)	82 (S)
8	L6 a	CH_2Cl_2	2	100 (30)	79 (S)
9	L6 a	CH_2Cl_2	0.75	73 (30)	23 (S)
10	L6 a	CH_2Cl_2	2	100 (30)	14 (S)

[a] All reactions were run at 23 °C; 0.5 mol% [{PdCl(η^3 -C₃H₅)}₂]; **S1** (0.5 mmol); BSA (*N*,*O*-bis(trimethylsilyl)acetamide) (1.5 mmol); dimethyl malonate (1.5 mmol). [b] Conversion measured by ¹H NMR spectroscopy. Reaction time shown in parentheses. [c] Enantiomeric excesses measured by HPLC. Absolute configuration shown in parentheses.

Although in toluene the enantioselectivity was higher than dichloromethane, the activity was much lower (Table 1, entries 3 and 6 vs. entries 1 and 4). Tetrahydrofuran yielded the lowest enantioselectivities of all three solvents (entries 2 and 5). We also found that an excess of ligand was not needed for enantioselectivities to be high (entries 1, 4, and 7–10). Interestingly, at higher ligand-to-palladium ratios, enantioselectivities were lower (entries 8 and 10 vs. entries 1 and 4). This is probably due to the fact that at a ligand-to-palladium ratio greater than 1, the phosphite–nitrogen ligands act as a monodentate ligand.^[11]

For comparison purposes, the rest of the ligands were tested using dichloromethane as a solvent and at a ligandto-palladium ratio of 1. Table 2 shows the results obtained when dimethyl malonate and benzylamine were used as nucleophiles. We found that enantioselectivities were highly affected by the bridge length, the substituents at the heterocyclic ring and in the alkyl backbone chain, and the substituents and configurations in the biaryl phosphite moieties (**a**– **g**). High activities (turnover frequencies (TOFs) up to $600 \text{ mol } \text{S1} \times (\text{mol } \text{Pd} \times \text{h})^{-1}$) and enantioselectivities (*ee* values up to 92%) were obtained for both enantiomers of the substitution products **15** and **16** using ligands **L1a** and **L5a**. Catalytic performance in the Pd-catalyzed allylic ami-

Table 2.	Selected results for	the Pd-catalyzed	allylic substitution	of S1 by	using ligand	library L1-L8a-g. ^[a]
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		H-Nu=H-CH(CO	$H-Nu = H-CH(COOMe)_2^{[a]}$		H-Nu=H-NHCH ₂ Ph ^[a]	
Entry	Ligand	Conv [%] (t [min]) ^[b]	ee [%] ^[c]	Conv [%] (t [min]) ^[b]	ee [%] ^{[4}	
1	L1a	100 (15) ^[d]	82 (S)	34 (360)	84 (R)	
2	L1b	99 (20)	81 (S)	37 (360)	80 (R)	
3	L1c	99 (15)	70 (S)	43 (360)	69 (R)	
4	L1d	100 (30) ^[d]	69 (S)	28 (360)	71 (R)	
5	L1e	$100(30)^{[d]}$	40 (S)	31 (360)	41 (R)	
6	L1 f	72 (30)	9 (S)	15 (360)	7(R)	
7	L1g	42 (30)	8 (R)	12 (360)	5 (S)	
8	L2a	80 (30)	43 (S)	24 (360)	32 (R)	
9	L3a	98 (30) ^[d]	79 (S)	36 (360)	82 (R)	
10	L4a	78 (30)	22(S)	19 (360)	25(R)	
11	L5a	100 (15)	81 (R)	32 (360)	84 (S)	
12	L6 a	$100(20)^{[d]}$	21 (S)	41 (360)	19 (R)	
13	L7a	100 (30) ^[d]	52 (S)	38 (360)	49 (R)	
14	L8a	100 (30)	51 (R)	33 (360)	50 (S)	
15 ^[e]	L1a	100 (360)	92 (<i>S</i>)	_	-	

[a] All reactions were run at 23 °C; 0.5 mol % [{PdCl(η^3 -C₃H₅)}₂]; dichloromethane as solvent; 1 mol % ligand. [b] Conversion measured by ¹H NMR spectroscopy. Reaction time shown in parentheses. [c] Enantiomeric excesses measured by HPLC. Absolute configuration shown in parentheses. [d] Isolated yields of **15** were >93 %. [e] Toluene as solvent.

nation of **S1** followed the same trend as for the allylic alkylation of **S1** (Table 2). As expected, however, the activity was lower than in the alkylation reaction of **S1**. The stereoselectivity of the amination was the same as for the alkylation reaction, though the Cahn–Ingold–Prelog (CIP) descriptor was inverted because the priority of the groups had changed.

The influence of the bridge length indicates that the use of ligands **L1** and **L5**, which form a six-membered chelate ring, provided higher enantioselectivity than the use of ligands **L6–L8**, which form a seven-membered chelate ring (Table 2, entries 1 and 11 vs. entries 12–14). In line with this, the increase of the rigidity of the ligand by replacing the hydrogen substituents in the alkyl backbone chain in ligand **L6a** with two methyl groups (ligand **L7a**) caused enantioselectivities to increase (Table 2, entries 12 vs. 13).

The effect of the substituents at the heterocyclic ring indicated that the presence of either bulky or electron-donating substituents decreased both activities and enantioselectivities (Table 2, entries 1, 8–10). The fact that enantiomeric excesses decrease when a bulky *tert*-butyl is present is due to the formation of Pd intermediate species with the phosphite–nitrogen ligand acting as a monodentate ligand (see below).

Regarding the effect of the substituents at the biphenyl phosphite moiety, we found that the presence of bulky *tert*butyl substituents at both the *ortho* and *para* positions is highly adventitious in terms of activity and enantioselectivity (Table 2, entry 1). Therefore, the presence of methoxy groups in the *para* position of the biphenyl moieties has a negative effect on activity, whereas the presence of trimethylsilyl substituents at the *ortho* positions has a negative effect on enantioselectivity (Table 2, entries 2 and 3 vs. entry 1). With ligands **L1d** and **L1e**, which contain different enantiomerically pure binaphthyl moieties, we found that there is a cooperative effect between the configuration of

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the biaryl moiety and the configuration of the ligand backbone on enantioselectivity. This led to a matched combination for ligand L1d, which contains (*S*)-binaphthyl an moiety (Table 2, entries 4 and 5). In addition, by comparing the results obtained using ligand L1c with those of the related binaphthyl ligands L1d and L1e (Table 2, entries 3-5), we can also conclude that the atropoisomeric biphenyl moiety in ligands L1a-c adopts an S configuration when coordinated in the Pd-π-allyl intermediate species.

To sum up, the best result was obtained with ligands **L1a** and **L5a**, which contain the optimal combination of ligand pa-

rameters (Table 2, entries 1 and 11). These findings clearly show the efficiency of highly modular scaffolds in ligand design. Enantioselectivity can be further improved by controlling not only the structural but also the reaction parameters. As expected, changing the solvent from dichloromethane to toluene increased enantioselectivity (*ee* values up to 92%, Table 2, entry 15).

Allylic substitution of S2 using dimethyl malonate as nucleophile: We also screened the phosphite-nitrogen ligand library L1–L8a–g in the allylic alkylation process of S2 using dimethyl malonate as nucleophile [Eq. (1a); R = iPr, X =OCO₂Et]. This substrate is more sterically demanding than substrate S1, which was used previously.^[1] If enantiomeric excesses are to be high, the ligand must create a slightly bigger chiral pocket (the chiral cavity in which the allyl is embedded) around the metal center to be able to accommodate the sterically demanding isopropyl substituents.^[1] Due to the flexibility conferred by the biaryl phosphite moiety, we expected to obtain good enantioselectivities for this substrate as well. Table 3 shows the most representative results. In general, the trends were the same as for the allylic substitution of S1. Again, both enantiomers of the alkylation product 17 were accessible in high enantioselectivities (ee values up to 93%) when catalyst precursors containing ligands L1a and L5a were used (Table 3, entries 1, 9, 13, and 14). As expected, the activities were lower than in the alkylation reaction of **S1**.^[1] The stereoselectivity of the alkylation of **S2** was the same as for the alkylation reaction of **S1**, though the CIP descriptor was inverted because of the change in the priority of the groups.

Allylic substitution of S3 using dimethyl malonate as nucleophile: We also tested ligands L1–L8a–g in the allylic substitution of the linear substrate S3 [Eq. (1a); R=Me, X= OAc]. Substrate S3 is less sterically demanding than sub-

Table 3. Selected results for the Pd-catalyzed allylic substitution of S2 by using ligand library $L1\!-\!L8a\!-\!g.^{[a]}$

Entry	Ligand	Conv [%] (t [h]) ^[b]	ee [%] ^[c]
1 ^[d]	L1 a	100 (24)	84 (R)
2	L1b	95 (24)	82 (R)
3	L1 c	100 (24)	73 (R)
4	L1 d	78 (24)	67 (R)
5	L1e	82 (24)	39 (R)
6	L2 a	75 (24)	47 (R)
7 ^[d]	L3 a	100 (24)	77 (R)
8	L4 a	69 (24)	60(R)
9	L5 a	100 (24)	83 (S)
10 ^[d]	L6 a	100 (24)	12(R)
11	L7 a	100 (24)	24(R)
12	L8 a	100 (24)	23(S)
13 ^[e]	L1 a	99 (48)	93 (R)
14 ^[e]	L5 a	94 (48)	92 (S)

[a] All reactions were run at 23 °C; 1 mol % [{PdCl(η^3 -C₃H₅)}₂]; dichloromethane as solvent; 2 mol % ligand. [b] Conversion measured by ¹H NMR spectroscopy. Reaction time shown in parentheses. [c] Enantiomeric excesses measured by ¹H NMR spectroscopy using [Eu(hfc)₃]. Absolute configuration shown in parentheses. [d] Isolated yields of **17** were >92 %. [e] Toluene as solvent.

strates **S1** and **S2**, used previously. There are therefore fewer successful catalyst systems for the Pd-catalyzed allylic substitution of this substrate than for the allylic substitution of hindered substrates such as **S1** and **S2**.^[3b,5,6e,11b,12] If enantiomeric excesses are to be high, the ligand must create a small chiral pocket around the metal center, mainly because of the presence of less sterically demanding methyl *syn* substituents.^[1] Due to the flexibility conferred by the biaryl phosphite moiety in combination with the possibility of changing the size of the chelate ring in the ligand, we expected to adequately modulate the chiral pocket to obtain good enantioselectivities for this substrate as well.

Preliminary investigations into the solvent and ligand-topalladium ratio revealed a different trend in solvent effect than with the previously tested substrates **S1** and **S2**. Enantioselectivities and activities were both at their highest when dichloromethane was used and the ligand-to-palladium ratio was 1 (see the Supporting Information).

Table 4 shows the results obtained using the ligand library L1-L8a-g in optimized conditions. We were able to finetune the ligands to obtain high activities and enantioselectivities (ee values up to 92%) in the alkylation of this substrate. Again, activities and enantioselectivities were affected by the bridge length, the substituent at the nitrogen heterocycle, and the substituents and configurations in the biaryl phosphite moiety. However, the effect of these parameters was different from their effect on substitution of hindered substrates S1 and S2. Enantioselectivities were best with ligands L7d and L8d (Table 4, entries 16, 17, and 19). Once again, it was possible to access both enantiomers of the substitution product 18. These results, which again clearly show the efficiency of using modular scaffolds in ligand design, are among the best reported for this type of unhindered substrate.^[3b,5,6e,11b,12]

Table 4. Selected results for the Pd-catalyzed allylic substitution of S3 by using ligand library $L1-L8\,a-g.^{\rm [a]}$

Entry	Ligand	Conv [%] (t [h]) ^[b]	ee [%] ^[c]
1 ^[d]	L1 a	100 (1)	59 (S)
2	L1b	100 (1)	67 (S)
3	L1 c	100 (1)	54 (S)
4 ^[d]	L1d	100 (2)	75 (S)
5 ^[d]	L1e	100 (2)	51 (S)
6	L1 f	82 (2)	30 (S)
7	L1g	63 (2)	21 (S)
8 ^[d]	L2 a	100 (2)	60(S)
9 ^[d]	L2 e	100 (2)	74 (S)
10 ^[d]	L3 a	100 (2)	59 (S)
11 ^[d]	L3e	100 (2)	75 (S)
12 ^[d]	L4 a	100 (2)	5(S)
13	L5a	100 (1)	57 (R)
14	L6 a	92 (2)	38 (S)
15	L7 a	93 (3)	80 (S)
16	L7d	100 (4)	87 (S)
17	L8 d	100 (4)	87 (R)
18 ^[d,e]	L1 d	100 (20)	84 (S)
19 ^[e]	L7 d	96 (24)	92 (<i>S</i>)

[a] All reactions were run at 23 °C; 0.5 mol % [{PdCl(η^3 -C₃H₅)}₂]; dichloromethane as solvent; 1 mol % ligand. [b] Conversion measured by GC. Reaction time shown in parentheses. [c] Enantiomeric excesses measured by GC. Absolute configuration shown in parentheses. [d] Isolated yields of **18** were >91 %. [e] T=0 °C.

Regarding the effect of ligand flexibility, in contrast to **S1** and **S2**, the highest enantioselectivities were obtained with ligands **L7** and **L8**, which form a seven-membered chelate ring and contain two methyl groups at the alkyl backbone chain. Concerning the effect of the substituents at the heterocyclic ring and the biaryl phosphite moiety, in contrast to **S1** and **S2**, the presence of aryl substituents in the heterocyclic moiety and a bulky (*S*)-binaphthyl phosphite moiety had a positive effect on enantioselectivities. In conclusion, our results indicate that both the size of the chelate ring and the flexibility of the biaryl phosphite moiety are the main ligand parameters that control the size of the chiral pocket to achieve high enantioselectivities.

Allylic alkylation of cyclic substrates S4–S6: With the unhindered cyclic substrates S4–S6, enantioselectivity is difficult to control, mainly because of the presence of less sterically demanding *anti* substituents. These *anti* substituents are thought to play a crucial role in the enantioselection observed with cyclic substrates in the corresponding Pd–allyl intermediates.^[1]

In this section, we show that the chiral ligand library L1– L8 a-g that was applied previously to the Pd-catalyzed allylic substitution of 1,3-disubstituted linear substrates (S1–S3) can also be used for cyclic substrates (*ee* values up to 88%). We tested three cyclic substrates [Eq. (1b)]: *rac*-3-acetoxycyclohexene (S4) (which is widely used as a model substrate), *rac*-3-acetoxycycloheptene (S5), and *rac*-3-acetoxycyclopentene (S6).

Preliminary investigations into the solvent effect and ligand-to-palladium ratio showed the same trends as with the previously tested unhindered linear substrate **S3**. The

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tradeoff between enantioselectivities and reaction rates was therefore optimum with dichloromethane and a ligand-topalladium ratio of 1 (see the Supporting Information).

Table 5 shows the results of using the ligand library L1– L8a–g under the optimized conditions. We also obtained high activities and enantioselectivities (up to 88%) in the al-

Table 5. Selected results for the Pd-catalyzed allylic substitution of S4–S6 by using ligand library L1–L8a–g.^[a]

Entry	Substrate	Ligand	Conv [%] (t [h]) ^[b]	ee [%] ^[c]
1	S4	L1 a	100 (6)	41 (S)
2	S4	L1b	100 (6)	59 (S)
3	S4	L1 c	100 (6)	43 (S)
4 ^[d]	S4	L1 d	100 (6)	80 (S)
5 ^[d]	S4	L1 e	100 (6)	85 (R)
6	S4	L2 a	100 (6)	41 (S)
7	S4	L3 a	100 (6)	45 (S)
8 ^[d]	S4	L3 e	100 (6)	84 (R)
9 ^[d]	S4	L4 a	100 (6)	60 (S)
10	S4	L5 a	100 (6)	39 (R)
11	S4	L6 a	100 (6)	13 (S)
12	S4	L7a	100 (6)	29 (S)
13	S4	L7d	100 (6)	63 (S)
14	S 5	L1d	100 (24)	83 (S)
15 ^[d]	S 5	L1 e	100 (24)	88 (R)
16	S 6	L1 d	100 (6)	72 (S)
17	S 6	L1 e	100 (6)	71 (R)

[a] All reactions were run at 23 °C; 0.5 mol % [{PdCl(η^3 -C₃H₅)}₂]; dichloromethane as solvent; 1 mol % ligand. [b] Conversion measured by GC. Reaction time shown in parentheses. [c] Enantiomeric excesses measured by GC. Absolute configuration shown in parentheses. [d] Isolated yields of **19** and **20** were >91%.

lylic substitution of the cyclic substrates S4-S6 using ligands L1d, L1e, and L3e. The results indicate that the effect of the ligand parameters on the catalytic performance are different from those observed for the linear substrates S1-S3. In contrast to the alkylation of unhindered linear S3, ligands that form a six-membered chelate ring (L1) provided higher enantioselectivities than those that form a seven-membered chelate ring (L6, L7). The results also indicate that the presence of an enantiopure bulky binaphthyl phosphite moiety (d and e) is therefore required for enantioselectivity to be high (Table 5, entries 4, 5, 8, 14, and 15). Interestingly, the sense of enantioselectivity is also governed by the configuration of the biaryl phosphite moiety. Thus, both enantiomers of the substitution products 19-21 can be accessed in high enantioselectivities by simply changing the configuration of the trimethylsilyl-substituted binaphthyl moiety (Table 5, entries 4 and 5).

In summary, the enantioselectivities obtained with ligands **L1d**, **L1e**, and **L3e** are among the best reported for this type of 1,3-disubstituted cyclic substrate.^[1e,3a,5a,11b,12a,13] Interestingly, compared with the hindered substrates **S1** and **S2** and in contrast to the unhindered linear substrate **S3**, the flexibility conferred by the biaryl phosphite moiety was enough to adequately control the size of the chiral pocket to achieve high enantioselectivities.

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Allylic substitution of unsymmetrical 1,3,3-trisubstituted allylic substrate: We also screened the ligands L1-L8a-g in the allylic substitution of rac-1,3,3-triphenylprop-2-enyl acetate (S7) and rac-1,1-diphenyl-1-hepten-3-yl acetate (S8) using dimethylmalonate as nucleophile [Eq. (2); R = R' =R'' = Ph for **S7** and R = R' = Ph, R'' = Me for **S8**]. These substrates are of synthetic interest because the substitution products formed in this way can easily be transformed into enantiomerically enriched acid derivatives and lactones.^[14] They are more sterically demanding than the previously used substrate S1,^[1] and it is therefore more difficult to achieve excellent enantioselectivities with them than with S1.^[12,15] Interestingly, with this P,N-ligand library, we obtained high enantiomeric excesses (ee values up to 96%) under standard reaction conditions. Although, as expected, the activities were lower than in the alkylation reaction of S1, they were much higher than those obtained with other successful ligands under similar reaction conditions.[15]

The results, summarized in Table 6, followed a trend similar to that of the more hindered substrates **S1** and **S2**. Thus, ligands **L1** and **L5**, which form a six-membered chelate ring and have a phenyl substituent at the nitrogen heterocycle, provided better enantioselectivities than ligands **L2–L4** and **L6–L8**. The presence of bulky substituents at the *ortho* positions of the biaryl phosphite moiety was necessary for enantioselectivities to be high. However, in contrast to **S1** and **S2**, enantioselectivities were less affected by the presence or lack of bulky substituents at the *para* positions of the biaryl phosphite moiety. Therefore, ligands **L1a–c** and **L5a–c** pro-

Table 6. Selected results for the Pd-catalyzed allylic substitution of S7 and S8 by using ligand library L1-L8a-g.^[a]

Entry	Ligand	S7 Conv [%] (<i>t</i> [h]) ^[b]	ee [%] ^[c]	S8 Conv [%] (<i>t</i> [h]) ^[b]	ee [%] ^[c]
1 ^[d]	L1a	92 (24)	88 (R)	100 (24)	95 (R)
2	L1b	88 (24)	95 (R)	100 (24)	94 (R)
3	L1 c	79 (24)	96 (R)	100 (24)	95 (R)
4	L1 d	46 (24)	42 (R)	53 (24)	32 (R)
5	L1 g	33 (24)	18 (R)	48 (24)	21(R)
6	L2 a	90 (24)	81 (R)	100 (24)	78 (R)
7	L3 a	88 (24)	90 (R)	100 (24)	85 (R)
8	L4 a	45 (24)	23 (R)	85 (24)	38 (R)
9	L5 a	88 (24)	90 (S)	100 (24)	95 (S)
10	L6 a	67 (24)	40 (R)	73 (24)	57 (R)
11	L7 a	29 (24)	30 (R)	46 (24)	44 (R)
12	L8 a	27 (24)	30 (S)	42 (24)	42 (<i>S</i>)

[a] All reactions were run at 23 °C; 1 mol % [{PdCl(η^3 -C₃H₅)}₂]; dichloromethane as solvent; 2 mol % ligand. [b] Conversion measured by ¹H NMR spectroscopy. Reaction time shown in parentheses. [c] Enantiomeric excesses measured by ¹H NMR spectroscopy. Absolute configuration shown in parentheses. [d] Isolated yields of **22** and **23** were >88 %. vided excellent enantiocontrol in the allylic substitution of the trisubstituted substrates **S7** and **S8**, and also gave access to both enantiomers of the alkylation products **22** and **23** (*ee* values up to 96%). These results are among the best reported for this class of substrate.^[15]

Allylic substitution of unsymmetrical 1- or 3-monosubstituted allylic substrates: To further study the potential of these readily available ligands, we tested L1–L8a–g in the regioand stereoselective allylic alkylation of 1-(1-naphthyl)allyl acetate (S9) and 1-(1-naphthyl)-3-acetoxyprop-1-ene (S10) with dimethyl malonate as nucleophile [Eq. (3)].



For these substrates, not only does the enantioselectivity of the process need to be controlled but regioselectivity is also a problem because a mixture of regioisomers can be obtained. Most Pd catalysts developed to date favor the formation of achiral linear product **25** rather than the desired branched isomer **24**.^[16] The development of highly regioand enantioselective Pd catalysts is therefore still important.^[5a,7,11b,17]

Table 7 summarizes the results obtained with the ligand library L1–L8a–g. High activities and enantioselectivities (of

Table 7. Selected results for the Pd-catalyzed allylic alkylation of monosubstituted substrates **S9** and **S10** by using ligand library **L1–L8a–g** under standard conditions.^[a]

Entry	Ligand	Substrate	Conv [%] (t [min]) ^[b]	24/25 ^[c]	ee [%] ^[d]
1	L1a	S9	100 (30)	70:30	20 (R)
2	L1b	S9	100 (30)	40:60	63 (R)
3	L1c	S9	100 (30)	55:45	70 (R)
4	L1 d	S9	100 (30)	50:50	78 (R)
5	L1 e	S9	100 (30)	55:45	34 (R)
6 ^[e]	L2 a	S9	100 (30)	80:20	45 (R)
7	L2 d	S9	100 (30)	75:25	88 (R)
8	L3 a	S9	100 (30)	75:25	19 (R)
9	L3 c	S9	100 (30)	70:30	73 (R)
10	L4 a	S9	100 (30)	75:25	25 (R)
11	L6 a	S9	100 (30)	45:55	2(S)
14	L7 a	S9	100 (30)	80:20	92 (R)
15	L7d	S9	100 (30)	60:40	90 (R)
16	L1 d	S10	100 (30)	50:50	79 (R)
17	L3c	S10	100 (30)	70:30	72 (R)
18	L2 d	S10	100 (30)	75:25	87 (R)
19	L7a	S10	100 (30)	80:20	92 (R)

[a] All reactions were run at 23 °C; 1 mol% [{PdCl($(\eta^3-C_3H_5)_2$]; dichloromethane as solvent; 2 mol% ligand; substrate (0.5 mmol); BSA (1.5 mmol); dimethyl malonate (1.5 mmol). [b] Reaction time in minutes shown in parentheses. [c] Percentage of branched (24) and linear (25) isomers. [d] Enantiomeric excesses of 24 determined by HPLC. Absolute configuration shown in parentheses. [e] Isolated yield of 24 was >72%.

up to 92%) combined with regioselectivities of up to 80% in favor of the branched product **24** were obtained under standard reaction conditions. The results indicate that the selectivity (regio- and enantioselectivity) is mainly affected by the ligand backbone, the substituent at the nitrogen heterocycle, and the substituents and configurations in the biaryl phosphite moiety. However, no general trend was seen. The tradeoff between regio- and enantioselectivities was best for ligand **L7a**, which forms a seven-membered chelate ring and has methyl substitutents at the alkyl-backbone chain and bulky *tert*-butyl groups at the *ortho* and *para* positions of the biphenyl phosphite moiety (Table 7, en-

tries 14 and 19). Again, these results are among the best reported for this type of sub-strate.^[5a,7,11b,17]

(3) Origin of enantioselectivity study of the Pd-π-allyl intermediates: To provide further insight into how ligand parameters affect catalytic performance, we studied the Pd-π-

allyl compounds **26–35** $[Pd(\eta^3-allyl)(L)]BF_4$ (L=L1–L8a–g), because they are key intermediates in the allylic substitution reactions studied.^[1] These ionic palladium complexes, which contain 1,3-diphenyl-, 1,3-dimethyl-, or cyclohexenylallyl groups, were prepared using the previously reported method from the corresponding Pd–allyl dimer and the appropriate ligand in the presence of silver tetrafluoroborate (Scheme 2).^[18] The complexes were characterized by ele-



Scheme 2. Preparation of $[Pd(\eta^3-allyl)(L)]BF_4$ complexes 26–35.

mental analysis and by ¹H, ¹³C, and ³¹P NMR spectroscopy. The spectral assignments (see the Experimental Section) were based on information from ¹H, ¹H, ³¹P, ¹H and ¹³C, ¹H correlation measurements in combination with ¹H, ¹H NOESY experiments. Unfortunately, we were unable to obtain crystals of sufficient quality to perform X-ray diffraction measurements. For some of the key Pd–allyl complexes, we also performed a DFT study.

Palladium 1,3-diphenylallyl complexes: When the phosphite-nitrogen ligand library **L1–L8a–g** was used in the allylic substitution of substrate **S1**, the catalytic results showed that enantioselectivity is highly affected by the ligand parameters. A six-membered chelate ring, phenyl substituents in the nitrogen heterocyclic ring, and bulky substituents at the *ortho* position of the biaryl moiety are therefore required if enantioselectivity is to be high. To understand this catalytic behavior, we studied the Pd- π -allyl complexes 26– 28, which contain ligands L1a, L6a, and L4a, respectively. Finally, with complexes 29 and 30, which contain ligands L1d and L1e, we studied the cooperative effect seen between the configuration of the ligand backbone and the configuration of the biaryl phosphite moiety.

The variable temperature (VT) NMR (30 to -80 °C) spectroscopic study of the Pd–allyl intermediate **26**, which contains ligand **L1a**, had a mixture of two isomers in equilibrium at a ratio of 9:1 (see the Experimental Section).^[19] Both isomers were unambiguously assigned by NMR spectroscopy (¹H, ³¹P, ¹³C, ¹H, ¹H, ¹G, and ¹H, ³¹P correlation and NOESY experiments) to the two *syn/syn endo* **A** and *exo* **B** isomers (Scheme 3). In both isomers, the NOE indicated in-



Scheme 3. Diastereoisomer Pd–allyl intermediates for **S1** with ligand **L1a**. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbon atoms are also shown.

teractions between the two terminal protons of the allyl group and also between the central allyl proton and orthohydrogen atoms of both phenyl groups of the allyl ligand, which clearly indicates a syn/syn disposition (Figure 1). Moreover, the central allyl proton showed a NOE interaction with the hydrogen of the CH-O group of the ligand backbone of the major isomer **A**, whereas in isomer **B** this interaction appeared with the hydrogen atoms of one tertbutyl group. These interactions can be explained by assuming a syn/syn endo disposition for isomer A and a syn/syn exo disposition for isomer B (Figure 1). We also carried out theoretical calculations at the DFT level for both isomers. Figure 1 shows these calculated structures and the relative values of the formation enthalpy, with isomer A being the most stable. The difference in the calculated formation enthalpy for the two isomers ($\Delta H = 1.8 \text{ kcal mol}^{-1}$) is in agreement with the population observed by NMR spectroscopy



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Figure 1. Calculated structures (DFT) for cationic species of complex $[Pd(\eta^3-1,3-diphenylallyl)(L1a)]BF_4$ (26) and their relative formation enthalpies. This figure also shows the relevant NOE contacts from the NOESY experiment.

of the different Pd-allyl intermediates formed in solution. For both isomers, the carbon NMR spectroscopic chemical shifts indicate that the more electrophilic allyl carbon terminus is trans to the phosphite moiety (Scheme 3). Assuming that the nucleophilic attack takes place at the more electrophilic allyl carbon terminus,^[1] the matching between enantiomeric excesses (82% S in product 15) and the diastereoisomeric Pd ratio (de = 80% (S)) indicate that the two isomers react at a similar rate. This is in agreement with the fact that the electrophilicities of the allylic terminal carbon atom trans to the phosphite are rather similar in both complexes ($\Delta(\delta^{13}C) = 0.5$ ppm). To prove this, we studied the reactivity of the Pd intermediates with sodium malonate at low temperature by in situ NMR spectroscopy (see the Supporting Information). Our results showed that the two isomers react at a similar rate.

The VT NMR spectroscopic study of Pd-allyl intermediate 27 containing ligand L6a, which forms a seven-membered chelate ring and provides lower enantioselectivity than Pd/L1a, also had a mixture of two syn/syn endo (A) and exo (B) isomers, but at a ratio of 1.25:1 (see the Experimental Section). Also, the more electrophilic allyl carbon terminus was trans to the phosphite moiety (Scheme 4). In contrast to the Pd/L1a catalytic system, the diastereoisomeric excess (de=11% (S)) of the Pd intermediates differed from the enantiomeric excess (21 % (S)) of alkylation product 15. As shown by NMR spectroscopy,^[20] isomer A reacts slightly faster than isomer \mathbf{B} ,^[21] thus explaining this difference. However, the lower enantioselectivity with this system than with the previous Pd/L1a catalytic system can mainly be attributed to the decrease in the relative amount of isomer A with respect to isomer B, compared with the population of the isomers (A and B) for complex 26.

Next, we studied the Pd–1,3-diphenylallyl intermediate **28** containing ligand **L4a**, which differs from **L1a** due to the presence of a bulky *tert*-butyl group at the oxazole moiety instead of a phenyl substituent and provides much lower enantioselectivity than the Pd/**L1a** catalyst. The VT NMR spectra indicated the presence of a mixture of two species at a ratio of 7:3 (see the Experimental Section). The major species were assigned by NOE to the *syn/syn endo* **A** isomer, whereas the minor species was attributed to com-



Scheme 4. Diastereoisomer Pd–allyl intermediates for S1 with ligand L6a. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbon atoms are also shown.



Scheme 5. Diastereoisomer Pd-allyl intermediates for S1 with ligand L4a. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbon atoms are also shown.

pound C, which contains two ligands coordinated in a monodentated fashion (Scheme 5).^[22] The formation of C is due to the fact that the replacement of the phenyl ozaxole substituents in ligand L1a by a bulky tert-butyl group caused greater steric interaction with one of the phenyl substitutents of substrate S1 in the related syn/syn exo B isomer observed in Pd/L1a complex 26 (Scheme 5). The formation of compound C minimizes this new steric interaction and explains why the expected syn/syn exo B isomer in solution was not detected (Scheme 5). Therefore, the fact that enantioselectivity was lower when the Pd/L4a catalyst was used (ee values up to 22%) than when the Pd/L1a catalyst was used (ee values up to 82%) may be due to the presence of Pd complex C. Complexes of this type are known to be faster and less enantioselective than their bidentate counterparts because they have more degrees of freedom.^[11b,23]

Finally, we studied the cooperative effect observed between the configuration of the ligand backbone and the con-

> figuration of the biaryl phosphite moiety with complexes 29 and 30, which contain ligands L1d and L1e, respectively. The VT NMR spectroscopic study indicated the presence of a mixture of two syn/syn endo (A) and exo (B) isomers in ratios of and 1:2.5, respectively 5:1 (Scheme 6). As for the 1,3-diphenylallyl intermediates discussed above, the NMR spectroscopic data showed that the more electrophilic allyl terminal carbon is trans to the phosphite moiety at the A isomers (29A and 30 A). For complex 29, our results indicated that the stereochemical outcome of the re-



Scheme 6. Diastereoisomer Pd-allyl intermediates for **S1** with a) ligand **L1d** and b) ligand **L1e**. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbon atoms are also shown.

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action (ee = 69% (S)) was mainly due to the diastereoisomeric excess (de=66% (S)) of the Pd complexes in solution, as observed for Pd/L1a. However, for complex 30, the diastereoisomeric excess (de = 43% (R)) did not match the enantioselectivity obtained (ee = 40% (S)). Therefore, we conclude that isomer 30A reacts faster than isomer 30B. A clear indication of this fact can be found in the higher electrophilicity of the allylic terminal carbon trans to the phosphite in **30A** than in **30B** ($\Delta(\delta^{13}C) = 4.6$ ppm). Although isomer 30 A has a faster reaction rate than isomer 29 A, this is not the reason why Pd/L1e causes lower enantioselectivity than Pd/L1d. Rather, the lower enantioselectivity of Pd/L1e is because the relative amount of isomer A respect to isomer **B** is much lower in Pd/L1e than in Pd/L1d. Therefore, as observed earlier, the presence of an S configuration at the biaryl phosphite moiety is necessary for good enantioselectivity. In addition, these results provide further evidence that the flexible biphenyl phosphite moieties in ligands L1a-c adopt an S configuration when coordinated to palladium.

In summary, the study of the Pd–1,3-diphenylallyl intermediate showed that for enantioselectivity to be high, the different ligand parameters need to be correctly combined to predominantly form one of the Pd isomers and also to avoid the formation of species

avoid the formation of species with ligands coordinated in a monodentated fashion.

Palladium-1,3-dimethylallyl

complexes: When the phosphite-nitrogen ligand library L1–L8a–g was used in the allylic substitution of substrate S3, the catalytic results revealed a different trend with regard to the effect of the ligand parameters than with the hindered substrate S1. A seven-membered chelate ring, the presence of methyl substituents at the alkyl backbone chain, and a bulky (S)-binaphthyl phosphite moiety are necessary for high enantioselectivity. To understand this catalytic behavior, we studied Pd-*π*-allyl complexes 31-33, which contain ligands L1a, L7a, and L7e, respectively. With ligands L1a and L7a, we studied the effect of the chelate ring size, whereas with ligand L7e we studied the configuration of the biaryl phosphite moiety.

The VT NMR (30 to -80 °C) spectroscopic study of Pd-allyl intermediate **31**, which contains

ligand L1a, had a mixture of three isomers in equilibrium at a ratio of 2.5:1:2 (see the Experimental Section). Isomers A and **B** were assigned by NOE to the two syn/syn endo **A** and exo B isomers, whereas isomer D was assigned to the syn/anti isomer (Scheme 7). For isomers A and B, the NOE indicated interactions between the two terminal protons of the allyl group, whereas for isomer **D** the central allyl proton showed a NOE interaction with the terminal allyl proton located trans to the oxazole group (Figure 2). Moreover, for isomers A and D, the central allyl proton showed a NOE interaction with the hydrogen of the CH-O group of the ligand, whereas in isomer **B** this interaction appeared with the hydrogen atoms of the tert-butyl group. These interactions can be explained by assuming a syn/syn endo disposition for isomer \mathbf{A} , a syn/syn exo disposition for isomer \mathbf{B} , and a syn/anti disposition for isomer D (Figure 2). The NOESY also indicated an exchange between the allylic terminal protons located trans to the oxazole moiety of isomers **B** and **D** (Figure 2). This confirms the $\eta^3 - \eta^1 - \eta^3$ movement for the exchange between isomers \mathbf{B} and $\mathbf{D}^{[24]}$ In addition, the fact that no other Hant-Hsyn exchange was observed indicates that the exchange took place by means of the selective opening of one of the terminal Pd-C bonds. The formation of isomer **D** is due to the fact that, in complex **B**, there was



Scheme 7. Diastereoisomer Pd-allyl intermediates for S3 with ligand L1a. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbon atoms are also shown.



Figure 2. Relevant NOE contacts from the NOESY experiment of the $[Pd(\eta^3-1,3-dimethylallyl)(L1a)]BF_4$ (31) isomers.

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an increase in the steric interaction between the oxazole phenyl group and one of the methyl substitutents of S3 due to the absence of the favorable π -stacking interaction observed in the related Pd-1,3-diphenylallyl complex 26. The formation of isomer **D** minimized this steric interaction (Scheme 7). Therefore, the open Pd-C bond belongs to the most electrophilic carbon atom containing the substituent that undergoes the biggest steric hindrance with the phenyl oxazole fragment. The NMR spectroscopic data also indicated that the most electrophilic allyl carbon terminus is trans to the phosphite moiety in syn/syn isomer A and in syn/syn isomer **B**, and that the allylic terminus carbon in isomer **D** is far less electrophilic ($\Delta(\delta^{13}C) > 11$ ppm). Assuming that the nucleophilic attack takes place at the most electrophilic allyl carbon terminus, on the basis of the observed stereochemical outcome of the reaction (59% S in product 18), and since the ee of alkylation product 18 differs from the diastereoisomeric excess (de = 41 % (S)) of the reacting Pd intermediates A and B, we conclude that isomer A reacts faster

than isomer **B**. This was confirmed by an in situ NMR spectroscopic study of the reactivity of the Pd intermediates with sodium malonate at low temperature. This study indicated that isomer **31A** reacts approximately 1.5 times faster than isomer **31B**. This is also consistent with the fact that, for both isomers, the most electrophilic allylic terminal carbon atom is the one *trans* to the phosphite in the major **A** isomer.

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The VT NMR spectroscopic study of Pd-allyl intermediate 32, which contains ligand L7a, which forms a seven-membered chelate ring and provides higher enantioselectivity than Pd/L1a, also had a mixture of three isomers at a ratio of 5:1:7 (see the Experimental Section). Isomers A, B, and D were assigned by NOE to the syn/syn endo, syn/syn exo, and syn/anti isomers, respectively (see Scheme 8). Again, the most electrophilic allyl carbon terminus was trans to the phosphite moiety in syn/syn isomer A and syn/syn isomer **B**, and the allylic terminus carbon in isomer D was far less electrophilic (Δ - $(\delta^{13}C) > 11 \text{ ppm})$. As observed for complex Pd/L1a (31), isomer A reacts faster than isomer B. The higher enantioselectivity with this system than with the Pd/L1a catalytic system discussed above can be attributed to the decrease in the relative amount of isomer **B** with respect to isomer **A** compared with the population of the isomers (**A** and **B**) for complex **31**. The decrease in the population of isomer **B** is due to the formation of a smaller chiral pocket with ligand L7a than with ligand L1a. This smaller chiral pocket creates a greater steric interaction between the thiazole phenyl group and one of the methyl substitutents of **S3**, which results in the preferential formation of the less electrophilic *syn/anti* isomer **D**.

Finally, the VT NMR spectroscopic study of Pd-allyl intermediate **33**, which contains ligand **L7d**, had a mixture of three isomers in a ratio of 10:1:9 (see the Experimental Section). Isomers **A**, **B**, and **D** were assigned by NOE to the *syn/syn endo, syn/syn exo*, and *syn/anti* isomers, respectively (Scheme 9). As for complexes **31** and **32**, the fastest-reacting isomer was the *syn/syn* isomer **A**, whereas isomer **D** was far less electrophilic and therefore did not play a direct role in



Scheme 8. Diastereoisomer Pd-allyl intermediates for **S3** with ligand **L7a**. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbon atoms are also shown.



Scheme 9. Diastereoisomer Pd-allyl intermediates for **S3** with ligand **L7d**. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbon atoms are shown.

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the enantiodiscrimination process. The higher enantioselectivity obtained with Pd/L7d than with the Pd/L7a catalytic system is due to the larger amount of the most reactive isomer (isomer A) than in complex 32.

In summary, for enantioselectivities to be high, the various ligand parameters need to be correctly combined to preferentially form the faster isomer **A**. The formation of this isomer is mainly governed by the size of the chelate ring and the configuration of the biaryl phosphite moiety.

Palladium-1,3-cyclohexenylallyl complexes: When the phosphite-nitrogen ligand library L1-L8a-g was used in the allylic substitution of cyclic sub-

strates **S5** and **S6**, the catalytic results showed that the effect of the ligand parameters on enantioselectivity was different from the effect observed in the substitution of the linear substrates **S1** and **S2**. The best results were obtained with ligands **L1d** and **L1e**, which form a six-membered chelate ring, have phenyl substituents in the nitrogen heterocyclic ring, and have bulky enantiopure binaphthyl phosphite moieties. Interestingly, the sense of enantioselectivity was controlled by the configuration of the biaryl phosphite moiety. So, to understand this catalytic behavior, we studied Pd– π allyl complexes **34** and **35**, which contain ligands **L1d** and **L1e**.

The VT NMR (35 to -80 °C) spectroscopic study of Pd intermediate **34**, which contains ligand **L1d**, showed a mixture of the two possible isomers at a ratio of 1:6, respectively (see Scheme 10). All isomers were unambiguously assigned



by NOE to the isomers **A** and **B** (Figure 3). Thus, for isomer **A**, the NOE indicated interactions between the hydrogen of the CH–O group and the central allyl proton, whereas for



Figure 3. Calculated structures (DFT) for cationic species of complex $[Pd(\eta^3-1,3-cyclohexenylallyl)(L1a)]BF_4$ (34) and their relative formation enthalpies. This figure also shows the relevant NOE contacts from the NOESY experiment.

isomer **B** this interaction appeared with one of the methylenic hydrogen atoms of the allyl ligand (Figure 3). The carbon NMR spectroscopic chemical shifts indicated that the most electrophilic allylic terminus carbon is trans to the phosphite moiety. The difference between the diastereoisomeric ratio and enantioselectivity observed in the alkylation of S5 (de = 70% (S) vs. ee = 80% (S)) indicated that the nucleophile reacts faster with the major **B** isomer. This was confirmed by the reactivity of the Pd intermediate with sodium malonate at low temperature by in situ NMR spectroscopy.^[25] We also carried out theoretical calculations at the DFT level. Figure 3 shows these calculated structures and their relative formation enthalpy values, with isomer **B** being the most stable. The difference in the calculated formation enthalpies of the two isomers ($\Delta H = 1.5 \text{ Kcal mol}^{-1}$) is in agreement with the population observed by NMR spectroscopy.

The VT NMR (35 °C to -80 °C) spectroscopic study of Pd intermediate 35, which contains ligand L1e, also showed a mixture of the two possible isomers (A and B), but at a ratio of 6.5:1, respectively (Scheme 11). The carbon NMR spectroscopic chemical shifts also indicated that the most electrophilic allylic terminus carbon is *trans* to the phosphite moiety. In contrast to Pd intermediate 34, the fastest-reacting isomer is A. This is in agreement with the fact that Pd/L1e provided a level of enantioselectivity similar to that of Pd/L1d, discussed above, but in the opposite enantiomer of the alkylation products 19 and 20.

Conclusion

Scheme 10. Diastereoisomer Pd-allyl intermediates for **S5** with ligand **L1d**. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbon atoms are also shown.

A new P,N-ligand library **L1–L8a–g** was synthesized for the Pd-catalyzed allylic substitution reactions of several substrates with different electronic and steric properties. These series of ligands have five main advantages: 1) they can be

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Scheme 11. Diastereoisomer Pd–allyl intermediates for **S5** with ligand **L1e**. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbon atoms are also shown.

prepared efficiently from readily available hydroxyl-oxazole/thiazole derivatives; 2) the hydroxyl-oxazole/thiazole cores are much more robust than the usually used hydroxyloxazoline ones; 3) the flexibility and larger bite angle created by the biaryl phosphite moiety and the different bridge lengths increase versatility; 4) the π -acceptor character of the phosphite moiety increases reaction rates; and 5) their modular nature enables the bridge length, the substituents at the heterocyclic ring and in the alkyl backbone chain, the configuration of the ligand backbone, and the substituents/ configurations in the biaryl phosphite moiety to be easily and systematically varied, so that activities and regio- and enantioselectivities can be maximized for each substrate as required. By carefully selecting the ligand components, therefore, high regio- and enantioselectivities (ee values up to 96%) and good activities were obtained in a broad range of mono-, di-, and trisubstituted linear hindered and unhindered substrates and cyclic substrates. Of particular note were the high regio- and enantioselectivities (up to 96% ee) combined with high activities obtained for the mono- and trisubstituted substrates S7-S10. In addition, for all substrates, both enantiomers of the substitution products were obtained with high enantioselectivities.

By studying the Pd–1,3-diphenylallyl, –1,3-dimethylallyl, and –1,3-cyclohexenylallyl intermediates by means of NMR spectroscopy and DFT calculations, we were able to better understand the observed catalytic behavior. For enantioselectivities to be high, we conclude that ligand parameters need to be correctly combined to predominantly form the Pd intermediate that has the fastest reaction with the nucleophile. We also found that the nucleophilic attack takes place predominantly at the allylic terminal carbon atom located *trans* to the phosphite moiety.

Experimental Section

General considerations: All reactions were carried out using standard Schlenk techniques under an atmosphere of argon. Solvents were purified and dried by standard procedures. Phosphorochloridites are easily prepared in one step from the corresponding biaryls.^[26] Hydroxyl-oxazoles 6 and 7 and hydroxyl-thiazole 13 were prepared as previously described.^[9] Racemic substrates S1-S10 were prepared as previously reported.^[27-30,14b] $[\{Pd(\eta^{3}-1,3-Ph_{2}-C_{3}H_{3})(\mu-Cl)\}_{2}],^{[31]} [\{Pd(\eta^{3}-1,3-Me_{2}-C_{3}H_{3})(\mu-Cl)\}_{2}],^{[32]} \text{ and }$ $[{Pd(\eta^3-cyclohexenyl)(\mu-Cl)}_2]^{[33]}$ were prepared as previously described. ¹H, ¹³C{¹H}, and ³¹P{¹H} NMR spectra were recorded using a 400 MHz spectrometer. Chemical shifts are relative to that of SiMe₄ (¹H and ¹³C) as internal standard or H₃PO₄ (³¹P) as external standard. ¹H, ¹³C, and ³¹P assignments were done based on ¹H,¹H gCOSY, ¹H,¹³C gHSQC, and ¹H,³¹P gHMBC experiments. Geometries of all substrates were optimized using the Jaguar program^[34] by applying the B3LYP hybrid density functional^[35] together with the LACVP** basis sets. Normal-mode analysis of stable structures revealed no imaginary frequencies, or a single imaginary frequency with negligibly low energy ($\tilde{\nu} < 100 \text{ cm}^{-1}$). LACVP in Jaguar defines a combination of the LANL2DZ basis set^[36] for palladium and the 6-31G basis set for other atoms.

General procedure for the preparation of hydroxyl–oxazoles 8 and 9: Diazodimedone (2.00 g, 12 mmol) and the corresponding *para*-substituted benzonitrile (60 mmol for 4-methylbenzonitrile and 14 mmol for 4-(trifluoromethyl)benzonitrile) were heated in an oil bath at 60 °C and Rh-(OAc)₂ (10 mg, 0.045 mmol) was added. The reaction was stirred for 1.5 h, and if starting material remained, another portion of Rh(OAc)₂ (10 mg, 0.045 mmol) was added. After an additional 1.5 h, the reaction was cooled to room temperature and purified by chromatography on silica (pentane/ethyl acetate 75:25 to 25:75) to give the ketone–oxazoles as white solids. Compound **3** (2.02 g, 66% yield): ¹H NMR (CDCl₃): δ = 1.21 (s, 6H; 2×CH₃), 2.41 (s, 3H; CH₃), 2.50 (s, 2H; CH₂), 2.90 (s, 2H; CH₂), 7.24–7.29 (m, 2H; Ar), 7.97–8.02 ppm (m, 2H; Ar). Compound **4** (1.53 g, 51% yield): ¹H NMR (CDCl₃): δ = 1.22 (s, 6H; 2×CH₃), 2.52 (s, 2H; CH₂), 2.93 (s, 2H; CH₂), 7.71–7.75 (m, 2H; Ar), 8.20–8.25 ppm (m, 2H; Ar).

For the reduction of the ketone-oxazoles, BH₃·SMe₂ (2.1 equiv) and (R)-Me-CBS (0.1 equiv) were dissolved in THF (5 mL for 4 mmol BH₃·SMe₂) at 0°C and stirred for 1 h. The temperature was raised to ambient and a solution of ketone (1 equiv) in THF/toluene (4:1 mL for 2 mmol ketone) was added over 2 h using a syringe pump. The reaction was stirred for an additional hour, cooled to 0°C, and quenched with methanol. The solvent was evaporated and the resulting oil was purified by chromatography on silica (pentane/ethyl acetate 25:75) to afford hydroxyl-oxazoles 8 and 9 as white solids (72% ee for 8 and 78% ee for 9). The enantiomeric excess could not be increased by recrystallization. The enantiomers were separated by chiral chromatography on semipreparative HPLC using a 250× 20 mm Chiralcel OD column (hexane/isopropanol 95:5, 5 mLmin⁻¹) to give ee > 99 %. Hydroxyl-oxazole 8: ¹H NMR (CDCl₃): $\delta = 1.03$ (s, 3H; CH₃), 1.19 (s, 3H; CH₃), 1.67 (dd, J=13.2, 8.0 Hz, 1H; CH₂), 1.99 (ddd, J=13.4, 6.0, 1.5 Hz, 1 H; CH₂), 2.38 (s, 3 H; CH₃-Ph), 2.45 (m, 1 H; CH₂), 2.59 (dd, J=16.4, 2.2 Hz, 1H; CH₂), 3.66 (brs, 1H; OH), 4.85 (m, 1H; CH), 7.18–7.28 (m, 2H; CH=), 7.87–7.95 ppm (m, 2H; CH=); ¹³C NMR $(CDCl_3): \delta = 24.5, 26.9, 30.6, 33.0, 35.6, 45.5, 62.9, 124.9, 126.1, 129.4,$ 136.0, 140.3, 147.3, 161.3 ppm; elemental analysis calcd (%) for C16H19NO2: C 74.68, H 7.44, N 5.44; found: C 74.47, H 7.50, N 5.43. Hydroxyl-oxazole 9: ¹H NMR (CDCl₃): $\delta = 1.05$ (s, 3H; CH₃), 1.21 (s, 3H; CH₃), 1.68 (dd, *J*=13.3, 8.1 Hz, 1H; CH₂), 2.02 (dd, *J*=13.3, 6.0 Hz, 1H; CH₂), 2.50 (d, J=16.8 Hz, 1 H; CH₂), 2.63 (d, J=16.8 Hz, 1 H; CH₂), 3.27 (brs, 1H; OH), 4.87 (m, 1H; CH), 7.63-7.74 (m, 2H; CH=), 8.07-8.18 ppm (m, 2H; CH=); 13 C NMR (CDCl₃): $\delta = 27.2$, 30.8, 33.3, 35.9, 45.7, 63.2, 126.0, 126.1, 126.6, 131.0, 137.0, 149.0, 160.0 ppm; elemental analysis calcd (%) for $C_{16}H_{16}F_3NO_2$: C 61.73, H 5.18, N 4.50; found: C 61.79, H 5.23, N 4.52.

Preparation of hydroxyl–oxazole 10: Diazodimedone (5.00 g, 30.1 mmol), pivalonitrile (12.5 g, 150 mmol), and Rh(OAc)₂ (27 mg, 0.12 mmol) were heated in an oil bath at 60 °C. The reaction was stirred for 1 h, cooled to room temperature, and purified by chromatography on silica (pentane/

ethyl acetate 75:25 to 25:75) to give ketone–oxazole **5** as a white solid (4.23 g, 68 % yield). ¹H NMR (CDCl₃): δ =1.16 (s, 6H; 2×CH₃), 1.39 (s, 9H; 3×CH₃), 2.41 (s, 2H; CH₂), 2.78 ppm (s, 2H; CH₂).

For the reduction of the ketone-oxazole, ketone (2.22 g, 10 mmol) was dissolved in 95% ethanol (125 mL), and NaBH₄ (1.14 g, 30 mmol) was added. The reaction was stirred and followed by TLC. The reaction was quenched with 1 M HCl and extracted with dichloromethane (2×25 mL). The organic phase was dried over MgSO4, filtered, and concentrated to give racemic hydroxyl-oxazole 10 as a white solid (1.94 g, 87% yield). The enantiomers were separated by chiral chromatography on semipreparative HPLC using a (250×20 mm) Chiralcel OD column (hexane/isopropanol 98:2, 5 mL min⁻¹) to give ee > 99 %. ¹H NMR (CDCl₃): $\delta = 0.96$ (s, 3H; CH₃), 1.11 (s, 3H; CH₃), 1.32 (s, 9H; CH₃, *t*Bu), 1.57 (dd, *J*=3.2, 8.0 Hz, 1H; CH₂), 1.86 (ddd, J=13.2, 5.9, 1.4 Hz, 1H; CH₂), 2.30 (dt, J= 16.2, 1.6 Hz, 1H; CH₂), 2.45 (dd, J=16.2, 2.1 Hz, 1H; CH₂), 4.52 (brs, 1H; OH), 4.74 ppm (m, 1H; CH); 13 C NMR (CDCl₃): $\delta = 26.8, 28.5, 30.5,$ 30.6, 32.9, 33.7, 35.4, 45.5, 62.0, 62.2, 134.2, 146.4, 170.4 ppm; elemental analysis calcd (%) for $C_{13}H_{21}NO_2$: C 69.92, H 9.48, N 6.27; found: C 69.89, H 9.52, N 6.22.

Preparation of hydroxyl-thiazole 14: CeCl₃·7H₂O (3.15 g) was dried in a vacuum oven at 120°C overnight. A flask with dry CeCl₃ (2.00 g, 8.10 mmol) was cooled under nitrogen in an ice bath, THF was added (30 mL), and the suspension was stirred at room temperature overnight. After cooling in an ice bath, a 3M solution of MeMgCl (4mL, 12.0 mmol) was added. Suspension was stirred for 2 h and a solution of thiazole ester^[9b] (0.65 g, 2.26 mmol) was added in THF (10 mL). After stirring overnight, the reaction was cooled in an ice bath and quenched by addition of saturated NH₄Cl solution (50 mL), and stirred for 1 h. Water (50 mL) was added, THF was evaporated, and the product was extracted with diethyl ether (2 times). After drying the organic phase over MgSO₄, filtering, and evaporation of the solvent, crude material was purified by chromatography on silica (pentane/ethyl acetate 100:0 to 95:5) to give hydroxyl-thiazole 14 as a white solid (0.44 g, 71 % yield). The resolution of enantiomers was achieved on semipreparative HPLC using a (250×20 mm) Chiralcel OD column (hexane/isopropanol 95:5, 5 mL min⁻¹) to give both enantiomers with ee > 99%. ¹H NMR (CDCl₃): $\delta = 1.01$ (s, 3H; CH₃), 1.32 (s, 3H; CH₃), 1.39 (m, 1H; CH₂), 1.74 (m, 1H; CH₂), 2.02-2.15 (m, 2H; CH₂), 2.73 (m, 1H; CH₂), 2.85 (m, 1H; CH₂), 2.99 (m, 1H; CH₂), 6.48 (brs, 1H; OH), 7.33-7.46 (m, 2H; CH=), 7.80–7.92 ppm (m, 2H; CH=); 13 C NMR (CDCl₃): δ =23.1, 23.8, 24.2, 26.2, 27.7, 48.4, 73.5, 126.1, 128.8, 129.8, 131.1, 133.1, 152.4, 164.2 ppm; elemental analysis calcd (%) for C₁₆H₁₉NOS: C 70.29, H 7.00, N 5.12; found: C 70.35, H 7.02, N 5.11.

General procedure for the preparation of phosphite–nitrogen ligands L1– L8a–g: Phosphorochloridite (2.2 mmol) produced in situ was dissolved in toluene (5 mL) and pyridine (0.36 mL, 4.6 mmol) was added. Hydroxyl– oxazole or hydroxyl–thiazole (2 mmol) was azeotropically dried with toluene (3×1 mL) and then dissolved in toluene (10 mL), to which pyridine (0.36 mL, 4.6 mmol) was added. The alcohol solution was transferred slowly at 0 °C to the solution of phosphorochloridite. The reaction mixture was warmed up to 80 °C and stirred overnight, and the pyridine salts were removed by filtration. Evaporation of the solvent gave a white foam, which was purified by flash chromatography (toluene/hexane/ NEt₃) to produce the corresponding ligand as a white powder.

Ligand L1a: Yield: 826 mg (60%); ³¹P NMR (C₆D₆): δ =148.7 ppm; ¹H NMR (C₆D₆): δ =0.57 (s, 3H; CH₃), 0.77 (s, 3H; CH₃), 1.23 (s, 9H; CH₃, *t*Bu), 1.25 (s, 9H; CH₃, *t*Bu), 1.57 (m, 1H; CH₂), 1.61 (s, 9H; CH₃, *t*Bu), 1.69 (s, 9H; CH₃, *t*Bu), 1.57 (m, 1H; CH₂), 1.92 (m, 1H; CH₂-C=), 2.08 (m, 1H; CH₂-C=), 5.53 (m, 1H; CH=O), 7.1–8.2 ppm (m, 9H; CH=); ¹³C NMR (C₆D₆): δ =27.7 (CH₃), 29.9 (CH₃), 31.9 (CH₃, *t*Bu), 33.1 (C, *t*Bu), 35.0 (CH₂-C=), 36.2 (CMe₂), 45.1 (CH₂), 67.8 (d, *J*(C,P)= 12.2 Hz; CH=O), 124–165 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₃H₅₆NO₄P: C 75.74, H 8.28, N 2.05; found: C 75.82, H 8.11, N 2.11.

Ligand L1b: Yield: 725 mg (58%); 31 P NMR (C₆D₆): δ =148.6 ppm; 1 H NMR (C₆D₆): δ =0.60 (s, 3H; CH₃), 0.82 (s, 3H; CH₃), 1.55 (s, 9H; CH₃, *t*Bu), 1.62 (s, 9H; CH₃, *t*Bu), 1.77 (m, 1H; CH₂), 1.80 (m, 1H; CH₂), 1.97 (m, 1H; CH₂–C=), 2.12 (m, 1H; CH₂–C=), 3.28 (s, 3H; CH₃–

O), 3.29 (s, 3H; CH₃–O), 5.52 (m, 1H; CH–O), 6.7–8.2 ppm (m, 9H; CH=); 13 C NMR (C₆D₆): $\delta = 27.8$ (CH₃), 29.2 (CH₃), 30.0 (CMe₂), 31.6 (CH₃, *t*Bu), 31.8 (CH₃, *t*Bu), 33.0 (C, *t*Bu), 35.6 (CH₂–C=), 45.1 (CH₂), 55.4 (CH₃–O), 55.5 (CH₃–O), 67.9 (d, *J*(C,P)=13 Hz; CH–O), 126–156 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₃₇H₄₄NO₆P: C 70.57, H 7.04, N 2.22; found: C 70.52, H 7.09, N 2.31.

Ligand L1c: Yield: 722 mg (59%); ³¹P NMR (C₆D₆): δ =147.8 ppm; ¹H NMR (C₆D₆): δ =0.44 (s, 9H; CH₃–Si), 0.55 (s, 9H; CH₃–Si), 0.59 (s, 3H; CH₃), 0.81 (s, 3H; CH₃), 1.58 (m, 1H; CH₂), 1.76 (m, 1H; CH₂), 1.96 (m, 1H; CH₂–C=), 2.13 (m, 1H; CH₂–C=), 5.50 (m, 1H; CH–O), 6.9–8.3 ppm (m, 11H; CH=); ¹³C NMR (C₆D₆): δ =0.8 (CH₃–Si), 0.9 (CH₃–Si), 27.9 (CH₃), 30.2 (CH₃), 33.2 (CMe₂), 35.9 (CH₂–C=), 45.3 (CH₂), 67.8 (d, *J*(C,P)=8.4 Hz; CH–O), 123–150 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₃₃H₄₀NO₄PSi₂: C 65.86, H 6.70, N 2.33; found: C 65.92, H 6.72, N 2.31.

Ligand L1d: Yield: 698 mg (49%); ³¹P NMR (C₆D₆): δ =146.3 ppm; ¹H NMR (C₆D₆): δ =0.45 (s, 3 H; CH₃), 0.54 (s, 9 H; CH₃–Si), 0.73 (s, 9 H; CH₃–Si), 0.82 (s, 3 H; CH₃), 1.57 (m, 1 H; CH₂), 1.88 (m, 1 H; CH₂), 2.11 (m, 2 H; CH₂–C=), 5.02 (m, 1 H; CH–O), 6.9–8.3 ppm (m, 15 H; CH=); ¹³C NMR (C₆D₆): δ =0.5 (CH₃–Si), 1.0 (CH₃–Si), 27.9 (CH₃), 29.7 (CH₃), 32.8 (CMe₂), 35.6 (CH₂–C=), 45.5 (CH₂), 70.2 (CH–O), 123–161 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₁H₄₄NO₄PSi₂: C 70.15, H 6.32, N 2.00; found: C 70.22, H 6.35, N 2.01.

Ligand L1e: Yield: 536 mg (39%); ³¹P NMR (C₆D₆): δ =147.4 ppm; ¹H NMR (C₆D₆): δ =0.33 (s, 3 H; CH₃), 0.55 (s, 9 H; CH₃–Si), 0.65 (s, 3 H; CH₃), 0.68 (s, 9 H; CH₃–Si), 1.34 (m, 1 H; CH₂), 1.61 (m, 1 H; CH₂), 1.89 (m, 1 H; CH₂–C=), 2.06 (m, 1 H; CH₂–C=), 5.49 (m, 1 H; CH–O), 6.9–8.3 ppm (m, 15 H; CH=); ¹³C NMR (C₆D₆): δ =0.3 (CH₃–Si), 0.4 (CH₃–Si), 26.5 (CH₃), 29.6 (CH₃), 32.4 (CMe₂), 35.1 (CH₂–C=), 44.3 (CH₂), 66.8 (d, *J*(C,P)=7.2 Hz; CH–O), 123–160 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₁H₄₄NO₄PSi₂: C 70.15, H 6.32, N 2.00; found: C 70.19, H 6.29, N 1.97.

Ligand L1f: Yield: 712 mg (64%); ³¹P NMR (C₆D₆): δ =138.7 ppm; ¹H NMR (C₆D₆): δ =0.42 (s, 3H; CH₃), 0.78 (s, 3H; CH₃), 1.48 (dd, ²J-(H,H) = 14 Hz, ³J(H,H) = 5.6 Hz, 1H; CH₂), 1.71 (dd, ²J(H,H) = 14 Hz, ³J-(H,H) = 5.6 Hz, 1H; CH₂), 1.84 (m, ²J(H,H) = 16 Hz, 1H; CH₂-C=), 2.06 (d, ²J(H,H) = 16 Hz, 1H; CH₂-C=), 5.34 (m, 1H; CH-O), 6.9–8.3 ppm (m, 17H; CH=); ¹³C NMR (C₆D₆): δ =28.0 (CH₃), 29.4 (CH₃), 32.7 (CH₂-C=), 35.7 (CMe₂), 45.2 (CH₂), 68.5 (d, J(C,P)=4.7 Hz; CH-O), 122–162 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₃₅H₂₈NO₄P: C 75.39, H 5.06, N 2.51; found: C 75.42, H 5.11, N 2.50.

Ligand L1g: Yield: 849 mg (76%); ³¹P NMR (C₆D₆): δ =153.1 ppm; ¹H NMR (C₆D₆): δ =0.51 (s, 3H; CH₃), 0.72 (s, 3H; CH₃), 1.50 (dd, ²J-(H,H)=12.4 Hz, ³J(H,H)=5.6 Hz, 1H; CH₂), 1.59 (dd, ²J(H,H)=12.4 Hz, ³J(H,H)=7.2 Hz, 1H; CH₂), 1.92 (m, ²J(H,H)=16.4 Hz, 1H; CH₂-C=), 2.01 (d, ²J(H,H)=16.4 Hz, 1H; CH₂-C=), 5.43 (m, 1H; CH-O), 6.8–8.3 ppm (m, 17H; CH=); ¹³C NMR (C₆D₆): δ =28.1 (CH₃), 29.7 (CH₃), 32.8 (CH₂-C=), 35.6 (CMe₂), 45.0 (d, J(C,P)=3.1 Hz; CH₂), 67.8 (d, J(C,P)=20.1 Hz; CH-O), 122–162 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₃₅H₂₈NO₄P: C 75.39, H 5.06, N 2.51; found: C 75.35, H 5.13, N 2.48.

Ligand L2a: Yield: 587 mg (42%); ³¹P NMR (C₆D₆): δ =148.0 ppm; ¹H NMR (C₆D₆): δ =0.78 (s, 3H; CH₃), 0.98 (s, 3H; CH₃), 1.44 (s, 9H; CH₃, *t*Bu), 1.46 (s, 9H; CH₃, *t*Bu), 1.79 (m, 1H; CH₂), 1.83 (s, 9H; CH₃, *t*Bu), 1.91 (s, 9H; CH₃, *t*Bu), 1.95 (m, 1H; CH₂), 2.19 (m, 2H; CH₂-C=), 2.28 (m, 3H; CH₃-Ph), 5.75 (m, 1H; CH–O), 7.1–8.4 ppm (m, 8H; CH=); ¹³C NMR (C₆D₆): δ =21.7 (CH₃-Ph), 27.8 (CH₃), 30.0 (CH₃), 31.9 (CH₃, *t*Bu), 32.0 (CH₃, *t*Bu), 33.1 (C, *t*Bu), 36.2 (CH₂-C=), 36.7 (CMe₂), 45.1 (CH₂), 67.9 (d, *J*(C,P)=17 Hz; CH–O), 124–164 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₄H₅₈NO₄P: C 75.94, H 8.40, N 2.01; found: C 75.89, H 8.41, N 2.0.5.

Ligand L2d: Yield: 698 mg (49%); ³¹P NMR (C₆D₆): δ =146.3 ppm; ¹H NMR (C₆D₆): δ =0.43 (s, 3 H; CH₃), 0.51 (s, 9 H; CH₃–Si), 0.70 (s, 9 H; CH₃–Si), 0.80 (s, 3 H; CH₃), 1.52 (m, 1 H; CH₂), 1.72 (m, 1 H; CH₂), 2.18 (m, 2 H; CH₂–C=), 2.23 (m, 3 H; CH₃–Ph), 5.11 (m, 1 H; CH–O), 6.9–8.3 ppm (m, 15H; CH=); ¹³C NMR (C₆D₆): δ =0.4 (CH₃–Si), 0.6 (CH₃–Si), 20.9 (CH₃–Ph), 27.8 (CH₃), 29.2 (CH₃), 33.8 (CMe₂), 35.8 (CH₂–C=), 45.6 (CH₂), 69.8 (CH–O), 123–161 ppm (aromatic carbon atoms); ele-

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mental analysis calcd (%) for $C_{42}H_{46}NO_4PSi_2$: C 70.46, H 6.48, N 1.96; found: C 70.52, H 6.49, N 2.00.

Ligand L2e: Yield: 623 mg (44%); ³¹P NMR (C₆D₆): δ =146.9 ppm; ¹H NMR (C₆D₆): δ =0.49 (s, 3H; CH₃), 0.510 (s, 9H; CH₃–Si), 0.66 (s, 9H; CH₃–Si), 0.76 (s, 3H; CH₃), 1.69 (m, 2H; CH₂), 2.21 (m, 2H; CH₂– C=), 2.25 (m, 3H; CH₃–Ph), 5.16 (m, 1H; CH–O), 6.9–8.3 ppm (m, 15H; CH=); ¹³C NMR (C₆D₆): δ =-0.2 (CH₃–Si), 0.1 (CH₃–Si), 21.1 (CH₃– Ph), 27.9 (CH₃), 29.5 (CH₃), 33.9 (CMe₂), 36.2 (CH₂–C=), 45.3 (CH₂), 70.1 (CH–O), 123–161 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₂H₄₆NO₄PSi₂: C 70.46, H 6.48, N 1.96; found: C 70.49, H 6.51, N 1.99.

Ligand L3a: Yield: 674 mg (45%); ³¹P NMR (C₆D₆): δ =147.8 ppm; ¹H NMR (C₆D₆): δ =0.61 (s, 3H; CH₃), 0.81 (s, 3H; CH₃), 1.28 (s, 9H; CH₃, *t*Bu), 1.29 (s, 9H; CH₃, *t*Bu), 1.60 (m, 1H; CH₂), 1.65 (s, 9H; CH₃, *t*Bu), 1.74 (s, 9H; CH₃, *t*Bu), 1.77 (m, 1H; CH₂), 1.97 (m, 1H; CH₂-C=), 2.12 (m, 1H; CH₂-C=), 5.55 (m, 1H; CH–O), 7.0–8.1 ppm (m, 8H; CH=); ¹³C NMR (C₆D₆): δ =27.6 (CH₃), 30.0 (CH₃), 31.9 (CH₃, *t*Bu), 32.0 (CH₃, *t*Bu), 33.1 (CMe₂), 35.0 (c, *t*Bu), 35.7 (c, *t*Bu), 36.2 (CH₂-C=), 45.0 (CH₂), 67.5 (d, *J*(C,P)=11.5 Hz; CH–O), 124–160 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₄H₅₅F₃NO₄P: C 70.47, H 7.39, N 1.87; found: C 70.57, H 7.42, N 1.89.

Ligand L3c: Yield: 593 mg (44%); ³¹P NMR (C₆D₆): δ =146.8 ppm; ¹H NMR (C₆D₆): δ =0.39 (s, 9H; CH₃–Si), 0.49 (s, 9H; CH₃–Si), 0.53 (s, 3H; CH₃), 0.77 (s, 3H; CH₃), 1.52 (m, 1H; CH₂), 1.69 (m, 1H; CH₂), 1.95 (m, 1H; CH₂–C=), 2.12 (m, 1H; CH₂–C=), 5.42 (m, 1H; CH–O), 6.9–8.1 ppm (m, 10H; CH=); ¹³C NMR (C₆D₆): δ =0.9 (CH₃–Si), 1.0 (CH₃–Si), 27.7 (CH₃), 30.3 (CH₃), 33.2 (CMe₂), 35.9 (CH₂–C=), 45.0 (CH₂), 67.5 (d, *J*(C,P)=6.1 Hz; CH–O), 124–160 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₃₄H₃₉F₃NO₄PSi₂: C 60.97, H 5.87, N 2.09; found: C 60.99, H 5.89, N 2.07.

Ligand L3e: Yield: 676 mg (43%); ³¹P NMR (C₆D₆): δ =146.3 ppm; ¹H NMR (C₆D₆): δ =0.43 (s, 3 H; CH₃), 0.51 (s, 9 H; CH₃–Si), 0.62 (s, 3 H; CH₃), 0.65 (s, 9 H; CH₃–Si), 1.34 (m, 1 H; CH₂), 1.56 (m, 1 H; CH₂), 1.84 (m, 1 H; CH₂–C=), 2.02 (m, 1 H; CH₂–C=), 5.42 (m, 1 H; CH–O), 6.8–8.1 ppm (m, 14H; CH=); ¹³C NMR (C₆D₆): δ =1.0 (CH₃–Si), 1.1 (CH₃–Si), 27.0 (CH₃), 30.3 (CH₃), 33.2 (CMe₂), 35.8 (CH₂–C=), 45.0 (CH₂), 67.2 (d, *J*(C,P)=5.3 Hz; CH–O), 123–160 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₂H₄₃F₃NO₄PSi₂: C 65.52, H 5.63, N 1.82; found: C 65.34, H 5.59, N 1.86.

Ligand L4a: Yield: 652 mg (49%); ³¹P NMR (C₆D₆): δ =148.0 ppm; ¹H NMR (C₆D₆): δ =0.59 (s, 3H; CH₃), 0.80 (s, 3H; CH₃), 1.28 (s, 9H; CH₃, *t*Bu), 1.29 (s, 9H; CH₃, *t*Bu), 1.40 (s, 9H; CH₃, *t*Bu), 1.26 (m, 1H; CH₂), 1.66 (s, 9H; CH₃, *t*Bu), 1.71 (s, 9H; CH₃, *t*Bu), 1.75 (m, 1H; CH₂), 1.98 (m, 1H; CH₂-C=), 2.14 (m, 1H; CH₂-C=), 5.52 (m, 1H; CH-O), 7.0–7.7 ppm (m, 4H; CH=); ¹³C NMR (C₆D₆): δ =27.8 (CH₃), 29.2 (CH₃), 29.9 (CH₃, *t*Bu), 32.0 (CH₃, *t*Bu), 32.2 (CH₃, *t*Bu), 33.1 (CMe₂), 34.4 (C, *t*Bu), 35.0 (C, *t*Bu), 36.1 (CH₂-C=), 36.2 (C, *t*Bu), 45.3 (CH₂), 68.1 (d, *J*-(C,P)=12 Hz; CH-O), 124–170 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₁H₆₀NO₄P: C 74.40, H 9.14, N 2.12; found: C 74.45, H 9.18, N 2.08.

Ligand L4c: Yield: 709 mg (61%); 31 P NMR (C₆D₆): $\delta = 148.2$ ppm; ¹H NMR (C₆D₆): $\delta = 0.49$ (s, 9H; CH₃-Si), 0.54 (s, 9H; CH₃-Si), 0.57 (s, 3H; CH₃), 0.78 (s, 3H; CH₃), 1.21 (s, 9H; CH₃, tBu), 1.45 (m, 1H; CH₂), 1.69 (m, 1H; CH₂), 1.96 (m, 1H; CH₂-C=), 2.19 (m, 1H; CH₂-C=), 5.48 (m, 1H; CH–O), 6.9–7.8 (m, 6H; CH=); 13 C NMR (C₆D₆): $\delta = 0.2$ (CH₃– Si), 0.4 (CH₃-Si), 27.9 (CH₃), 29.8 (CH₃), 30.2 (CH₃, tBu), 33.2 (CMe₂), 36.3 (CH₂-C=), 45.5 (CH₂), 68.3 (d, J(C,P)=10 Hz; CH-O), 123-155 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C31H44NO4PSi2: C 63.99, H 7.62, N 2.41; found: C 64.03, H 7.65, N 2.39. **Ligand L5a**: Yield: 843 mg (62%); 31 P NMR (C₆D₆): $\delta = 148.7$ ppm; ¹H NMR (C₆D₆): $\delta = 0.57$ (s, 3H; CH₃), 0.77 (s, 3H; CH₃), 1.23 (s, 9H; CH₃, tBu), 1.25 (s, 9H; CH₃, tBu), 1.57 (m, 1H; CH₂), 1.61 (s, 9H; CH₃, *t*Bu), 1.69 (s, 9H; CH₃, *t*Bu), 1.73 (m, 1H; CH₂), 1.92 (m, 1H; CH₂-C=), 2.08 (m, 1H; CH₂-C=), 5.53 (m, 1H; CH-O), 7.1-8.2 ppm (m, 9H; CH=); 13 C NMR (C₆D₆): δ = 27.7 (CH₃), 29.9 (CH₃), 31.9 (CH₃, tBu), 33.1 (C, tBu), 35.0 (CH₂-C=), 36.2 (CMe₂), 45.1 (CH₂), 67.8 (d, J(C,P) =12.2 Hz; CH-O), 124-165 ppm (aromatic carbon atoms); elemental analysis calcd (%) for $\rm C_{43}H_{56}NO_4P\colon C$ 75.74, H 8.28, N 2.05; found: C 75.72, H 8.31, N 2.01.

Ligand L6a: Yield: 738 mg (54%); ³¹P NMR (C_6D_6): $\delta = 137.9$ ppm; ¹H NMR (C_6D_6): $\delta = 1.43$ (s, 9H; CH₃, *tBu*), 1.48 (s, 9H; CH₃, *tBu*), 1.76 (m, 2H; CH₂), 1.82 (s, 9H; CH₃, *tBu*), 1.87 (s, 9H; CH₃, *tBu*), 2.00 (m, 2H; CH₂-CH), 2.46 (m, 2H; CH₂-C=), 3.36 (m, 1H; CH), 4.48 (m, 1H; CH₂-O), 4.83 (m, 1H; CH₂-O), 7.2–8.2 ppm (m, 9H; CH=); ¹³C NMR (C_6D_6): $\delta = 21.4$ (CH₂), 23.9 (CH₂-C=), 26.1 (CH₂-CH), 31.8 (CH₃, *tBu*), 35.0 (C, *tBu*), 36.1 (C, *tBu*), 39.1 (CH), 67.3 (CH₂-O), 125–165 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₂H₅₄NO₃PS: C 73.76, H 7.96, N 2.05; found: C 73.86, H 7.99, N 2.08.

Ligand L6b: Yield: 598 mg (48%); ³¹P NMR (C₆D₆): δ =137.5 ppm; ¹H NMR (C₆D₆): δ =1.33 (m, 2H; CH₂), 1.54 (s, 9H; CH₃, *t*Bu), 1.59 (s, 9H; CH₃, *t*Bu), 1.82 (m, 2H; CH₂-CH), 2.27 (m, 2H; CH₂-C=), 3.15 (m, 1H; CH), 3.29 (s, 3H; CH₃-O), 3.33 (s, 3H; CH₃-O), 4.33 (m, 1H; CH₂-O), 4.71 (m, 1H; CH₂-O), 6.7–8.1 ppm (m, 9H; CH=); ¹³C NMR (C₆D₆): δ =21.7 (CH₂), 23.9 (CH₂-C=), 26.3 (CH₂-CH), 31.3 (CH₃, *t*Bu), 31.4 (CH₃, *t*Bu), 35.9 (C, *t*Bu), 36.0 (C, *t*Bu), 39.2 (CH), 55.4 (CH₃-O), 67.7 (CH₂-O), 112–165 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₃₆H₄₂NO₅PS: C 68.44, H 6.70, N 2.22; found: C 68.48, H 6.72, N 2.20.

Ligand L6c: Yield: 640 mg (53%); ³¹P NMR (C₆D₆): δ =136.5 ppm; ¹H NMR (C₆D₆): δ =0.44 (s, 9H; CH₃–Si), 0.52 (s, 9H; CH₃–Si), 1.36 (m, 1H; CH₂), 1.56 (m, 1H; CH₂), 1.79 (m, 2H; CH₂–CH), 2.27 (m, 2H; CH₂–C=), 3.15 (m, 1H; CH), 4.15 (m, 1H; CH₂–O), 4.68 (m, 1H; CH₂–O), 6.9–8.2 ppm (m, 11H; CH=); ¹³C NMR (C₆D₆): δ =0.7 (CH₃–Si), 0.8 (CH₃–Si), 21.8 (CH₂), 24.2 (CH₂–C=), 26.5 (CH₂–CH), 39.3 (CH), 67.8 (CH₂–O), 125–165 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₃₂H₃₈NO₃PSSi₂: C 63.65, H 6.34, N 2.32; found: C 63.68, H 6.36, N 2.35.

Ligand L6d: Yield: 647 mg (47%); ³¹P NMR (C₆D₆): δ =136.2 ppm; ¹H NMR (C₆D₆): δ =0.58 (s, 9H; CH₃–Si), 0.59 (s, 9H; CH₃–Si), 1.36 (m, 2H; CH₂), 1.72 (m, 2H; *CH*₂–CH), 2.32 (m, 2H; CH₂–C=), 2.94 (m, 1H; CH), 3.96 (m, 1H; CH₂–O), 4.76 (m, 1H; CH₂–O), 6.9–8.3 ppm (m, 15H; CH=); ¹³C NMR (C₆D₆): δ =-0.3 (CH₃–Si), -0.1 (CH₃–Si), 21.3 (CH₂), 23.2 (CH₂–C=), 25.8 (*C*H₂–CH), 38.4 (d, *J*(C,P)=3.2 Hz; CH), 67.1 (CH₂–O), 122–164 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₀H₄₂NO₃PSSi₂: C 68.24, H 6.01, N 1.99; found: C 68.21, H 5.98, N 1.97.

Ligand L6e: Yield: 605 mg (43%); ³¹P NMR (C_6D_6): $\delta = 136.3$ ppm; ¹H NMR (C_6D_6): $\delta = 0.61$ (s, 9H; CH₃-Si), 0.73 (s, 9H; CH₃-Si), 1.31 (m, 1H; CH₂), 1.45 (m, 1H; CH₂), 1.63 (m, 1H; CH₂-CH), 1.88 (m, 1H; CH2-CH), 2.16 (m, 2H; CH2-C=), 3.21 (m, 1H; CH), 4.16 (m, 1H; CH2-O), 4.43 (m, 1H; CH2-O), 6.8-8.4 ppm (m, 15H; CH=); ¹³C NMR $(C_6D_6): \delta = -0.2 \text{ (CH}_3 - \text{Si}), -0.1 \text{ (CH}_3 - \text{Si}), 20.6 \text{ (CH}_2), 23.0 \text{ (CH}_2 - \text{C}_2),$ 25.6 (CH₂-CH), 38.1 (d, J(C,P)=4.0 Hz; CH), 67.1 (CH₂-O), 122-164 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C40H42NO3PSSi2: C 68.24, H 6.01, N 1.99; found: C 68.27, H 6.03, N 2.02. **Ligand L7a**: Yield: 598 mg (42%); 31 P NMR (C₆D₆): $\delta = 151.4$ ppm; ¹H NMR (C₆D₆): $\delta = 1.19$ (s, 3H; CH₃), 1.24 (s, 9H; CH₃, *t*Bu), 1.26 (s, 9H; CH₃, tBu), 1.50 (s, 9H; CH₃, tBu), 1.53 (s, 3H; CH₃), 1.62 (s, 9H; CH₃, tBu), 1.89 (m, 2H; CH₂), 2.12 (m, 2H; CH₂-CH), 2.28 (m, 2H; CH₂-C=), 3.16 (m, 1H; CH), 7.0-8.0 ppm (m, 9H; CH=); ¹³C NMR (C₆D₆): δ = 22.4 (CH₂), 24.3 (CH₂-C=), 26.4 (CH₂-CH), 27.9 (CH₃), 30.4 (CH₃), 31.4 (CH₃, tBu), 31.6 (CH₃, tBu), 31.8 (CH₃, tBu), 35.1 (C, tBu), 36.3 (C, tBu), 36.4 (C, tBu), 40.3 (CH), 71.0 (CMe2), 124-166 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C44H58NO3PS: C 74.23, H 8.21, N 1.97; found: C 74.31, H 8.24, N 1.93.

Ligand L7d: Yield: 638 mg (42%); ³¹P NMR (C₆D₆): δ =143.2 ppm; ¹H NMR (C₆D₆): δ =0.53 (s, 9H; CH₃–Si), 0.56 (s, 9H; CH₃–Si), 1.14 (s, 3H; CH₃), 1.50 (s, 3H; CH₃), 1.77 (m, 2H; CH₂), 2.09 (m, 2H; CH₂– CH), 2.25 (m, 2H; CH₂–C=), 3.17 (m, 1H; CH), 7.0–8.0 ppm (m, 15H; CH<C=>); ¹³C NMR (C₆D₆): δ =-0.3 (CH₃–Si), -0.1 (CH₃–Si), 22.1 (CH₂), 23.8 (CH₂–C<C=>), 25.9 (CH₂–CH), 27.8 (CH₃), 29.9 (CH₃), 39.9 (CH), 70.3 (CH₂–O), 122–166 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₄H₅₀NO₃PSSi₂: C 69.53, H 6.63, N 1.84; found: C 69.55, H 6.64, N 1.80.

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Ligand L8a: Yield: 798 mg (56%); ³¹P NMR (C_6D_6): δ =151.4 ppm; ¹H NMR (C_6D_6): δ =1.24 (s, 9H; CH₃, tBu), 1.26 (s, 9H; CH₃, tBu), 1.50 (s, 9H; CH₃, tBu), 1.53 (s, 3H; CH₃), 1.62 (s, 9H; CH₃, tBu), 1.89 (m, 2H; CH₂), 2.12 (m, 2H; *CH*₂-CH), 2.28 (m, 2H; CH₂-C=), 3.16 (m, 1H; CH), 7.0-8.0 ppm (m, 9H; CH=); ¹³C NMR (C_6D_6): δ =22.4 (CH₂), 24.3 (CH₂-C=), 26.4 (*CH*₂-CH), 27.9 (CH₃), 30.4 (CH₃), 31.4 (CH₃, tBu), 31.6 (CH₃, tBu), 31.8 (CH₃, tBu), 35.1 (C, tBu), 36.3 (C, tBu), 36.4 (C, tBu), 40.3 (CH), 71.0 (CMe₂), 124-166 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₄H₅₈NO₃PS: C 74.23, H 8.21, N 1.97; found: C 74.28, H 8.21, N 1.99.

Ligand L8d: Yield: 808 mg (50%); ³¹P NMR (C₆D₆): δ =143.1 ppm; ¹H NMR (C₆D₆): δ =0.49 (s, 9H; CH₃–Si), 0.59 (s, 9H; CH₃–Si), 1.19 (s, 3H; CH₃), 1.53 (s, 3H; CH₃), 1.74 (m, 2H; CH₂), 2.04 (m, 2H; CH₂–CH), 2.21 (m, 2H; CH₂–C=), 3.32 (m, 1H; CH), 7.0–8.0 ppm (m, 15H; CH=); ¹³C NMR (C₆D₆): δ =-0.1 (CH₃–Si), 0.1 (CH₃–Si), 21.9 (CH₂), 23.4 (CH₂–C=), 26.3 (CH₂–CH), 27.9 (CH₃), 30.2 (CH₃), 39.3 (CH), 69.9 (CH₂–O), 122–166 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₄H₃₀NO₃PSSi₂: C 69.53, H 6.63, N 1.84; found: C 69.57, H 6.66, N 1.86.

General procedure for the preparation of $[Pd(\eta^3-allyl)(L)]BF_4$ complexes 26–35: The corresponding ligand (0.05 mmol) and the complex $[\{Pd(\mu-Cl)(\eta^3-1,3-allyl)\}_2]$ (0.025 mmol) were dissolved in CD_2Cl_2 (1.5 mL) at room temperature under argon. AgBF₄ (9.8 mg, 0.5 mmol) was added after 30 min and the mixture was stirred for 30 min. The mixture was then filtered over Celite under argon and the resulting solutions were analyzed by NMR spectroscopy. After the NMR spectroscopic analysis, the complexes were precipitated adding hexane as pale yellow solids.

Complex 26: Isomer A (90%): ³¹P NMR (CD₂Cl₂, 263 K): $\delta = 135.6$ ppm (s, 1P); ¹H NMR (CD₂Cl₂, 263 K): $\delta = 0.97$ (s, 3H; CH₃), 1.05 (s, 3H; CH₃), 1.21 (s, 9H; CH₃, tBu), 1.25 (s, 9H; CH₃, tBu), 1.42 (s, 9H; CH₃, tBu), 1.59 (s, 9H; CH₃, tBu), 1.72 (m, 1H; CH₂), 1.84 (m, 1H; CH₂), 1.96 (m, 1H; CH2-C=), 2.23 (m, 1H; CH2-C=), 5.14 (m, 1H; CH allyl trans to N), 5.39 (m, 1H; CH-O), 5.99 (m, 1H; CH allyl trans to P), 6.55 (m, 1H; CH allyl central), 6.7-8.0 ppm (m, 19H; CH=); ¹³C NMR (C₆D₆, 263 K): δ=26.3 (CH₃), 29.8 (CH₃), 31.3 (CH₃, tBu), 31.5 (CH₃, tBu), 32.5 (C, tBu), 34.8 (CH2-C=), 35.6 (CMe2), 42.5 (CH2), 70.7 (br; CH-O), 81.4 (m; CH allyl trans to N), 95.9 (m; CH trans to P), 112.5 (m; CH allyl central), 125–164 ppm (aromatic carbon atoms). Isomer **B** (10%): 31 P NMR $(CD_2Cl_2, 263 \text{ K}): \delta = 140.4 \text{ ppm} (s, 1P); {}^{1}\text{H NMR} (CD_2Cl_2, 263 \text{ K}): \delta =$ 0.99 (s, 3H; CH₃), 1.03 (s, 3H; CH₃), 1.14 (s, 9H; CH₃, tBu), 1.17 (s, 9H; CH₃, tBu), 1.40 (s, 9H; CH₃, tBu), 1.55 (s, 9H; CH₃, tBu), 1.85 (m, 1H; CH₂), 1.92 (m, 1H; CH₂-C=), 2.10 (m, 1H; CH₂-C=), 4.84 (m, 1H; CH allyl trans to N), 5.34 (m, 1H; CH-O), 5.21 (m, 1H; CH allyl trans to P), 6.45 (m, 1H; CH allyl central), 6.7-8.0 ppm (m, 19H; CH=); ¹³C NMR (C₆D₆, 263 K): δ = 26.8 (CH₃), 30.1 (CH₃), 31.4 (CH₃, tBu), 31.6 (CH₃, tBu), 32.0 (C, tBu), 35.1 (CH2-C=), 35.8 (CMe2), 42.3 (CH2), 69.7 (br; CH-O), 75.0 (m; CH allyl trans to N), 95.4 (m; CH trans to P), 112.7 (m; CH allyl central), 125-164 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C58H69BF4NO4PPd: C 65.20, H 6.51, N 1.31; found: C 65.43, H 6.61, N 1.35.

Complex 27: Isomer A (56%): ³¹P NMR (CD₂Cl₂, 243 K): $\delta = 132.5$ ppm (s, 1P); ¹H NMR (CD₂Cl₂, 243 K): $\delta = 1.45$ (s, 9H; CH₃, tBu), 1.49 (s, 9H; CH₃, tBu), 1.65 (s, 9H; CH₃, tBu), 1.69 (s, 9H; CH₃, tBu), 1.82 (m, 2H; CH₂), 2.03 (m, 2H; CH₂-CH), 2.56 (m, 2H; CH₂-C=), 3.43 (m, 1H; CH), 4.56 (m, 1H; CH2-O), 4.93 (m, 1H; CH allyl trans to N), 4.98 (m, 1H; CH₂-O), 5.87 (m, 1H; CH allyl trans to P), 6.33 (m, 1H; CH allyl central), 7.0–8.4 ppm (m, 19H; CH=); 13 C NMR (CD₂Cl₂, 243 K): $\delta =$ 22.1 (CH₂), 24.2 (CH₂-C=), 25.9 (CH₂-CH), 31.8-32.9 (br; CH₃, tBu), 35.0 (C, tBu), 35.8 (C, tBu), 39.3 (CH), 69.1 (CH2-O), 80.4 (m; CH allyl trans to N), 98.3 (m; CH trans to P), 112.1 (m; CH allyl central), 125-165 ppm (aromatic carbon atoms). Isomer **B** (44%): 31 P NMR (CD₂Cl₂, 243 K): $\delta = 136.9$ ppm (s, 1 P); ¹H NMR (CD₂Cl₂, 243 K): $\delta = 1.42$ (s, 9 H; CH₃, tBu), 1.52 (s, 9H; CH₃, tBu), 1.72 (s, 18H; CH₃, tBu), 1.89 (m, 2H; CH₂), 2.12 (m, 2H; CH₂-CH), 2.63 (m, 2H; CH₂-C=), 3.49 (m, 1H; CH), 4.53 (m, 1H; CH2-O), 4.88 (m, 1H; CH allyl trans to N), 5.13 (m, 1H; CH2-O), 5.45 (m, 1H; CH allyl trans to P), 6.28 (m, 1H; CH allyl central), 7.0–8.4 ppm (m, 19H; CH=); 13 C NMR (CD₂Cl₂, 243 K): $\delta =$ 22.3 (CH₂), 24.5 (CH₂-C=), 26.1 (CH₂-CH), 31.8-32.9 (br; CH₃, tBu), 35.3 (C, *t*Bu), 35.5 (C, *t*Bu), 39.2 (CH), 70.0 (CH₂–O), 77.2 (m; CH allyl *trans* to N), 96.2 (m; CH *trans* to P), 112.0 (m; CH allyl central), 125–165 ppm (aromatic carbon atoms); elemental analysis calcd (%) for $C_{57}H_{67}BF_4NO_3PPdS$: C 63.96, H 6.31, N 1.31; found: C 64.03, H 6.32, N 1.33.

Complex 28: Isomer A (70%): ³¹P NMR (CD₂Cl₂, 233 K): $\delta = 139.2$ ppm (s, 1P); ¹H NMR (CD₂Cl₂, 233 K): $\delta = 0.62$ (s, 3H; CH₃), 0.89 (s, 3H; CH₃), 1.31 (s, 18H; CH₃, tBu), 1.43 (s, 9H; CH₃, tBu), 1.54 (s, 9H; CH₃, tBu), 1.69 (m, 1H; CH₂), 1.74 (s, 9H; CH₃, tBu), 1.86 (m, 1H; CH₂), 2.01 (m, 1H; CH₂-C=), 2.24 (m, 1H; CH₂-C=), 5.43 (m, 1H; CH allyl trans to N), 5.57 (m, 1H; CH-O), 6.02 (m, 1H; CH allyl trans to P), 6.21 (m, 1H; CH allyl central), 6.8-8.0 ppm (m, 14H; CH=); ¹³C NMR (CD₂Cl₂, 233 K): δ=28.1 (CH₃), 29.4 (CH₃), 30.2-32.5 (CH₃, tBu), 33.5 (CMe₂), 34.5 (C, tBu), 34.7 (C, tBu), 35.2 (C, tBu), 36.2 (CH2-C=), 36.4 (C, tBu), 45.5 (CH₂), 68.4 (m; CH–O), 81.4 (m; CH allyl trans to N), 95.9 (m; CH trans to P), 112.3 (m; CH allyl central), 124-170 ppm (aromatic carbon atoms). Product C (30%): ³¹P NMR (CD₂Cl₂, 233 K): $\delta = 143.1$ ppm (s, 1P); ¹H NMR (CD₂Cl₂, 233 K): $\delta = 0.59$ (s, 6H; CH₃), 0.84 (s, 6H; CH₃), 1.29 (s, 18H; CH₃, tBu), 1.33 (s, 18H; CH₃, tBu), 1.59 (s, 36H; CH₃, tBu), 1.64 (m, 2H; CH₂), 1.71 (s, 18H; CH₃, tBu), 1.81 (m, 2H; CH₂), 2.06 (m, 2H; CH2-C=), 2.18 (m, 2H; CH2-C=), 5.32 (m, 1H; CH allyl), 5.52 (m, 2H; CH-O), 5.59 (m, 1H; CH allyl), 6.19 (m, 1H; CH allyl central), 6.8-8.0 ppm (m, 18H; CH=); 13 C NMR (CD₂Cl₂, 233 K): δ = 28.2 (CH₃), 29.6 (CH₃), 30.2-32.5 (CH₃, tBu), 33.7 (CMe₂), 34.3 (C, tBu), 34.9 (C, tBu), 35.0 (C, tBu), 35.5 (C, tBu), 36.3 (CH2-C=), 36.9 (C, tBu), 45.3 (CH2), 69.2 (m; CH-O), 99.7 (br; CH allyl), 112.5 (m; CH allyl central), 124-170 ppm (aromatic carbon atoms); elemental analysis calcd (%) for $0.7\,C_{56}H_{73}BF_4NO_4PPd+0.3\,C_{97}H_{133}BF_4N_2O_8P_2Pd\colon C\ 65.79,\ H\ 7.36,\ N\ 0.87;$ found: C 65.87, H 7.42, N 0.92.

Complex 29: Isomer A (83%): ³¹P NMR (CD₂Cl₂, 273 K): $\delta = 138.1$ ppm (s, 1P); ¹H NMR (C₆D₆, 273 K): $\delta = 0.22$ (s, 9H; CH₃-Si), 0.75 (s, 9H; CH₃-Si), 1.04 (s, 3H; CH₃), 1.18 (s, 3H; CH₃), 1.92 (m, 1H; CH₂), 2.21 (m, 1H; CH₂), 2.53 (m, 2H; CH₂-C=), 4.97 (m, 1H; CH allyl trans to N), 5.38 (m, 1H; CH-O), 6.06 (m, 1H; CH allyl trans to P), 6.42 (m, 1H; CH allyl central), 6.6–8.3 ppm (m, 25H; CH=); ^{13}C NMR (C_6D_6, 273 K): $\delta = 0.7$ (CH₃-Si), 1.0 (CH₃-Si), 28.2 (CH₃), 32.9 (CMe₂), 35.2 (CH₂-C=), 42.5 (d, *J*(C,P)=1.2 Hz; CH₂), 70.5 (d, *J*(C,P)=7.2 Hz; CH–O), 75.4 (m; CH allyl trans to N), 96.7 (m; CH allyl trans to P), 112.4 (m; CH allyl central), 121-165 ppm (aromatic carbon atoms). Isomer B (17%): ³¹P NMR (CD₂Cl₂, 273 K): $\delta = 140.5$ ppm (s, 1 P); ¹H NMR (C₆D₆, 273 K): $\delta = 0.56$ (s, 9H; CH₃-Si), 0.69 (s, 9H; CH₃-Si), 1.04 (s, 3H; CH₃), 1.07 (s, 3H; CH₃), 1.70 (m, 1H; CH₂), 1.98 (m, 1H; CH₂), 2.21 (m, 2H; CH₂-C=), 5.19 (m, 1H; CH allyl trans to N), 5.32 (m, 1H; CH allyl trans to P), 5.42 (m, 1H; CH-O), 6.49 (m, 1H; CH allyl central), 6.6-8.3 ppm (m, 25H; CH=); ¹³C NMR (C_6D_6 , 273 K): $\delta = 1.2$ (CH₃-Si), 1.5 (CH₃-Si), 26.3 (CH₃), 29.5 (CH₃), 33.4 (CMe₂), 35.0 (CH₂-C=), 42.7 (d, J(C,P)= 3.2 Hz; CH₂), 68.5 (d, J(C,P)=7.2 Hz; CH-O), 70.7 (m; CH allyl trans to N), 96.1 (m; CH allyl trans to P), 110.7 (m; CH allyl central), 121-165 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₅₆H₅₇BF₄NO₄PPdSi₂: C 61.80, H 5.28, N 1.29; found: C 61.92, H 5.34, N 1.33.

Complex 30: Isomer A (28%): ³¹P NMR (CD₂Cl₂, 223 K): $\delta = 140.6$ ppm (s, 1P); ¹H NMR (C₆D₆, 223 K): $\delta = 0.04$ (s, 9H; CH₃–Si), 0.65 (s, 9H; CH₃-Si), 0.98 (s, 3H; CH₃), 1.20 (s, 3H; CH₃), 1.64 (m, 1H; CH₂), 2.03 (m, 1H; CH₂), 2.54 (m, 2H; CH₂-C=), 4.80 (m, 1H; CH allyl trans to N), 5.33 (m, 1H; CH-O), 5.77 (m, 1H; CH allyl trans to P), 6.17 (m, 1H; CH allyl central), 6.4–8.4 ppm (m, 25H; CH=); ¹³C NMR (C₆D₆, 223 K): $\delta = -0.5$ (CH₃-Si), 1.1 (CH₃-Si), 25.1 (CH₃), 30.2 (CH₃), 33.5 (CMe₂), 34.8 (CH₂-C=), 42.6 (d, J(C,P)=10.2 Hz; CH₂), 69.3 (CH-O), 71.1 (d, J-(C,P)=8.0 Hz; CH allyl trans to N), 98.4 (m; CH allyl trans to P), 110.9 (m; CH allyl central), 120-165 ppm (aromatic carbon atoms). Isomer B (72 %): ³¹P NMR (CD₂Cl₂, 223 K): $\delta = 141.1$ ppm (s, 1 P); ¹H NMR (C₆D₆, 223 K): δ=0.25 (s, 9H; CH₃-Si), 0.69 (s, 9H; CH₃-Si), 1.07 (s, 3H; CH₃), 1.14 (s, 3H; CH₃), 1.67 (m, 1H; CH₂), 2.18 (m, 1H; CH₂), 2.62 (m, 2H; CH2-C=), 4.74 (m, 1H; CH allyl trans to N), 5.30 (m, 1H; CH allyl trans to P), 5.37 (m, 1H; CH-O), 6.21 (m, 1H; CH allyl central), 6.4-8.4 ppm (m, 25 H; CH=); 13 C NMR (C₆D₆, 223 K): $\delta = 0.3$ (CH₃-Si), 1.4 (CH₃-Si), 25.2 (CH₃), 30.8 (CH₃), 33.5 (CMe₂), 34.8 (CH₂–C=), 43.0 (d, J(C,P) =

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9.4 Hz; CH₂), 69.2 (CH–O), 70.5 (d, J(C,P) = 9.6 Hz; CH allyl *trans* to N), 93.0 (d, J(C,P) = 36.8 Hz; CH allyl *trans* to P), 111.2 (m; CH allyl central), 120–165 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₅₆H₅₇BF₄NO₄PPdSi₂: C 61.80, H 5.28, N 1.29; found: C 61.91, H 5.37, N 1.34.

Complex 31: Isomer A (46%): ³¹P NMR (CD₂Cl₂, 295 K): $\delta = 134.3$ ppm (s, 1P); $^1\!\mathrm{H}\,\mathrm{NMR}$ (CD_2Cl_2, 295 K): $\delta\!=\!0.84$ (m, 3H; CH_3 allyl), 1.12 (m, 3H; CH₃ allyl), 1.15 (s, 3H; CH₃), 1.29 (s, 3H; CH₃), 1.35 (s, 9H; CH₃, tBu), 1.39-1.42 (s, 18H; CH₃, tBu), 1.59 (s, 9H; CH₃, tBu), 1.95 (m, 1H; CH₂), 2.35 (m, 1H; CH₂), 2.69 (m, 1H; CH₂-C=), 2.73 (m, 1H; CH₂-C=), 4.12 (m, 1H; CH allyl trans to N), 5.03 (m, 1H; CH allyl trans to P), 5.36 (m, 1H; CH allyl central), 5.46 (m, 1H; CH-O), 7.1-8.1 ppm (m, 9H; CH=); ¹³C NMR (CD₂Cl₂, 295 K): $\delta = 16.3$ (CH₃ allyl), 19.4 (CH₃ allyl), 26.8 (CH₃), 29.9 (CH₃), 31.4–32.1 (CH₃, tBu), 32.5–33.6 (C, tBu), 35.2 (CH2-C=), 36.0 (CMe2), 42.9 (br; CH2), 70.6 (br; CH-O), 72.3 (m; CH allyl trans to N), 107.3 (m; CH trans to P), 115.3 (m; CH allyl central), 123–164 ppm (aromatic carbon atoms). Isomer B (18%): ³¹P NMR $(CD_2Cl_2, 295 \text{ K}): \delta = 135.2 \text{ ppm} (s, 1 \text{ P}); {}^{1}\text{H NMR} (CD_2Cl_2, 295 \text{ K}): \delta =$ $0.90 \ (m, 3H; CH_3 \ allyl), 1.05 \ (m, 3H; CH_3 \ allyl), 1.18 \ (s, 3H; CH_3), 1.28$ (s, 3H; CH₃), 1.39-1.42 (s, 18H; CH₃, tBu), 1.49 (s, 9H; CH₃, tBu), 1.53 (s, 9H; CH₃, tBu), 1.98 (m, 1H; CH₂), 2.22 (m, 1H; CH₂), 2.73 (m, 1H; CH2-C=), 2.78 (m, 1H; CH2-C=), 4.50 (m, 1H; CH allyl trans to N), 4.73 (m, 1H; CH allyl trans to P), 5.42 (m, 1H; CH allyl central), 5.52 (m, 1H; CH-O), 7.1-8.1 ppm (m, 9H; CH=); ¹³C NMR (CD₂Cl₂, 295 K): $\delta = 16.2$ (CH₃ allyl), 17.5 (CH₃ allyl), 27.3 (CH₃), 29.4 (CH₃), 31.4-32.1 (CH₃, tBu), 32.5-33.6 (C, tBu), 35.4 (CH₂-C=), 36.3 (CMe₂), 42.9 (br; CH₂), 70.2 (br; CH-O), 74.5 (m; CH allyl trans to N), 106.3 (m; CH trans to P), 116.8 (m; CH allyl central), 123-164 ppm (aromatic carbon atoms). Isomer **D** (36%): ³¹P NMR (CD₂Cl₂, 295 K): $\delta = 134.7$ ppm (s, 1P); ¹H NMR (CD₂Cl₂, 295 K): $\delta = 0.54$ (m, 3H; CH₃ allyl), 0.95 (m, 3H; CH₃ allyl), 1.20 (s, 3H; CH₃), 1.22 (s, 3H; CH₃), 1.39–1.42 (s, 9H; CH₃, tBu), 1.45 (s, 9H; CH₃, tBu), 1.54 (s, 9H; CH₃, tBu), 1.60 (s, 9H; CH₃, *t*Bu), 1.98 (m, 1H; CH₂), 2.20 (m, 1H; CH₂), 2.78 (m, 1H; CH₂-C=), 2.82 (m, 1H; CH2-C=), 3.61 (m, 1H; CH allyl trans to N), 3.87 (m, 1H; CH allyl trans to P), 5.23 (m, 1H; CH allyl central), 5.71 (m, 1H; CH-O), 7.1–8.1 ppm (m, 9H; CH=); ¹³C NMR (CD₂Cl₂, 295 K): δ=17.5 (CH₃ allyl), 18.8 (CH3 allyl), 27.5 (CH3), 29.4 (CH3), 31.4-32.1 (CH3, tBu), 32.5-33.6 (C, tBu), 35.5 (CH2-C=), 36.2 (CMe2), 42.9 (br; CH2), 69.8 (br; CH-O), 70.6 (m; CH allyl trans to N), 95.0 (m; CH trans to P), 118.6 (m; CH allyl central), 123-164 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C48H65BF4NO4PPd: C 61.06, H 6.94, N 1.48; found: C 61.11, H 6.96, N 1.49.

Complex 32: Isomer A (40%): ³¹P NMR (CD₂Cl₂, 253 K): $\delta = 127.4$ ppm (s, 1P); ¹H NMR (CD₂Cl₂, 253 K): $\delta = 0.63$ (m, 3H; CH₃ allyl), 0.85 (m, 3H; CH₃ allyl), 1.19 (s, 3H; CH₃), 1.26 (s, 3H; CH₃), 1.37 (s, 9H; CH₃, tBu), 1.39 (s, 9H; CH₃, tBu), 1.44 (s, 9H; CH₃, tBu), 1.51 (s, 9H; CH₃, tBu), 1.85 (m, 2H; CH₂), 2.06 (m, 2H; CH₂-CH), 2.22 (m, 2H; CH₂-C=), 2.73 (m, 1H; CH2-C=), 3.42 (m, 1H; CH allyl trans to N), 4.26 (m, 1H; CH-O), 4.56 (m, 1H; CH allyl trans to P), 4.91 (m, 1H; CH allyl central), 7.1–8.2 ppm (m, 9H; CH=); 13 C NMR (CD₂Cl₂, 253 K): $\delta = 17.0$ (CH₃ allyl), 18.1 (CH₃ allyl), 21.0 (CH₂), 23.8 (CH₂-C=), 26.0 (CH₂-CH), 26.1 (CH₃), 29.5 (CH₃), 31.4-32.2 (CH₃, tBu), 35.1-36.2 (C, tBu), 46.5 (CH), 64.8 (m; CH allyl trans to N), 78.3(CMe22), 107.8 (m; CH trans to P), 115.8 (m; CH allyl central), 125-170 ppm (aromatic carbon atoms). Isomer **B** (8%): 31 P NMR (CD₂Cl₂, 253 K): $\delta = 129.1$ ppm (s, 1P); ¹H NMR (CD₂Cl₂, 253 K): $\delta = 0.53$ (m, 3H; CH₃ allyl), 0.74 (m, 3H; CH₃ allyl), 1.15 (s, 3H; CH₃), 1.31 (s, 3H; CH₃), 1.32 (s, 9H; CH₃, tBu), 1.53 (s, 9H; CH₃, tBu), 1.62 (s, 9H; CH₃, tBu), 1.71 (s, 9H; CH₃, tBu), 1.87 (m, 2H; CH₂), 2.06 (m, 2H; CH₂-CH), 2.22 (m, 2H; CH₂-C=), 2.73 (m, 1H; CH₂-C=), 3.48 (m, 1H; CH allyl trans to N), 3.71 (m, 1H; CH allyl trans to P), 4.11 (m, 1H; CH-O), 5.07 (m, 1H; CH allyl central), 7.1-8.2 ppm (m, 9H; CH=); 13 C NMR (CD₂Cl₂, 253 K): $\delta = 17.2$ (CH₃ allyl), 18.6 (CH₃ allyl), 20.7 (CH₂), 23.8 (CH₂-C=), 25.7 (CH₂-CH), 26.8 (CH₃), 29.1 (CH₃), 31.4–32.2 (CH₃, tBu), 35.1–36.2 (C, tBu), 46.7 (CH), 65.7 (m; CH allyl trans to N), 78.1 (CMe2), 106.7 (m; CH trans to P), 115.9 (m; CH allyl central), 125–170 ppm (aromatic carbon atoms). Isomer ${\bf D}$ (52%): ³¹P NMR (CD₂Cl₂, 253 K): $\delta = 128.0$ ppm (s, 1 P); ¹H NMR $(CD_2Cl_2, 253 \text{ K}): \delta = 0.59 \text{ (m, 3H; CH}_3 \text{ allyl}), 0.88 \text{ (m, 3H; CH}_3 \text{ allyl}),$ 1.09 (s, 3H; CH₃), 1.21 (s, 3H; CH₃), 1.35 (s, 9H; CH₃, *t*Bu), 1.46 (s, 9H; CH₃, *t*Bu), 1.50 (s, 9 H; CH₃, *t*Bu), 1.78 (s, 9 H; CH₃, *t*Bu), 1.94 (m, 2 H; CH₂), 2.06 (m, 2 H; *CH*₂–CH), 2.22 (m, 2 H; CH₂–C=), 2.73 (m, 1 H; CH₂–C=), 3.18 (m, 1 H; CH allyl *trans* to N), 3.83 (m, 1 H; CH allyl *trans* to P), 4.05 (m, 1 H; CH–O), 4.89 (m, 1 H; CH allyl central), 7.1–8.2 ppm (m, 9 H; CH=); ¹³C NMR (CD₂Cl₂, 253 K): δ =18.0 (CH₃ allyl), 18.3 (CH₃ allyl), 21.1 (CH₂), 23.8 (CH₂–C=), 25.9 (CH₂–CH), 26.7 (CH₃), 30.2 (CH₃), 31.4-32.2 (CH₃, *t*Bu), 35.1–36.2 (C, *t*Bu), 46.5 (CH), 66.8 (m; CH allyl *trans* to N), 78.2 (CMe₂), 95.4 (m; CH *trans* to P), 115.7 (m; CH allyl central), 125–170 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C₄₉H₆₇BF₄NO₃PPdS: C 60.40, H 6.93, N 1.44; found: C 60.37, H 6.90, N 1.43.

Complex 33: Isomer A (45%): ³¹P NMR (CD₂Cl₂, 253 K): $\delta = 132.2$ ppm (s, 1P); ¹H NMR (CD₂Cl₂, 253 K): $\delta = 0.42$ (m, 9H; CH₃-Si), 0.60 (m, 9H; CH3-Si), 0.68 (m, 3H; CH3 allyl), 0.87 (m, 3H; CH3 allyl), 1.21 (s, 3H; CH₃), 1.28 (s, 3H; CH₃), 1.84 (m, 2H; CH₂), 2.11 (m, 2H; CH₂-CH), 2.24 (m, 2H; CH2-C=), 2.69 (m, 1H; CH2-C=), 3.89 (m, 1H; CH allyl trans to N), 4.13 (m, 1H; CH-O), 4.26 (m, 1H; CH allyl trans to P), 4.98 (m, 1H; CH allyl central), 7.0-8.4 ppm (m, 15H; CH=); ¹³C NMR $(CD_2Cl_2, 253 \text{ K}): \delta = 0.8 (CH_3-Si), 1.4 (CH_3-Si), 16.5 (CH_3 allyl), 19.4$ (CH₃ allyl), 21.5 (CH₂), 24.2 (CH₂-C=), 26.3 (CH₂-CH), 26.6 (CH₃), 29.9 (CH₃), 45.8 (CH), 73.1 (m; CH allyl trans to N), 76.4 (CMe₂), 104.3 (m; CH trans to P), 115.3 (m; CH allyl central), 125-170 ppm (aromatic carbon atoms). Isomer **B** (5%): ³¹P NMR (CD₂Cl₂, 253 K): $\delta = 136.3$ ppm (s, 1P); ¹H NMR (CD₂Cl₂, 253 K): $\delta = 0.39$ (m, 9H; CH₃-Si), 0.57 (m, 9H; CH₃-Si), 0.74 (m, 3H; CH₃ allyl), 1.04 (m, 3H; CH₃ allyl), 1.24 (s, 3H; CH₃), 1.31 (s, 3H; CH₃), 1.86 (m, 2H; CH₂), 2.11 (m, 2H; CH₂-CH), 2.24 (m, 2H; CH2-C=), 2.69 (m, 1H; CH2-C=), 3.90 (m, 1H; CH allyl trans to N), 4.10 (m, 1H; CH-O), 4.15 (m, 1H; CH allyl trans to P), 4.95 (m, 1H; CH allyl central), 7.0-8.4 ppm (m, 15H; CH=); ¹³C NMR $(CD_2Cl_2, 253 \text{ K}): \delta = 0.5 (CH_3-Si), 1.2 (CH_3-Si), 16.2 (CH_3 allyl), 19.4$ (CH₃ allyl), 21.6 (CH₂), 24.1 (CH₂-C=), 26.2 (CH₂-CH), 26.7 (CH₃), 30.1 (CH₃), 45.6 (CH), 74.0 (m; CH allyl trans to N), 76.3 (CMe₂), 102.1 (m; CH trans to P), 116.1 (m; CH allyl central), 125-170 ppm (aromatic carbon atoms). Isomer **D** (50%): ³¹P NMR (CD₂Cl₂, 253 K): $\delta =$ 134.6 ppm (s, 1P); ¹H NMR (CD₂Cl₂, 253 K): δ =0.51 (m, 9H; CH₃-Si), 0.58 (m, 9H; CH₃-Si), 0.75 (m, 3H; CH₃ allyl), 0.82 (m, 3H; CH₃ allyl), 1.27 (s, 3H; CH₃), 1.41 (s, 3H; CH₃), 1.84 (m, 2H; CH₂), 2.11 (m, 2H; *CH*₂–CH), 2.24 (m, 2H; CH₂–C=), 2.69 (m, 1H; CH₂–C=), 3.64 (m, 1H; CH allyl trans to N), 3.98 (m, 1H; CH allyl trans to P), 4.18 (m, 1H; CH-O), 4.79 (m, 1H; CH allyl central), 7.0-8.4 ppm (m, 15H; CH=); ¹³C NMR (CD₂Cl₂, 253 K): $\delta = 0.9$ (CH₃-Si), 1.0 (CH₃-Si), 16.4 (CH₃ allyl), 19.2 (CH3 allyl), 21.7 (CH2), 24.0 (CH2-C=), 26.1 (CH2-CH), 26.9 (CH₃), 30.2 (CH₃), 45.9 (CH), 71.3 (m; CH allyl trans to N), 76.3 (CMe₂), 96.1 (m; CH trans to P), 115.5 (m; CH allyl central), 125-170 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C47H55BF4NO3PPdSSi2: C 56.77, H 5.58, N 1.41; found: C 56.73, H 5.55, N 1.38.

Complex 34: Isomer A (15%): ³¹P NMR (CD₂Cl₂): $\delta = 139.1$ ppm (s, 1P); ¹H NMR (CD₂Cl₂): $\delta = 0.32$ (s, 9H; CH₃-Si), 0.44 (s, 9H; CH₃-Si), 1.09 (s, 3H; CH₃), 1.16 (m, 3H; CH₃), 1.39 (m, 2H; CH₂, allyl), 1.49 (m, 2H; CH₂), 1.72 (m, 4H; CH₂ allyl), 1.92 (m, 1H; CH₂-CH), 2.14 (m, 1H; CH2-CH), 2.72 (CH2-C=), 4.94 (m, 1H; CH allyl trans to N), 5.49 (m, 1H; CH-O), 5.79 (m, 1H; CH allyl central), 5.98 (m, 1H; CH allyl trans to P), 6.8–8.4 ppm (m, 15 H; CH=); 13 C NMR (CD₂Cl₂): $\delta = 0.3$ (CH₃–Si), 19.1 (CH2 allyl), 26.3 (CH2 allyl), 28.2 (CH3), 28.7 (CH3), 31.0 (CH2 allyl), 33.0 (CMe2), 35.5 (CH2-C=), 42.2 (m; CH2), 69.7 (CH-O), 70.6 (m; CH allyl trans to N), 103.2 (d, J(C,P)=38.2 Hz; CH allyl trans to P), 112.7 (m; CH allyl central), 121-167 ppm (aromatic carbon atoms). Isomer **B** (85%): 31 P NMR (CD₂Cl₂): $\delta = 138.7$ ppm (s, 1P); 1 H NMR $(CD_2Cl_2): \delta = 0.37$ (s, 9H; CH₃-Si), 0.52 (s, 9H; CH₃-Si), 0.94 (m, 2H; CH₂, allyl), 1.14 (s, 3H; CH₃), 1.23 (s, 3H; CH₃), 1.42 (m, 4H; CH₂ and CH₂ allyl), 1.62 (m, 2H; CH₂ allyl), 2.03 (m, 1H; CH₂-CH), 2.26 (m, 1H; CH2-CH), 2.75 (CH2-C=), 3.72 (m, 1H; CH allyl trans to N), 5.30 (m, 1H; CH allyl central), 5.42 (m, 1H; CH allyl trans to P), 5.52 (m, 1H; CH–O), 6.8–8.4 ppm (m, 15H; CH=); 13 C NMR (CD₂Cl₂): $\delta = 0.9$ (CH₃-Si), 1.1 (CH₃-Si), 20.2 (CH₂ allyl), 25.5 (CH₂ allyl), 26.5 (CH₃), 29.1 (CH₃), 30.9 (CH₂ allyl), 33.6 (CMe₂), 35.5 (CH₂-C=), 43.7 (CH₂), 69.9 (m; CH-O), 70.4 (m; CH allyl trans to N), 106.2 (d, J(C,P) = 38.2 Hz; CH allyl trans to P), 111.3 (m; CH allyl central), 121-167 ppm

(aromatic carbon atoms); elemental analysis calcd (%) for $C_{47}H_{53}BF_4NO_4PPdSi_2$: C 57.82, H 5.47, N 1.43; found: C 57.91, H 5.51, N 1.45.

Complex 35: Isomer A (87%): ³¹P NMR (CD₂Cl₂): $\delta = 141.6$ ppm (s, 1P); ¹H NMR (CD₂Cl₂): $\delta = 0.53$ (s, 9H; CH₃-Si), 0.56 (s, 9H; CH₃-Si), 0.89 (m, 2H; CH₂, allyl), 1.15 (s, 3H; CH₃), 1.21 (m, 2H; CH₂, allyl), 1.32 (s, 3H; CH₃), 1.36 (m, 2H; CH₂), 1.45 (m, 2H; CH₂ allyl), 1.96 (m, 1H; CH2-CH), 2.17 (m, 1H; CH2-CH), 2.72 (CH2-C=), 3.87 (m, 1H; CH allyl trans to N), 5.21 (m, 1H; CH allyl central), 5.32 (m, 1H; CH-O), 5.36 (m, 1H; CH allyl *trans* to P), 6.8-8.4 ppm (m, 15H; CH=); ¹³C NMR $(CD_2Cl_2): \delta = 0.9 (CH_3-Si), 1.1 (CH_3-Si), 20.2 (CH_2 allyl), 25.5 (CH_2)$ allyl), 26.5 (CH₃), 29.1 (CH₃), 30.9 (CH₂ allyl), 33.6 (CMe₂), 35.5 (CH₂-C=), 43.7 (CH₂), 68.6 (m; CH allyl trans to N), 69.9 (m; CH-O), 107.6 (d, J(C,P)=38.2 Hz; CH allyl trans to P), 111.3 (m; CH allyl central), 121–167 ppm (aromatic carbon atoms). Isomer **B** (13%): 31 P NMR $(CD_2Cl_2): \delta = 140.3 \text{ ppm}$ (s, 1P); ¹H NMR $(CD_2Cl_2): \delta = 0.54$ (s, 9H; CH₃-Si), 0.59 (s, 9H; CH₃-Si), 1.16 (s, 3H; CH₃), 1.24 (m, 4H; CH₃ and CH₂ allyl), 1.49 (m, 1H; CH₂ allyl), 1.67 (m, 1H; CH₂-CH), 1.93 (m, 1H; CH₂ allyl), 2.08 (m, 1H; CH₂-CH), 2.72 (CH₂-C=), 4.69 (m, 1H; CH allyl trans to N), 4.84 (m, 1H; CH-O), 5.18 (m, 1H; CH allyl trans to P), 5.41 (m, 1H; CH allyl central), 6.8-8.4 ppm (m, 15H; CH=); ¹³C NMR (CD₂Cl₂): $\delta = 1.2$ (CH₃-Si), 1.6 (CH₃-Si), 20.2 (CH₂ allyl), 25.5 (CH₂ allyl), 26.6 (CH₃), 29.2 (CH₃), 30.8 (CH₂ allyl), 33.4 (CMe₂), 35.5 (CH2-C=), 45.6 (CH2), 62.4 (m; CH-O), 68.6 (m; CH allyl trans to N), 104.7 (d, J(C,P)=34.6 Hz; CH allyl trans to P), 112.4 (m; CH allyl central), 121–167 ppm (aromatic carbon atoms); elemental analysis calcd (%) for C47H53BF4NO4PPdSi2: C 57.82, H 5.47, N 1.43; found: C 57.87, H 5.49, N 1.44.

Study of the reactivity of the [Pd(η^3 -allyl)(L)]BF₄ with sodium malonate by in situ NMR spectroscopy:^[37] A solution of in situ prepared [Pd(η^3 -allyl)(L)]BF₄ (L=phosphite-nitrogen ligand, 0.05 mmol) in CD₂Cl₂ (1 mL) was cooled in the NMR spectroscopy tube at -80 °C. At this temperature, a solution of cooled sodium malonate (0.1 mmol) was added. The reaction was then followed by ³¹P NMR spectroscopy. The relative reaction rates were calculated using a capillary containing a solution of triphenylphosphine in CD₂Cl₂ as external standard.

Typical procedure of allylic alkylation of disubstituted linear (S1 and S3) and cyclic (S4-S6) substrates: A degassed solution of [PdCl(n³-C₃H₅)]₂ (0.9 mg, 0.0025 mmol) and the corresponding phosphite-nitrogen ligand (0.0055 mmol) in dichloromethane (0.5 mL) was stirred for 30 min. Subsequently, a solution of the corresponding substrate (0.5 mmol) in dichloromethane (1.5 mL), dimethyl malonate (171 µL, 1.5 mmol), N,Obis(trimethylsilyl)acetamide (370 µL, 1.5 mmol), and the corresponding base (5 mg) were added. The reaction mixture was stirred at room temperature. After the desired reaction time, the reaction mixture was diluted with Et₂O (5 mL) and saturated aqueous NH₄Cl (25 mL) was added. The mixture was extracted with Et₂O (3×10 mL) and the extract dried over MgSO₄. For substrate S1, solvent was removed and conversion was measured by ¹H NMR spectroscopy. To determine the ee by HPLC (Chiralcel OD, 0.5 % 2-propanol/hexane, flow 0.5 mLmin⁻¹), a sample was filtered over basic alumina using dichloromethane as the eluent.[38] For substrates S3-S6, conversion and enantiomeric excess were determined by GC.[24]

Typical procedure of allylic alkylation of disubstituted linear (S2), 1,3,3trisubstituted (S7 and S8), and monosubstituted (S9 and S10) substrates: A degassed solution of [{PdCl(η^3 -C₃H₅)}₂] (1.8 mg, 0.005 mmol) and the corresponding phosphite–nitrogen ligand (0.011 mmol) in dichloromethane (0.5 mL) was stirred for 30 min at room temperature. Subsequently, a solution of substrate (0.5 mmol) in dichloromethane (1.5 mL), dimethyl malonate (171 µL, 1.5 mmol), *N*,*O*-bis(trimethylsilyl)acetamide (370 µL, 1.5 mmol), and KOAc (5 mg) were added. After 2 h at room temperature, the reaction mixture was diluted with Et₂O (5 mL) and saturated aqueous NH₄Cl (25 mL) was added. The mixture was extracted with Et₂O (3×10 mL) and the extract dried over MgSO₄. Solvent was removed and conversion and regioselectivity were measured by ¹H NMR spectroscopy. For substrate **S2**, enantiomeric excess was determined by ¹H NMR spectroscopy using [Eu(hfc)₃] as resolving agent (hfc= 6,6,7,7,8,8,8-heptafluoro-2,2'-dimethyl-1,3,5- hydroxymethylene(+) canfore).^[15b] For substrates **S7** and **S8**, to determine the *ee* by HPLC (Chiralcel OJ, 13 % 2-propanol/hexane, flow 0.5 mLmin⁻¹), a sample was filtered over basic alumina using dichloromethane as the eluent.^[24] For substrates **S9** and **S10**, to determine the *ee* by HPLC (Chiralcel OJ, 13 % 2-propanol/hexane, flow 0.7 mLmin⁻¹), a sample was filtered over basic alumina using dichloromethane as the eluent.^[39]

Typical procedure of allylic amination of disubstituted linear substrate S1: A degassed solution of $[{PdCl(\eta^3-C_3H_5)}_2]$ (0.9 mg, 0.0025 mmol) and the corresponding phosphite–nitrogen ligand (0.0055 mmol) in dichloromethane (0.5 mL) was stirred for 30 min. Subsequently, a solution of the corresponding substrate (0.5 mmol) in dichloromethane (1.5 mL) and benzylamine (131 µL, 1.5 mmol) were added. The reaction mixture was stirred at room temperature. After the desired reaction time, the reaction mixture was diluted with Et₂O (5 mL) and saturated NH₄Cl (aq) (25 mL) was added. The mixture was extracted with Et₂O (3 × 10 mL) and the extract dried over MgSO₄. Solvent was removed and conversion was measured by ¹H NMR spectroscopy. To determine the *ee* by HPLC (Chiralcel OJ, 13 % 2-propanol/hexane, flow 0.5 mLmin⁻¹), a sample was filtered over silica using 10% Et₂O/hexane mixture as the eluent.^[38]

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