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# STEREOSELECTIVE SYNTHESIS OF LEWIS-ASSOCIATED TRISACCHARIDES AS E-SELECTIN INHIBITORS

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**Abstract**: Three types of Lewis-associated trisaccharides [the Le<sup>a</sup> analogs, their epimers with respect to the fucose residue (the 1c-epi-Le<sup>a</sup> analogs), and the Le<sup>x</sup> analogs] were synthesized in a stereoselective manner. Not only the Le<sup>a</sup> analogs but also the 1c-epi-Le<sup>a</sup> analogs inhibited E-selectin-mediated neutrophil accumulation into pleural cavity in lipoteichoic acid-treated mice, with the trend being Le<sup>a</sup> > 1c-epi-Le<sup>a</sup> > Le<sup>x</sup>.

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## Introduction

Selectins are calcium dependent lectins that regulate neutrophil rolling in the early step of the inflammatory response. Sialyl Le<sup>x</sup> (SLe<sup>x</sup>) and Sialyl Le<sup>a</sup> (SLe<sup>a</sup>) represent the sialylated and fucosylated oligosaccharides identified as ligands recognized by the selectins.<sup>1</sup>) Identification of the minimum carbohydrate structure required for selectin binding should therefore provide a means to design more potent inhibitors of selectin mediated-cell adhesion.

Previous structure-functional studies have suggested that both the fucose (Fuc) and sialic acid carboxylate moieties are essential for high binding ability with E-selectin.<sup>2,3</sup> Replacement of the sialic acid moiety of SLe<sup>x</sup> or SLe<sup>a</sup> with a sulfate group has also provided analogs which inhibit E-selectin-mediated adhesion.<sup>4</sup> These findings have focused intense attention on the search of SLe<sup>x</sup> mimetics that would maintain high affinity while using a simpler structure.<sup>5</sup> Moreover, a synthesized SLe<sup>a</sup> tetrasaccharide analog has been reported to show higher inhibitory activity than the reducing tetrasaccharide SLe<sup>x</sup> in vitro.<sup>3</sup> The sulfated Le<sup>a</sup> tetra- and pentasaccharides were also reported to have higher affinity than the corresponding SLe<sup>x</sup> derivatives in vitro.<sup>4a</sup> These results suggest that Le<sup>a</sup> structures may be more potent inhibitors of E-selectin-mediated cell adhesion than Le<sup>x</sup>, although NMR and molecular modeling studies have demonstrated conformational similarities between the two structures.<sup>6</sup> A more interesting observation was that non-sialylated trisaccharide Le<sup>a</sup> exhibited a slight

[Figure 1]



2-Naph; 2-naphthyl

inhibitory effect against E-selectin.<sup>3)</sup> This finding suggests that the Le<sup>a</sup> trisaccharide alone might contain the minimal structural features required for inhibitors of E-selectin. However, there are few reports concerning investigation on structure-activity relationships of Le<sup>a</sup> analogs.<sup>4a)</sup> Therefore, our current interest in this area is to identify the requirement of the following two moieties of Le<sup>a</sup> structure; one is the configuration of the Fuc residue that is revealed to be essential for recognition by E-selectin and the other is the substituent on the glucosamine (GlcN) nitrogen. Transformation of these moieties has not been studied concerning Le<sup>a</sup> structure, although *N*-modification of the GlcN concerning SLe<sup>x</sup> analogs was reported to improve the ability to inhibit E-selectin-mediated adhesion.<sup>7, 8</sup>) Herein, we report the stereoselective preparation of the *N*-modified Le<sup>a</sup> analogs (3 and 4) and their epimers with respect to the Fuc residue (the 1c-epi-Le<sup>a</sup> analogs, 1 and 2), and their inhibitory activity both on human E-selectin-mediated cellular adhesion *in vitro*<sup>9</sup> and on an inflammatory lung injury animal model<sup>10</sup>).

### Synthesis

The crucial step in the synthesis of the 1c-epi-Le<sup>a</sup> and Le<sup>a</sup> trisaccharide analogs (1, 2, 3, and 4) is stereoselective introduction of the Fuc residue to disaccharide intermediate 10 by using two differentially protected trichloroacetimidates  $(11 \text{ and } 20)^{11,12}$ . An allyloxycarbonyl group (alloc) on 10 was used to protect the GlcN nitrogen and allow for the later introduction of alternate acyl groups.

Preparation of the common intermediate 10 is shown in Scheme 1. Compound 8 was prepared in 98% overall yield from tetraacetate  $7^{13}$ ) by glycosylation with EtOH in the presence of TMSOTf followed by hydrolysis with NaOMe and treatment with benzaldehyde dimethyl acetal. The condensation of the resulting 4,6-O-benzylidene acetal 8 with protected galactosyl bromide in the presence of Hg(CN)<sub>2</sub> afforded disaccharide 9 in 61% yield. Selective opening of the benzylidene acetal of 9 produced the desired glycosyl acceptor 10 in 73% vield.<sup>14</sup>)





a) EtOH, TMSOTf (cat.), MS4A, CICH<sub>2</sub>CH<sub>2</sub>Cl (100%); b) NaOMe, MeOH (98%); c) PhCH(OMe)<sub>2</sub>, *p*-TosOH, MeCN (100%); d) tetra-*O*-acetyl-α-D-galactosyl bromide, Hg(CN)<sub>2</sub>, HgBr<sub>2</sub>, MS4A, CICH<sub>2</sub>CH<sub>2</sub>Cl (61%); e) NaBH<sub>3</sub>CN, TMSCI, THF, 0°C→rt (73%).

For preparation of the 1c-epi-Le<sup>a</sup> analogs (1 and 2), stereoselective introduction of the Fuc residue onto the hydroxyl group of 10 was achieved by applying the Schmidt's procedure<sup>12</sup>) with tri-O-acetyl fucose  $\alpha$ -trichloroacetimidate (11)<sup>11</sup>) in the presence of TMSOTf, as shown in Scheme 2. This fucosylation provided the expected  $\beta$ -glycosylated trisaccharide 12 in 76% yield accompanying with 10% of the corresponding  $\alpha$ -isomer. The alloc group on 12 was removed by treatment with Pd(PPh<sub>3</sub>)4 in the presence of polymethylhydrosiloxane (PMHS) to afford amine 13 in 90% yield. *N*-Acetylation of 13 afforded 14, which was then deprotected utilizing benzylic hydrogenation to afford 15. After peracetylation of compound 15 for purification, compound

16 was provided in overall yield of 89% from 14.<sup>15</sup>) Compound 16 was deprotected under basic conditions to provide the desired acetamide analog 1 in 95% yield. In the case of naphthamide analog 2, the amino group of 17 was selectively acylated with 2-naphthoyl chloride in the presence of NaHCO3.



j) NaOMe, MeOH (93%).

Preparation of the N-acylated Le<sup>a</sup> analogs (3 and 4) is shown in Scheme 3. Reaction of tri-O-benzyl fucose  $\alpha$ -trichloroacetimidate (20)<sup>12</sup>) with acceptor 10 afforded the desired  $\alpha$ -glycosylated trisaccharide 21 in 80% yield accompanying with 7% of the corresponding  $\beta$ -anomer. These results are consistent with Schmidt et al., who did not mention about formation of  $\beta$ -fucoside product during fucosylation of lactose derivative.<sup>12</sup>) The desired products 3 and 4 were synthesized from 21 using the similar conditions as described above.



d) 10%Pd-C, HCO<sub>2</sub>NH<sub>4</sub>, EtOH, reflux; e) Ac<sub>2</sub>O, DMAP (cat.), pyridine (74% yield from 23); f) NaOMe, MeOH (100%); g) 10%Pd-C, HCO2NH4, EtOH, reflux; h) 2-naphthoyl chloride, NaHCO3, CH2Cl2; i) Ac2O, DMAP (cat.), pyridine (44% yield from 22); i) NaOMe, MeOH (92%).

The Le<sup>x</sup> analogs (5 and 6) were prepared from compound 29,<sup>8)</sup> as shown in Scheme 4. Removal of the alloc group from 29, regioselective deprotection of the 3-O-acetyl group and regioselective reprotection of the amino group by benzyloxycarbonyl chloride (Z-Cl) afforded 32 in 75% overall yield. Stereoselective

fucosylation using fucosyl fluoride 33 provided trisaccharide 34 in 78% yield. Trisaccharide 34 was converted into the desired Le<sup>x</sup> analogs 5 and 6 by N-modification followed by transformation of 2-(trimethylsilyl)ethyl (SE) glycoside into ethyl glycoside and deprotection by the similar method as we previously reported for synthesis of SLe<sup>x</sup> analogs.<sup>8)</sup>



n) NaOMe, MeOH then H2O (93%).

## **Biological Activity**

These trisaccharides 1 - 6 were evaluated for their ability to inhibit the adhesion of HL-60 cells to purified recombinant human E-selectin in vitro<sup>9</sup>), as shown in Table 1. Among the six compounds, only compounds 3 and 4, the Le<sup>a</sup> analogs, were found to inhibit E-selectin-mediated adhesion, with IC50 values of 1.4 mM and 2.0 mM, respectively. However, the 1c-epi-Le<sup>a</sup> and Le<sup>x</sup> analogs (1, 2, 5, and 6) failed to inhibit the cell adhesion at concentrations up to 6.6 mM. In comparison, a SLe<sup>x</sup> analog was reported to inhibit the cell adhesion with the IC50 value of approximately 1 mM<sup>8</sup>) under the same conditions.<sup>9</sup>) These results indicate that the inhibitory potency of the Le<sup>a</sup> analogs (3 and 4) was roughly equivalent to that of the SLe<sup>x</sup>, in vitro. Considering the Nsubstituent in the GlcN moiety, naphthamide analog 4 demonstrated approximately the same activity as the corresponding acetamide analog 3, although previous results have indicated that naphthamide substitution on  $SLe^{x}$  increased the inhibitory potency in this assay as much as ten fold.<sup>7,8)</sup> To clarify the observed differences. we are now investigating further structure-activity relationships on N-substituents and conformational analysis.

We examined the in vivo effects of these trisaccharides 1 - 6 on lipoteichoic acid (LTA)-induced murine pleurisy model, in which E-selectin has been demonstrated as playing a significant role.<sup>10)</sup> Each compound was administered intravenously at a dose of 30 mg/kg. The inhibitory effects of these compounds were shown in Table 1. Among the acetamide analogs, compound 3, the Le<sup>a</sup> analog, and compound 1, the 1c-epi-Le<sup>a</sup> analog, were the most potent inhibiting LTA-induced neutrophil accumulation by 51% and 37%, respectively. Compound 5, the Le<sup>x</sup> analog, also inhibited by 31%, but not as strongly as the Le<sup>a</sup> analog 3. A similar inhibitory trend was also observed in the series of naphthamide analogs 2, 4, and 6. Namely, compound 4, the Le<sup>a</sup> analog, and compound 2, the 1c-epi-Le<sup>a</sup> analog, inhibited the neutrophil accumulation by 62% and 49%,

respectively. However, compound 6, the Le<sup>x</sup> analog, did not have any effects at a dose of 30 mg/kg. From these results, we can conclude that the *in vivo* results are consistent with the *in vitro* results following a trend in which the Le<sup>a</sup> analogs are the most potent inhibitors, Le<sup>a</sup> > 1c-epi-Le<sup>a</sup> > Le<sup>x</sup>. In addition, the inhibitory potency of the Le<sup>a</sup> analogs was approximately equal to that of a SLe<sup>x</sup> analog which showed 52% inhibition in this model.<sup>10</sup>)

Compound		in vitro <sup>a</sup> )	in vivo b)
No.	Structure	IC50 (mM)	inhibition (%)
1	1c-epi-Le <sup>a</sup> (acetamide)	>6.6 <sup>c)</sup>	37
3	Le <sup>a</sup> (acetamide)	1.4	51
5	Le <sup>x</sup> (acetamide)	>6.6 <sup>c)</sup>	31
2	lc-epi-Le <sup>a</sup> (naphthamide)	>6.6 <sup>c)</sup>	49
4	Le <sup>a</sup> (naphthamide)	2.0	62
6	Le <sup>X</sup> (naphthamide)	>6.6 <sup>c)</sup>	3

[Table 1] The in vitro and in vivo ability of the synthesized trisaccharides as E-selectin inhibitors.

a) The IC50 value means the 50% inhibitory concentration for cell adhesion of HL-60 to purified recombinant human E-selectin. For details, see reference 7.

b) Each compound was administered intravenously at a dose of 30 mg/kg.

Each data represents the mean of 6-8 determinations. For details, see reference 9.

c) Maximum concentration in use.

In conclusion, the Le<sup>a</sup> trisaccharides were found to be potent E-selectin inhibitors *in vitro* and *in vivo*. And moreover, the 1c-epi-Le<sup>a</sup> trisaccharides were found to show inhibitory activities *in vivo*. This finding suggests that E-selectin binding requirements for the Le<sup>a</sup> structure is not so rigid for the Fuc moiety which may allow for the design of more potent selectin inhibitors. Our synthetic approach provides a facile access to these analogs allowing both modifications on the Fuc or the GlcN moieties. Further investigation on these and other modifications are currently on going.

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### **References and notes**

- 1) Boschelli, D., H., Drugs Fut., 1995, 20, 805.
- 2) a) Rmphal, j., Y.; Zheng, Z.-L.; Perez, C.; Waiker, L., E.; DeFrees, S, A.; Gaeta, F., C., A., J. Med. Chem., 1994, 37, 3459. b) Brandkley, B., K.; Kiso, M.; Abbas, S.; Nikrad, P.; Srivasatava, S.; Foxall, C.; Oda, Y.; Hasegawa, A., Glycobiology, 1993, 3, 633.
- 3) Nelson, R. M.; Dolich, S.; Arffo, A.; Cecconi, O.; Bevilacqua, M. P., J. Clin. Invest., 1993, 91, 1157.
- 4) a) Yuen, C.-T.; Bezouska, K.; O'Brien, J.; Stoll, M.; Lemoine, R.; Lubineau, A.; Kiso, M.; Hasegawa, A.; Bockovich, N., J.; Nicolaou, K. C.; Feizi, T., J. Biol. Chem., 1994, 269, 1595. b) Lubineau, A.; Gallic, J. L.; Lemoine, R., J. Chem. Soc., Chem. Commun., 1993, 1419.
- 5) For recent studies on SLe<sup>x</sup> mimetics, see; Birkbeck, A. A.; Ley, S. V.; Prodger, J. C., *BioMed. Chem.* Lett., **1995**, *5*, 2637.
- 6) a) Tyrrell, D; James, P.; Rao, C.; Abbas, S.; Dasgupta, F.; Nashed, M.; Hasegawa, A.; Kiso, M.; Asa, D.; Kidd, J.; Brandley, B., K., Proc. Natl. Acad. Sci. USA, 1991, 88, 10372. b) Berg, E. L.; Robinson, M. K.; Mansson, O.; Butcher, E. C.; Magnani, J. L., J. Biol. Chem., 1991, 266, 14869.
- 7) Ramphal, J. Y.; Hiroshige, M.; Lou, B.; Guadino, J.; Hayashi, M.; Chen, S. M.; Chiang, L. C.; Gaeta, F. C. A.; DeFrees, S. A., J. Med. Chem., 1996, 39, 1357.
- 8) Hayashi, M.; Tanaka, M.; Itoh, M.; Miyauchi, H., J. Org. Chem., 1996, 61, 2938.
- 9) DeFrees, S. A.; Kosch, W.; Way, W.; Paulson, J.C.; Sabesan, S.; Halcomb, R. L.; Huang, D.-H.; Ichikawa, Y.; Wong, C.-H., J. Am. Chem. Soc., 1995, 117, 66.
- 10) Tojo, S.; Koike, H.; Yokota, S.; Hamazume, Y.; Natsume, Y.; Phillips, L.; Paulson, J.; Hayashi, M. and Morooka, S., submitted.
- 11) a)Schmidt, R., R.; Wegmann, B.; Jung, K.-H., Liebigs Ann. Chem., 1991, 121. b)Schmidt, R. R., Angew. Chem. Int. Ed. Engl., 1986, 25, 212.
- 12) Schmidt, R. R.; Toepfer, A., Tetrahedron Lett, 1991, 32, 3353.
- 13) Boullanger, P.; Banoub, J.; Descotes, G., Can. J. Chem., 1987, 259, 1343.
- 14) Garegg, P. J.; Hultberg, H.; Wallin, S., Carbohydr. Res., 1982, 108, 97.
- 15) Selected <sup>1</sup>H-NMR data for structural determination of **16** (270MHz,CDCl<sub>3</sub>); δ5.89 (1H, d, J=8.6 Hz, NHAc), 4.96 (1H, d, J=7.9 Hz, H-1b or H-1c), 4.80 (1H, d, J=7.3 Hz, H-1b or H-1c), and 4.65 (1H, d, J=5.3 Hz, H-1a).

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