A New Approach to Tubacin

Jian Hong*, Xin Xu, Debasis Das*, Pengyu Yang, Shu-Hui Chen and Ge Li

Division of Discovery Chemistry Service, WuXi AppTec Inc., 288 Fute Zhong Road, Waigaoqiao Free Trade Zone, Shanghai 200131, P.R., China

Received May 15, 2009: Revised December 11, 2009: Accepted December 18, 2009

Abstract: A new simple synthesis of tubacin is reported. It entails the formation of the *syn*-1,3-diol unit and its stereoselective ketalization with a functionalized benzaldehyde derivative.

Keywords: Tubacin, histone deacetylases, allyl borane reaction, stereoselective ketal formation, solution phase synthesis, hydroxyamide synthesis.

INTRODUCTION

Every year throughout the world, around 10 million people are diagnosed with cancer. Cancer cells are highly dependent on protein degradation due to cell cycle, hypermutation and chromosomal rearrangement. Histone deacetylases (HDACs) [1] are nuclear zinc-dependent enzymes which catalyze deacetylation of N-acetyl-lysine residues and play an important role in a number of biological processes such as gene expression [2] and cell cycle. Compounds which inhibit HDACs can suppress the expression of certain genes thus resulting in an antitumor effect [3]. Therefore in recent years, a key success in cancer research was the discovery of novel HDAC inhibitors [4]. This resulted in the launch of a non-selective HDAC inhibitor Zolinza[®] by Merck Co. Still much research is being focused on the discovery of new selective HDAC inhibitors [5].



Fig. (1).

Identified recently as a selective HDAC6 inhibitor by high-throughput synthesis [6], tubacin (1) (Fig. 1) does not

affect the level of histone acetylation in gene expression and cell cycle. As selective inhibitors of HDAC6 are used in the treatment of protein degradation disorders [7], tubacin may have therapeutic applications as antimetastatic and antiangiogenic agent. Since the discovery of tubacin, its demand has been increasing and the compound is used as a reference for the design of new HDAC inhibitors with enhanced activity and selectivity.

Challenges in the tubacin synthesis are the construction of the three-substituted 1,3-dioxane ring with all substituents in *syn* dispositions and finding a short route to its *syn*-1,3diol unit. Schreiber and coworkers described a synthesis of tubacin on a solid support [8]. Herein we report our facile route to tubacin relying upon a simple formation of *syn*-1,3diol unit and its stereoselective ketalization with a functionalized benzaldehyde derivative.

RESULTS AND DISCUSSION

Our route to tubacin is depicted in Scheme 1. Thus, the commercially available 1,4-benzenedimethanol (2) was selectively mono-TBDPS-protected and oxidized to 3 in 92% The reaction of (-)-B-allyldiisopinoyield. campheylborane ((-)-Ipc₂BAll) [9] with the para-substituted benzaldehyde 3 led to the desired S-homoallylic alcohol 4 in 65% yield and 96% ee. Instead of targeting the chiral pure syn-epoxide, we have prepared epoxide 5 as a mixture of diastereomers via m-CPBA oxidation [10]. This mixture was then reacted with 4,5-diphenyloxazolin-2-thione [11] affording diastereomers 6a (42%) and 6b (36%) [12] which were separated by preparative HPLC. The absolute stereochemistry of the syn-1,3-diol **6a** was unambiguously determined based on ¹³C NMR analysis of the corresponding acetonide **7a** [13]. The ¹³C NMR chemical shifts of the two methyls and the central quaternary carbon of 19.1, 29.3 and 98.8 ppm, respectively, support a chair conformation and confirm the syn-1,3-orientation compared to the antidiastereomer 7b which has a boat conformation (with 24.1, 24.2 and 100.5 ppm, respectively).

Progressing towards tubacin (Scheme 2), the synthesis of fragment 9 was accomplished in good yield by coupling 4-aminobenzyl alcohol (8) with octanoyl monomethyl ester

^{*}Address correspondence to these authors at the Division of Discovery Chemistry Service, WuXi AppTec Inc., 288 Fute Zhong Road, Waigaoqiao Free Trade Zone, Shanghai 200131, P.R.China; Tel: +86-21-58682139, +86-21-58683340;

E-mails: hong_jian@wuxiapptec.com, debasis_das@wuxiapptec.com



Scheme 1. Reagents and conditions: (a) TBDPSCl, imidazole, DMF, 50 °C, 16 h, 42%; (b) MnO₂, CH₂Cl₂, rt, 12 h, 92%; (c) (-)-Ipc₂BAll, Dioxane, Et₂O, -100 °C, 2 h, 65%; (d) *m*-CPBA, CHCl₃, rt, 24 h, 75%; (e) 4,5-diphenyloxazole-2(3H)-thione, DIEA, DMF, rt, 78%; (f) Me₂C(OMe)₂, TsOH, rt, 12 h, 76-80%.



Scheme 2. Reagents and conditions: (a) Methyl 8-chloro-8-oxooctanoate, Et₃N, CH₂Cl₂, rt, 10 h, 61%; (b) MnO₂, CH₂Cl₂, rt, 16 h, 90%; (c) HC(OMe)₃, TsOH, MeOH, reflux, 2 h, 94%; (d) TBAF, THF; (e) 10, PPTS, DMF, rt, 12 h, 74%; (f) NH₂OH.HCl, KOH, MeOH, rt, 46%.

acid chloride, followed by oxidation. However, as the condensation of diol **6a** with aldehyde **9** did not proceed well, the aldehyde was converted into its acetal **10**. Several failed attempts to couple **6a** with **10** under different conditions pushed us to resort to **11** which was prepared by TBAF-deprotection of **6a**. Thus, acetal **10** was easily coupled with **11** in the presence of pyridinium *p*-toluenesulfonate (PPTS) to afford the desired compound **12** in 74% yield. Interestingly, the ketal formation was completely stereoselective as only product **12** was formed. The absolute configuration of the newly generated chiral center was

established by NOE experiment [14]. Finally, treatment of **12** with hydroxylamine led smoothly to tubacin in a moderate 46% yield [15].

CONCLUSION

In summary, we have achieved the synthesis of tubacin in a simple and efficient way. This new synthetic route is based on the stereoselective preparation of the *syn*-1,3-diol fragment of tubacin and on its successful stereoselective ketalization with a benzaldehyde derivative. Our simple approach could be extended to the synthesis of other tubacin analogues such as tropcin and histacin.

REFERENCES AND NOTES

- [1] For a review, see: Ropero, S.; Esteller, M. The role of histone deacetylases (HDACs) in human cancer. *Mol. Oncol.*, **2007**, *1*, 19.
- (a) Wolffe, A. P. Sinful repression. *Nature*, **1997**, *387*, 16; (b) Cheung, W. L.; Briggs, S. D.; Allis, C. D. Acetylation and chromosomal functions. *Curr. Opin. Cell. Biol.*, **2000**, *12*, 326.
- [3] (a) Miller, T. A.; Witter, D. J.; Belvedere, S. Histone deacetylase inhibitors. J. Med. Chem., 2003, 46, 5097; (b) Muri, E. M. F.; Nieto, M. I.; Sindelar, R. D.; Williamson, J. D. Hydroxamic acids as pharmacological agents. Curr. Med. Chem., 2002, 9, 1631; (c) Yang, K.; Lou, B. Molecular diversity of hydroxamic acids. part 1. solution-and solid-phase synthesis. Mini-Rev. Med. Chem., 2003, 3, 349; (d) Iizuka, M.; Smith, M. M. Functional consequences of histone modifications. Curr. Opin. Genet. Dev., 2003, 13, 154; (e) Minucci, S.; Pelicci, P. G. Histone deacetylase inhibitors and the promise of epigenetic (and more) treatments for cancer. Nat. Rev. Cancer, 2006, 6, 38; (f) Hildmann, C.; Wegener, D.; Riester, D.; Hempel, R.; Schober, A.; Meraner, J.; Giurato, L.; Guccione, S.; Nielsen, T. K.; Ficner, R.; Schwienhorst, A. Substrate and inhibitor specificity of class 1 and class 2 histone deacetylases. J. Biotech., 2006, 124, 258.
- [4] Weidle, U. H; Grossmann, A. Inhibition of histone deacetylases: a new strategy to target epigenetic modifications for anticancer treatment. *Anticancer Res.*, 2000, 20, 1471.
- For new HDAC inhibitors, see: (a) Methot, J. L.; Chakravarty, P. [5] K.; Chenard, M.; Close, J.; Cruz, J. C.; Dahlberg, W. K.; Fleming, J.; Hamblett, C. L.; Hamill, J. E.; Harrington, P.; Harsch, A.; Heidebrecht, R.; Hughes, B.; Jung, J.; Kenific, C. M.; Kral, A. M.; Meinke, P. T.; Middleton, R. E.; Ozerova, N.; Sloman, D. L.; Stanton, M. G.; Szewczak, A. A.; Tyagarajan, S.; Witter, D. J.; Secrist J. P. Miller, T. A. Exploration of the internal cavity of histone deacetylase (HDAC) with selective HDAC1/HDAC2 inhibitors (SHI-1:2). Bioorg. Med. Chem. Lett., 2008, 18, 973; (b) Andrews, D. M.; Gibson, K. M.; Graham, M. A.; Matusiak, Z. S.; Roberts, C. A.; Stokes, E. S. E.; Brady M. C.; Chresta, C. M. Design and campaign synthesis of pyridine-based histone deacetylase inhibitors. Bioorg. Med. Chem. Lett., 2008, 18, 2525 (c) Estiu, G.; Greenberg, E.; Harrison, C. B.; Mazitschek, R.; Bradner, J. E.; Wiest, O. On the structural origin of selectivity in class II selective histone deacetylase inhibitors. J. Med. Chem., 2008, 51, 2898 and references cited therein.
- [6] (a) Haggarty, S. J.; Koeller, M. K.; Wong, J. C.; Butcher, R. A.; Schreiber, S. L. Multidimensional chemical genetic analysis of diversity oriented synthesis-derived deacetylase inhibitors using cell-based assays. *Chem. Biol.*, **2003**, *10*, 383; (b) Wong, J. C.; Hong, R.; Schreiber, S. L. Structural biasing elements for in-cell histone deacetylase paralog selectivity. *J. Am. Chem. Soc.*, **2003**, *125*, 5586.
- [7] (a) Hideshima, T.; Bradner, J. E.; Wong, J.; Chauhan, D.; Richardson, P.; Schreiber, S. L.; Anderson, K.C. Small-molecule inhibition of proteasome and aggresome function induces synergistic antitumor activity in multiple myeloma. *Proc. Natl. Acad. Sci. USA*, 2005, 102, 8567; (b) Itoh, Y.; Suzuki, T.; Kouketsu, A.; Suzuki, N.; Maeda, S.; Yoshida, M.; Nakagawa, H.; Miyata, N. Design, synthesis, structure--selectivity relationship, and effect on human cancer cells of a novel series of histone deacetylase 6-selective inhibitors. *J. Med. Chem.*, 2007, 50, 5425.
- [8] (a) Sternson, S. M.; Wong, J. C.; Grozinger, C. M.; Schreiber, S. L. Synthesis of 7200 small molecules based on a substructural analysis of the histone deacetylase inhibitors trichostatin and troposin. *Org. Lett.*, **2001**, *3*, 4239; (b) Schreiber, S. L.; Sternson, S. M.; Wong, J. C.; Grozinger, C. M.; Haggarty, S. J. *US patent* 2004/0072849.
- [9] (a) Racherla, U. S.; Brown, H. C. Chiral synthsis via organoboranes. 27. Remarkably rapid and exceptionally enantioselective (approaching 100%ee) allylboration of representative aldehydes at -100.degree under new, salt-free conditions. J. Org. Chem., 1991, 56, 401; (b) Racherla, U. S.; Liao, Y.; Brown, H. C. Chiral synthsis via organoboranes. 36. Exceptionally enantioselective allylborations of representative heterocyclic aldehydes at -100.degree.C under new, salt-free conditions. J. Org. Chem., 1992,

57, 6614; (c) Yamamoto, Y.; Asao, N. Selective reactions using allylic metals. *Chem. Rev.*, **1993**, *93*, 2207; (d) Cossy, J.; Willis, C.; Bellosta, V.; Bouzbouz, S. Enantioselective Allyltitanation. Synthesis of (+)-Sedamine. *Synlett*, **2000**, 1461; (e) Cossy, J.; Willis, C.; Bellosta, V.; Bouzbouz, S. Enantioselective allyltitanations and metathesis reactions: application to the synthesis of piperidine alkaloids (+)-Sedamine and (-)-Prosophylline. *J. Org. Chem.*, **2002**, *67*, 1982 and references cited therein.

- [10] Felpin, F.-X.; Lebreton, J. A highly stereoselective asymmetric synthesis of (-)-Lobeline and (-)-Sedamine. J. Org. Chem., 2002, 67, 9192.
- [11] Harris, N. V.; Smith, C.; Asthon, M. J.; Bridge, A. W.; Bush, R. C.; Coffee, D. I. D.; Harper, M. F.; Lythgoe, D. J.; Newton, C. G. Riddell, D. Acyl-CoA:cholesterol O-acetyl transferase (ACAT) inhibitors.1. 2-(alkylthio)-4,5-diphenyl-1H-imidazoles as potent inhibitors of ACAT. J. Med. Chem., 1992, 35, 4384.
- Preparation of 6: To a solution of 4,5-diphenyloxazole-2(3H)-[12] thione (1.0 g, 4.0 mmol) in DMF (5 mL), diisopropyl ethyl amine (DIEA) (1.5 mL) was added followed after 15 min by compound 5 (1.5 g, 3.5 mmol) in DMF (5 mL). The mixture was stirred at room temperature for 12h then poured onto water (100 mL) and extracted with ethyl acetate (3x50 mL). The organic phase was washed with brine and dried over anhydrous Na2SO4. The solvent was removed in vacuo and the residue was passed through a silica gel pad (eluting with petroleum ether: ethyl acetate = 1:1) to give mixture 6 (1.85 g, 78% yield) as a yellow oil. Preparative HPLC afforded pure diastereomers **6a** and **6b**. **6a**: ¹H NMR (400 MHz, CDCl₃): $\delta =$ 1.16 (s, 9H), 1.92-2.19 (m, 2H), 3.30-3.45 (m, 2H), 4.37-4.44 (m,1H), 4.78 (s, 2H), 5.05 (dd, 1H, $J_1 = 2.4$, $J_2 = 11.6$ Hz), 7.34-7.42 (m, 16H), 7.53-7.57 (m, 2H), 7.64-7.70 (m, 6H). **6b**: ¹HNMR (400 MHz, CDCl₃): $\delta = 1.09$ (s, 9H), 1.99-2.17 (m, 2H), 3.35-3.45 (m, 2H), 4.38-4.43 (m, 1H), 4.77 (s, 2H), 5.12 (dd, 1H, $J_1 = 2.8$, J_2 = 8 Hz), 7.34-7.45 (m, 16H), 7.50-7.57 (m, 2H), 7.65-7.72 (m, 6H).
- [13] a) Rychnovsky, S. D.; Rogers, B. N.; Richardson, T. Configurational assignment of polyene macrolide antibiotics using ¹³C]acetonide analysis. Acc. Chem. Res., **1998**, 31, 9; (b) Lira, R.; Roush, W. R. Stereoselective synthesis of the C91)-C(19) fragment of Tetrafibricin. Org. Lett., 2007, 9, 533; (c) Migita, A.; Shichijo, Y.; Oguri, H.; Watanabe, M.; Tokiwano, T.; Oikawa, H. Stereocontrolled synthesis of prelasalocid, a key precursor proposed in the biosynthesis of polyether antibiotic lasalocid A. Tetrahedron Lett., 2008, 49, 1021; The stereo-chemistry of diol 6a was determined to be syn by ¹H and ¹³C NMR analyses of the corresponding acetonide 7a. Preparation of 7a and 7b: To a stirred solution of 6a (or 6b) (0.2 g, 0.3 mmol) in acetone (10 mL) was added 2,2-dimethoxypropane (1.0 mL, 10 mmol), TsOH (0.01 g, 0.05 mmol) and the mixture was left to stir at room temperature for ~24 h. Acetone was evaporated under reduced pressure, and the residue was dissolved in ethyl acetate (20 mL), washed with saturated NaHCO3 solution (5 mL), brine (5 mL) and dried on anhydrous Na2SO4. The solution was concentrated and the residue was purified by column chromatography to give the desired ketal **7a** (or **7b**). **7a** (0.18 g, 80% yield). ¹H NMR (400 MHz, CDCl₃): δ = 1.09 (s, 9H), 1.52 (s, 3H), 1.58 (s, 3H), 1.63 (m, 1H), 1.99 (m, 1H), 3.39-3.41 (m, 2H), 4.37-4.44 (m, 1H), 4.76 (s, 2H), 4.95 (dd, 1H, J₁ = 11.6, J₂ = 2.4 Hz), 7.30-7.45 (m, 16H), 7.55-7.57 (m, 2H), 7.64-7.72 (m, 6H). ¹³C NMR (100 MHz, CDCl₃): δ = 158.2, 146.4, 139.8, 139.7, 135.6, 134.7, 132.7, 131.3, 128.9, 127.9, 127.8, 127.7, 127.4, 127.1, 126.9, 125.5, 125.3, 125.1, 98.8, 70.4, 67.9, 64.5, 37.3, 37.0, 29.3, 26.1, 19.1, 18.5. **7b** (0.16 g, 76% yield). ¹HNMR (400 MHz, CDCl₃): $\delta = 1.09$ (s, 9H), 1.47 (s, 6H), 2.07-2.26 (m, 2H), 3.39-3.57 (m, 2H), 4.37-4.40 (m, 1H), 4.76 (s, 2H), 4.93-4.97 (m, 1H), 7.32-7.45 (m, 16H), 7.53-7.59 (m, 2H), 7.64-7.73 (m, 6H). ³C NMR (100 MHz, CDCl₃): $\delta = 158.3$, 146.3, 139.7, 139.6, 135.5, 134.7, 132.7, 131.3, 128.9, 128.1, 127.9, 127.8, 127.7, 127.6, 127.4, 127.2, 127.1, 126.9, 125.6, 125.5, 125.4, 125.3, 125.2, 125.1, 124.7, 100.5, 67.4, 65.6, 64.4, 38.0, 36.8, 26.0, 24.2, 24.1, 18.5.

29.3 ppm $0 \stackrel{}{\underset{6}{\overset{6}{\overset{0}}}} 0$ 98.8 ppm 100.5 ppm 24.2 ppm 19.1 ppm 24.1 ppm

A New Approach to Tubacin

[14] Synthesis of 12: To a solution of compound 11 (100 mg, 0.23 mmol) in DMF (3.0 mL) were added compound 10 (337 mg, 1.0 mmol) and pyridinium p-toluenesulfonate (PPTS) (100 mg, 0.40 mmol). The mixture was stirred at room temperature for 12 h. Water (30 mL) was added and the mixture was extracted with ethyl acetate (3x50 mL). The organic phase was washed with brine and dried over anhydrous Na₂SO₄. The solvent was removed in vacuo and the residue was purified by column chromatography on silica gel (ethyl acetate: $CH_2Cl_2 = 1:6$, then, petroleum ether: ethyl acetate = 1:1) to afford ester 12 (103 mg, 74% yield) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): $\delta = 1.35-1.36$ (m, 4H), 1.61-1.71 (m, 4H), 1.82 (m,1H), 2.06-2.12 (m, 2H), 2.31 (t, 4H, J = 7.2 Hz), 3.51 (d, 2H, J = 6 Hz), 3.67 (s, 3H), 4.36 (m, 1H), 4.66 (s, 2H), 4.90 (d, 1H, J = 11.2 Hz), 5.71 (s, 1H), 7.32-7.41 (m, 10H), 7.51-7.56 (m, 6H), 7.61-7.69 (m, 2H).



Characteristic NOEs of 12

[15] Synthesis of tubacin (1): To hydroxylamine hydrochloride (3.0 g, 43 mmol) in methanol (7.5 mL) KOH (2.4 g, 43 mmol) in methanol (14 mL) was added. The mixture was stirred at room temperature for 1 h and filtered. The prepared NH2OH solution (4.0 mL, 2.0 M in methanol, 8.0 mmol) was added to compound 12 (0.27 g, 0.40 mmol), followed by a KOH solution (0.8 ml, 1.0 M in methanol, 0.8 mmol). The mixture was stirred at room temperature for 16 h then concentrated. Water (10 mL) was added to the residue and extracted with ethyl acetate (3x20 mL). The organic phase was dried over anhydrous Na2SO4, concentrated and the residue purified by chromatography on silica gel (MeOH: $CH_2Cl_2 = 1:30$ then 1:10) affording tubacin (110 mg, 46% yield) as a white solid. 1H NMR (400 MHz, CD₃OD): δ = 1.37-1.38 (m, 4H), 1.61-1.69 (m, 4H), 1.81 (m, 1H), 2.03 (m, 1H), 2.09 (t, 2H, J = 7.2 Hz), 2.33 (t, 2H, J = 7.2), 3.52 (m, 2H), 4.38 (m, 1H), 4.58 (s, 2H), 4.95 (dd, 1H, $J_1 =$ 1.8, J₂ = 11.0 Hz), 5.71 (s, 1H), 7.32-7.40 (m, 12H), 7.44-7.48 (m, 6H). ESI-MS m/z calcd for C41H43N3O7S: 721.28; found: 722.3 $(M+1)^{+}$.