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Studies on Antitumor-active 2,3-Dioxopiperazine Derivatives. II.¹⁾ Synthesis and Structure-Antitumor Activity Relationship of 1-Benzyl-2,3-dioxopiperazine Derivatives

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2,3-Dioxopiperazine derivatives, which are antitumor agents of a new type, were synthesized and the structure-activity relationship was investigated from the viewpoint of lipophilic-hydrophilic balance. It was found that 1-(4-diethylaminobenzyl)-4-n-hexyl-2,3-dioxopiperazine (12n) possessed significant in vitro cytotoxicity and in vivo antitumor activity against transplanted tumor, but this in vivo antitumor activity did not reflect the in vitro cytotoxicity. The metabolism of 12n in rats and mice was then studied. It was found that the Et₂N-group of 12n was easily metabolized to an AcNH-group in vivo.

Keywords—new type of antitumor agent; 2,3-dioxopiperazine derivatives; structure-activity relationship; cytotoxicity; antitumor activity; Ehrlich ascites carcinoma; HeLa S3 cell; metabolism

In the preceding paper,¹⁾ we reported that antitumor agents of a new type could be developed from 2,3-dioxopiperazine derivatives. It has already been reported that highly lipophilic polyene compounds such as vitamin A, its analogs and coenzyme Q, possess tumor preventive effect or antitumor effect.²⁾ On the other hand, the 2,3-dioxopiperazinyl group is known to be a hydrophilic group.³⁾

This work was undertaken from the viewpoint of lipophilic-hydrophilic balance in an attempt to develop a new antitumor agent having a 2,3-dioxopiperazine moiety. In the present paper we wish to report the antitumor activity of 1-(4-diethylaminobenzyl)-4-n-hexyl-2,3-dioxopiperazine (12n) and some problems regarding its *in vivo* activity.

First, in order to find antitumor-active 2,3-dioxopiperazine derivatives having polyene groups, 1-(2,4-hexadien-1-yl)-2,3-dioxopiperazine derivatives (1a, b) were synthesized by two methods, as shown in Chart 1: method A via the intermediate (2b) and method B through the cyclization of (5a, b) to form the 2,3-dioxopiperazine ring, followed by introduction of a 2,4-hexadienyl group. In method A, the yield in the removal of the 2,4-dimethoxybenzyl protecting group^{4,5)} of the amide nitrogen of the 2,3-dioxopiperazine ring was poor, and synthetically method B was superior to method A.

The minimum inhibitory concentrations (MIC values) against HeLa S3 cells and Ehrlich ascites carcinoma (EAC) cells of 1a, b, 2b, and 3 were tested. Among them, only 2b showed high cytotoxicity (MIC: 3.13 µg/ml against HeLa S3 and EAC cells). This result suggests that the 2,4-dimethoxybenzyl group contributes to the cytotoxicity and that the polyene group does not participate (Table I). Next, the relation between the nature of 4-substituents of 1-(2,4-dimethoxybenzyl)-2,3-dioxopiperazine derivatives (2) and cytotoxicity was investigated. Compounds (2c—n) were synthesized by the same method as 2b. The cytotoxicity of 2 was greatest when the carbon number of the 4-substituent was 6. Introduction of polar groups such as OH, NH₂, and COOH, into the 4-substituent decreased the cytotoxicity. 2b and 2c showed the highest cytotoxicity (Table II).

In order to find more effective compounds than 2b, c, chemical modification of the A,

method B

Chart 1

Table I. Structure-Cytotoxicity Relationship of 1-(2,4-Hexadien-1-yl)-2,3-dioxopiperazine Derivatives

| Compd. No. | R | MIC $(\mu g/ml)^{a}$ | | | |
|---------------|--|----------------------|------|--|--|
| No. | K | HeLaS3 | EAC | | |
| 3 | Н | >100 | >100 | | |
| 1a | CH ₃ | 25 | 25 | | |
| 1b | CH₃ \ | 50 | 50 | | |
| 2b | OCH ₃ CH ₃ O-CH ₂ - | 3.13 | 3.13 | | |

a) Microplate method.

Incubation medium: Eagle's MEM supplemented with 20% calf serum.

Inoculum size: 2×10^4 cells/ml. Incubation period: 4 days.

Determination: Giemsa staining.

B, and C moieties of 2e (Fig. 1) was carried out and the structure-activity relationship was investigated. In the A moiety, the substituent effect on the benzene ring was studied. Compounds (12a—q) were synthesized by method A from substituted benzaldehydes (7) or by method B from substituted benzyl halides 13 (Chart 2). Table III shows the *in vitro* cytotoxicities of 12a—q and 2e. Among them, the 2,5-dimethoxy compound (12d), 2,3,4-trimethoxy compound (12h) and 4-diethylamino compound (12n) showed high cytotoxicities, as did the 2,4-dimethoxy compound (2e).

These four compounds showed cytotoxic effects against HeLa and Ehrlich cells in vitro, but only 12n showed antitumor activity against transplanted EAC; the other three compounds showed no therapeutic effect (Table IV). It may be concluded from the results of this investigation that the 4-diethylamino group is the preferred substituent on the benzene ring. The 4-diethylaminophenyl group was therefore chosen as the A moiety.

Next, the B moiety in Fig. 1 was investigated. Compounds (14a—d) were synthesized by method shown in Chart 3. In particular, it is of interest that 14a was easily obtained by

Table II. Structure-Cytotoxicity Relationship of 1-(2,4-Dimethoxybenzyl)-2,3-dioxopiperazine Derivatives

| Compd. No. | n | $\mathrm{MIC}(\mu\mathrm{g/ml})^{a)}$ | | |
|---------------|--|---------------------------------------|------|--|
| | R | HeLaS3 | EAC | |
| 2a | Н | 100 | 100 | |
| 2c | -CH ₂ CH=CH ₂ | 100 | 100 | |
| 2d | n-C ₄ H ₉ | 12.5 | 12.5 | |
| 2e | n-C ₆ H ₁₃ | 3.13 | 3.13 | |
| 2 f | n-C ₇ H ₁₅ | 6.25 | 6.25 | |
| $2\mathbf{g}$ | n-C ₈ H ₁₇ CH ₃ CH ₃ | 25 | 25 | |
| 2h | CH ₃ CH ₃ | 25 | 25 | |
| 2i | -CH ₂ - | 6.25 | 6.25 | |
| 2 j | $-CH_2CH_2$ | 12.5 | 12.5 | |
| 2k | $-(CH_2)_6Br$ | 12.5 | 6.25 | |
| 21 | -(CH ₂) ₆ OH | 50 | 100 | |
| 2m | $-(CH_2)_6NH_2$ | 25 | 12.5 | |
| 2n | -CH ₂ -CH=CH-CH=C(CH ₃)COOH | 100 | 100 | |
| 2b | CH ₃ | 3.13 | 3.13 | |

a) See the legend to Table I.



$$R_3$$
 R_4
 R_5
 R_4
 R_5
 R_6
 R_7
 R_8
 R_8
 R_8
 R_8
 R_9
 R_9

12g, n, o, 18a-k

12k

method B

Chart 2

the reaction of 1-n-hexyl-2,3-dioxopiperazine with an aromatic iodine compound in the presence of K_2CO_3 and $Cu.^4$) Table V shows the cytotoxicities of 14a—d and 12n against HeLa S3 and EAC cells. 12n, in which the B moiety is $-CH_2$ -, showed the highest cytotoxicity. Thus, the 4-diethylaminobenzyl group was chosen as the AB moiety.

$$\begin{array}{c|c} CH_3O & O & O \\ \hline CH_3O - CH_2 - N & N - C_6H_{13} \\ \hline A & B & C \\ \end{array} \textbf{2e}$$

Fig. 1. Moieties of **2e** That were separately subjected to Chemical Modification

Investigation of the C moiety shown in Fig. 1 was next carried out. Compounds 18a—k were synthesized by method A as shown in Chart 2. Table VI shows the structure-antitumor activity relationships in vitro and in vivo of 18a—k and 12n. Among 18a—g and 12n, 12n, 18c, and 18d, in which the carbon number of the C moiety was 6, showed cytotoxicity. However, among these three compounds, only 12n showed in vivo antitumor activity against EAC (i.p.—i.p.); the other two compounds showed no therapeutic effect. Among 18h—k, 18h showed cytotoxicity equal to that of 12n, but no in vivo antitumor activity was found. Further, 18j and 18k showed lower cytotoxicities than 12n but their in vivo antitumor activities were

Table III. Structure-Cytotoxicity Relationship of 1-Benzyl-4-n-hexyl-2,3-dioxopiperazine Derivatives

$$\begin{array}{c} O & O \\ \hline R-CH_2-N & N-C_6H_{13} \end{array}$$

| Compd. | · D | R $\frac{\mathrm{MIC}(\mu\mathrm{g/ml})^{a)}}{\mathrm{Compd.}}$ | | R | MIC(μg | $\mathrm{MIC}(\mu\mathrm{g/ml})^{a}$ | |
|-------------|--|---|------|------|---|--------------------------------------|------|
| No. | K | HeLaS3 | EAC | No. | K | HeLaS3 | EAC |
| 12a | OCH ₃ | 50 | 25 | 12j | НО- | 100 | 100 |
| 12b | CH ₃ O-CH ₃ OCH ₃ | 50 | 50 | 12k | НО- | 50 | 50 |
| 12c | OCH ₃ | 100 | 100 | 121 | CI- | 100 | 50 |
| 2 e | CH ₃ O-CH ₃ | 3.13 | 3.13 | 12m | Cl-Cl | 25 | 12.5 |
| 12d | CH ₃ O | 6.25 | 6.25 | 12 n | $(C_2H_5)_2N-$ | 3.13 | 3.13 |
| 12e | OCH ₃ | 100 | 50 , | 120 | $(C_2H_5)_2N N(C_2H_5)_2N-$ |) ₂ 50 | 50 |
| 12 f | CH ₃ O CH ₃ O OCH ₃ | 50 | 50 | 12p | (C ₂ H ₅) ₂ N | 50 | 50 |
| 12g | CH ₃ O-CH ₃ | 100 | 50 | 12q | $(C_2H_5)_2NCH_2-$ | 100 | 100 |
| 12h | CH ₃ O OCH ₃ CH ₃ O CH ₃ O | 3.13 | 3.13 | | | | |
| 12i | CH ₃ O — —————————————————————————————————— | 100 | 50 | | | | |

a) See the legend to Table I.

similar to that of 12n. These results showed that the *in vitro* cytotoxicity did not correlate with the *in vivo* antitumor activity. This might be due to pharmacodynamic factors, so the metabolism of the basic compound 12n in rats and mice was investigated (Fig. 2). At 5, 15, 30 min and 1 hr after intraperitoneal administration of 12n to mice or rats, metabolites in

Table IV. Structure-Antitumor Activity Relationship of 1-Benzyl-4-n-hexyl-2,3-dioxopiperazine Derivatives

| - | R. , | In vitro | | In vivo | |
|---------------|------------------------------------|------------------------------------|-----------------------------------|---------------|---------------|
| Compd. No. | ~~~ | MIC(μg/ml) ^{α)} HeLaS3 | $\widehat{\mathrm{LD}_{50}^{b)}}$ | EAC(i.p. | $-i.p.)^{c)}$ |
| | \/ | песаза | (mg/kg) | Dose(mg/kg) | T/C(%) |
| | ,OCH₃ | | | | |
| 2e | CH ₃ O- | 3.13 | 200 | 40×7 | 110 |
| | CH ₃ O OCH ₃ | | | | |
| 12h | CH ₃ O- | 3.13 | 200 | 40×7 | 107 |
| 40.4 | OCH_3 | c or | 000 | 407 | 101 |
| 12d | CH ₃ O | 6.25 | 200 | 40×7 | 101 |
| 12n | $(C_2H_5)_2N$ | 3.13 | 100 | 40×7 | 157 |
| 1211 | (02115)21 | 0.10 | 100 | 10 / 1 | 101 |

- a) See the legend to Table I.
- Animals; SLC-ICR (9) mice, 6 weeks old, 2 mice/group. Treatment: i.p.
- Observation period: 1 week.

 c) Animals: SLC-ICR (2) mice, 6 weeks old, 4 or 5 mice/group.
 Treatment: from day 1 to day 7
 Inoculum size: EAC 1×10⁶ cells/mouse, i.p.

 $T/C(\%) = \frac{\text{mean survival days of treated}}{\text{mean survival days of control}} \times 100$

the blood, intraperitoneal cavity, liver, urine and feces were qualitatively studied by thin-layer chromatography (TLC). The metabolites of 12n in rats were the same as those in mice. 12n disappeared from the blood at 30 min after administration. 12n could not be detected in urine and feces at 15 min after administration, and three metabolites (19, 20, 21) were observed.

TABLE V. Structure-Cytotoxicity Relationship of 1-n-Hexyl-2,3-dioxopiperazine Derivatives

$$(C_2H_5)_2N X-N$$
 $N-C_6H_{13}$

| Compd. | -X- | $\mathrm{MIC}(\mu\mathrm{g/ml})^{a}$ | | | |
|--------|------------------|--------------------------------------|------|--|--|
| No. | 28 | HeLaS3 | EAC | | |
| 14a | | 12.5 | 50 | | |
| 14b | -CO- | 100 | 100 | | |
| 12n | -CH ₂ | 3.13 | 3.13 | | |
| 14c | $-CH_2CH_2-$ | 12.5 | 50 | | |
| 14d | -CH=CH- | 25 | 25 | | |

a) See the legend to Table I.

Table VI. Structure-Antitumor Activity Relationship of 1-(4-Diethylaminobenzyl)-2,3-dioxopiperazine Derivatives

| | | In vitroa) | | In vivo | | | |
|---------------|--|----------------------|--|-----------------|---------------|--|--|
| Compd. No. | R | MIC(μg/ml) HeLaS3 | $LD_{50}^{b)}$ (mg/kg) | EAC(i.p) | $-i.p.)^{c)}$ | | |
| | | | (| Dose (mg/kg) | T/C (%) | | |
| 18a | Н | 100 | | | | | |
| 18b | C_2H_5 | 100 | | | production . | | |
| 12 n | C ₆ H ₁₃ | 3.13 | 100 | 40×7 | 157 | | |
| 18c | -< > | 12.5 | - | 40×7 | 102.6 | | |
| 18d | ∕\/\CH ₃ | 6.25 | arrangem. | 40×7 | 107.7 | | |
| 18e | $-(CH_2)_2NH_2$ | 25 | 200300 | 40×7 | 134 | | |
| 18 f | $-(CH_2)_3NH_2$ | >100 | and defining the same of the s | 40×7 | 116.7 | | |
| 18 g | -(CH2)6NH2 OCH ₃ | 12.5 | 100 | 40×7 | 113 | | |
| 18h | N=\OCH ₃ OH | 1.56—3.13 | e manager | 40 × 7 | 121 | | |
| 18i | -\bigcap \bigcap \bigc | 100 | | | | | |
| 18 j | -CH2\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\ | 25 | 500 | 100 × 7 | 159.5 | | |
| 18k | -CH₂∕N | 25 | 500 | 100×7 | 164.5 | | |

a) See the legend to Table I.

b,c) See the legend to Table IV.

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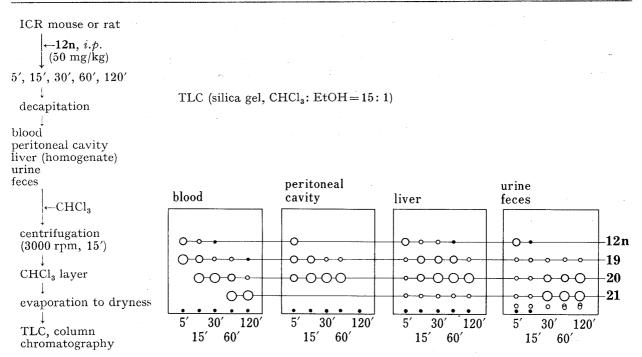
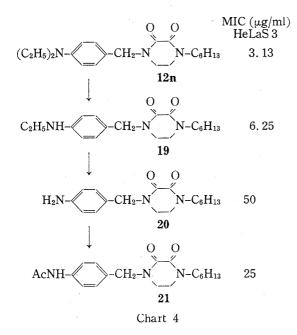


Fig. 2. Metabolism of 12n in Rats and Mice



rved. These metabolites were separated and purified by column chromatography. It was found that 19 was an EtNH compound, 20 was an NH₂ compound and 21 was an AcNH compound. Their structures were confirmed by comparison of the infrared (IR) and nuclear magnetic resonance (NMR) spectra, and the Rf values on TLC of 19, 20, and 21 with those of authentic samples synthesized independently. These metabolites showed lower cytotoxicity than 12n. It may be concluded that 12n was easily metabolized to 21 through the pathway shown in Chart 4, inactivated and excreted.

The results of this investigation suggest that a more potent antitumor agent which possesses high *in vivo* antitumor activity reflecting the *in vitro* cytotoxicity can be

developed if the metabolism of the Et₂N-moiety of 12n can be suppressed. Studies along this line are in progress.

Experimental⁶⁾

Cell Culture of HeLa S3 and Ehrlich Cells⁷⁾——In each case 5×10^4 cells/bottle were inoculated in a 5 ml culture bottle containing 3 ml of Eagle's minimum essential medium (MEM) supplemented with 10% calf serum and cultured at 37° in a CO₂-incubator for 5 days. Cells which grew on a glass surface, after removal of the culture solution, in PBS (–) (Ca²+ and Mg²+ are free) containing 0.02% ethylenediaminetetraacetic acid (EDTA) were incubated for 5—7 min and then an equal volume of 0.2% trypsin was added. The clusters of cells were dissociated by pipetting. The cell suspension was added to a centrifuge tube containing 5 ml of Eagle's MEM and then centrifuged at 1000 rpm for 5 min. The precipitate was added to fresh medium. The resulting cell suspension was used for tests.

Cytotoxic Effects on HeLa and Ehrlich Cells in Microplate Tests7)——Microplates (Sumitomo Bakelite

Co.) having 3×12 chambers per plate flattened on the bottom were used. Test compounds were dissolved in 20% dimethyl sulfoxide (DMSO) and sterilized by filtration through a Millipore filter, and then they were diluted with culture medium in 2 fold serial dilutions (12 steps). The concentrations of the compounds were from $100~\mu\text{g/ml}$ to $0.05~\mu\text{g/ml}$. Test compounds dissolved in culture medium (0.1 ml) and cell suspension (0.1 ml, 2×10^4 cells/ml) in one chamber were cultured at 37° in a CO₂-incubator for 5 days. The culture medium was removed and the cells were washed twice with Hanks' salt solution. The cells were fixed for 5 min with 95% EtOH and stained with Giemsa solution for 15 min. The inhibition of cell growth could be observed by measuring the degree of cell staining on microplates macroscopically, because chambers in which the cell growth was inhibited were not stained. MIC values (minimum inhibitory concentration) were determined as the minimum concentrations of the compounds at which the cell growth was inhibited.

Acute Toxicity to Mice—One group, consisting of two 5-week-old female SLC-ICR mice weighing of 20 ± 1 g, received test compounds dissolved or suspended in saline or saline containing 0.3% carboxymethyl cellulose (CMC) intraperitoneally. At 7 days after administration, the number of deaths was noted and the LD₅₀ was calculated.

Antitumor Activity against Ehrlich Ascites Carcinoma (i.p.-i.p.)—EAC cells $(1\times10^6$ cells/head/0.2 ml saline) were intraperitoneally transplanted into one group containing of five 6-week-old female ICR mice weighing of 22 ± 1 g. Test compounds suspended in 0.3% CMC saline or dissolved in saline were intraperitoneally administered daily from the day after transplantation for 7 days. Antitumor activity (T/C%) was evaluated from the following formula.

$$T/C(\%) = \frac{\text{mean survival days of treated group}}{\text{mean survival days of control group}} \times 100$$

1-(2,4-Dimethoxybenzyl)-4-(2,4-hexadien-1-yl)-2,3-dioxopiperazine (2b)——A solution of 1-(2,4-dimethoxybenzyl)-2,3-dioxopiperazine⁵⁾ (2a) (10 g) in dimethylformamide DMF (70 ml) was added dropwise to a suspension of NaH (purity 50%, 1.8 g) in DMF (20 ml) over a period of 10 min at room temperature. The mixture was stirred at 50—60° for 30 min, then a solution of 2,4-hexadien-1-yl bromide (7.0 g) in DMF (20 ml) was added. The reaction mixture was stirred at 50—60° for 1 hr and evaporated to dryness in vacuo. The residue was dissolved in CHCl₃ (50 ml), washed with H_2O and dried over MgSO₄. Removal of the solvent in vacuo left a brown oil, which was chromatographed on silica gel with CHCl₃ to give 2b as a liquid. Yield 10.8 g (81.7%). IR $v_{\rm max}^{\rm mex}$ cm⁻¹: 1668 (C=O). NMR (CDCl₃) δ : 1.72 (3H, d, J=5.5 Hz, CH₃), 3.38 (4H, bs, piperazine ring 5 and 6 CH₂), 3.77 (6H, s, $2 \times {\rm OCH_3}$), 4.01 (2H, d, J=6.5 Hz, CH₂), 4.54 (2H, s, CH₂), 5.23—6.14 (4H, m, -CH=CH-CH=CH-), 6.27—6.66 (2H, m, benzene ring 3 and 5 CH), 7.15 (1H, d, J=9 Hz, benzene ring 6 CH).

1-(2,4-Hexadien-1-yl)-2,3-dioxopiperazine (3)—Method A: CF₃COOH (50 ml) was added to a solution of **2b** (5.0 g) in anisole (50 ml) at room temperature. After being stirred at room temperature for 1 hr and at 50—60° for 1 hr, the whole was evaporated to dryness in vacuo. The residue was washed with iso-Pr₂O and dissolved in CHCl₃ (50 ml). The solution was washed with H₂O and dried over anhydrous MgSO₄. After removal of the solvent, the residue was chromatographed on silica gel with CHCl₃-EtOH to give 3 (0.2 g, 7.1%) as colorless crystals of mp 138—139° (iso-PrOH). IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 1695, 1665 (C=O). NMR (DMSO- d_6) δ : 1.71 (3H, d, J=4.2 Hz, CH₃), 3.42 (4H, bs, piperazine ring 5 and 6 CH₂), 4.00 (2H,-d, J=6.6 Hz, CH₂), 5.20—6.30 (4H, m, -CH=CH-CH=CH-), 8.42 (1H, bs, NH). Anal. Calcd for C₁₀H₁₄N₂O₂: C, 61.83; H, 7.27; N, 14.22. Found: C, 61.68; H, 7.19; N, 14.08.

Method B: 2,4-Hexadien-1-yl bromide (14.9 g) was added dropwise to a solution of ethylenediamine (22.2 g) in EtOH (200 ml) over a period of 30 min at -30° , and the mixture was refluxed for 4 hr. After removal of the solvent in vacuo, H_2O (20 ml) and NaOH (10 g) were added to the residue and the separated oil was distilled to give N-(2,4-hexadien-1-yl)ethylenediamine (5a) (6.3 g, 48.7%) as a pale yellow liquid of bp 200—230° (15 mmHg). This liquid (6.3 g) and diethyl oxalate (6.56 g) were added dropwise at the same time to EtOH (60 ml) under ice-cooling over a period of 20 min. The reaction mixture was refluxed for 5 hr. Removal of the solvent in vacuo left a pale brown solid, which was chromatographed on silica gel with CHCl₃-EtOH to give 3 (3.5 g, 40%) as colorless crystals of mp 140° (iso-PrOH). The IR and NMR spectra and Rf value on TLC were identical with those of the product obtained by method A.

Compounds 5b and 6 were similarly synthesized, as described in an earlier paper.8)

1-n-Octyl-4-(2,4-hexadien-1-yl)-2,3-dioxopiperazine (1b)——Compound 1b was obtained from 1-n-octyl-2,3-dioxopiperazine (6) and 2,4-hexadien-1-yl bromide by the procedure used to prepare 2b. mp 81—83° (iso-Pr₂O-petr. ether). IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 1659 (C=O). NMR (CDCl₃) δ : 0.86 (3H, m, CH₃), 1.00—1.80 (12H, m, 6×CH₂), 1.73 (3H, d, J=4.2 Hz, CH₃), 3.31—3.53 (6H, m, piperazine ring 5 and 6 CH₂ and NCH_2 -), NCH_2 -),

4.07 (2H, d,
$$J=6.0~{\rm Hz}$$
, $NCH_2CH=$), 5.10—6.40 (4H, m, $-CH=CH-CH=CH-$). Anal. Calcd for $C_{18}H_{30}-CH=CH=CH-$

 N_2O_2 : C, 70.55; H, 9.87; N, 10.94. Found: C, 70.68; H, 9.93; N, 11.10.

Compound 1a was similarly obtained. mp 136—138° (AcOEt). IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 1660, 1640 (C=O). Anal. Calcd for $C_{16}H_{22}N_2O_2$: C, 70.05; H, 8.80; N, 10.21. Found: C, 70.21; H, 8.11; N, 10.30.

1-(2,4-Dimethoxybenzyl)-4-n-hexyl-2,3-dioxopiperazine (2e)—Compound 2e was obtained from 2,4-dimethoxybenzaldehyde by the procedure used to prepare 2b. mp 77—79° (AcOEt-iso-Pr₂O). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1658 (C=O). NMR (CDCl₃) δ : 0.81 (3H, m, CH₃), 1.00—1.80 (8H, m, 4×CH₂), 3.20—3.60 (6H, m, piperazine ring 5 and 6CH₂ and $NC\underline{H}_2$ -C₅H₁₁), 3.78 (6H, s, 2×OCH₃), 4.56 (2H, s, —C \underline{H}_2 -N),

6.25—6.45 (2H, m, benzene ring 3 and 5 CH), 7.17 (1H, d, J=8.5 Hz, benzene ring 6 CH). Anal. Calcd for $C_{19}H_{28}N_2O_4$: C, 65.49; H, 8.10; N, 8.04. Found: C, 65.26; H, 8.13; N, 7.94.

The following compounds (Table II) were similarly synthesized (Table VII).

TABLE VII

| Compd. | mp (°C) (Recryst. solv.) | Formula | Analysis(%) Calcd (Found) | | | $rac{	ext{NMR (solvent)}}{\delta 	ext{ (ppm)}}$ |
|------------|---|-----------------------------------|---------------------------|--------------|---------------|--|
| | 3014.) | | С | Н | N | |
| 2c | 115 —116.5 (AcOEt) | $C_{16}H_{20}N_2O_4$ | 63.14 (62.92 | 6.62 6.60 | 9.21 9.02) | (CDCl ₃) 3.42 (4H, bs), 3.79 (6H, s), 4.04 (2H, d), 4.60 (2H, s), 5.05 (1H, m), 5.27 (1H, m), 5.70 (1H, m), 6.30—6.47 (2H, m), 7.18 (1H, d) |
| 2d | $76 - 78$ (AcOEt-iso- Pr_2O) | $C_{17}H_{24}N_2O_4$ | 63.73 (63.87 | 7.55 7.58 | 8.74 8.77) | (CDCl ₃) 0.85 (3H, m), 1.00—1.80 (4H, m), 3.35—3.55 (6H, m), 3.78 (6H, s), 4.59 (2H, s), 6.30—6.48 (2H, m), 7.18 (1H, d) |
| 2f | 81.5— 83.5 (AcOEt- iso-Pr ₂ O) | $\mathrm{C_{20}H_{30}N_{2}O_{4}}$ | 66.27 (65.95 | | 7.73 7.70) | (CDCl ₃) 0.84 (3H, t), 1.00—1.68 (10H, m), 3.11—3.58 (6H, m), 3.74 (6H, s), 4.52 (2H, s), 6.18—6.48 (2H, m), 7.10 (1H, d) |
| 2g | 75 - 76 (AcOEt-iso-Pr ₂ O) | $\mathrm{C_{21}H_{32}N_2O_4}$ | 66.99 (66.91 | | 7.44 7.49) | (CDCl ₃) 0.84 (3H, m), 1.00—1.80 (12H, m), 3.30—3.50 (6H, m), 3.76 (6H, s), 4.55 (2H, s), 6.26—6.45 (2H, m), 7.17 (1H, d) |
| 2h | Oil | | | | | (CDCl ₃) 0.67—1.02 (3H, m), 1.07—1.50 (4H, m), 1.59 (3H, s), 1.67 (3H, s), 3.44 (6H, bs), 3.76 (3H, s), 3.79 (3H, s), 3.91—4.27 (1H, m), 4.58 (2H, s), 4.87—5.27 (1H, m), 6.23—6.54 (2H, m), 7.19 (1H, d) |
| 2i | 93 - 95 (AcOEt- iso-Pr ₂ O) | ${ m C_{20}H_{28}N_{2}O_{4}}$ | 66.64 (66.64 | | 7.77 7.68) | (DMSO-d ₆) 1.00—2.10 (11H, m), 3.22 (2H, d), 3.46 (4H, bs), 3.79 (3H, s), 3.80 (3H, s), 4.49 (2H, s), 6.43—6.56 (2H, m), 7.12 (1H, d) |
| 2 j | 108 —110 (EtOH) | $C_{21}H_{24}N_2O_4$ | 68.46 (68.02 | | 7.60 7.53) | (CDCl ₃) 2.81 (2H, t), 3.16 (4H, s), 3.59 (2H, t), 3.70 (6H, s), 4.47 (2H, s), 6.26— 6.69 (2H, m), 6.89—7.19 (6H, m) |
| 2k | 75 — 76 (iso-PrOH— iso-Pr $_2$ O) | $\mathrm{C_{19}H_{27}BrN_2O_4}$ | 53.40 (53.59 | 6.36 6.48 | 6.55 6.59) | (CDCl ₃) 1.10—2.10 (8H, m), 3.22—3.57 (8H, m), 3.75 (6H, s), 4.54 (2H, s), 6.24—6.43 (2H, m), 7.12 (1H, d) |

^{1-(2,4-}Dimethoxybenzyl)-4-(6-hydroxy-1-hexyl)-2,3-dioxopiperazine (21)——Compound 2k (2.0 g) and AcONa (0.6 g) were added to DMF (10 ml) and the reaction mixture was refluxed for 3 hr. After removal of the solvent *in vacuo*, the residue was dissolved in AcOEt (50 ml) and the solution was washed with H₂O and dried over MgSO₄. The solid obtained by removing the solvent was dissolved in MeOH (10 ml). NaOMe (0.4 g) was added to the above solution and the whole was stirred for 12 hr at room temperature. After removal of the solvent *in vacuo*, CHCl₃ was added to the residue and the extract was washed with H₂O and dried over MgSO₄. The oil obtained by removing the solvent was chromatographed on silica gel with CHCl₃-

EtOH to afford 21 as a liquid. Yield 1.13 g (66%). IR $v_{\text{max}}^{\text{neat}}$ cm⁻¹: 1650 (C=O). NMR (CDCl₃) δ : 1.06—1.82 (8H, m, 4×CH₂), 2.82 (1H, s, OH), 3.12—3.58 (8H, m, piperazine ring 5 and 6 CH₂, $N-CH_2(CH_2)_5-$ and

 $-(CH_2)_5-C\underline{H}_2OH)$, 3.70 (6H, s, $2\times OCH_3$), 4.49 (2H, s, $-(EH_2)_5-(EH_2)_5$), 6.18—6.40 (2H, m, benzene ring 3 and 5 CH), 7.08 (1H, d, J=9 Hz, benzene ring 6 CH).

1-(6-Amino-1-hexyl)-4-(2,4-dimethoxybenzyl)-2,3-dioxopiperazine Hydrochloride (2m·HCl)—2m·HCl was similarly obtained by the method described in the previous paper, i.e., Gabriel's primary amine synthetic method. mp 59—62°. IR ν_{\max}^{RBr} cm⁻¹: 1650 (C=O). NMR (CDCl₃-DMSO- d_6) δ : 0.85—1.95 (8H, m, 4×CH₂), 2.65—3.02 (2H, m, -(CH₂)₅CH₂NH₂), 3.02—3.60 (6H, m, piperazine ring 5 and 6 CH₂ and $NCH_2(CH_2)_5$ -), 3.75 (6H, s, 2×OCH₃), 4.43 (2H, s, -CH₂N), 6.29—6.70 (2H, m, benzene ring C)

3 and 5 CH), 7.05 (1H, d, J=8.5 Hz, benzene ring 6 CH), 7.70—8.65 (2H, bs, NH₂). Anal. Calcd for C₁₉H₃₀-ClN₃O₄: C, 57.06; H, 7.56; N, 10.51. Found: C, 56.74; H, 7.80; N, 10.17.

0.55 g) was added to a solution of 2a (3.0 g) in DMF (30 ml) and after the mixture had been stirred for 1 hr at 50-60°, 1-bromo-2,2-diethoxyethane (2.9 g) was added dropwise at room temperature. After removal of the solvent in vacuo, the residue was dissolved in $\mathrm{CH_2Cl_2}$ (50 ml) and the solution was washed with $\mathrm{H_2O}$ and dried over MgSO4. After removal of the solvent under reduced pressure, 6 N HCl (50 ml) was added to the residue and the whole was stirred for 2 hr at 50-60°. The reaction mixture was extracted with AcOEt (50 ml), washed with satd. aq. NaCl and dried over MgSO4. Removal of the solvent by evaporation gave 1-(2,4-dimethoxybenzyl)-4-formylmethyl-2,3-dioxopiperazine (2.1 g) as crystals. The crystals were added to a solution of 3-ethoxycarbonyl-2-buten-1-yltriphenylphosphonium bromide (3.0 g) and NaOMe (0.38 g) in abs. CH_2Cl_2 (30 ml) and the reaction mixture was refluxed for 4 hr. H_2O (30 ml) was added. The separated organic layer was washed with satd. aq. NaCl and dried over MgSO4. After removal of the solvent, the residue was chromatographed on silica gel with CHCl3 to give an oil. 6 n HCl (20 ml) was added to the oil and the whole was refluxed for 5 hr. The reaction mixture was adjusted to pH 8 with NaHCO3 and washed with AcOEt (30 ml). The $\rm H_2O$ layer was adjusted to pH 1 with $\rm 6\,N$ HCl and extracted with AcOEt (30 ml). The extract was washed with satd. aq. NaCl and dried over MgSO4. After removal of the solvent, the residue was recrystallized from iso-PrOH. Yield 1.41 g (32%). mp 153—155°. IR $v_{\rm max}^{\rm KBT}$ cm $^{-1}$: 1670 (C=O). NMR (CDCl $_3$) δ : 1.93 (3H, s, CH $_3$), 3.43 (4H, bs, piperazine ring 5 and 6 CH $_2$), 3.77 (6H, $-CH_2\dot{N}$), 5.45—6.20 (3H, m, $N-CH_2CH=$), 4.59 (2H, s, \langle s, $2 \times OCH_3$), 4.00-4.45 (2H, m,

 $-C\underline{H}=C\underline{H}-C\underline{H}=C(CH_3)-)$, 6.20—6.65 (2H, m, benzene ring 3 and 5 CH), 7.23 (1H, d, J=9 Hz, benzene ring 6 CH), 9.04 (1H, bs, COOH). Anal. Calcd for $C_{20}H_{24}N_2O_6$: C, 61.85; H, 6.23; N, 7.21. Found: C, 61.62; H, 6.46; N, 6.81.

1-n-Hexyl-4-(4-methoxybenzyl)-2,3-dioxopiperazine (12b)——Compound 12b was synthesized from 1-n-hexyl-2,3-dioxopiperazine and 4-methoxybenzyl chloride by method B shown in Chart 2. mp 137—138° (AcOEt). IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 1660 (C=O). Anal. Calcd for $C_{18}H_{26}N_2O_3$: C, 67.90; H, 8.23; N, 8.80. Found: C, 67.87; H, 8.27; N, 8.76.

The following compounds were similarly obtained (Table VIII).

TABLE VIII

| Compd. | mp (°C) (Recryst. solvent) | Formula | Analysis (%) Calcd (Found) | | | NMR (solvent) δ (ppm) |
|--------|--|-------------------------------|----------------------------|--------------|---------------|---|
| | | | С | H | N | |
| 12a | 75 - 76 (AcOEt-iso-Pr ₂ O) | ${ m C_{18}H_{26}N_2O_3}$ | 67.90 (67.74 | 8.23 8.13 | 8.80 8.64) | (CDCl ₃) 0.85 (3H, t), 1.01—1.81 (8H, m), 3.21—3.61 (6H, m), 3.79 (3H, s), 4.63 (2H, s), 4.66—6.99 (2H, m), 7.04—7.41 (2H, m) |
| 12c | 72 - 74 (AcOEt-iso-Pr ₂ O) | $C_{19}H_{28}N_2O_4$ | 65.49 (65.57 | 8.10 7.99 | 8.04 8.03) | (CDCl ₃) 0.86 (3H, t), 1.04—1.84 (8H, m), 3.14—3.54 (6H, m), 3.80 (3H, s), 3.83 (3H, s), 4.67 (2H, s), 6.60— 6.97 (3H, m) |
| 12d | 61 — 64 (AcOEt- iso-Pr ₂ O) | $\mathrm{C_{19}H_{28}N_2O_4}$ | 65.49 (65.54 | 8.10 8.02 | 8.04 7.63) | (CDCl ₃) 0.86 (3H, m), 1.00—1.90 (8H, m), 3.30—3.50 (6H, m), 3.71 (3H, s), 3.76 (3H, s), 4.63 (2H, s), 6.76— 6.91 (3H, m) |

| Compd. | mp (°C) (Recryst. | (Recryst. Formula | | | %) | $egin{array}{c} 	ext{NMR} & 	ext{(solvent)} \ \delta & 	ext{(ppm)} \end{array}$ |
|-------------|--|-------------------------------|-----------------|--------------|-----------------|---|
| 2.0. | solvent) | | ć | Н | N | |
| 12e | 101 —104 (AcOEt— iso-Pr ₂ O) | $C_{19}H_{28}N_2O_4$ | 65.49 (65.24 | | 8.04 7.78) | (CDCl ₃) 0.87 (3H, m), 1.00—1.70 (8H, m), 3.10—3.70 (6H, m), 3.79 (6H, s), 4.77 (2H, s), 6.50 (2H, d), 7.19 (1H, d) |
| 12f | 85 - 87 (AcOEt-iso-Pr ₂ O) | $C_{19}H_{28}N_2O_4$ | 65.49 (65.59 | 8.10 8.23 | 8.04 8.11) | (CDCl ₃) 0.85 (3H, m), 1.10—1.80 (8H, m), 3.20—3.60 (6H, m), 3.83 (6H, s), 4.55 (2H, s), 6.80 (3H, bs) |
| 12h | 111.5—113 (AcOEt– iso-Pr ₂ O) | $\mathrm{C_{20}H_{30}N_2O_5}$ | 63.47 (63.31 | 7.99 7.83 | 7.40 7.20) | (CDCl ₃) 0.86 (3H, t), 1.03—1.81 (8H, m), 3.11—3.64 (6H, m), 3.83 (3H, s), 3.86 (3H, s), 3.88 (3H, s), 4.61 (2H, s), 6.64 (1H, d), 7.03 (1H, d) |
| 12 i | 97 - 98 (AcOEt-iso-Pr ₂ O) | $C_{20}H_{30}N_2O_5$ | 63.47 (63.52 | | 7.40 7.37) | (CDCl ₃) 0.86 (3H, m), 1.00—1.80 (8H, m), 3.38—3.64 (6H, m), 3.82 (9H, s), 4.58 (2H, s), 6.48 (2H, s) |
| 121 | 103 —105 (AcOEt) | $C_{17}H_{23}ClN_2O_2$ | 63.25 (63.40 | 7.18 7.32 | 8.68 8.86) | (CDCl ₃) 0.88 (3H, m), 1.05—1.85 (8H, m), 3.31—3.65 (6H, m), 4.62 (2H, s), 7.25 (4H, s) |
| 12m | 106 —108 (EtOH) | $C_{17}H_{22}Cl_2N_2O_2$ | 57.15 (57.09 | 6.21 6.27 | 7.84 7.87) | (CDCl ₃) 0.85 (3H, m), 1.05—1.85 (8H, m), 3.32—3.60 (6H, m), 4.72 (2H, s), 7.10—7.45 (3H, m) |
| 12p | Oil | | | | | (CDCl ₃) 1.04 (6H, t), 0.71—1.82 (11H, m), 3.30 (4H, q), 3.39—3.95 (6H, m), 6.63—7.11 (4H, m) |
| 12q• HCl | 188 —190 (MeOH) | $C_{22}H_{36}ClN_3O_2$ | 62.24 (63.74 | | 10.89 10.09) | (DMSO- d_6) 0.85 (3H, m), 1.01—1.83 (8H, m), 1.12 (6H, t), 3.21—3.68 (10H, m), 4.62 (2H, s), 4.48 (2H, s), 6.50 (2H, d), 7.10 (2H, d) |

1-(4-Diethylaminobenzyl)-4-n-hexyl-2,3-dioxopiperazine (12n)—4-Diethylaminobenzaldehyde (22.2 g) and N-acetylethylenediamine (12.8 g) in benzene (130 ml) were refluxed azeotropically for 4 hr. After removal of the solvent in vacuo, the residue was dissolved in MeOH (160 ml) and NaBH₄ (6.2 g) was added in portions under ice-cooling. The mixture was stirred for 3 hr under ice-cooling, then the solvent was removed. The residual oil in CHCl₃ (100 ml) was washed with H₂O and dried over MgSO₄. After removal of the solvent, the residual oil was taken up in 20% HCl (110 ml) and refluxed for 1 hr. The reaction mixture was made strongly basic with KOH and extracted with benzene (100 ml). The organic layer was dried over MgSO₄ and evaporated to dryness to give an oil, which was distilled to afford N-(4-diethylaminobenzyl)-ethylenediamine (10n) as a liquid of 165—176° (2—3 mmHg). Yield 26.8 g (96.6%). Solutions of 10n (26.8 g) in EtOH (20 ml) and of diethyl oxalate (17.7 g) in EtOH (20 ml) were added dropwise to EtOH (180 ml) over a period of 1 hr under ice-cooling and the whole was refluxed for 3 hr. Next, 80 ml of EtOH was removed and remaining reaction mixture was left to stand at room temperature. Separated crystals were collected and recrystallized from EtOH to give 1-(4-diethylaminobenzyl)-2,3-dioxopiperazine (11n) as colorless crystals of mp 214.5—215.5° (18.9 g, 56.7%). IR $v_{\rm max}^{\rm KBT}$ cm⁻¹: 3180, 3125 (NH), 1650 (C=O). Anal. Calcd for C₁₅H₂₁N₃O₂: C, 65.43; H, 7.69; N, 15.26. Found: C, 65.46; H, 7.61; N, 15.22.

The following compounds were similarly obtained. 12g: mp 91—92° (AcOEt-iso-Pr₂O). NMR (CDCl₃)

 $\delta\colon 0.86$ (3H, t), 1.00—1.69 (8H, m), 3.00—3.56 (6H, m), 3.73 (6H, s), 3.76 (3H, s), 4.64 (2H, s), 6.04 (2H, s). Anal. Calcd for $C_{20}H_{30}N_2O_5\colon C$, 63.47; H, 7.99; N, 7.40. Found: C, 63.31; H, 8.00; N, 7.31. 12o: oil. NMR (CDCl₃) $\delta\colon 0.85$ (3H, m), 1.11 (12H, t), 1.00—1.70 (8H, m), 2.90 (8H, q), 3.20—3.60 (6H, m), 4.67 (2H, s), 6.36 (2H, m), 7.00 (1H, m).

1-(2,4-Dihydroxybenzyl)-4-n-hexyl-2,3-dioxopiperazine (12k)—Compound 2e (2.0 g) in abs. CH_2Cl_2 (20 ml) was treated with BBr₂ (4.32 g) at -20 to -30° and the solution was stirred for 1 hr at room temperature. MeOH (5 ml) was added to the reaction mixture and the solvent was evaporated off. The residue was dissolved in AcOEt (30 ml) and the solution was washed with satd. aq. NaCl and dried over MgSO₄. After removal of the solvent in vacuo, the residue was recrystallized from AcOEt. Yield 0.77 g (42%). mp 131—133°. IR $\nu_{\text{max}}^{\text{kBr}}$ cm⁻¹: 1620 (C=O). Anal. Calcd for $C_{17}H_{24}N_2O_4$: C, 63.73; H, 7.55; N, 8.74. Found: C, 63.64; H, 7.54; N, 8.62. Compound 12j was similarly obtained. mp 82—87° (CHCl₃). IR $\nu_{\text{max}}^{\text{kBr}}$ cm⁻¹: 1653 (C=O). NMR (DMSO- d_6) δ : 0.86 (3H, t), 1.02—1.79 (8H, m), 3.09—3.69 (6H, m), 4.47 (2H, s), 6.71 (2H, d), 7.11 (2H, d), 9.34 (1H, s). Anal. Calcd for $C_{17}H_{24}N_2O_3$: C, 67.08; H, 7.95; N, 9.20. Found: C, 66.96; H, 8.12; N, 9.16.

1-(4-Diethylaminophenyl)-4-n-hexyl-2,3-dioxopiperazine (14a)——1-n-Hexyl-2,3-dioxopiperazine (1.7 g), 4-diethylaminoiodobenzene (3.1 g), K₂CO₃ (1.5 g), and activated Cu⁹) (as a catalyst) were added to DMF (20 ml) and the mixture was refluxed for 5 hr. H₂O (50 ml) was added and the reaction mixture was extracted with AcOEt. The organic layer was washed with satd. aq. NaCl and dried over MgSO₄. After removal of the solvent *in vacuo*, the residue was washed with iso-Pr₂O and recrystallized from AcOEt. Yield 1.5 g (50.7%). mp 114—115.5°. IR ν_{max} cm⁻¹: 1680 (C=O). NMR (CDCl₃) δ: 3.32 (4H, q, J=7 Hz, 2×CH₂), 3.39—3.97 (6H, m, piperazine ring 5 and 6 CH₂ and NCH₂-), 6.56 (2H, d, J=9 Hz, benzene ring 3 and 5

CH), 7.09 (2H, d, J=9 Hz, benzene ring 2 and 6 CH). Anal. Calcd for $C_{20}H_{31}N_3O_2$: C, 69.53; H, 9.05; N, 12.16. Found: C, 69.43; H, 9.02; N, 12.28.

5 CH), 7.01 (2H, d, J=9 Hz, benzene ring 2 and 6 CH). Anal. Calcd for $C_{22}H_{35}N_3O_2$ ·HCl: C, 64.45; H, 8.85; N, 10.25. Found: C, 64.23; H, 8.83; N, 10.03.

Compound 14b was similarly obtained. mp 115—116.5° (AcOEt-iso-Pr₂O). IR ν_{\max}^{KBr} cm⁻¹: 1680, 1650 (C=O). NMR (CDCl₃) δ : 0.89 (3H, m), 1.16 (6H, t), 3.40 (4H, q), 3.46 (2H, t), 3.80 (4H, m), 6.72 (2H, d), 7.59 (2H, d). Anal. Calcd for C₂₁H₃₁N₃O₃: C, 67.53; H, 8.37; N, 11.25. Found: C, 67.33; H, 8.32; N, 11.12.

1-[2-(4-Diethylaminophenyl)vinyl]-4-n-hexyl-2,3-dioxopiperazine (14d)—1-n-Hexyl-2,3-dioxopiperazine (10.0 g) in 37% formalin (50 ml) was refluxed for 30 min. The reaction mixture was evaporated to dryness and the residue was extracted with MeOH. After removal of the solvent, the residual oil was added to SOCl₂ (30 ml) and the mixture was refluxed for 30 min. The residue obtained by removal of excess SOCl₂ was dissolved in benzene (100 ml) and a solution of Ph₃P (13.2 g) in benzene (30 ml) was added to the above solution, then the whole was refluxed for 3 hr. The separated oil was isolated by decantation and washed with iso-Pr₂O to afford (4-n-hexyl-2,3-dioxo-1-piperazinyl)methyl triphenylphosphonium chloride (17) as hygroscopic crystals (11.0 g, 43%). NaOMe (1.9 g) was added to 17 (8.6 g) in CH₂Cl₂ (100 ml) at -10° . The mixture was stirred at -10° for 10 min, then a solution of 4-diethylaminobenzaldehyde (3.0 g) in CH₂Cl₂ (10 ml) was added and the whole was refluxed for 2 hr. The reaction mixture was extracted with 1 n HCl (30 ml). The extract was neutralized with NaHCO₃ and extracted with CH₂Cl₂. The organic layer was washed with H₂O and dried over MgSO₄. After removal of the solvent, the residue was chromatographed on silica gel to give 14d as an oil (2.82 g, 45%). IR v_{max}^{nest} cm⁻¹: 1660 (C=O). NMR (CDCl₃) δ : 0.87 (3H, m, CH₃), 1.25 (6H, t, J=6.5 Hz, 2×CH₃CH₂N-), 1.10—1.80 (8H, m, 4×CH₂), 3.32 (4H, q, J=6.5 Hz, 2×CH₃CH₂N), 3.37—3.70 (6H, m, piperazine ring 5 and 6 CH₂ and NCH₂-), 6.05 (1H, d, J=9.5 Hz,

—CH=CHN), 6.60 (2H, m, benzene ring 3 and 5 CH), 6.69 (1H, d, J=9.5 Hz, —CH=CH), 7.37 (2H, m, benzene ring 2 and 6 CH).

1-(4-Diethylaminobenzyl)-4-(2-pyridylamino)-2,3-dioxopiperazine (18k)—Compound 18k was obtained by a method similar to that described for 12n. mp 95° (AcOEt-iso-Pr₂O). IR ν_{\max}^{KBr} cm⁻¹: 1670 (C=O). NMR (CDCl₃) δ : 1.14 (6H, t, J=7 Hz, $2\times$ CH₃), 2.70—3.00 (8H, m, $4\times$ CH₂), 4.52 (2H, s, CH₂), 4.72 (2H, s, CH₂), 6.57 (2H, d, J=8 Hz, benzene ring $2\times$ CH), 6.90—7.45 (4H, m, benzene ring $2\times$ CH and pyridine ring $2\times$ CH), 7.65 (1H, m, pyridine ring CH), 8.45 (1H, m, pyridine ring CH). Anal. Calcd for C₂₁H₂₆N₄O₂: C, 68.83; H, 7.15; N, 15.29. Found: C, 68.64; H, 7.16; N, 15.15.

The following compounds were similarly obtained (Table IX).

TABLE IX

| Compd. No. | mp°C (Recryst. solvent) | Formula | | Analysis (%) Calcd (Found) | | $rac{	ext{NMR (solvent)}}{\delta 	ext{ (ppm)}}$ |
|---------------|--|-------------------------------|-----------------|----------------------------|-----------------|---|
| 18b | 134—135 (AcOEt- | $C_{17}H_{25}N_3O_2$ | 67.30 (67.56 | | 13.85 13.69) | (CDCl ₃) 1.15 (9H, t), 3.32 (4H, q), 3.40 (4H, |
| 18c | iso-Pr ₂ O) 159—161 (AcOEt) | $\mathrm{C_{21}H_{31}N_3O_2}$ | 70.55 (70.57 | | 11.75 11.92) | s), 3.46 (2H, q), 4.50 (2H, s), 6.55 (2H, d), 7.07 (2H, d) (CDCl ₃) 1.14 (6H, t), 1.24—1.94 (10H, m), 3.00—3.79 (8H, m), 4.16—4.66 (1H, |
| 18d | Oil | | | | | m), 4.55 (2H, s), 6.59 (2H, d), 7.13 (2H, d) (CDCl ₃) 1.14 (6H, t), 1.76 (3H, d), 3.10—3.50 (8H, m), 4.01 (2H, d), 4.50 (2H, s), 5.00—6.30 (4H, m), 6.53 (2H, d), |
| 18h | 160—161 (EtOH) | $C_{21}H_{27}N_5O_4$ | 61.00 (61.15 | 6.58 6.63 | 16.94 16.79) | 7.05 (2H, d) (CDCl ₃) 1.14 (6H, t), 3.30 (4H, q), 3.30—3.67 (2H, m), 3.90 (6H, s), 4.15—4.40 (2H, m), 4.57 (2H, s), 6.56 (2H, d), 7.12 (2H, d), 7.37 (1H, s) |
| 18j | 150—152 (iso-PrOH) | $\mathrm{C_{21}H_{26}N_4O_2}$ | 68.83 (68.81 | 7.15 7.17 | 15.29 15.30) | (CDCl ₃) 1.13 (6H, t), 3.31 (4H, q), 3.35 (4H, bs), 4.50 (2H, s), 4.61 (2H, s), 6.53 (2H, d), 7.05 (2H, d), 7.24 (1H, m), 7.63 (1H, m), 8.45 (2H, m) |

1-(2-Aminoethyl)-4-(diethylaminobenzyl)-2,3-dioxopiperazine Hydrochloride (18e•2HCl)—18e·HCl was obtained by the method described in the previous paper, 3b) i.e., Gabriel's primary amine synthetic method. mp 225—226° (EtOH). IR $r_{\rm max}^{\rm KBr}$ cm⁻¹: 1665 (C=O). Anal. Calcd for $C_{17}H_{26}N_4O_2\cdot 2HCl$: C, 52.18; H, 7.21; N, 14.32. Found: C, 52.02; H, 7.19; N, 14.30.

The following compounds were similarly obtained. 18f ·HCl: mp 174—175° (EtOH). NMR (DMSOd₆) δ : 0.85—1.87 (8H, m), 2.71—3.15 (2H, m), 3.15—3.70 (10H, m), 4.30 (2H, s), 6.51 (2H, d), 7.08 (2H, d). Anal. Calcd for C₁₉H₃₀N₄O₂·2HCl: C, 54.41; H, 7.69; N, 13.36. Found: C, 54.18; H, 7.62; N, 13.22. (18g): amorphous solid. NMR (CDCl₃) δ : 0.88—1.90 (14H, m), 2.60—3.10 (2H, m), 3.10—3.62 (10H, m), 4.33 (2H, s), 6.54 (2H, d), 7.10 (2H, d). Anal. Calcd for C₂₁H₃₄N₄O₂: C, 67.34; H, 9.15; N, 14.96. Found: C, 67.25; H, 9.13; N, 14.62.

1-(4-Diethylaminobenzyl)-4-(2,6-dihydroxy-4-pyrimidinyl)-2,3-dioxopiperazine (18i)—A solution of 18h (1.0 g) in abs. CH₂Cl₂ (20 ml) was added dropwise to a solution of BBr₃ (0.6 g) in abs. CH₂Cl₂ (20 ml) at -30° over a period of 10 min. After being stirred at room temperature for 2 hr and refluxed for 30 min, the reaction mixture was cooled to 0—5°. MeOH (30 ml) was added under ice-cooling and the reaction mixture was stirred for 30 min at the same temperature. After removal of the solvent, CHCl₃ (30 ml) and H₂O (30 ml) were added to the residue. The H₂O layer was neutralized with satd. aq. NaHCO₃ and extracted with CHCl₃. The extract was dried over MgSO₄ and evaporated to dryness. The residue was chromatographed on silica gel with EtOH-CHCl₃ (1: 5) to give 18i (0.2 g, 21.5%) of mp>270°. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1685, 1655 (C=O). NMR (CDCl₃) δ : 1.11 (6H, t, J=7.5 Hz, 2×CH₃), 3.22 (4H, q, J=7.5 Hz, 2×CH₂), 3.40—3.70 (2H, m, piperazine ring CH₂), 4.10—4.40 (2H, m, piperazine ring CH₂), 4.96 (2H, s, CH₂), 6.49 (2H, d, J=8.5 Hz, benzene ring 3 and 5 CH), 7.00 (1H, s, pyrimidine ring CH), 7.20 (2H, d, J=8.5 Hz, benzene ring 2 and 6 CH), 8.14 (1H, bs, OH), 8.71 (1H, bs, OH). Anal. Calcd for C₁₉H₂₃N₅O₄: C, 59.21; H, 6.02; N, 18.17. Found: C, 59.02; H, 5.98; N, 19.97.

The Metabolism of 12n in Mice——Compound 12n (50 mg/kg) was suspended in 0.3% CMC saline and intraperitoneally administered to 6-week-old ICR female mice in which the urethral meatus was closed. Six groups, consisting of 2 mice per group, were used. At 5, 15, 30, 60, and 120 min after the administration, mice were decapitated and the blood, peritoneal cavity, liver, and urine were treated as follows. CHCl₃ (2 ml) and 10% NaHCO₃ were added to 0.4 ml of the blood. After centrifugation at 3000 rpm for 15 min, the CHCl₃ layer was evaporated to dryness and the residue was subjected to thin-layer chromatography with CHCl₃-MeOH (15: 1). In the case of the peritoneal cavity, after being washed with H₂O (5 ml), it was treated by the same method as the blood. In the case of the liver, H₂O (3 ml) was added and the mixture was homogenized. The procedure was the same as that for the blood. Urine was taken from the bladder and extracted with 3 ml of CHCl₃. The CHCl₃ layer was evaporated to dryness and the residue was tested by

thin-layer chromatography on silica gel with CHCl₃-MeOH (15:1). One group was treated in the same way without the administration of test compounds; this was used as a control group.

Separation of the Metabolites of 12n——The first group of Wistar strain rats, consisting of 7 rats, received 12n (10 mg/ml/head) suspended in 0.3% carboxymethyl cellulose (CMC) saline intraperitoneally 3 times at 3 hr intervals and urine and feces were collected at 17 hr after administration. The second group, consisting of 14 Wistar rats, received 12n (20 mg/ml/head) suspended in 0.3% CMC saline intraperitoneally at 4 hr intervals and urine and feces were collected at 17 hr after administration. The third group, consisting of 15 rats, received 12n (10 mg/ml/head) suspended in 0.3% CMC saline intraperitoneally at 4 hr intervals and urine and feces were collected at 17 hr after administration. The above urine and feces of groups 1—3 were combined and H_2O was added. They were adjusted to pH 8.0 with 10% NaHCO₃ and extracted with CHCl₃. The CHCl₃ layer was dried over MgSO₄ and the solvent was removed. The residue was chromatographed on silica gel. 19 was eluted with CHCl₃ and 20 and 21 (52 mg) were eluted with CHCl₃-MeOH (50: 1). The physical properties of these products were as follows: 19; mp $109-110^\circ$. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3340 (NH), 1660 (C=O). 20; mp $99-101^\circ$. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3345, 3420 (NH), 1665 (C=O). 21; mp $178-179^\circ$ (AcOEt). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1660 (C=O). NMR (CDCl₃) δ : 0.84 (3H, m, CH₃), 1.06—1.75 (8H, m, $4 \times \text{CH}_2$), 2.16 (3H, s, COCH₃), 3.16—3.56 (6H, m, piperazine ring 5 and 6 CH₂ and NCH₂-), 4.49 (2H, s, CH₂), 6.99 (2H, d, $-\frac{1}{2}$)

J=9 Hz, benzene ring $2\times CH$), 7.39 (2H, d, J=9 Hz, benzene ring $2\times CH$), 8.86 (1H, bs, NH).

1-(4-Aminobenzyl)-4-n-hexyl-2,3-dioxopiperazine (20)—1-n-Hexyl-4-(4-nitrobenzyl)-2,3-dioxopiperazine (4.5 g), obtained from 4-nitrobenzyl bromide and 1-n-hexyl-2,3-dioxopiperazine by the method described for 12b, was suspended in 50% $\rm H_2O-EtOH$ (150 ml) and Zn powder (22.5 g) was added. A solution of $\rm CaCl_2$ (4.5 g) in $\rm H_2O$ (5 ml) was added to the mixture and the whole was refluxed for 2 hr. The reaction mixture was filtered off and the filtrate was evaporated to dryness in vacuo. The residue was extracted with AcOEt (50 ml) and the extract was washed with $\rm H_2O$ and dried over MgSO₄. Removal of the solvent gave a solid which was recrystallized from iso-PrOH. Yield 2.7 g (65.9%). mp 99—101.5°. The IR spectrum of the synthesized product 20 was identical with that of one of the metabolites of 12n. Anal. Calcd for $\rm C_{17}H_{25}N_3O_2$: C, 67.30; H, 8.31; N, 13.85. Found: C, 67.40; H, 8.36; N, 13.68. NMR (CDCl₃) δ : 0.86 (3H, m, CH₃), 1.00—1.82 (8H, m, $\rm 4 \times CH_2$), 3.36 (6H, m, $\rm 3 \times CH_2$), 3.57—3.92 (2H, m, NH₂), 4.48 (2H, s, CH₂), 6.54 (2H, d, $\rm J=8$ Hz, benzene ring 2×CH).

1-(4-Ethylaminobenzyl)-4-n-hexyl-2,3-dioxopiperazine (19)—A suspension of 20 (2.7 g), EtI (1.1 ml), and Na₂CO₃ (1.4 g) in H₂O (30 ml) and EtOH (50 ml) was refluxed for 5 hr, then evaporated to dryness in vacuo. The residue was extracted with CHCl₃ (50 ml) and the extract was dried over MgSO₄. Removal of the solvent left a solid, which was chromatographed on silica gel with benzene-AcOEt (3:1), and then recrystallized from AcOEt-iso-Pr₂O to give colorless crystals of mp 109—110.5°. Yield 1.5 g (52%). The IR spectrum was identical with that of the metabolite 19. Anal. Calcd for $C_{19}H_{29}N_3O_2$: C, 68.85; H, 8.82; N, 12.68. Found: C, 68.82; H, 8.96; N, 12.62.

1-(4-Acetylaminobenzyl)-4-n-hexyl-2,3-dioxopiperazine (21)——Ac₂O (0.56 ml) was added to a solution of 20 (1.5 g) in MeOH (30 ml). The mixture was stirred for 3 hr at room temperature, then the solvent was evaporated off under reduced pressure and the residue was recrystallized from iso-PrOH-iso-Pr₂O to give 21 (1.0 g, 59%). mp 178—179°. The IR and NMR spectra of the product were identical with those of one of the metabolites of 12n. Anal. Calcd for $C_{19}H_{27}N_3O_3$: C, 66.06; H, 7.88; N, 12.16. Found: C, 66.20; H, 7.95; N, 12.03.

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