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An enantioselective synthesis of the $C_{24}-C_{40}$ fragment of (-)-pulvomycin†

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Sandra Börding and Thorsten Bach*

The C₂₄-C₄₀ fragment of (-)-pulvomycin was prepared in enantiomerically pure form using a concise synthesis method (15 linear steps from D-fucose, 6.8% overall yield) featuring a diastereoselective addition to an aldehyde, a \beta-selective glycosylation and a Stille crosscoupling as the key steps.

The antibiotic pulvomycin was first isolated in 1957 from a Streptomyces species but due to the limited analytical data no structure was assigned to the compound. In 1963, Akita et al. isolated a natural product from Streptomyces albosporeus var. labilomyceticus, which they called labilomycin and which was later shown to be identical to pulvomycin.³ Extensive analytical work by Smith et al. revealed the constitution of the natural product (Fig. 1) as well as the absolute and relative configuration at most stereogenic centers except for C₃₂ and C₃₃. The assignment was confirmed and the complete configuration was eventually proven by a crystal structure (1.4 Å resolution) of pulvomycin with the bacterial elongation factor Tu (EF-Tu).5 It is well established that pulvomycin is a potent inhibitor of EF-Tu and it therefore represents a promising lead compound for the development of new antibiotics.⁶

While synthetic reports on pulvomycin are scarce, the biosynthesis of the pulvomycin aglycone has been elucidated by labeling experiments.⁷ Our own interest in pulvomycin was triggered by our previous studies on the synthesis8 and antibiotic activity⁹ of thiazole peptides, such as the GE factors and the amythiamicins. It has been shown that the EF-Tu binding site of pulvomycin is in close proximity to the binding site of thiazole peptides. 10 The synthesis of pulvomycin and pulvomycin analogues might consequently help to further investigate the many facets of EF-Tu activity. 11 Apart from its biological activity, pulvomycin presents itself as a formidable synthetic challenge due to its complex and labile structure. In this communication

Lehrstuhl für Organische Chemie I, Technische Universität München, 85747 Garching, Germany. E-mail: thorsten.bach@ch.tum.de; Fax: +49 89 28913315: Tel: +49 89 28913330

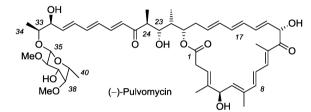


Fig. 1 Structure and compound numbering of (–)-pulvomycin.

TBDPSO cross-coupling

MeO 40 33

TBDPSO
$$glycosylation$$
 $glycosylation$

TBDPSO $glycosylation$
 $glycosylation$

Scheme 1 Retrosynthetic disconnection of the title compound 1 leading to D-fucose (2) as an appropriate carbohydrate substrate.

we disclose the enantioselective synthesis of a suitably protected C₂₄-C₄₀ fragment 1 (Scheme 1) of pulvomycin.

Retrosynthetically, it was envisioned that ketone 1 (TBDPS = tert-butyldiphenylsilyl) could be derived from commercially available p-fucose (2), which shows the correct configuration at the stereogenic centers (C₃₆-C₃₉) of the pyranose ring. In order to establish the desired β-configuration at the glycosidic center an appropriate neighbouring group, e.g. an acetate, was required (at carbon atom C₃₆)¹² and the methyl ether linkage was to be introduced after glycosylation. There was precedence for the differentiation of the two equatorial hydroxy groups at C₃₆ and C₃₇ of D-fucose. 13

Regarding the C24-C34 fragment, it seemed best to assemble the triene¹⁴ after the glycosylation step by an appropriate crosscoupling reaction, e.g. between C29 and C30. The stereogenic center at C_{33} appeared to be accessible from the chiral pool, e.g. from lactic acid, while the adjacent stereogenic center was to be introduced by a diastereoselective reaction.

The acetylation of D-fucose (2) (Scheme 2) proceeded quantitatively delivering the tetraacetate as an α/β -mixture ($\alpha/\beta = 95/5$)

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2
$$\xrightarrow{\text{(py)}}$$
 $\xrightarrow{\text{(CH}_2\text{Cl}_2)}$ $\xrightarrow{\text{89}\%}$ $\xrightarrow{\text{AcO}}$ $\xrightarrow{\text{OAc}}$ $\xrightarrow{\text{OAc}}$ $\xrightarrow{\text{SEt}}$ $\xrightarrow{\text{AcO}}$ $\xrightarrow{\text{OAc}}$ $\xrightarrow{\text{SEt}}$ $\xrightarrow{\text{AcO}}$ $\xrightarrow{\text{OAc}}$ $\xrightarrow{\text{SEt}}$ $\xrightarrow{\text{AcO}}$ $\xrightarrow{\text{OAc}}$ $\xrightarrow{\text{SEt}}$ $\xrightarrow{\text{AcO}}$ $\xrightarrow{\text{OAc}}$ $\xrightarrow{\text{SEt}}$ $\xrightarrow{\text{SET}}$ $\xrightarrow{\text{AcO}}$ $\xrightarrow{\text{OAc}}$ $\xrightarrow{\text{SET}}$ $\xrightarrow{\text$

Scheme 2 Synthesis of the protected glycosyl donor **5** from D-fucose (**2**). DMAP = 4-(N,N-dimethylamino)pyridine, py = pyridine, im = imidazole.

of anomers. 13 Conversion to the required thioacetal 3 proceeded best in our hands with ethanethiol and BF3. OEt2 in CH₂Cl₂, ¹⁵ which delivered depending on the reaction conditions and on the reaction scale variable amounts of separable α/β -isomers (see the ESI† for further details).

Since the relative configuration at the anomeric center was irrelevant for the desired glycosylation reaction, the α/β -mixture of 3 was taken into the four-step procedure previously described for the selective preparation of alcohol β -4¹³ and it furnished the desired product 4 as an α/β -mixture ($\alpha/\beta \cong 50/50$) in a total yield of 60% over six steps from p-fucose (2). Conversion of the equatorial alcohol 4 to silvl ether 5 required elevated temperature (60 °C) and a prolonged reaction time (3 d).

As mentioned above, it was planned to introduce the stereogenic center at C₃₂ by a diastereoselective reaction induced by the adjacent stereogenic center at the carbon atom C₃₃. Surprisingly, the reduction of a (S)-lactate-derived, para-methoxybenzyl (PMB) protected alkynyl ketone¹⁶ produced the desired alcohol 7 either in low yields or with insufficient diastereoselectivity (see the ESI† for further details). As an alternative approach, (S)-lactate-derived aldehyde 6^{17} was alkynylated with TMSacetylene under chelation control¹⁸ yielding alcohol 7 and its epimer epi-7 in 81% yield and in a diastereomeric ratio (d.r.) of 87/13 (Scheme 3). The diastereomerically pure product 7 was isolated in 65% yield.

Protection of the secondary alcohol proceeded smoothly at ambient temperature and the PMB group was cleaved oxidatively with 2,3-dicloro-5,6-dicyanobenzoguinone (DDQ)¹⁹ to deliver alcohol 8. The enantiomeric excess (ee) of alcohol 8 was established by chiral HPLC analysis and comparison with a racemic sample (see the ESI† for further details). Gratifyingly, the glycosylation reaction, when performed with N-iodosuccinimide (NIS) and trifluoromethanesulfonic acid (HOTf) as activating agents, ²⁰ delivered a single diastereomerically pure product 9, which was shown to have the desired β-configuration.²¹ Reductive removal of the acetyl groups with diisobutylaluminium hydride (DIBAL-H)²² produced 1,3-diol 10, which was converted into the respective dimethylether 11 upon treatment with an excess (10 equiv.) of Meerwein salt and proton sponge [1,8-bis(dimethylamino)naphthalene].²³ Less electrophilic methylating reagents (MeOTf, MeI) in combination with appropriate bases failed to react or led to substrate decomposition. Selective desilylation of the alkyne was achieved with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU).²⁴

With alkyne 12 in hand, various approaches to potential cross-coupling substrates were pursued. It was found that the

Scheme 3 Assembly of the $C_{28}-C_{40}$ fragment \emph{via} glycosylation of enantiomerially pure (≥98% ee) alcohol 8 with glycosyl donor 5.

Pd-catalyzed hydrostannylation with Bu₃SnH²⁵ can be successfully performed with alkyne 12 delivering stannane 13 in 44% yield (Scheme 4). Iodide 14 was obtained from stannane 13 upon treatment with iodine in dichloromethane (85% yield).²⁶ The alkyne hept-3-en-1-yne-5-ol²⁷ seemed to be the most suitable precursor for iodide 15 and stannane 16. The compound was available from bis-1,4-(trimethylsilyl)buta-1,3-diyne in four steps and an overall yield of 53% (see the ESI† for further details). Stannylation of hept-3-en-1-vne-5-ol with Bu₃SnH was readily achieved employing the Cu-based protocol of Betzer et al. 28 to deliver stannane 16 in 79% yield. As for 14, iodide 15 was generated by iodo-de-stannylation employing iodine in dichloromethane (79% yield).

While attempted Stille cross-coupling reactions²⁹ of stannane 13 and iodide 15 failed, the desired C-C bond formation proceeded smoothly, when performed with the carbohydrate building block as the electrophile. Iodide 14 and stannane 16 underwent a clean cross-coupling employing Pd(MeCN)₂Cl₂

Scheme 4 Stille cross-coupling of building blocks 13 and 15 as key step for the assembly of the title compound.

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(10 mol%) as the catalyst.³⁰ Alcohol **17** was obtained in 87% yield and was immediately further oxidized to the desired ketone by treatment with an excess (30 equiv.) of MnO₂. Despite a pronounced long wavelength absorption ($\lambda_{\rm max}=308$ nm, $\varepsilon=28\,035$ M⁻¹ cm⁻¹ in MeCN), trienone **1** appears to be more stable than alcohol **17** ($\lambda_{\rm max}=271$ nm, $\varepsilon=39\,350$ M⁻¹ cm⁻¹ in MeCN; shoulder at $\lambda_{\rm max}=282$ nm, $\varepsilon=31\,180$ M⁻¹ cm⁻¹) and could be stored for one week at -25 °C in the dark.

In summary, the enantiomerically pure western fragment 1 of (–)-pulvomycin was synthesized in 15 linear steps. The fragment comprises the carbohydrate part (labilose, C_{35} – C_{40}) of the natural product and one of its three triene components (C_{24} – C_{34}). Should an aldol-type reaction of fragment 1 with a suitable Eastern fragment not be successful, stannane 13 and iodide 14 offer suitable options to connect the protected glycoside fragment to the rest of the molecule.

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Notes and references

- 1 M. Zief, R. Woodside and H. Schmitz, *Antibiot. Chemother.*, 1957, 7, 384–386.
- 2 (a) E. Akita, K. Maeda and H. Umezawa, J. Antibiot., Ser. A, 1963, 16, 147–151; (b) E. Akita, K. Maeda and H. Umezawa, J. Antibiot., Ser. A, 1964, 17, 200–217.
- 3 J. L. Schwartz, M. Tishler, B. H. Arison, H. M. Shafer and S. Omura, J. Antibiot., 1976, 29, 236–241.
- 4 R. J. Smith, D. H. Williams, J. C. J. Barna, I. R. McDermott, K. Haegele, F. Piriou, J. Wagner and W. Higgins, *J. Am. Chem. Soc.*, 1985, 107, 2849–2857.
- 5 A. Parmeggiani, I. M. Krab, S. Okamura, R. C. Nielsen, J. Nyborg and P. Nissen, *Biochemistry*, 2006, 45, 6846–6857.
- 6 For a recent review, see: K. M. G. O'Connell, J. T. Hodgkinson, H. F. Sore, M. Welch, G. P. C. Salmond and D. R. Spring, Angew. Chem., Int. Ed., 2013, 52, 10706–10733.
- 7 N. D. Priestley and S. Gröger, J. Org. Chem., 1995, 60, 4951-4953.
- 8 (a) O. Delgado, H. M. Müller and T. Bach, Chem. Eur. J., 2008, 14, 2322–2339; (b) C. Ammer and T. Bach, Chem. Eur. J., 2010, 16, 14083–14093.
- 9 S. Gross, F. Nguyen, M. Bierschenk, D. Sohmen, T. Menzel, I. Antes, D. N. Wilson and T. Bach, *ChemMedChem*, 2013, 8, 1954–1962.
- 10 A. Parmeggiani, I. M. Krab, S. Okamura, R. C. Nielsen, J. Nyborg and P. Nissen, *Biochemistry*, 2006, 45, 6846–6857.
- 11 Reviews: (a) R. Berisio, A. Ruggiero and L. Vitagliano, Isr. J. Chem., 2010, 50, 71–79; (b) A. Parmeggiani and P. Nissen, FEBS Lett., 2006, 580, 4576–4581.
- 12 (a) R. U. Lemieux and J.-I. Hayami, Can. J. Chem., 1965, 43, 2162–2173; (b) G.-J. Boons, Contemp. Org. Synth., 1996, 3, 173–200; (c) T. K. Lindhorst, Essentials of Carbohydrate Chemistry and Biochemistry, Wiley-VCH, Weinheim, 3rd edn, 2007, pp. 157–208.

- 13 D. Comegna, E. Bedini and M. Parrilli, *Tetrahedron*, 2008, **64**, 3381–3391.
- 14 For selected recent total syntheses of naturally occurring conjugated (E,E,E)-trienes, see: (a) D. J. Del Valle and M. J. Krische, J. Am. Chem. Soc., 2013, 135, 10986–10989; (b) C. Jahns, T. Hoffmann, S. Müller, K. Gerth, P. Washausen, G. Höfle, H. Reichenbach, M. Kalesse and R. Müller, Angew. Chem., Int. Ed., 2012, 51, 5239–5243; (c) M. Yoshino, K. Eto, K. Takahashi, J. Ishihara and S. Hatakeyama, Org. Biomol. Chem., 2012, 10, 8164–8174; (d) P. G. E. Craven and R. J. K. Taylor, Tetrahedron Lett., 2012, 53, 5422–5425; (e) H. J. Jessen, A. Schumacher, F. Schmid, A. Pfaltz and K. Gademann, Org. Lett., 2011, 13, 4368–4370; (f) D. Amans, V. Bellosta and J. Cossy, Chem. Eur. J., 2009, 15, 3457–3473; (g) M. T. Crimmins, H. S. Christie, A. Long and K. Chaudhary, Org. Lett., 2009, 11, 831–834; (h) I. S. Mitchell, G. Pattenden and J. Stonehouse, Org. Biomol. Chem., 2005, 3, 4412–4431.
- 15 P. Sjölin, S. K. George, K.-E. Bergquist, S. Roy, A. Svensson and J. Kihlberg, J. Chem. Soc., Perkin Trans. 1, 1999, 1731–1742.
- 16 The ketone was prepared from literature known Weinreb amide (V. Convertino, P. Manini, W. B. Schweizer and F. Diederich, Org. Biomol. Chem., 2006, 4, 1206–1208) by substitution with the respective magnesium acetylide (see the ESI† for further details).
- 17 W. Yu, Y. Zhang and Z. Jin, Org. Lett., 2001, 3, 1447-1450.
- 18 K. T. Mead, Tetrahedron Lett., 1987, 28, 1019-1022.
- 19 Y. Oikawa, T. Yoshioka and O. Yonemitsu, Tetrahedron Lett., 1982, 23, 885–888.
- 20 (a) G. H. Veeneman, S. H. van Leeuwen and J. H. van Boom, Tetrahedron Lett., 1990, 31, 1331-1334; (b) P. Konradsson, U. E. Udodong and B. Fraser-Reid, Tetrahedron Lett., 1990, 31, 4313-4316.
- 21 ¹H-NMR data of glycoside 9 (500 MHz, CDCl₃): δ (ppm) = 0.09 [s, 9H; Si(CH₃)₃], 0.97 [s, 9H; C(CH₃)₃], 0.96–0.99 (m, 3H; H₃-40), 1.05 [s, 9H; C(CH₃)₃], 1.30 (d, ${}^{3}J$ = 6.3 Hz, 3H; H₃-34), 1.46 (s, 3H; C(O)CH₃-36), 2.12 [s, 3H; C(O)CH₃-38], 3.05 (q, ${}^{3}J$ = 6.4 Hz, 1H; H-39), 3.41 (qd, ${}^{3}J$ = 6.3, 3.9 Hz, 1 H; H-33), 3.44 (d, ${}^{3}J$ = 8.1 Hz, 1 H; H^β-35), 3.60 (dd, ${}^{3}J$ = 9.8, 3.3 Hz, 1H; H-37), 4.19 (d, ${}^{3}J$ = 3.9 Hz, 1H; H-32), 4.79 (d, ${}^{3}J$ = 3.3 Hz, 1H; H-38), 5.06 (dd, ${}^{3}J$ = 9.8, 8.1 Hz, 1H; H-36), 7.23–7.27 (m, 4H; H_{arom}), 7.30–7.48 (m, 10H; H_{arom}), 7.55–7.58 (m, 2H; H_{arom}), 7.60–7.65 (m, 4H; H_{arom}), 7.69–7.73 (m, 2H; H_{arom}).
- 22 F. E. McDonald and M. Wu, Org. Lett., 2002, 4, 3979-3981.
- 23 B. Wang, T. M. Hansen, T. Wang, D. Wu, L. Weyer, L. Ying, M. M. Engler, M. Sanville, C. Leitheiser, M. Christmann, Y. Lu, J. Chen, N. Zunker, R. D. Cink, F. Ahmed, C.-S. Lee and C. J. Forsyth, *J. Am. Chem. Soc.*, 2010, 133, 1484–1505.
- 24 C.-E. Yeom, M. J. Kim, W. Choi and B. M. Kim, Synlett, 2008, 565–568.
- 25 (a) H. X. Zhang, F. Guibe and G. Balavoine, J. Org. Chem., 1990, 55, 1857–1867; (b) J. R. Frost, C. M. Pearson, T. N. Snaddon, R. A. Booth and S. V. Ley, Angew. Chem., Int. Ed., 2012, 51, 9366–9371.
- 26 R. Alvarez, M. Herrero, S. López and A. R. de Lera, *Tetrahedron*, 1998, 54, 6793–6810.
- 27 K. Green, J. W. Keeping and V. Thaller, J. Chem. Res., Synop., 1985, 103; K. Green, J. W. Keeping and V. Thaller, J. Chem. Res., Miniprint, 1985, 1260–1267.
- 28 J.-F. Betzer, F. Delaloge, B. Muller, A. Pancrazi and J. Prunet, J. Org. Chem., 1997, 62, 7768–7780.
- Reviews: (a) J. K. Stille, Angew. Chem., Int. Ed., 1986, 25, 508-524;
 (b) T. N. Mitchell, Synthesis, 1992, 803-815;
 (c) V. Farina, V. Krishnamurthy and W. J. Scott, Org. React., 1997, 50, 1-652.
- 30 J. K. Stille and B. L. Groh, J. Am. Chem. Soc., 1987, 109, 813-817.