TERPENE AND LIGNAN GLYCOSIDES FROM PLUCHEA INDICA

TAKETO UCHIYAMA, TOSHIO MIYASE, AKIRA UENO and KHAN USMANGHANI*

School of Pharmaceutical Sciences, University of Shizuoka, 395, Yada, Shizuoka 422, Japan; *Department of Pharmacognosy, Faculty of Pharmacy, University of Karachi, Karachi 32, Pakistan

(Received 14 May 1990)

Key Word Index-Pluchea indica; Compositae; monoterpene glycoside; sesquiterpene glycoside; lignan glycoside.

Abstract—Investigation of the roots of *Pluchea indica* afforded a new monoterpene glycoside, plucheoside C, three new eudesmane-type sesquiterpenes, plucheols A, B, plucheoside E and three new lignan glycosides, plucheosides D_1 , D_2 , D_3 together with a known eudesmane-type sesquiterpene. The structures of new compounds were elucidated on the basis of chemical and spectral data.

INTRODUCTION

In connection with a study on the terpenic glycosides in some plants of the Compositae, we have investigated *Pluchea indica*. In a previous paper [1], we reported the isolation and structural elucidation of the constituents which were isolated from its aerial parts. Now, we report the structures of four new terpenoids and three new lignan glycosides which were isolated from its roots.

RESULTS AND DISCUSSION

The careful separation of the polar fraction of P. *indica* afforded eight polar compounds. The known compound was identified as pterocarptriol (3) [2] by comparison of spectral data with those reported for this compound.

Plucheoside C (1), amorphous powder, $[\alpha]_D - 72.7^\circ$ showed $[M + Na]^+$ at m/z 467 in the positive FAB mass spectrum. The ¹H NMR spectrum indicated the presence of an aryl methyl group $[\delta 2.32 \ (br s)]$, an aryl isopropyl group $[\delta 1.24; 1.27 \ (each 3H, d, J = 7 Hz); 3.71 \ (1H, m)]$ and two anomeric protons $[\delta 5.46 \ (1H, d, J = 8 Hz); 5.70 \ (1H, d, J = 3 Hz)]$. The ¹³C NMR spectrum showed that 1 was a glycoside of thymol 2 having a glucose and an apiose as the sugar moiety. The C-6 of glucose was shifted downfield by *ca* 6 ppm, suggesting that an apiose was attached to the C-6 of glucose [3].

Plucheol A (4), amorphous powder, $[\alpha]_D + 14.2^\circ$ showed a molecular ion at m/z 254 in the EI mass spectrum. The ¹H NMR spectrum indicated the presence of three methyl groups $[\delta 0.86 \ (3H, s); 1.39; 1.40 \ (each 3H, s)]$, two carbinyl protons $[\delta 4.11 \ (1H, dt, J = 9, 5 \text{ Hz}); 4.34 \ (1H, d, J = 9 \text{ Hz})]$ and exomethylene protons $[\delta 4.97; 5.87 \ (each 1H, br s)]$. These data were identical with those of an aglycone of atractyloside G [4], but this was the first isolation from plants.

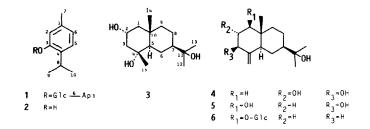
Plucheol B (5), amorphous powder, $[\alpha]_D + 43.9^{\circ}$ showed a molecular ion at m/z 254 in the EI mass spectrum. The ¹H NMR spectrum was similar to that of 4 except for two carbinyl protons. One hydroxyl group was assigned as 3β -equatorial because of a down-field shift of an olefinic proton [δ 5.80 (1H, br s)], as found in the spectrum of compound 4, and the large coupling constants of the carbinyl proton (dd, J = 11, 5 Hz). The other hydroxyl group was assigned as 1 β -equatorial because of an upfield shift ($\Delta - 2.9$ ppm) of the C-9 compared with that of 4 and from the splitting pattern of the carbinyl proton (dd, J = 12, 4 Hz).

Plucheoside E (6), amorphous powder, $[\alpha]_D - 2.8^{\circ}$ showed $[M + Na]^+$ at m/z 423 and $[M + H]^+$ at m/z 401 in the positive FAB mass spectrum. The ¹H NMR spectrum indicated the presence of three methyl groups $[\delta 0.89; 1.34; 1.35$ (each 3H, s)], a methyne proton $[\delta 3.78$ (1H, dd, J = 11, 4 Hz)], exomethylene protons $[\delta 4.62; 4.79]$ (each 1H, br s)] and an anomeric proton $[\delta 4.92$ (1H, d, J = 8 Hz)]. Acid hydrolysis gave glucose as the sugar moiety and in the ¹³C NMR spectrum, 21 carbon signals, including six carbon signals due to a glucopyranosyl moiety, were observed. The C-9 was shifted upfield as in compound 5 and compared with that of 4. Thus the glucosyl residue was decided to be 1β .

Plucheoside D_1 (7), amorphous powder, $[\alpha]_D - 116.3^\circ$ showed $[M + Na]^+$ at m/z 541 and $[M + H]^+$ at m/z 519 in the positive FAB mass spectrum. The UV spectrum showed the absorption maximum at 342 nm. The ¹H NMR spectrum indicated the presence of two methoxyl groups [δ 3.68; 3.84 (each 3H, s)], an anomeric proton [$\delta 4.92$ (1H, d, J = 8 Hz)], an α,β -unsaturated aldehyde group [δ 6.81 (1H, dd, J = 16, 8 Hz); 7.30 (1H, d, J = 16 Hz); 9.76 (1H, d, J = 8 Hz)] and a methyne proton $[\delta 5.99 (1H, d, J=6 Hz)]$. These data suggest that this compound is a dihydrobenzofuran-type lignan glycoside [5]. Acid hydrolysis gave glucose. In the ¹³C NMR spectrum, the C-9 was shifted at δ 71.2 while an anomeric carbon was at δ 104.8, and thus the glucosyl linkage was at C-7. The CD spectrum of 7a, obtained by reduction with LiAlH₄, showed negative Cotton effects $[\theta]_{267}$ - 5900 and $[\theta]_{287}$ - 6600, suggesting a 7S,8R configuration [6]

The ¹H NMR spectrum of plucheoside D_2 (8) was very similar to that of 7, except for the presence of two equivalent methoxyl groups $[\delta 3.73 \ (6H, s)]$ and two equivalent aromatic protons $[\delta 7.39 \ (2H, s)]$.

Plucheoside D₃ (9), amorphous powder, $[\alpha]_D - 56.8^{\circ}$ showed $[M + Na]^+$ at m/z 557 and $[M + H]^+$ at m/z 534 in the positive FAB mass spectrum. Its UV spectrum



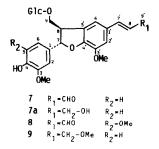


Table 1. ¹³C NMR spectral data of compounds 1, 2, 4-9

С	1†	1*	2*	4†	5†	6†	7†	8†	9†
Aglycone	moiety								
1 (1')	136.8	137.5	‡	49.8	77.2	85.3	132.5 (128.7)	‡ (128.7)	132.7 (132.7)
2 (2')	116.6	117.6	117.4	73.5	42.9	28.7	111.1 (113.6)	105.0 (113.6)	111.1 (112.0)
3 (3')	154.8	156.1	156.0	79.6	70.9	34.6	148.9 (145.3)	149.3 (145.1)	149.0 (145.0)
4 (4')	132.9	136.6	136.8	152.3	154.5	150.1	‡ (152.1)	131.4 (152.0)	148.2 (152.1)
5 (5')	126.7	126.7	126.1	48.5	45.9	48.5	116.6 (130.6)	149.3 (130.5)	116.6 (133.3)
6 (6')	121.4	124.3	‡	22.6	23.1	22.8	119.8 (119.8)	105.0 (119.6)	119.7 (116.5)
7 (7')	21.0	21.4	21.3	49.3	50.2	49.6	89.5 (153.8)	89.7 (153.3)	88.7 (131.4)
8 (8')	27.5	27.2	26.7	25.6	25.4	25.1	51.6 (126.8)	51.6 (126.8)	52.2 (129.9)
9 (9')	23.1	23.5	23.2	41.3	38.4	37.9	71.2 (193.5)	71.1 (193.3)	71.8 (73.4)
10	23.1	23.6	23.4	35.5	41.1	40.3	. ,		. ,
11				71.5	71.7	72.2			
12				27.7	27.8	27.6			
13				28.1	28.3	28.2			
14				17.8	11.3	11.6			
15				104.8	103.8	106.7			
OMe							56.0, 56.3	56.3, 56.5, 56.5	56.0, 56.4, 57.7
Sugar mo	oiety								
Glc-1	103.2	103.4				102.4	104.8	104.7	105.0
2	75.0	75.0				75.4	75.1	75.0	75.1
3	78.3	78.8				78.7	78.7	78.6	78.7
4	71.7	71.6				71.6	71.7	71.7	71.8
5	76.9	77.2				78.2	78.7	78.6	78.7
6	68.7	68.2				63.3	62.9	62.8	62.9
Api-1	110.9	111.0							
2	78.2	78.0							
3	80.5	80.5							
4	75.1	75.0							
5	65.9	65.8							

Run at 22.5 MHz in * methanol- d_4 and † pyridine- d_5 solution. ‡ Overlapped with solvent.

showed the absorption maximum at 278 nm. The ¹H and ¹³C NMR spectra indicated the presence of an aliphatic methoxyl group [δ 3.34 (3H, s); 57.7], two aromatic methoxyl groups [δ 3.68; 3.86 (each 3H, s), 56.0; 56.4] and two *trans*-olefinic protons [δ 6.28 (1H, dt, J = 16, 6 Hz); 6.68 (1H, d, J = 16 Hz)]. The CD spectrum showed negative Cotton effects [θ]₂₆₈ - 7800 and [θ]₂₈₈ - 10 100 and suggested that compound 9 has the 75,8*R*-configuration.

EXPERIMENTAL

¹H and ¹³C NMR spectra were recorded at 89.55 and 22.5 MHz, respectively, and also at 399.65 MHz. TMS was used as int. standard.

Plant material. Pluchea indica (L.) Less. was collected in Drigh Road, Karachi, Pakistan in 1988. Plants were identified by Prof. Dr S. I. Ali and Prof. Dr Muhammad Qaiser (Department of Botnay, University of Karachi).

Extraction and isolation. Dried roots (4.5 kg) were extracted $\times 2$ with MeOH under reflux. The extract was concd under red. pres. and the residue suspended in H₂O. This suspension was extracted with Et₂O. The H₂O layer was passed through an Amberlite XAD-2 column and the MeOH eluate concd under red. pres. The residue (43 g) was chromatographed on a silica gel column with CHCl₃-MeOH (19:1-4:1) and semi-prep. HPLC [ODS, H₂O-MeCN (23:2-7:3)] to give 15 mg 1, 4 mg 3, 2 mg 4, 10 mg 5, 21 mg 6, 14 mg 7, 24 mg 8, 9 mg 9 and 9 mg 10.

Plucheoside C (1). Amorphous powder, $[\alpha]_{b}^{22} - 72.7^{\circ}$ (MeOH; c 1.50). FABMS m/z 467 $[M + Na]^+$. ¹H NMR pyridine $d_5:\delta$ 1.24; 1.27 (each 3H, d, J = 7 Hz, H_3-9/H_3-10), 2.32 (3H, br s, H_3-7), 3. 71 (1H, m, H-8), 4.18 (2H, s, H_2-5 of apiose), 4.34; 4.57 (each 1H, d, J = 10 Hz, H-4 of apiose), 4.75 (1H, d, J = 3 Hz, H-2 of apiose), 5.46 (1H, d, J = 8 Hz, H-1 of Glc), 5.70 (1H, d, J = 3 Hz, H-1 of Api). ¹H NMR (CD₃OD): δ 1.16 (6H, d, J = 7 Hz, H_3-9 ; H_3-10), 2.30 (3H, br s, H_3-7), 4.96 (1H, d, J = 3 Hz, H-1 of Api), 6.78 (1H, br d, J = 8 Hz, H-6), 6.95 (1H, br s, H-2), 7.08 (1H, d, J = 8 Hz, H-5). ¹³C NMR: Table 1.

Plucheol A (4). Amorphous powder, $[\alpha]_D^{21} + 14.2^{\circ}$ (MeOH; c 0.67). EIMS m/z (rel. int.): 254 [M]⁺ (5), 236 [M - H₂O]⁺ (30), 218 [M - 2H₂O]⁺ (50), 175 (100). ¹H NMR(pyridine-d₅): δ 0.86 (3H, s, H₃-14), 1.39; 1.40 (each 3H, s, H₃-12/H₃-13), 4.11 (1H, dt, J = 9, 5 Hz, H-2), 4.34 (1H, d, J = 9 Hz, H-3), 4.97; 5.87 (each 1H, br s, H-15). ¹³C NMR: Table 1.

Plucheol B (5). Amorphous powder, $[\alpha]_D^{21} + 43.9^{\circ}$ (MeOH; c 1.71). EIMS m/z (rel. int.): 254 [M]⁺ (6), 236 [M-H₂O]⁺ (7), 218 [M-2H₂O]⁺ (13), 160 (100). ¹H NMR (pyridine-d₅): δ 1.08 (3H, s, H₃-14), 1.43 (6H, s, H₃-12; H₃-13), 3.79 (1H, dd, J = 12, 4 Hz, H-1), 4.50 (1H, dd, J = 11, 5 Hz, H-3), 4.99; 5.80 (each 1H, br s, H-15). ¹³C NMR: Table 1.

Plucheoside E (6). Amorphous powder, $[\alpha]_D^{21} - 2.8^{\circ}$ (MeOH; c 1.09). FABMS m/z: 423 [M + Na]⁺, 401 [M + 1]⁺. ¹H NMR (pyridine-d₅): δ 0.89 (3H, s, H₃-14), 1.34; 1.35 (each 3H, s, H₃-12/H₃-13), 3.78 (1H, dd, J = 11, 4 Hz, H-1), 4.62; 4.79 (each 1H, br s, H-15), 4.92 (1H, d, J = 8 Hz, H-1 of Glc). ¹³C NMR : Table 1.

Plucheoside D_1 (7). Amorphous powder, $[\alpha]_{D}^{21}-116.4^{\circ}$ (MeOH; c0.64). FABMS m/z 541 $[M+Na]^+$, 519 $[M+1]^+$. UV λ_{max}^{MeOH} nm (log ε): 342 (4.45). CD (MeOH; c0.06) $[\theta]$ (nm): +28 500 (236), -15 500 (255), +3100 (279), -16 600 (338). ¹H NMR (pyridine- d_5): δ 3.68; 3.84 (each 3H, s, OMe), 4.92 (1H, d, J = 8 Hz, H-1 of glucose), 5.99 (1H, d, J = 6 Hz, H-7), 6.81 (1H, dd, J = 16, 8 Hz, H-8'), 7.30 (1H, d, J = 16 Hz, H-7'), 9.76 (1H, d, J = 8 Hz, H-9'). ¹³C NMR: Table 1. Plucheoside D_2 (8). Amorphous powder, $[\alpha]_D^{21} - 67.5^{\circ}$ (MeOH; c 0.40). FABMS m/z 549 $[M + 1]^+$. UV λ_{MeOH}^{MeOH} nm (log ε): 341 (4.33). CD (MeOH; c 0.06) $[\partial]$ (nm): + 23 300 (240), -9600 (257), -4600 (281), -13 700 (337). ¹H NMR (pyridine- d_5): δ 3.73 (6H, s, 2-OMe), 3.82 (3H, s, OMe), 4.96 (1H, d, J = 8 Hz, H-1 of Glc), 6.01 (1H, d, J = 6 Hz, H-7), 6.80 (1H, dd, J = 16, 8 Hz, H-8'), 7.15 (1H, d, J = 16 Hz, H-7'), 7.39 (2H, s, H-2; H-6), 9.75 (1H, d, J = 8 Hz, H-9'). ¹³C NMR: Table 1.

Plucheoside D_3 (10). Amorphous powder, $[\alpha]_D^{21} - 56.8^{\circ}$ (MeOH; c 0.22). FABMS m/z 557 $[M + Na]^+$, 534 $[M + H]^+$. UV λ_{mex}^{MeOH} nm (log ε): 278 (4.36). CD (MeOH; c 0.06) $[\theta]$ (nm): -7800 (268), -10100 (288). ¹H NMR (C₅D₅N): δ 3.34 (3H, s, OMe at C-9'), 3.68; 3.86 (each 3H, s, OMe), 4.96 (1H, d, J = 8 Hz, H-1 of Glc), 6.00 (1H, d, J = 6 Hz, H-7), 6.28 (1H, dt, J = 16, 6 Hz, H-8'), 6.68 (1H, d, J = 16 Hz, H-7'). ¹³C NMR: Table 1.

LiAlH₄ reduction of 7. Compound 7 (2 mg) was dissolved in THF (1 ml) and reduced with LiAlH₄ (3 mg) at -20° for 1 hr. The reaction mixt. was acidified with dil. HCl and passed through a Diaion HP-20 column. The adsorbed material was eluted with MeOH after washing with H₂O. The MeOH eluate was purified by HPLC [Develosil Ph-7; H₂O-MeOH (3:2)] to give 7a (1 mg) as amorphous powder. ¹H NMR (pyridine-d₅): δ 3.68; 3.85 (each 3H, s, OMe), 4.98 (1H, d, J = 8 Hz, H-1 of Glc), 6.01 (1H, d, J = 6 Hz, H-7), 6.57 (1H, dt, J = 16, 5 Hz, H-8'), 6.87 (1H, d, J = 16 Hz, H-7'), 7.14 (1H, br s, H-2'), 7.18 (1H, d, J = 8 Hz, H-5), 7.20 (1H, br s, H-6'), 7.25 (1H, dd, J = 8, 2 Hz, H-6), 7.36 (1H, d, J = 2 Hz, H-2). CD (MeOH; c 0.06 [θ] (nm): -5900 (267), -6600 (287).

Acid hydrolysis of glycosides 6–9. A soln of each glycoside (ca 0.1 mg) in 5% H₂SO₄ (2 drops) was heated at 100° for 30 min. The soln was passed through an Amberlite IR-45 column and concd to give a residue which was reduced with NaBH₄ (*ca* 1 mg) for 1 hr at room temp. The reaction mixt. was passed through an Amberlite IR-120 column and concd to dryness. Boric acid was removed by dist. with MeOH and the residue acetylated with Ac₂O (1 drop) and pyridine (1 drop) at 100° for 1 hr. The reagents were evapd *in vacuo*. Glucitol acetate was detected by GC from all glycosides. GC conditions: capillary column SP-2380, 0.25 mm × 30 m; column temp. 250°; carrier gas N₂; R_t 11.5 min.

Acknowledgement—We thank Dr M. Uchida, the Central Analytical Laboratory of this school for measurement of mass spectra.

REFERENCES

- 1. Uchiyama, T., Miyase, T., Ueno, A. and Usmanghani, K. (1989) Phytochemistry 28, 3369.
- Nanayakkara, N. P., Kinghorn, A. D. and Farnsworth, N. R. (1986) J. Chem. Res. (S), 454.
- Miyase, T., Ueno, A., Takizawa, N., Kobayashi, H. and Oguchi, H. (1987) Chem. Pharm. Bull. 35, 3713.
- Yahara, S., Higashi, T., Iwaki, K., Nohara, T., Marubayashi, N., Ueda, I., Kohda, H., Goto, K., Izumi, H., Nuno, M., Katsuki, S., Isoda, S. and Satake, M. (1989) *Chem. Pharm. Bull.* 37, 2995.
- Miyase, T., Ueno, A., Takizawa, N., Kobayashi, H. and Oguchi, H. (1989) Phytochemistry 28, 3483.
- Binns, A. N., Chen, R. H., Wood, H. N. and Lynn, D. G. (1987) Proc. Natl Acad. Sci. U.S.A. 84, 980.