



Transition metal-carbonyl labeling reagent containing iodoacetamido function: $\text{CpFe}(\text{CO})_2[\eta^1\text{-}N(1)\text{-}4\text{-iodoacetamidophthalimidato}]$ - synthesis and reaction with the phenytoin and ethosuximide anions

Konrad Kowalski and Janusz Zakrzewski*

Department of Organic Chemistry, University of Łódź, 90-136 Łódź, Narutowicza 68, Poland

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Abstract—The first transition metal-carbonyl complex containing iodoacetamido function, $\text{CpFe}(\text{CO})_2[\eta^1\text{-}N(1)\text{-}4\text{-iodoacetamidophthalimidato}]$ was synthesized and used for labeling of two drugs: phenytoin and ethosuximide. © 1998 Published by Elsevier Science Ltd. All rights reserved

Keywords: metal-carbonyl; iodoacetamido; phenytoin anion; ethosuximide anion

Transition metal carbonyl complexes having substituents able to react selectively with functional groups present in molecules of biological and pharmacological interest, currently attract considerable interest as IR-detectable labeling reagents [1]. The detection of the bioconjugates is feasible in a pico- or subpicomole scale by virtue of very strong absorption bands corresponding to the $\nu(\text{CO})$ modes, appearing at $2100\text{--}1850\text{ cm}^{-1}$ i.e. in the region which is generally free from other interfering absorptions, even in aqueous solutions and biological systems [2,3]. This approach was successfully applied for the labeling of haptens in immunoassays (CMIA-carbonylmetalloimmunoassay) [2], for labeling of steroid hormones for the study of hormone-receptor interaction [4], and labeling of oligonucleotides [5,6]. Most of the labeling procedures described up to now is based on organometallic complexes containing active *N*-hydroxy-succinimide ester function which reacts with the amino group present in the biomolecule and forming the amido bond. Another approach uses organometallic complexes having pyrylium system [7], or iminoester [8], as a reactive function (target function- NH_2).

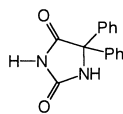
We have recently reported the synthesis of

$\text{CpFe}(\text{CO})_2(\eta^1\text{-}N\text{-maleimidato})$, which contains a maleimide system reactive towards thiol groups and can be used for labeling of e.g. cysteine-containing peptides or proteins [9]. However, we have demonstrated that it can also label histidine and (in basic medium) lysine residues [10]. Another approach developed recently in our laboratory in collaboration with the Prof. Jaouen's group is the use of the $\text{CpFe}(\text{CO})_2$ - complexes of phthalimide containing in the 3- or 4-position isothiocyanato functions reactive towards amino groups [11].

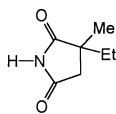
In this communication we report the synthesis of the first metallocarbonyl marker containing the $\text{CpFe}(\text{CO})_2(\eta^1\text{-}N\text{-phthalimidato})$ system and iodoacetamido group as the reactive function. This function is believed to be specific for thiols (cysteines), although it can also react with methionines (MeS-), histidines (imidazole) and lysines (NH_2) [12,13]. However, prompted by recent interest in the development of organometallic tracers for immunoassays of anti-epileptic and sedative drugs [2,14,15], we decided to study the reactivity of our reagent toward anions of two important representatives of this group of drugs: phenytoin (5,5-diphenylhydantoin) and ethosuximide (2-methyl-2-ethylsuccinimide).

We synthesized the bioconjugates of these products which are of interest as potential markers in carbonylmetalloimmunoassays (CMIA).

* Author to whom correspondence should be addressed.



phenytoin



ethosuximide

Experimental

All reactions were carried out under argon. Dichloromethane was distilled from CaH_2 . $\text{CpFe}(\text{CO})_2[\eta^1\text{-}N(1)\text{-}4\text{-aminophthalimidato}]$ (**1**) was prepared according to the earlier published procedure [11]. All reagents and other solvents (reagent grade) were used as received from Aldrich or Fluka. Column chromatographies were carried out using Kieselgel 60 (230–400 mesh ASTM) purchased by Merck. Instrumentation: IR-Specord 75 IR; ^1H NMR: Varian Gemini 200BB (200 MHz); FAB MS: Finnigan MAT 95. IR spectra were recorded in CHCl_3 , ^1H -NMR in CDCl_3 and referenced to internal TMS, FAB spectra in *m*-nitrobenzyl alcohol matrices.

$\text{CpFe}(\text{CO})_2(\eta^1\text{-}N(1)\text{-}4\text{-iodoacetamidophthalimidato})$ (**2**)

To a solution of **1** (310 mg, 0.92 mmol) in CH_2Cl_2 (5 ml) DCC (206 mg, 1.0 mmol) and iodoacetic acid (186 mg, 1.0 mmol) were added. After 1 h stirring at r.t. the solid formed was filtered off and the filtrate evaporated to dryness. Column chromatography (eluent CHCl_3) and crystallization (CH_2Cl_2 -ether) gave **2** as yellow solid. Yield 386 mg (83%). IR (cm^{-1}): 2050 and 1995 cm^{-1} (Fe–CO), 1690 cm^{-1} (CO–amide), 1655 (CO–phthalimide), 1535 (amide-II). ^1H NMR (CHCl_3): 8.10, b, 1H, NH; 7.76, d ($J = 8.0$ Hz), 1H and 7.58, d ($J = 8.0$ Hz), 1H, H-5 and H-6; 7.69, s, 1H, H-3; 5.12, s, 5H, Cp; 3.90, s, 2H, CH_2 . Residual ether (0.2 molecule) shows signals at 3.49, q and 1.22, t.

Analysis: Found: C, 40.61; H, 2.37; N, 5.83; I, 24.75. Calc. for $\text{C}_{17}\text{H}_{11}\text{N}_2\text{O}_5\text{IFe} \times 0.2$ $\text{C}_4\text{H}_{10}\text{O}$: C, 40.35; H, 2.19; N, 5.54; I, 25.08.

Reaction of **2** with *Na*-phenytoin

A solution of **2** (102 mg, 0.20 mmol) and phenytoin (51 mg, 0.20 mmol) in 0.05 M NaOMe in MeOH (4 ml) was incubated at 50°C for 24 h, cooled to r.t. and poured to water (20 ml). The yellow precipitate was filtered off, dried under vacuum and recrystallized from CH_2Cl_2 –pentane. Yield 107 mg (85%). IR (cm^{-1}): 2045, 1990 (Fe–CO); 1780, 1725 (phenytoin and amide CO); 1645 (phthalimide CO); ^1H -NMR: 9.11, bs, 1H (NH); 7.3–7.8, m, 13H (aromatic Hs); 5.07, s, 5H (Cp); 4.34, s, 2H (CH_2); FAB MS [positive ions, *m/e* (intensity)]: 631 (24%), $\text{M} + \text{H}$; 575 (56%),

$\text{M} + \text{H} - 2\text{CO}$. Elemental analysis: Found: C 61.08; H 3.72; N 8.82. Calc. for $\text{C}_{32}\text{H}_{22}\text{N}_4\text{O}_7\text{Fe}$: C 60.97; H 3.52; N 8.89.

Reaction of **2** with *Na*-ethosuximide

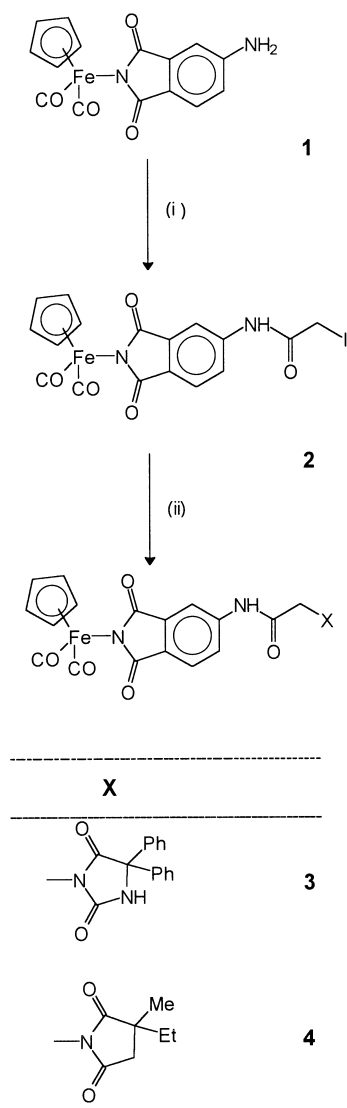
A solution of **2** (102 mg, 0.20 mmol) and ethosuximide (30 mg, 0.21 mmol) in 0.05 M NaOMe in MeOH (4 ml) was incubated at 50°C for 2 d, cooled to r.t., poured to water (20 ml) and extracted with dichloromethane (3×5 ml). The combined extracts were dried with sodium sulphate and evaporated to dryness to give yellow oil. The oil was dissolved in ether (~ 20 ml), diluted with hexanes and slowly concentrated *in vacuo* to give a yellow solid (53 mg, 51%). IR (cm^{-1}): 2045, 1990, (Fe–CO); 1775, 1720 (amide and succinimide CO); 1645 (phthalimide CO); ^1H -NMR: 8.32 bs, 1H, NH; 7.72, d ($J = 8$ Hz), 1H and 7.54, d ($J = 8$ Hz), 1H, H-5 and H-6; 7.67, s, 1H, H-3; 5.12, s, 5H, Cp; 4.38, s, 2H, COCH_2 ; 2.77, d ($J = 18$ Hz) 1H and 2.54, d ($J = 18$ Hz), 1H, succinimide ring CH_2 , 1.82 m, 2H CH_2 (in Et); 1.38, s, 3H, Me; 0.94, t ($J = 7$ Hz), 3H, CH_3 (in Et); FAB MS [positive ions, *m/e* (intensity)]: 520 (26%), $\text{M} + \text{H}$; 464 (51%), $\text{M} + \text{H} - 2\text{CO}$. Elemental analysis: Found: C 55.68; H 4.12; N, 8.32. Calc. for $\text{C}_{24}\text{H}_{21}\text{N}_3\text{O}_7\text{Fe}$: C 55.51; H 4.08; N, 8.09.

RESULTS AND DISCUSSION

Our results are summarized in Scheme 1.

We have found that readily accessible 4-aminophthalimidato complex **1** [11] readily reacts with iodoacetic acid and *N,N'*-dicyclohexylcarbodiimide (DCC) in dichloromethane at room temperature to give the 4-iodoacetamido derivative **2** in high yield. Its structure was confirmed by spectroscopic as well as elemental analyses data. The amino group in **1** can be therefore acylated under mild conditions and **1** is a potential labeling reagent for biomolecules containing carboxylic functions. This possibility, however, will be a subject of further studies.

The iodoacetamido complex **2** readily reacts with sodium salts of phenytoin and ethosuximide (prepared by dissolving of the corresponding NH acids in methanol containing the equimolar amount of sodium methoxide) to afford conjugates **3** and **4**, respectively. The structures of these complexes were confirmed by spectroscopic and elemental analysis data. Their FAB mass spectra (*meta*-nitrobenzyl alcohol as a matrix) display, in the positive ions mode, peaks corresponding to $\text{M} + \text{H}$ and $\text{M} + \text{H} - 2\text{CO}$ ions and in the negative ions mode to $\text{M} - \text{H}$ and $\text{M} - \text{H} - 2\text{CO}$ ions. ^1H NMR spectra of conjugates **3** and **4** show characteristic singlet of $-\text{COCH}_2\text{N}-$ protons (4.44 ppm for **3** and 4.38 ppm for **4**), shifted downfield in comparison to the signal of the $-\text{COCH}_2\text{I}$ protons in **2** (3.90 ppm). The complexes **3** and **4** are yellow air-stable microcrystalline solids, soluble in polar organic



solvents (ethanol, dimethylsulfoxide, dimethylformamide), in chloroform, and practically insoluble in water. Their IR spectra show very strong absorption bands at around 2050 cm^{-1} and 1990 cm^{-1} i.e. in

Compound **2** as containing the iodoacetamido function is also a potential label for biomolecules containing HS-groups [12,13]. Feasibility of such labeling is currently under study in our laboratory.

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