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Letter

Total Synthesis of Naphterpin and Marinone Natural Products

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S Supporting Information

ABSTRACT: A concise and divergent strategy for the synthesis of the naphterpin and marinone meroterpenoid families has been developed. The approach features a succession of pericyclic reactions—an aromatic Claisen rearrangement, a retro- 6π -electrocyclization, and two Diels–Alder reactions— which facilitated the first total synthesis of naphterpin itself in



five steps from 2,5-dimethoxyphenol, alongside similar syntheses of 7-demethylnaphterpin and debromomarinone. Late-stage oxidation and bromination reactions were also investigated, leading to the first total syntheses of naphterpins B and C and isomarinone.

Marine and terrestrial strains of *Streptomyces* bacteria have been an excellent source of new natural products with novel structures and potent bioactivity,¹ including naphthoquinone meroterpenoids such as the naphterpins (Figure 1a),



Figure 1. THN meroterpenoids of interest in this work: (a) the naphterpins, (b) the naphthgeranines, (c) the marinones.

the naphthgeranines (Figure 1b), and the marinones (Figure 1c). Naphterpin (1) was the first natural product discovered in this family, isolated by Seto et al. in 1990 from *Streptomyces* sp. CL-190 that was collected in Ishigaki Island, Okinawa.² The isolation of the oxidized derivatives naphterpin B (2) and naphterpin C (3) from the same microbial source was reported later in 2005.³ Naphthgeranine A (4, also independently reported as 7-demethylnaphterpin⁴) was isolated alongside several oxidized derivatives such as naphthgeranine B (5) and naphthgeranine C (6) by Zeeck in 1991.⁵ Debromomarinone

(7) and marinone (8) were isolated by Fenical and coworkers from an actinomycete derived from marine sediment in $1992_{,}^{6}$ with isomarinone (9) reported in $2000.^{7}$

Previous 13 C labeling studies by Seto et al.⁸ and Moore et al.⁹ have indicated that the naphterpins and marinones are biosynthetically derived from 1,3,6,8-tetrahydroxynaphthalene (THN, **10**) and either geranyl or farnesyl diphosphate, respectively, although the precise biosynthesis was not immediately obvious (Scheme 1). We recently proposed a

Scheme 1. Proposed Biosynthesis and Previous Biomimetic Synthesis of the Naphterpins and Marinones



biosynthetic pathway to the naphterpins and marinones in which the THN ring system undergoes oxidation via "cryptic chlorination" catalyzed by vanadium-dependent haloperoxidases (VHPOs), despite the absence of chlorine in these natural products.¹⁰ This biosynthetic speculation was originally based on analogies to the merochlorin¹¹ and napyradiomycin¹² families of marine *Streptomyces* meroterpenoids, which we have previously shown to be biosynthesized via the VHPO-catalyzed dearomatization of THN derivatives. On the basis of our

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biosynthetic studies, we achieved a biomimetic synthesis of 7demethylnaphterpin (4) and debromomarinone (7) from the protected THN derivative 11.¹⁰ We also synthesized several proposed biosynthetic intermediates, which were used to decipher the reactivity of two VHPO enzymes from the putative marinone gene cluster that chlorinate and dearomatize prenylated THN derivatives. However, because of the complexity of the biosynthetic pathway, our biomimetic route to 7-demethylnaphterpin (4) and debromomarinone (7) was 15 steps from commercially available (3,5dimethoxyphenyl)acetic acid (Scheme 1). The biomimetic strategy was too long and too linear to allow the synthesis of further oxidized or brominated members of the naphterpin/ marinone family.

We have therefore devised a simpler, nonbiomimetic retrosynthetic analysis of the naphterpins that is not constrained by the use of a THN building block (Scheme 2). We





envisaged that the cis-fused cyclohexane and pyran rings of **1** and **4** could be formed by an intramolecular hetero-Diels– Alder reaction of a reactive enone, which is unveiled via a latestage retro- 6π -electrocyclization (and alkene isomerization) of naphthoquinones **12a/b**. This endgame was also used in our previous synthesis.¹⁰ However, the disconnection of **12a/b** from the simple Brassard dienes¹³ **13a/b** and quinone **14** via an intermolecular Diels–Alder reaction¹⁴ (and aerobic oxidation) had the potential to significantly shorten our previous biomimetic synthesis and therefore allow divergent access to oxidized and brominated family members. A related intermolecular Diels–Alder reaction was used in Nakata's total synthesis of the napyradiomycin A80915G^{12a} and also in Tapia's synthetic study toward the naphterpins (but no naphterpin total syntheses were achieved by this route).¹⁵

As shown in Scheme 3, our synthesis of the naphterpin family began with the commercially available 2,5-dimethoxyphenol (15),¹⁶ which was coupled to propargylic carbonate 16¹⁷ in the presence of DBU and copper(II) chloride¹⁸ to give 17. The thermal rearrangement of 17 gave chromene 18 in high yield via a sequence of aromatic Claisen rearrangement to give an *o*-allenyl phenol, followed by a 1,5-hydrogen shift to give an *o*-quinone methide and a final oxa- 6π -electrocyclization.¹⁹ Chromene 18 was then oxidized to give quinone 14 using PhI(O₂CCF₃)₂.²⁰ A regioselective intermolecular Diels– Alder reaction between 14 and diene 13a²¹ proceeded spontaneously at room temperature to give naphthoquinone 12a (presumably via the aerobic oxidation of an intermediate hydroquinone). Heating 12a with MgBr₂ in MeCN induced a retro- 6π -electrocyclization and alkene isomerization to give a reactive enone, which then underwent an intramolecular Diels-Alder reaction to form naphterpin (1). The labile C-8 methyl ether was also cleaved under these mild conditions. The epoxidation of naphterpin with *m*-CPBA gave a separable 3:1 mixture of epoxides 19 and 20. Unexpectedly, the major epoxide diastereomer 19 results from the electrophilic attack on the concave face of the $\Delta^{12,13}$ alkene of **1**. However, Brase et al. observed a similar ratio of endo and exo epoxide products in the epoxidation of a related meroterpenoid system.²² The relative configuration of 19 was unambiguously assigned via single-crystal X-ray crystallography, whereas 20 was assigned using nuclear Overhauser effect (NOE) data and a comparison of key ³J coupling constants with computational predictions.²³ The ring opening of epoxides 19 and 20 with lithium diisopropylamide (LDA) then gave naphterpin B (2) and naphterpin C (3), respectively, presumably by an E1cB mechanism that involves an intermediate o-quinone methide enolate, with the retention of the stereochemistry at C-13. The relative configuration of naphterpin C was originally assigned by Seto via a rotating frame Overhauser enhancement spectroscopy (ROESY) correlation between Me-20 and Me-18. Replacing diene 13a with $13b^{24}$ in the intermolecular Diels-Alder reaction with quinone 14 allowed the synthesis of 12b, followed by 7-demethylnaphterpin (4) upon Lewis-acidcatalyzed rearrangement. To date, we have been unable to convert 4 into more oxidized members of the naphthgeranine family, such as naphthgeranines B and C (5 and 6), via allylic oxidations.

The synthetic strategy was also applicable to the marinone family by replacing the prenyl propargylic carbonate 16 with geranyl propargylic carbonate 21,²⁵ which was coupled to 15 to give 22 (Scheme 4). The thermal aromatic Claisen rearrangement of 22, followed by oxidation gave quinone 23, which underwent a facile intermolecular Diels-Alder reaction with 13b to give naphthoquinone 24. The conversion of 24 to debromomarinone (7) was then achieved via the now familiar MgBr₂-catalyzed cascade of retro- 6π -electrocyclization and the intramolecular hetero-Diels-Alder reaction. Finally, debromomarinone was brominated using 1.5 equiv N-bromosuccinimide (NBS) in CH₂Cl₂ at 0 °C. Under these conditions, we observed the exclusive bromination of 7 at C-7 to give isomarinone (9) in 38% isolated yield. We observed no bromination at C-5 and therefore no formation of marinone (8). Heteronuclear multiple bond correlation (HMBC) correlations confirmed the original assignment of the isomarinone bromination pattern. In nature, the selective bromination of 7 at C-5 to give marinone (8) might occur under the control of a vanadium-dependent haloperoxidase enzyme, or it could occur at an earlier point in the biosynthetic pathway.

While searching online databases for further members of the naphterpin family of meroterpenoids, we found a report of "naphthoquinone C" (25), isolated from a marine-derived *Penicillium* sp. (Figure 2).²⁶ Given the likely biosynthetic origin of 25 from THN and geranyl diphosphate, we thought that its structure was probably misassigned. Indeed, the comparison of the ¹H NMR spectrum of 25 recorded in CD₃OD showed an excellent match with the corresponding data for the previously reported naphthgeranine C (6).^{5a,27} Unfortunately, the ¹³C NMR spectra of the two natural products were recorded in different solvents; nevertheless, we suggest that naphthoqui-



Scheme 3. Divergent Total Synthesis of Naphterpin, Naphterpin B, Naphterpin C, and 7-Demethylnaphterpin





none C has been incorrectly assigned, and that it is probably reisolated naphthgeranine C. 28 This proposal is supported by the computational prediction of the 13 C NMR spectra of



Figure 2. Suggested structure revision of naphthoquinone C.

structures 6 and 25 and the comparison to the natural product data.²³

In conclusion, we have developed a concise, modular strategy for the synthesis of the naphterpin and marinone families of marine merotepenoids. This has resulted in the divergent synthesis of six natural products, with naphterpin, naphterpin B, naphterpin C, and isomarinone all synthesized for the first time. The synthetic strategy features a sigmatropic rearrangement, an electrocyclization and two Diels-Alder reactions, thus showcasing the power of pericyclic reactions in the construction of complex natural products.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.9b03095.

Experimental procedures and analytical data for all new compounds (PDF)

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Accession Codes

CCDC 1944007 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) (a) Fenical, W.; Jensen, P. R. Nat. Chem. Biol. 2006, 2, 666.
(b) Gulder, T. A. M.; Moore, B. S. Curr. Opin. Microbiol. 2009, 12, 252.

(2) Shin-ya, K.; Imai, S.; Furihata, K.; Hayakawa, Y.; Kato, Y.; Vanduyne, G. D.; Clardy, J.; Seto, H. J. Antibiot. **1990**, 43, 444.

(3) For the isolation of naphterpins B and C, see: (a) Takagi, H.; Motohashi, K.; Miyamoto, T.; Shin-ya, K.; Furihata, K.; Seto, H. J. Antibiot. 2005, 58, 275. More recently, naphterpins D and E were reported. See: Park, J.-S.; Kwon, H. C. Mar. Drugs 2018, 16, 90.

(4) Shin-ya, K.; Shimazu, A.; Hayakawa, Y.; Seto, H. J. Antibiot. 1992, 45, 124.

(5) For the isolation of naphthgeranines A–E, see: (a) Wessels, P.; Gohrt, A.; Zeeck, A.; Drautz, H.; Zähner, H. J. Antibiot. **1991**, 44, 1013. For the isolation of naphthgeranine F, see: (b) Volkmann, C.; Jen, U. H.; Zeeck, A.; Fiedler, H.-P. J. J. Antibiot. **1995**, 48, 522. For the isolation of 12-hydroxy-naphthgeranine A and isonaphthgeranine C, see: (c) Lu, C.; Yang, C.; Xu, Z. Rec. Nat. Prod. **2016**, 10, 437.

(6) Pathirana, C.; Jensen, P. R.; Fenical, W. Tetrahedron Lett. **1992**, 33, 7663.

(7) Hardt, I. H.; Jensen, P. R.; Fenical, W. Tetrahedron Lett. 2000, 41, 2073.

(8) Shin-ya, K.; Furihata, K.; Hayakawa, Y.; Seto, H. Tetrahedron Lett. **1990**, 31, 6025.

(9) Kalaitzis, J. A.; Hamano, Y.; Nilsen, G.; Moore, B. S. Org. Lett. 2003, 5, 4449.

(10) Murray, L. A. M.; McKinnie, S. M. K.; Pepper, H. P.; Erni, R.; Miles, Z. D.; Cruickshank, M. C.; Lopez-Perez, B.; Moore, B. S.; George, J. H. Angew. Chem., Int. Ed. **2018**, *57*, 11009.

(11) For the isolation of the merochlorins, see: (a) Kaysser, L.; Bernhardt, P.; Nam, S.-J.; Loesgen, S.; Ruby, J. G.; Skewes-Cox, P.; Jensen, P. R.; Fenical, W.; Moore, B. S. J. Am. Chem. Soc. 2012, 134, 11988. For biosynthetic studies on the merochlorins, see: (b) Teufel, R.; Kaysser, L.; Villaume, M. T.; Diethelm, S.; Carbullido, M. K.; Baran, P. S.; Moore, B. S. Angew. Chem., Int. Ed. 2014, 53, 11019.
(c) Diethelm, S.; Teufel, R.; Kaysser, L.; Moore, B. S. Angew. Chem., Int. Ed. 2014, 53, 11023. For biomimetic synthetic studies on the merochlorins, see: (d) Pepper, H. P.; George, J. H. Angew. Chem., Int. Ed. 2013, 52, 12170. (e) Meier, R.; Strych, S.; Trauner, D. Org. Lett. 2014, 16, 2634. (f) Yang, H.; Liu, X.; Li, Q.; Li, L.; Zhang, J.-R.; Tang, Y. Org. Biomol. Chem. 2016, 14, 198. (g) Pepper, H. P.; George, J. H. Synlett 2015, 26, 2485. (h) Lopez-Perez, B.; Pepper, H. P.; Ma, R.; Fawcett, B. J.; Pehere, A. D.; Wei, Q.; Ji, Z.; Polyak, S. W.; Dai, H.; Song, F.; Abell, A. D.; Zhang, L.; George, J. H. ChemMedChem 2017, 12, 1969. For an asymmetric synthesis of merochlorin A, see: (i) Brandstatter, M.; Freis, M.; Huwyler, N.; Carreira, E. M. Angew. Chem., Int. Ed. **2019**, 58, 2490.

(12) For biosynthetic and synthetic studies on the napyradiomycins, see: (a) Takemura, S.; Hirayama, A.; Tokunaga, J.; Kawamura, F.; Inagaki, K.; Hashimoto, K.; Nakata, M. *Tetrahedron Lett.* **1999**, 40, 7501. (b) Tatsuta, K.; Tanaka, Y.; Kojima, M.; Ikegami, H. *Chem. Lett.* **2002**, 31.14 (c) Snyder, S. A.; Tang, Z.-Y.; Gupta, R. J. Am. Chem. Soc. **2009**, 131, 5744. (d) Bernhardt, P.; Okino, T.; Winter, J. M.; Miyanaga, A.; Moore, B. S. J. Am. Chem. Soc. **2011**, 133, 4268. (e) Miles, Z. D.; Diethelm, S.; Pepper, H. P.; Huang, D. M.; George, J. H.; Moore, B. S. Nat. Chem. **2017**, 9, 1235. (f) Moore, B. S. Synlett **2018**, 29, 401. (g) McKinnie, S. M. L.; Miles, Z. D.; Jordan, P. A.; Awakawa, T.; Pepper, H. P.; Murray, L. A. M.; George, J. H.; Moore, B. S. J. Am. Chem. Soc. **2018**, 140, 17840. (h) Schnell, S. D.; Linden, A.; Gademann, K. Org. Lett. **2019**, 21, 1144. (i) Landry, M. L.; McKenna, G. M.; Burns, N. Z. J. Am. Chem. Soc. **2019**, 141, 2867.

(13) (a) Savard, J.; Brassard, P. Tetrahedron Lett. 1979, 20, 4911.
(b) Courchesne, M.; Brassard, P. J. Nat. Prod. 1993, 56, 722.

(14) For a review of the use of quinones as dienophiles in Diels-Alder reactions applied to total synthesis, see: Nawrat, C. C.; Moody, C. J. Angew. Chem., Int. Ed. 2014, 53, 2056.

(15) Tapia, R. A.; Alegria, L.; Valderrama, J. A.; Cortes, M.; Pautet, F.; Fillion, H. *Tetrahedron Lett.* **2001**, *42*, 887.

(16) The synthesis of 2,5-dimethoxyphenol (15) from 2,5dimethoxybenzaldehyde was also conducted. Wriede, U.; Fernandez, M.; West, K. F.; Harcour, D.; Moore, H. W. J. Org. Chem. 1987, 52, 4485.

(17) (a) Tsuji, J.; Sugiura, T.; Minami, I. Synthesis 1987, 1987, 603.
(b) Gassner, C.; Hesse, R.; Schmidt, A. W.; Knolker, H.-J. Org. Biomol. Chem. 2014, 12, 6490.

(18) Godfrey, J. D.; Mueller, R. H.; Sedergran, T. C.; Soundararajan, N.; Colandrea, V. J. *Tetrahedron Lett.* **1994**, 35, 6405.

(19) For a similar aromatic Claisen rearrangement, see: (a) Tisdale,
E. J.; Chowdhury, C.; Vong, B. G.; Li, H.; Theodorakis, E. A. Org. Lett. 2002, 4, 909. For mechanistic discussion of this reaction, see:
(b) Rhoads, S. J.; Raulins, N. R. Org. React. 1975, 22, 1.

(20) For a similar oxidation using $PhI(O_2CCF_3)_2$, see: Nicolaou, K. C.; Sasmal, P. K.; Xu, H.; Namoto, K.; Ritzen, A. Angew. Chem., Int. Ed. 2003, 42, 4225.

(21) Nawrat, C. C.; Lewis, W.; Moody, C. J. J. Org. Chem. 2011, 76, 7872.

(22) Glaser, F.; Brohmer, M. C.; Hurrle, T.; Nieger, M.; Brase, S. *Eur. J. Org. Chem.* **2015**, 2015, 1516.

(23) See the Supporting Information for a full presentation of molecular modeling and NMR predictions.

(24) Barker, D.; Brimble, M. A.; Do, P.; Turner, P. Tetrahedron 2003, 59, 2441.

(25) Tumma, N.; Jacolot, M.; Jean, M.; Chandrasekhar, S.; van de Weghe, P. *Synlett* **2012**, *23*, 2919.

(26) Li, X.; Zheng, Y.; Sattler, I.; Lin, W. Arch. Pharmacal Res. 2006, 29, 942.

(27) See the Supporting Information for comparison tables of natural product NMR data alongside 1D and 2D NMR spectra.

(28) For a recent account of natural product structure revisions inspired by biosynthetic speculation, see: Brown, P. D.; Lawrence, A. L. Nat. Prod. Rep. 2017, 34, 1193.