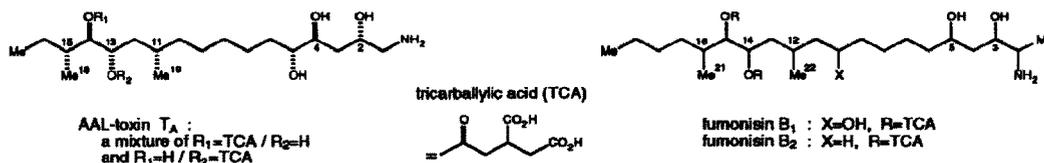


Relative and Absolute Stereochemistry of the Fumonisin B₂ Backbone

Jean-Christophe Harmange, Craig D. Boyle, and Yoshito Kishi*

Department of Chemistry, Harvard University
12 Oxford Street, Cambridge, Massachusetts 02138, U.S.A.**Abstract:** The relative and absolute stereochemistry of the backbone of fumonisin B₂ is established to be **7a**.

Fumonisin B₂ is a mycotoxin produced by the corn pathogen *Fusarium moniliforme*.¹ They are known to be carcinogenic and directly linked to human esophageal cancer. Coupled with its contamination in some commercially based corn products, this biological activity has drawn worldwide attention to the fumonisin family of mycotoxins.² Many comparisons have been made between fumonisins and AAL toxins produced by the tomato fungus *Alternaria alternata* f. sp. *lycopersici*.³ In particular, both families exhibit cross-bioactivity and have been shown to inhibit sphingolipid biosynthesis.^{2,4} Fumonisin B₂ and AAL toxins bear striking structural similarity. Their gross structures have been determined, but their relative and absolute stereochemistry remains unknown.^{1,3} We have recently established the relative and absolute stereochemistry of the backbone of AAL toxin T_A by using a stepwise approach: (1) determination of the relative stereochemistry of the left and right halves independently, (2) differentiation of the two possible diastereomers, corresponding to **4a** and **6a** in the fumonisin B₂ series, by the ¹H NMR spectra in the presence of an achiral shift reagent, and (3) determination of the absolute stereochemistry of the amino alcohol, corresponding to **4a** vs. **7a** in the fumonisin B₂ series, by the ¹H NMR spectra in the presence of a chiral shift reagent.⁵



The striking similarity in their gross structures may suggest that fumonisins and AAL toxins are biosynthesized via related pathways, yielding the same stereochemical array on their backbones. Indeed, the ¹H NMR characteristics reported for the C12-C16 moiety of N-acetyl fumonisin B₁ methyl ester¹ compare amazingly well with those for the corresponding portion of the peracetate prepared from the amino alcohol of AAL toxin T_A. This fact convincingly argues that the stereochemistry of the left half of fumonisins relates to that of AAL toxins. Then, it is tempting to suggest that the stereochemistry at the C3 and C5 positions of fumonisin B₁ or B₂ corresponds to that at the C2 and C4 positions of AAL toxin T_A. Comparing the ¹³C NMR data of the C1-C4 portion of N-acetyl fumonisin B₁ methyl ester with the acetates derived from 2-aminotetradeca-5,7-dien-3-ols,⁶ the relative stereochemistry at the C2 and C3 positions appears to be syn. On the basis of these considerations and

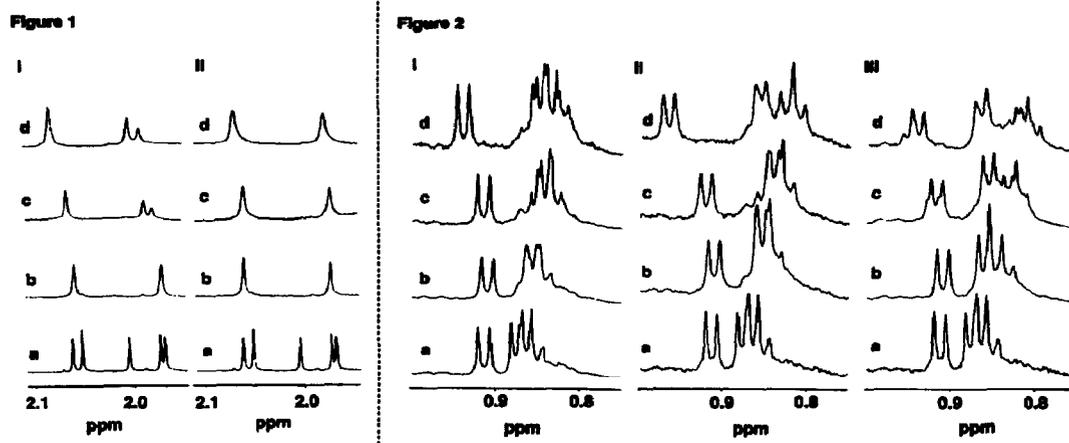


Figure 1. Acetate group region of ^1H NMR (500 MHz, CDCl_3). I: $\text{Eu}(\text{fod})_3$ titration of a 2:1 mixture of **6b** and **7b**. II: $\text{Eu}(\text{fod})_3$ titration of a 2:1 mixture of **4b** and **7b**. (a) 0 eq. $\text{Eu}(\text{fod})_3$. (b) ca. 0.2 eq. $\text{Eu}(\text{fod})_3$. (c) ca. 0.6 eq. $\text{Eu}(\text{fod})_3$. (d) ca. 0.8 eq. $\text{Eu}(\text{fod})_3$. *Note:* Upon addition of ca. 0.2 eq. $\text{Eu}(\text{fod})_3$, three out of the five acetate peaks broadened significantly and shifted downfield beyond 2.1 ppm.

Figure 2. Methyl group region of ^1H NMR (500 MHz, CDCl_3). I: $\text{Eu}(\text{fod})_3$ titration of a 2:1 mixture of **6b** and **7b**. II: $\text{Eu}(\text{fod})_3$ titration of a 2:1 mixture of **4b** and **7b**. III: (+)- $\text{Eu}(\text{hfc})_3$ titration of a 2:1 mixture of **4b** and **7b**. (a) 0 eq. EuR_3 . (b) ca. 0.4 eq. EuR_3 . (c) ca. 0.8 eq. EuR_3 . (d) ca. 1.2 eq. EuR_3 .

Further studies on the stereochemistry of the tricarballic acid moiety of fumonisins and AAL toxins are in progress in our laboratories.

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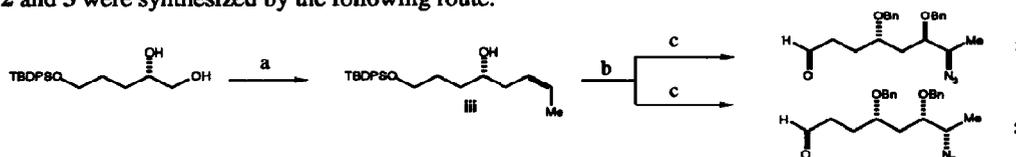
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7. **1** was synthesized by the route used to make a similar intermediate for AAL toxin⁵, using (S)-(-)-2-methyl-1-hexanol instead of (S)-(-)-2-methyl-1-butanol. (S)-(-)-2-Methyl-1-hexanol was prepared from the procedure given in Kato, M.; Mori, K. *Agric. Biol. Chem.* **1985**, *49*, 2479.
8. Inspection of ref. 6 shows that when the C2 amine and C3 hydroxyl group are syn, the C1 methyl of the peracetate derivative has a ¹³C NMR chemical shift of ~18 ppm. However, when C2 and C3 are anti, the chemical shift is ~14 ppm. Fumonisin B₂ peracetate **7b** and model compounds **i** and **ii** all have a C1 methyl chemical shift at ~18 ppm in their ¹³C NMR spectra.

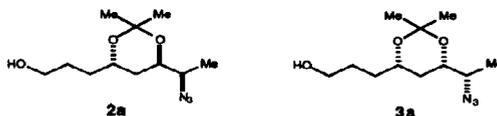


9. **2** and **3** were synthesized by the following route:



Reagents and conditions: (a) (1) NaH, THF, then Ts-imid. (2) *cis*-propenylmagnesium bromide, CuI, THF. (b) (1) NaH, BnBr, TBAI, THF/DMF. (2) MCPBA, CH₂Cl₂. (3) NaN₃, TBAI, DMF/HOCH₂CH₂OCH₃/DME/H₂O. (4) NaH, BnBr, TBAI, THF/DMF, then separation of diastereomers via SiO₂ column chromatography. (c) (1) TBAF, THF. (2) (COCl)₂, DMSO, CH₂Cl₂, Et₃N.

The starting diol was synthesized in 6 steps from L-glutamic acid by a modified Larcheveque procedure: Larcheveque, M.; Lalonde, J. *Tetrahedron* **1984**, *40*, 1061. The stereochemistry of **2** and **3** was determined by comparing the ¹³C NMR data of derivatives **2a** and **3a** as reported by Rychnovsky, S. D.; Rogers, B.; Yang, G. *J. Org. Chem.* **1993**, *58*, 3511.



10. The authentic amino alcohol was prepared from natural fumonisin B₂, purchased from the South African Research Council, Tygerberg, South Africa. Thus, fumonisin B₂ was treated with boiling NaOH to afford the free amine alcohol **7a**. Subsequent treatment with conc. HCl in MeOH provided the HCl salt **7a·HCl**, and treatment with acetic anhydride and pyridine gave the acetate **7b**.
11. The enantiomer of **2** was synthesized from **iii** via Mitsunobu inversion, followed by the sequence given in ref. 9.
12. ¹H NMR (500 MHz, D₂O) of **4a·HCl**, **6a·HCl** and **7a·HCl**: δ 0.76 (3H, t, *J* = 7.0 Hz), 0.77 (3H, d, *J* = 6.7 Hz), 0.80 (3H, d, *J* = 6.7 Hz), 0.98 (2H, m), 1.0-1.6 (22H, m), 1.18 (3H, d, *J* = 6.7 Hz), 3.13 (1H, m), 3.20 (1H, dd, *J* = 5.2, 6.4 Hz), 3.66 (2H, m), 3.74 (1H, m). **5a·HCl**: 0.77 (3H, t, *J* = 7.0 Hz), 0.78 (3H, d, *J* = 6.7 Hz), 0.82 (3H, d, *J* = 6.7 Hz), 0.99 (2H, m), 1.0-1.7 (22H, m), 1.19 (3H, d, *J* = 6.8 Hz), 3.20 (2H, m), 3.63 (1H, ddd, *J* = 3.4, 7.2, 10.2 Hz), 3.68 (1H, ddd, *J* = 1.8, 5.1, 10.3 Hz), 3.78 (1H, m).
13. ¹H NMR (500 MHz, CDCl₃) of **4b**, **6b** and **7b**: δ 0.86 (3H, t, *J* = 7.1 Hz), 0.88 (3H, d, *J* = 6.4 Hz), 0.91 (3H, d, *J* = 6.8 Hz), 1.0-1.8 (24H, m), 1.08 (3H, d, *J* = 6.8 Hz), 1.97 (3H, s), 1.97 (3H, s), 2.00 (3H, s), 2.05 (3H, s), 2.06 (3H, s), 4.13 (1H, m), 4.86 (1H, m), 4.88 (1H, dd, *J* = 3.3, 8.5 Hz), 4.95 (1H, m), 5.12 (1H, ddd, *J* = 2.7, 3.1, 10.8 Hz), 5.53 (1H, d, *J* = 9.3 Hz).

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