

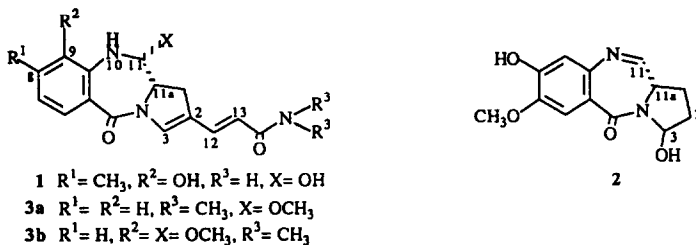
Enantioselective Synthesis of (+) Porphthramycin B and its 9-Demethoxy Analogue

Nicole Langlois*, Florence Favre and Anne Rojas

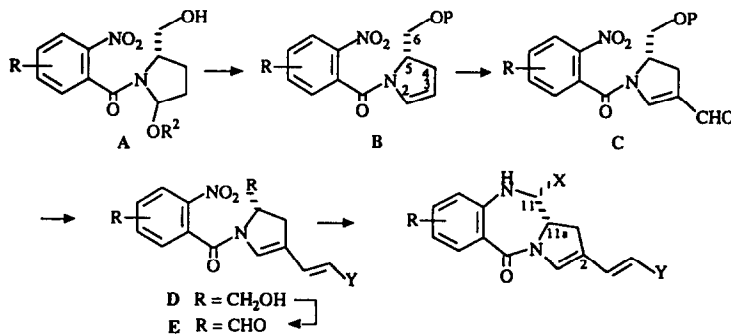
Institut de Chimie des Substances Naturelles, C.N.R.S., 91198 Gif-sur-Yvette, France

Abstract. (+) 9-Demethoxyporphthramycin B **3a** and (+) porphthramycin B **3b** were synthesized from (S) pyroglutamic acid through a versatile and efficient route.¹

Pyrrolo[2,1-c][1,4]benzodiazepines of the anthramycin **1**² group are antitumor antibiotics produced by various *Streptomyces* which mediate their activities by covalent binding with DNA.³ Their 11a*S* absolute configuration is important for their antitumor properties. Indeed, this configuration provides these compounds with a twist along the molecule which allows their interaction with DNA.⁴



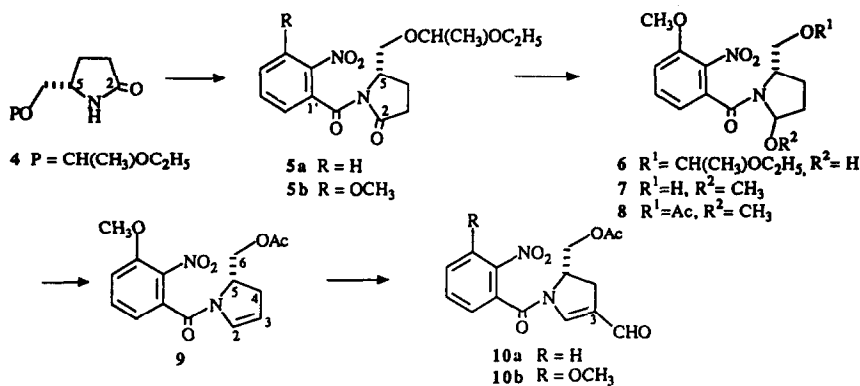
(S) Pyroglutamic acid is a convenient chiral starting material to synthesize natural enantiomers of this family of antitumor compounds, as first exemplified some years ago by the synthesis of neothramycins **2** and analogues.⁵ The (5*S*) *N*-2 nitrobenzoyl-5-hydroxymethyl-2-methoxy pyrrolidines of general formula **A** used in this work were shown to be also versatile intermediates in the synthesis of other pyrrolo [2,1-c][1,4]benzodiazepines including a dihydropyrrole ring and a conjugated double bond in a side chain attached to C-2, as outlined in general Scheme 1.



Scheme 1

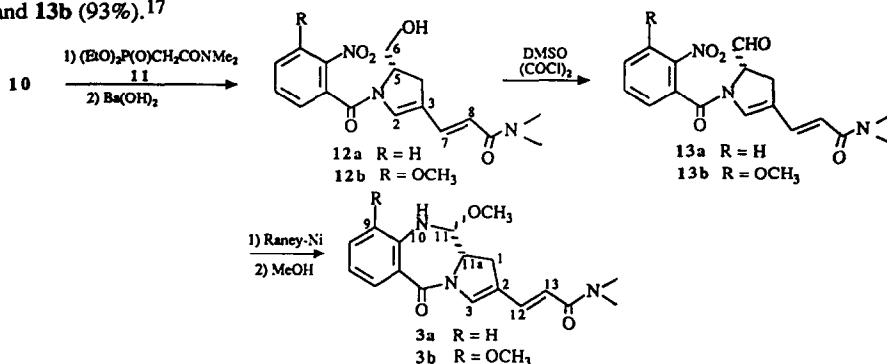
This synthetic strategy involves the preparation of the enamidoaldehydes **C** which allow the introduction of the unsaturated side chain *via* Wittig-type reactions to **D**.^{1,6} The recently reported synthesis of porothramycin **B 3b**⁷ using the same type of intermediate⁸ prompts us to disclose our own results in the same area,^{1a} and we describe here the short total syntheses of 9-demethoxy porothramycin **B 3a** and porothramycin **B 3b**.

The imide **5b**, prepared (95% yield)⁹ from 3-methoxy-2-nitrobenzoyl chloride and (5*S*) 5-ethoxyethoxymethyl-2-pyrrolidone **4**, as previously described for **5a**⁵, was reduced by DIBAL-H. The addition of a hexane solution of DIBAL-H to a THF solution of **5b** was required in this case to lead to a partial regioselective reduction to **6** (68%). These compounds were converted⁵ to 9-methoxy *ortho*nitrobenzamides **7** (83%)¹⁰ which was quantitatively acetylated to **8** (Scheme 2). Elimination reaction by heating a toluene solution of **8** in the presence of quinolinium camphorsulfonate (QCS)⁶ provided the enamide **9**¹¹ in 87% yield. The formyl group was introduced at C-3 through a Vilsmeier-Haack reaction affording the key enamidoaldehyde intermediate **10b** (87%).¹²



Scheme 2

The *N,N*-dimethylacrylamide side chain, common to the target molecules **3a** and **3b** was elaborated through a Wittig-Horner reaction with the suitably functionalized diethylphosphonate **11**¹³ (Scheme 3). The products of these reactions could be directly saponified with 1N Ba(OH)₂, respectively to the alcohols **12a** (97% from **10a**) and **12b** (non optimized 78% yield from **10b**)¹⁴. Swern oxidation¹⁵ of these primary alcohols **12** required the use of diisopropylethylamine as a base¹⁶ to avoid partial racemization of the aldehydes **13a** (96%) and **13b** (93%).¹⁷



Scheme 3

The reduction of the aromatic nitro group with an excess of Raney-nickel at room temperature and cyclization to the pyrrolo[2,1-c][1,4]benzodiazepine skeleton were directly followed by treatment with a very diluted solution of trifluoroacetic acid (0.002%) in CH_2Cl_2 -MeOH (9:1)¹⁸, providing respectively 9-demethoxyporothramycin B **3a** (65%), and porothramycin B **3b** (non optimized 45%).¹⁹

These results show the versatility of our synthetic strategy to prepare antitumor antibiotics of the anthramycin group as well as 11aS enantiomers of structural analogues, the biological activities of which will be reported elsewhere.

Acknowledgments : We thank the Ligue Nationale Française contre le Cancer and the Association de la Recherche sur le Cancer for financial support.

References and Notes

1. a) Langlois, N.; Favre, F.; Tempête-Gaillourdet, C.; Werner, G. Patent Fr. n° 91 05636, 7.05.1991; WO 92 19,620 (*Chem. Abs.* 1993, 118, 147588m). b) Langlois, N. Communication at "Natural Products and Drugs" Symposium, Marion Merrell Dow Research Institute, Strasbourg, France, 4-5 September 1992. c) Favre, F. Ph.D Thesis, University of Paris-Sud, Orsay, 1992, order n° 2502.
2. a) Leimgruber, W.; Stefanovic, V.; Schenker, F.; Karr, A.; Berger, J. *J. Am. Chem. Soc.* **1965**, *87*, 5791-5792. b) Leimgruber, W.; Batcho, A.D.; Schenker, F. *J. Am. Chem. Soc.* **1965**, *87*, 5793-5795. c) Leimgruber, W.; Batcho, A.D.; Czajkowski, R.C. *J. Am. Chem. Soc.* **1968**, *90*, 5641-5643. d) Peña, M.R.; Stille, J.K. *J. Am. Chem. Soc.* **1989**, *111*, 5417-5424.
3. a) Hurley, L.H. *J. Antibiotics*, **1977**, *30*, 349-370. b) Hurley, L.H.; Reck, T.; Thurston, D.E.; Langley, D.R. *Chem. Res. Toxicol.* **1988**, *1*, 258-268 and references therein.
4. a) Mostad, A.; Rømming, C.; Storm, B. *Acta Chemica Scandinavica* **1978**, *B32*, 639-645. b) Petrusek, R.L.; Anderson, G.L.; Garner, T.F.; Fannin, Q.L.; Kaplan, D.J.; Zimmer, S.G.; Hurley, L.H. *Biochemistry* **1981**, *20*, 1111-1119.
5. Andriamialisoa, R.Z.; Langlois, N. *Tetrahedron Lett.* **1986**, *27*, 1149-1152.
6. Langlois, N.; Favre, F. *Tetrahedron Lett.* **1991**, *32*, 2233-2236.
7. Tsunakawa, M.; Kamei, H.; Konishi, M.; Miyaki, T.; Oki, T.; Kawaguchi, H. *J. Antibiotics* **1988**, *41*, 1366-1372.
8. Fukuyama, T.; Liu, G.; Linton, S.D.; Lin S.-C.; Nishino, H. *Tetrahedron Lett.* **1993**, *34*, 2577-2580.
9. **5b** : IR (ν cm^{-1}) : 3000, 1750, 1680 ; ^1H NMR (200MHz, CDCl_3 , $\delta=0$: TMS, J Hz) : 7.52 (dd, 1H, $J_{4',5'}$ = 7.5, C-5'-H), 7.15 and 6.91 (d + m, 2H, $J_{4',5'}$ = 7.5, C-4'-H and C-6'-H), 4.74 (m, 1H, OCHO), 4.62 (m, 1H, C-5-H), 3.96 (s, 3H, C-3'-OCH₃), 3.87-3.52 (m, 4H, C-6-H₂ and OCH₂), 2.8-2.3 (m, 4H, C-4-H₂ and C-3-H₂), 1.32 (d, 3H, CHCH₃), 1.22 (m, 3H, CH₂CH₃) ; MS (CI, isobutane, m/z) : 367 (MH^+), 321 (100%), 295, 277 ; HRMS (m/z) : (M^+) 320.1507, calcd. for $\text{C}_{17}\text{H}_{22}\text{NO}_5$: 320.1498 .
10. **7b** (major diast.) : $[\alpha]_D = -282$ (c = 0.39, CHCl_3) ; IR : 3420, 2940, 1650, 1580, 1535 ; ^1H NMR (250MHz) : 7.55 (dd, 1H, $J_{4',5'}$ = 7.5, C-5'-H), 7.15 (m, 2H, C-4'-H and C-6'-H), 4.68 (d, 1H, $J_{2,3}$ = 3.8, C-2-H), 4.35 (m, 1H, C-5-H), 3.96 (s, 3H and m, 1H, C-3'-OCH₃ and C-6-Ha), 3.70 (m, 1H, C-6-Hb), 3.07 (s, 3H, C-2-OCH₃), 2.3-1.7 (m, 4H, C-3-H₂ and C-4-H₂) ; MS (m/z) : 310 (M^+), 309, 292, 291, 279, 278, 197, 180 (100%) ; Analysis : $\text{C}_{14}\text{H}_{18}\text{N}_2\text{O}_6$, calcd.% : C = 54.19, H = 5.85, N = 9.03, found : C = 54.01, H = 5.60, N = 8.99.
11. **9** : $[\alpha]_D = -222$ (c = 0.48, CHCl_3 , lit⁸ = -211, c = 0.56) ; IR : 2940, 1740, 1650, 1535 ; ^1H NMR (200 MHz) : 7.54 (dd, 1H, $J_{4',5'}$ = 7.5, C-5'-H), 7.16 and 7.02 (2d, 2H, $J_{4',5'}$ = 7.5, C-4'-H and C-6'-H), 6.17 (m, 1H, C-2-H), 5.19 (m, 1H, C-3-H), 4.85 (m, 1H, C-5-H), 4.43 (dd, 1H, $J_{5,6a}$ = 5, $J_{6a,6b}$ = 12, C-6-Ha), 4.22 (dd, 1H, $J_{5,6b}$ = 4 and $J_{6a,6b}$ = 12, C-6-Hb), 3.97 (s, 3H, C-3'-OCH₃), 2.93 (m, 1H, $J_{4a,4b}$ = 16, C-4-Ha), 2.50 (dd, 1H, $J_{4a,4b}$ = 16, C-4-Hb), 2.11 (s, 3H, OCOCH₃) ; MS : 320 (M^+), 279, 180 (100%), 106, 80, 76 ; Analysis : $\text{C}_{15}\text{H}_{16}\text{N}_2\text{O}_6$, calcd.% : C = 56.25, H = 5.04, N = 8.75, found : C = 56.18, H = 5.07, N = 8.57.
12. **10b** : $[\alpha]_D = -254$ (c = 0.35, CHCl_3) ; IR : 3400, 2950, 1750, 1655, 1610 ; ^1H NMR (200 MHz) : 9.53 (s, 1H, CHO), 7.61 (dd, 1H, C-5'-H), 7.25 (d, 1H, C-6'-H ou C-4'-H), 7.04 (d, 2H, C-4'-H ou C-6'-H and C-2-H), 4.94 (m, 1H, C-5-H), 4.55 (bd, 1H, $J_{6a,6b}$ = 12, C-6-Ha), 4.19 (dd, 1H, $J_{6a,6b}$ = 12 et $J_{5,6b}$ = 4, C-6-Hb), 3.98 (s, 1H, C-3'-OCH₃), 3.08 (dd, 1H, $J_{4a,4b}$ = 16.5, $J_{4a,5}$ = 10, C-4-Ha), 2.77 (dd, 1H, $J_{4a,4b}$ = 16.5,

- $J_5, 4b = 4.5$, C-4-Hb), 2.08 (s, 3H, OCOCH₃); ¹³C NMR (50 MHz): 185.8 (CHO), 170.8 (OCO), 163.8 (NCO), 152.3 (C-3'), 145.0 (C-2), 142.7 (C-3), 133.0 (C-5'), 130.3 and 127.0 (C-1' and C-2'); 119.1 and 115.7 (C-4' and C-6'), 63.5 (C-6), 58.2 and 56.9 (ArOCH₃ and C-5), 28.8 (C-4), 20.6 (COCH₃); MS: 348 (M⁺), 279, 265, 180 (100%), 164, 133, 106, 78, 76; HRMS: (M⁺) 348.0957, calcd. for C₁₆H₁₆N₂O₇: 348.0958.
13. a) Bartlett, P.A.; Carruthers, N.I.; Winter, B.M.; Long, K.P. *J. Org. Chem.* **1982**, *47*, 1284-1291.
14. a) **12a**: [α]_D = -114 (c = 0.66, CHCl₃); IR: 3390, 2930, 1635, 1605; ¹H NMR (200MHz): 8.26 (d, 1H, J_{3',4'} = 8, C-3'-H), 7.80 and 7.68 (2dd, 2H, C-4'-H and C-5'-H), 7.50 (d, 1H, J_{5',6'} = 8, C-6'-H), 7.22 (d, 1H, J_{7,8} = 15, C-7-H), 6.21 (s, 1H, C-2-H), 6.14 (d, 1H, J_{7,8} = 15, C-8-H), 4.86 (m, 1H, C-5-H), 4.0 (m, 2H, C-6-H₂), 3.10 (bs, 1H, OH), 3.01 (s + m, 7H, N(CH₃)₂ and C-4-Ha), 2.65 (dd, 1H, J_{4a,4b} = 15, J_{4b,5} = 4.5, C-4-Hb); SM: 345 (M⁺), 327, 280, 269, 208, 150 (100%), 120, 72; HRMS: (M⁺) 345.1308, calcd. for C₁₇H₁₉N₃O₅: 345.1317. b) **12b**: [α]_D = -111 (c = 0.76, CHCl₃); IR: 3390, 2925, 1635, 1600, 1580, 1530; ¹H NMR (200 MHz): 7.59 (dd, 1H, J_{4',5'} = J_{5',6'} = 7.5, C-5'-H), 7.29 (d, 1H, J_{7,8} = 15, C-7-H); 7.21 and 7.02 (2d, 2H, J_{4',5'} = J_{5',6'} = 7.5, C-4'-H and C-6'-H), 6.50 (s, 1H, C-2-H), 6.15 (d, 1H, J_{7,8} = 15, C-8-H), 4.75 (m, 1H, C-5-H), 3.97 (s, 3H, C-3'-OCH₃), 3.90 (m, 2H, C-6-H₂), 3.05 (s + m, 7H, N(CH₃)₂ and C-4-Ha), 2.65 (dd, 1H, J_{4a,4b} = 16, J_{4b,5} = 4, C-4-Hb); MS: 375 (M⁺), 357, 331, 310, 180 (100%), 120, 76, 72; HRMS: (M⁺) 375.1424, calcd. for C₁₈H₂₁N₃O₆: 375.1431.
15. Mancuso, A.J.; Huang, S.L.; Swern, D. *J. Org. Chem.* **1978**, *43*, 2480-2482.
16. a) Walba, D.M.; Thurmes, W.N.; Haliwanger, R.C. *J. Org. Chem.* **1988**, *53*, 1046-1056. b) Tidwell, T.T. *Synthesis* **1990**, 857-870.
17. a) **13a**: [α]_D = -126 (c = 0.96, CHCl₃); IR: 3300, 2920, 1720 weak ($\pm$ hydrated depending on the isolation conditions), 1630, 1595; ¹H NMR (250 MHz): 9.90 (s, 1H, CHO), 8.26 (dd, 1H, J_{3',4'} = 7.5, C-3'-H), 7.82 and 7.70 (2dd, 2H, J_{3',4'} = J_{4',5'} = J_{5',6'} = 7.5, C-4'-H and C-5'-H), 7.57 (d, 1H, J_{5',6'} = 7.5, C-6'-H), 7.23 (d, 1H, J_{7,8} = 15, C-7-H); 6.33 (s, 1H, C-2-H), 6.14 (d, 1H, J_{7,8} = 15, C-8-H), 5.20 (dd, 1H, J = 11.5 and 5, C-5-H), 3.08 and 3.00 (2s + m, 8H, N(CH₃)₂ and C-4-H₂); MS: 343 (M⁺), 269, 220, 208, 150 (100%); HRMS: (M⁺) 343.1147, calcd. for C₁₇H₁₇N₃O₅: 343.1161. b) **13b**: [α]_D = -110 (c = 0.40, CHCl₃); IR: 3300, 2980, 1730, 1640; ¹H NMR (200MHz): 9.75 (s, 1H, CHO), 7.27 (d, 1H, J_{7,8} = 15, C-7-H), 7.58, 7.25 (masked) and 7.10 (aromatic H), 6.65 (bs, 1H, C-2-H), 6.16 (d, 1H, J_{7,8} = 15, C-8-H), 5.12 (m, 1H, C-5-H), 3.99 (s, 3H, C-3'-OCH₃), 3.17 and 3.10 (2s + m, 8H, N(CH₃)₂ and C-4-H₂); MS: 373 (M⁺), 344, 310, 299, 180 (100%); HRMS: (M⁺) 373.1277, calcd. for C₁₈H₁₉N₃O₆: 373.1273.
18. Langlois, N.; Andriamialisoa, R. Z. *Heterocycles* **1989**, *29*, 1529-1536.
19. a) **3a**: mp inst. (dec.): 228 °C (MeOH); [α]_D = +775 (c = 0.2, CH₂Cl₂); IR: 3320, 2924, 2850, 1620; ¹H NMR (400MHz): 8.03 and 6.65 (2d, 2H, J_{6,7} = J_{8,9} = 7.5, C-6-H and C-9-H), 7.30 and 6.87 (2dd, 2H, J_{6,7} = J_{7,8} = J_{8,9} = 7.5, C-7-H and C-8-H), 7.53 (s, 1H, C-3-H), 7.52 (d, 1H, J_{12,13} = 15, C-12-H), 6.06 (d, 1H, J_{12,13} = 15, C-13-H), 5.34 (d, 1H, J_{10,11} = 5.6, N-10-H), 4.61 (d, 1H, J_{10,11} = 5.6, C-11-H), 4.30 (dd, 1H, J_{1a,11a} = 10.5, J_{1b,11a} = 4.5, C-11a-H), 3.38 (s, 3H, OCH₃), 3.16 (masked m, 1H, C-1-Ha), 3.11 (s, 3H, NMe), 3.05 (s, 3H, NMe), 2.87 (dd, 1H, J_{1a,1b} = 15.5 et J_{1b,11a} = 4.5, C-1-Hb); MS: 327 (M⁺, very weak), 295 (100%), 251, 249 (100%), 223, 120; HRMS: (M⁺ - CH₃OH) 295.1321, calcd. for C₁₇H₁₇N₃O₂: 295.1321. b) **3b**: mp inst. (dec.): 164-7 °C (MeOH-AcOEt); [α]_D = +622 (c = 0.2, CHCl₃), comparison of ¹H NMR, MS and HRMS data.⁷

(Received in France 29 April 1993; accepted 27 May 1993)