

Benzopyrans as selective estrogen receptor β agonists (SERBAs). Part 4: Functionalization of the benzopyran A-ring

Bryan H. Norman,^{a,*} Timothy I. Richardson,^a Jeffrey A. Dodge,^a Lance A. Pfeifer,^a Gregory L. Durst,^a Yong Wang,^a Jim D. Durbin,^a Venkatesh Krishnan,^b Sean R. Dinn,^c Shengquan Liu,^c John E. Reilly^c and Kendal T. Ryter^c

^aDiscovery Chemistry Research, Eli Lilly and Company, Lilly Corporate Center, Indianapolis, IN 46285, USA

^bMusculoskeletal Research, Eli Lilly and Company, Lilly Corporate Center, Indianapolis, IN 46285, USA

^cAlbany Molecular Research, Inc., 21 Corporate Circle, PO Box 15098 Albany, New York, NY 12212, USA

Received 20 April 2007; revised 4 July 2007; accepted 6 July 2007

Available online 13 July 2007

Abstract—Benzopyrans are selective estrogen receptor (ER) β agonists (SERBAs), which bind the ER receptor subtypes α and β in opposite orientations. We have used structure based drug design to show that this unique phenomena can be exploited via substitution at the 8-position of the benzopyran A-ring to disrupt binding to ER α , thus improving ER β subtype selectivity. X-ray cocrystal structures with ER α and ER β are supportive of this approach to improve selectivity in this structural class.

© 2007 Elsevier Ltd. All rights reserved.

Estrogen receptors (ERs, subtypes α and β) are nuclear hormone receptors with diverse tissue distribution and biological function.¹ The understanding of ER ligands, agonists and antagonists, is a fairly mature science, dating back over the past several decades.² More recently, the development of selective estrogen receptor modulators (SERMs) have impacted several disease targets, primarily in women's health.³ From a historical perspective, most of the ER pharmacology to date has focused on ER α . However, since its discovery in 1996,⁴ much effort has gone into understanding the relative importance of ER β .^{5,6} One important ER β responsive tissue is the prostate.⁷ We have recently described novel benzopyrans which are selective ER β agonists (SERBAs) displaying striking effects on rodent prostates in models of benign prostatic hyperplasia (BPH).⁸ As part of this program, we have tried to improve ER β selectivity through the use of structure based drug design.⁹ In this paper, we describe our approaches to selectivity improvements in the benzopyran A-ring.¹⁰

In recent years, we and others have utilized structure based drug design (SBDD) tools to assist our understanding of ER-ligand binding, with the hopes of

improving subtype specificity.^{11,12} Specifically, protein crystallography has been instrumental in providing information regarding the binding modes of ligands within the ligand binding pockets (LBP) of ER α and ER β . As we have already shown, benzopyrans, such as **1**, are unique, in that they bind to ER α and ER β with different orientations (a 180° rotation around the bis-phenol axis). This is illustrated in Figure 1, which shows an overlay of the X-ray cocrystal structures of **1**/ER α (blue) and **1**/ER β (green). The residues lining the LBP are also shown in the overlay, with phenol-binding residues and α versus β residue changes highlighted. It should be noted that the D-ring phenol binds within the Arg–Glu network in both ER α (Arg394–Glu353) and ER β (Arg346–Glu305). Similarly, the A-ring phenol forms a hydrogen bond to the imidazole ring in His525 in ER α (His475 in ER β). We and others have shown that these interactions are critical to the overall binding affinity of ER ligands. Structure modifications which compromise either interaction are expected to significantly diminish ER binding affinity.

The 'ligand rotation' phenomenon noted in Figure 1 provides a unique opportunity to improve ER β specificity, which does not focus completely on exploiting the conservative residue changes within the LBP (M421(α) \rightarrow I373(β); L384(α) \rightarrow M336(β)). Since the benzopyran scaffold already possesses very high affinity

Keywords: Estrogen receptor β ; Prostate; Crystallography.

* Corresponding author. Tel.: +1 317 277 2235; fax: +1 317 277 3652; e-mail: norman@lilly.com

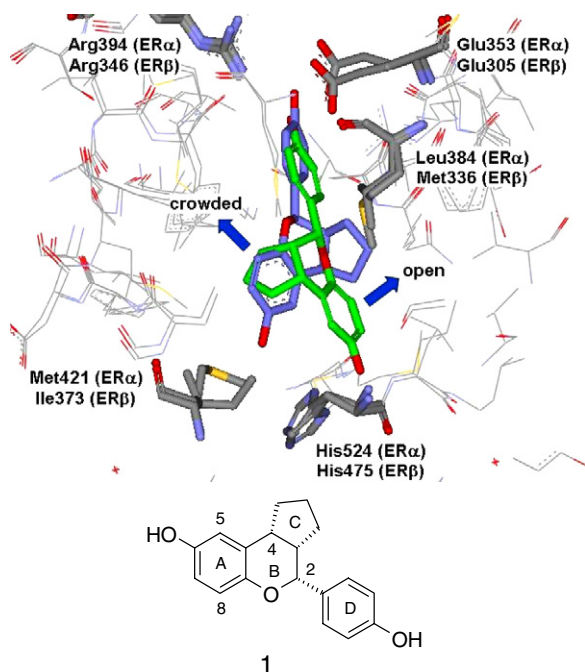
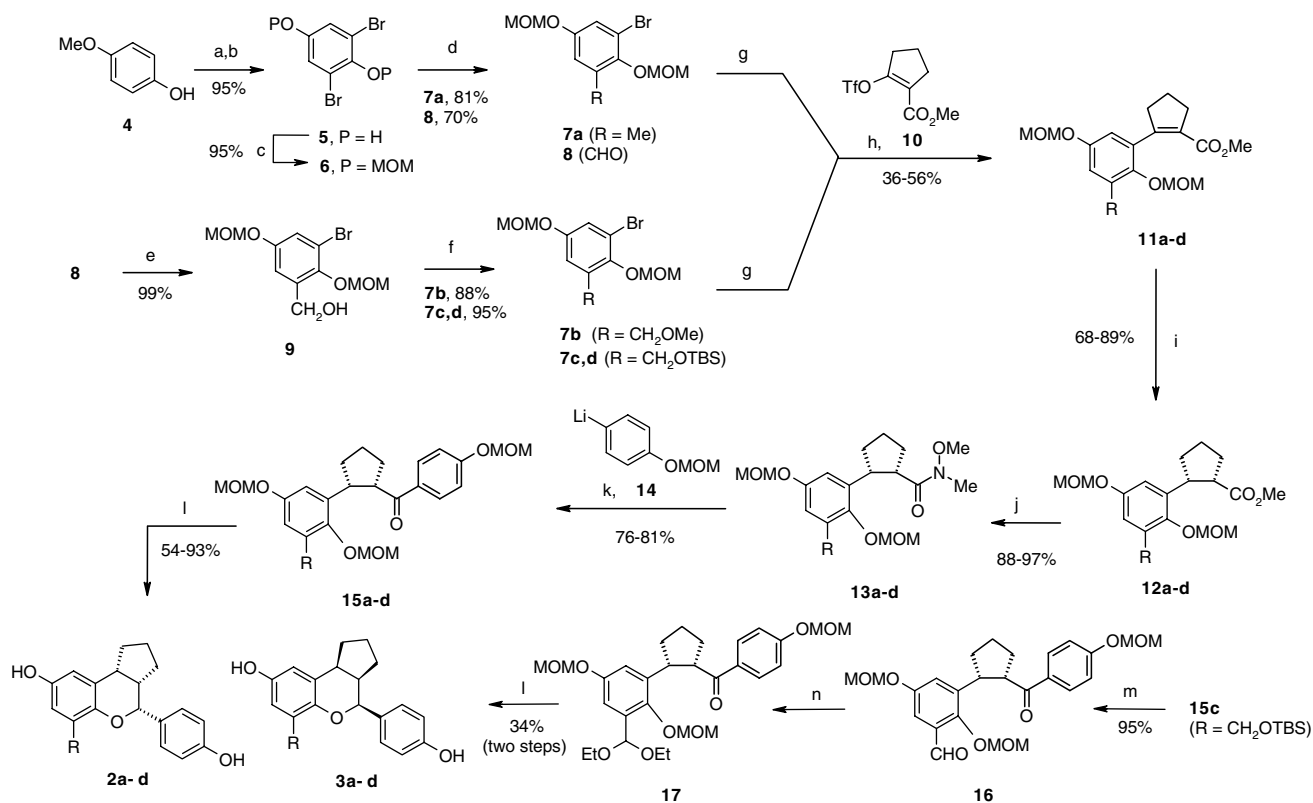


Figure 1. Overlay of 1 with ER α and ER β .

for ER β , the task of improving ER β selectivity becomes an exercise in selectively disrupting ER α binding. Figure 1

shows that substitution at the 8 position of the benzopyran should have minimal effect on ER β binding (note the 'open' area), as compared to the 8 position of the benzopyran in the ER α structure (crowded). When we prepared the 8-methyl substituted analog of 1, we observed no effect on binding affinity to either receptor subtype. Therefore, steric interactions alone are insufficient and it is always possible that the binding modes are altered when substitution at this position is made. Here we describe the effect of substitutions at the 8 position that do disrupt ER α binding to a greater extent than ER β binding, thereby improving ER β selectivity.

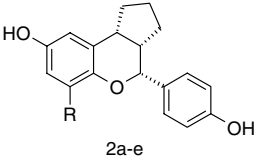
The synthesis of the 8-methyl substituted analogs took advantage of the reductive cyclization route reported earlier and is described in Scheme 1.^{8,13} 4-Methoxyphenol was treated with benzyltrimethylammonium tribromide to provide the dibromide. Demethylation with trimethylsilyl iodide gave hydroquinone 5, which was protected as the bis-methoxymethyl ether 6 using sodium hydride and chloromethyl methyl ether. Ortho mono-lithiation of protected hydroquinone 6 with *s*BuLi for phenyllithium for 8 followed by quenching of the anion with iodomethane or DMF provided 7a and 8, respectively. 8 could be elaborated to 7b and 7c via reduction to the benzyl alcohol 9, followed by alkylation with iodomethane (for 7b) or protection as the TBS ether (for 7c). A second ortho lithiation followed



Scheme 1. Reagents and conditions: (a) Benzyltrimethylammonium tribromide, CH₂Cl₂, MeOH; (b) TMSI, CH₃CN; (c) NaH, DMF, MOMCl; (d) *s*BuLi, THF; MeI for 7a or PhLi, THF; DMF for 8; (e) NaBH₄, EtOH; (f) NaH, DMF, MeI for 7b or TBSCl, DMF, imidazole for 7c; (g) i—PhLi, THF, ii—ZnCl₂; (h) 10, Pd(PPh₃)₄; (i) H₂, Pd/C, EtOH; (j) NH(OMe)Me, THF, *i*-PrMgCl; (k) 14, THF; (l) i—HCl, MeOH, NaBH₃CN, ii—Chiral separation; (m) i—TBAF, THF; ii—NaOCl, Bu₄NBr; (n) H₂, PtO₂, EtOH.

by coupling with enol triflate **10** using Negishi conditions gave the unsaturated esters **11a–c**. Unsaturated esters **11a–c** were hydrogenated over palladium on carbon to give **12a–c**, and then transformed into racemic Weinreb amides **13a–c**. The Weinreb amides **13** were reacted with lithiated *p*-bromophenyl methoxymethyl ether **14** to give racemic ketones **15a–c**. Deprotection and cyclization of ketones **15** under acidic conditions was followed by reduction with sodium cyanoborohydride under acidic conditions to give racemic benzopyrans **2** and **3** (a, b, and d analogs), which were separated into their individual enantiomers using chiral preparative HPLC. Benzopyrans **2c** and **3c** were prepared from **15c** under slightly different conditions. Specifically, **15c** was deprotected using TBAF and oxidized with bleach to give benzaldehyde **16**. This material was subjected to hydrogenation conditions in an attempt to selectively reduce the aldehyde in the presence of the ketone. However, these conditions did not result in reduction, but merely generated diethyl acetal **17**. Reductive cyclization of **17** using conditions described above gave ethoxymethyl benzopyrans **2c** and **3c**, which were separated into their individual enantiomers using chiral preparative HPLC. The *2R,3R,4R* enantiomer (isomer **2**, as drawn) was found to have the higher binding affinity for all analogs.

Table 1. A-ring modifications: ER α and ER β binding data



Compound	R	ER α (nM) K_i^a	ER β (nM) K_i^a	Ratio
1	H	2.29 (± 1.52)	0.17 (± 0.094)	14
2a	Me	2.82 (± 2.01)	0.26 (± 0.091)	11
2b	CH ₂ OMe	11.0 (± 5.64)	0.28 (± 0.02)	43
2c	CH ₂ OEt	27.2	0.745	36
2d	CH ₂ OH	147	5.97	25

^a Standard errors shown in parentheses. Others not tested in duplicate. Assay conditions described in Ref. 8.

The A-ring, position 8 effects on ER α and ER β binding are shown in Table 1. The addition of a methyl group at the 8-position caused no change in binding affinity to either receptor subtype; however, when larger, more polar groups were added, the ER α binding affinity decreased significantly. We attribute this finding to the limited space available (steric effect) and the nature of the surrounding residues in this area of the LBP, which are very hydrophobic (polarity effect). Perhaps ligands such as **2a** maintain good ER α binding due to the plasticity of the ER LBPs,¹⁴ which may accommodate hydrophobic substituents residing in the highly hydrophobic pocket. Alternatively, a larger, more polar group, such as alkoxymethyl or hydroxymethyl is tolerated in the hydrophobic LBP only when adequate space exists between the ligand polar groups and the hydrophobic residues. In this series, it appears that **2b** (methoxymethyl substitution) provides the best mix of polarity and steric bulk to disrupt ER α binding, while maintaining very good binding to ER β , thus improving ER β selectivity to 43-fold.

With these results in hand, we sought to test our hypothesis with X-ray crystal structures of **2b** with both ER α and ER β (Fig. 2).¹⁵ The similarity of binding modes, relative to **1**, is demonstrated by overlaying the structures of **2b** with the structures of **1** in both ER α and ER β . This result clearly shows (b) that **2b** binds to ER β in an identical manner, relative to **1**/ER β . However, as shown in Figure 2a, the methoxymethyl group in **2b** forces the A-ring out of its optimal position, thereby disrupting the important A-ring phenol-His524 hydrogen bonding interaction. We believe that this is responsible for the ~4-fold decrease in binding affinity of **2b**, relative to **1**.

In conclusion, we have shown that the introduction of A-ring substitution to the 8 position of benzopyrans results in diminished ER α binding, while maintaining strong ER β binding. We have shown that this is due to the fact that compounds in this class bind to the ER subtypes with different poses, and that substitution at the 8 position is less tolerated in the ER α LBP, relative to ER β . Finally, the 8-methoxymethyl shows improved binding selectivity (43-fold) in this class of selective ER β agonists.

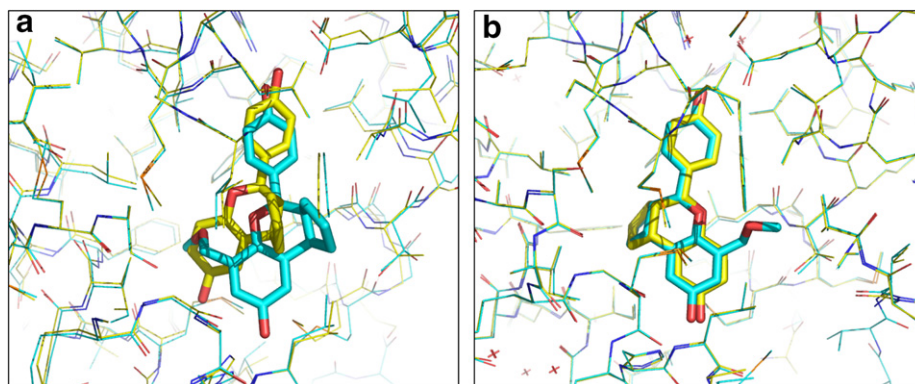


Figure 2. Overlay of **1** and **2b** with ER α (a) and ER β (b).

References and notes

1. Katzenellenbogen, J. A.; Katzenellenbogen, B. S. *Chem. Biol.* **1996**, *3*, 529.
2. (a) Ullrich, J. W.; Miller, C. P. *Exp. Opin. Ther. Pat.* **2006**, *16*, 559; (b) Wallace, O. B.; Richardson, T. I.; Dodge, J. A. *Curr. Top. Med. Chem.* **2003**, *3*, 1663; (c) Miller, C. P. *Curr. Pharm. Des.* **2002**, *8*, 2089.
3. Palkowitz, A. D.; Glasebrook, A. L.; Thrasher, K. J.; Hauser, K. L.; Short, L. L.; Phillips, D. L.; Muehl, B. S.; Sato, M.; Shetler, P. K.; Cullinan, G. J.; Pell, T. R.; Bryant, H. U. *J. Med. Chem.* **1997**, *40*, 1407.
4. (a) Mosselman, S.; Polman, J.; Dijkema, R. *FEBS Lett.* **1996**, *392*, 49; (b) Kuiper, G. G. J. M.; Enmark, E.; Pelto Huikko, M.; Nilsson, S.; Gustafsson, J. A. *Proc. Natl. Acad. Sci. U.S.A.* **1996**, *93*, 5925.
5. Harris, H. A.; Albert, L. M.; Leathurby, Y.; Malamas, M. S.; Mewshaw, R. E.; Miller, C. P.; Kharode, Y. P.; Marzolf, J.; Komm, B. S.; Winneker, R. C.; Frail, D. E.; Henderson, R. A.; Zhu, Y.; Keith, J. C. *Endocrinology* **2003**, *144*, 4241.
6. Hillisch, A.; Peters, O.; Kosemund, D.; Muller, G.; Walter, A.; Schneider, B.; Redderson, G.; Elger, W.; Fritzemeier, K. *Mol. Endocrinol.* **2004**, *18*, 1599.
7. McPherson, S. J.; Ellem, S. J.; Simpson, E. R.; Patchev, V.; Fritzemeier, K. H.; Risbridger, G. P. *Endocrinology* **2007**, *148*, 566.
8. Norman, B. H.; Dodge, J. A.; Richardson, T. I.; Borromeo, P. S.; Lugar, C. W.; Jones, S. A.; Chen, K.; Wang, Y.; Durst, G. L.; Barr, R. J.; Montrose-Rafizadeh, C.; Osborne, H. E.; Amos, R. M.; Guo, S.; Boodhoo, A.; Krishnan, V. *J. Med. Chem.* **2006**, *49*, 6155.
9. Richardson, T. I.; Norman, B. H.; Lugar, C. W.; Jones, S. A.; Wang, Y.; Durbin, J. D.; Krishnan, V.; Dodge, J. A. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 3570.
10. Richardson, T. I.; Dodge, J. A.; Durst, G. L.; Pfeifer, L. A.; Shah, J.; Wang, Y.; Durbin, J. D.; Krishnan, V.; Norman, B. H. *Bioorg. Med. Chem. Lett.*, in press, doi:10.1016/j.bmcl.2007.06.052.
11. Sun, J.; Baudry, J.; Katzenellenbogen, J. A.; Katzenellenbogen, B. S. *Mol. Endocrinol.* **2003**, *17*, 247; Compton, D. R.; Sheng, S.; Carlson, K. E.; Rebacz, N. A.; Lee, I. Y.; Katzenellenbogen, B. S.; Katzenellenbogen, J. A. *J. Med. Chem.* **2004**, *47*, 5872.
12. Manas, E. S.; Unwalla, R. J.; Xu, Z. B.; Malamas, M. S.; Miller, C. P.; Harris, H. A.; Hsiao, C.; Akopian, T.; Hum, W. T.; Malakian, K.; Wolfrom, S.; Bapat, A.; Bhat, R. A.; Stahl, M. L.; Somers, W. S.; Alvarez, J. C. *J. Am. Chem. Soc.* **2004**, *126*, 15106; Mewshaw, R. E.; Edsall, R. J.; Yang, C.; Manas, E. S.; Xu, Z. B.; Henderson, R. A.; Keith, J. C., Jr.; Harris, H. A. *J. Med. Chem.* **2005**, *48*, 3953.
13. Experimental details have been described in Ref. 8 (suppl. material), WO 2004094401 and WO 2003044006.
14. Pike, A. C. W.; Brzozowski, A. M.; Hubbard, R. E.; Bonn, T.; Thorsell, A.; Engström, O.; Ljunggren, J.; Gustafsson, J.; Carlquist, M. *EMBO* **1999**, *18*, 4608.
15. Atomic coordinates for ER α and ER β complexes with **2b** have been deposited into the Protein Data Bank, with accession codes of 2qe4 and 2jj3, respectively.