

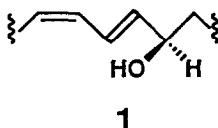
A CONCISE SYNTHESIS OF (*R*)-HYDROXY-*E,Z*-DIENE FATTY ACIDS:
PREPARATION OF 12(*R*)-HETE, TETRA α NOR-12(*R*)-HETE, AND
13(*R*)-HODE

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Summary: Chiral *E*-enals derived from functionalized 2-deoxy-D-ribofuranoses by ylide-induced β -elimination were exploited for the synthesis of fatty acid metabolites containing the (*R*)-hydroxy-*E,Z*-diene subunit.

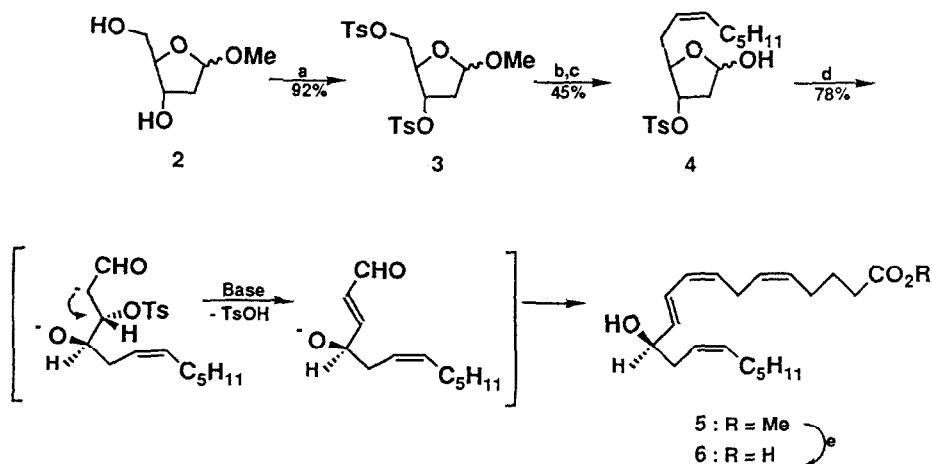
The enzymatic conversion of polyunsaturated fatty acids to conjugated *E,Z*-dienols, e.g., arachidonic and linoleic acids to hydroxyeicosatetraenoic (HETE) and hydroxyoctadecadienoic (HODE) acids, respectively, is a prominent pathway to bioactive metabolites in plants and animals¹. While the *S*-antipode generally prevails, recent studies have revealed several *R*-stereospecific systems². Moreover, the *R*-series metabolites frequently display pharmacologic profiles that are qualitatively and/or quantitatively different from their *S*-counterparts. 12(*R*)-HETE (**6**), for instance, is a more potent chemotactic and chemokinetic factor than 12(*S*)-HETE³; additionally, it inhibits Na⁺/K⁺-ATPase activity in mammalian tissue whereas the 12(*S*)-enantiomer does not⁴. As endogenous constituents of the cornea, **6** and its metabolite **9** have been postulated to have a regulatory role in ocular function⁵. Likewise, both enantiomers of 13-HODE are naturally occurring⁶. The *S*-isomer has attracted considerable attention as an inhibitor of platelet and cancer cell adhesion to endothelium⁷ and as a self-defense substance against rice-blast disease⁸. However, comparatively little is known about 13(*R*)-HODE (**11**), due in part to its limited availability from natural sources.



To satisfy the urgent need for sufficient amounts of enantiomerically homogenous standards⁹ for biological evaluation, we describe herein a convergent strategy for the preparation of fatty acid metabolites containing the (*R*)-hydroxy-*E,Z*-diene subunit **1**¹⁰ and illustrate its versatility in concise syntheses of **6**, **9**, and **11**.

Methyl furanoside **2**, obtained¹¹ as an anomeric mixture from commercial 2-deoxy-D-ribose (95%), was readily converted to lactol **4**¹² via selective coupling of the bis-tosylate **3**¹³ with the higher order cuprate generated¹⁴ from (*Z*)-1-iodo-1-heptene (**7**) in Et₂O and subsequent mild acid hydrolysis (Scheme I). Creation of the *E,Z*-diene exploited a facile, ylide-induced elimination of tosylate from the open-chain tautomer of **4** under the conditions utilized for Wittig olefination. *In situ* condensation of the resultant *E*-enal with 7-carbomethoxyhepta-3(*Z*)-en-1-ylidenetriphenylphosphorane¹⁴ (**8**) furnished 12(*R*)-HETE methyl ester (**5**), identical in all respects with an authentic sample¹⁵. Saponification afforded the corresponding free acid **6**.

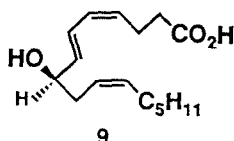
Scheme I



^aTsCl, C₅H₉N/CH₂Cl₂ (4:5), 0°C, 12 h. ^b**7**, BuLi, Et₂O, -40°C, 20 min; CuCN, -40°C ~ 0°C, 4h.

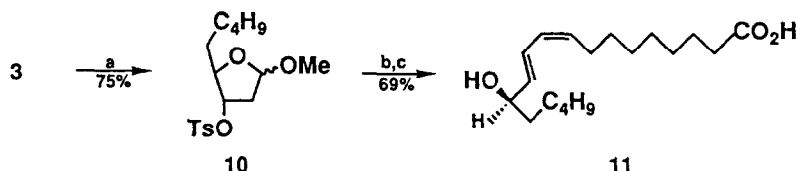
^cAcOH/THF/H₂O (2:1:1), 70°C, 8h. ^d**8** (3 equiv), THF/HMPA (10:1), -20°C, 1h. ^eLiOH, MeOH, 24°C, 4h; HCl.

Analogous treatment of **4** with 3-carboxypropylidenetriphenylphosphorane¹⁶ (3.5 equiv) yielded 8(*R*)-hydroxyhexadeca-4(*Z*),6(*E*),10(*Z*)-trienoic acid (**9**) as a colorless oil. Its chromatographic (capillary GC, HPLC) and mass spectroscopic characteristics were identical with the major metabolite of 12(*R*)-HETE produced by freshly isolated corneal tissue and cells in culture⁵.



Extension of the general theme to 13(*R*)-HODE (**11**) required homologation of **3** to tosylate **10** with dibutyl copper lithium in Et₂O (Scheme II). Successive methyl lactol hydrolysis, generation of the corresponding *E*-enal as above, and *in situ* olefination using 8-carboxyoctylidenetriphenylphosphorane^{16,17} (**12**) smoothly evolved **11** whose spectral and chromatographic properties were comparable with a 13(*S*)-standard¹⁷.

Scheme II



^aBuLi, CuI, Et₂O, -40°C, 3h. ^bAcOH/THF/H₂O (1:1:1), 70°C, 12h. ^c**12** (4 equiv), THF/HMPA (10:1), -20°C, 30 min.

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12. Satisfactory spectral data were obtained for all new compounds using chromatographically homogeneous samples.
13. ¹H NMR (CDCl₃, 250 MHz) of 3 (less polar isomer): δ 2.16-2.24 (m, 2H), 2.50 (s, 6H), 3.23 (s, 3H), 3.92 (dd, J=6, 10 Hz, 1H), 3.97 (dd, J=5.0, 10 Hz, 1H), 4.17-4.24 (m, 1H), 4.86-4.94 (m, 1H), 5.03 (dd, J=3, 4.5 Hz, 1H), 7.36 (d, J=7 Hz, 4H), 7.75 (d, J=7 Hz, 4H). 3 (more polar isomer): δ 2.10-2.22 (m, 2H), 2.50 (s, 6H), 3.30 (s, 3H), 3.96 (dd, J=1.5, 3 Hz, 1H), 4.13 (dd, J=1.5, 6 Hz, 1H), 4.30-4.36 (m, 1H), 4.78-4.84 (m, 1H), 4.96 (d, J=6 Hz, 1H), 7.36 (d, J=7 Hz, 4H), 7.75 (d, J=7 Hz, 4H). 4 (less polar isomer): δ 0.90 (t, J=7 Hz, 3H), 1.20-1.48 (m, 6H), 1.88-2.06 (m, 2H), 2.13-2.32 (m, 4H), 2.46 (s, 3H), 3.34 (s, 3H), 4.12-4.20 (m, 1H), 4.54-4.63 (m, 1H), 4.87 (dd, J=1.5, 5 Hz, 1H), 5.16-5.30 (complex m, 1H), 5.39-5.52 (complex m, 1H), 7.32 (d, J=7 Hz, 2H), 7.79 (d, J=7 Hz, 2H). 4 (more polar isomer): δ 0.90 (t, J=7 Hz, 3H), 1.20-1.48 (m, 6H), 1.88-2.08 (m, 2H), 2.16-2.38 (m, 4H), 2.47 (s, 3H), 3.36 (s, 3H), 4.00-4.07 (m, 1H), 4.62-4.79 (m, 1H), 5.05 (d, J=5 Hz, 1H), 5.16-5.30 (complex m, 1H), 5.40-5.52 (complex m, 1H), 7.37 (d, J=7 Hz, 2H), 7.80 (d, J=7 Hz, 2H). 9 methyl ester: δ 0.89 (t, J=7 Hz, 3H), 1.20-1.42 (m, 6H), 1.55-1.72 (m, 2H), 1.93-2.11 (m, 2H), 2.29-2.59 (m, 4H), 3.68 (s, 3H), 4.15-4.28 (m, 1H), 5.30-5.67 (complex m, 3H), 5.73 (dd, J=6, 16 Hz, 1H), 6.02 (apparent t, J=11 Hz, 1H), 6.54 (dd, J=11, 15 Hz, 1H). 10 (less polar isomer): δ 0.90 (t, J=7 Hz, 3H), 1.16-1.45 (m, 6H), 1.68-2.23 (m, 4H), 2.45 (s, 3H), 3.35 (s, 3H), 4.12-4.25 (m, 1H), 4.53-4.62 (m, 1H), 4.84 (dd, J=1.5, 5 Hz, 1H), 7.35 (d, J=7 Hz, 2H), 7.75 (d, J=7 Hz, 2H).
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