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Abstract: The 8-membered ring framework of vinigrol, a unique tricyclic diterpene isolated as a novel antihypertensive compound from a culture of Virgaria nigra, was efficiently synthesized employing an SmI2induced intramolecular Barbier coupling.

Vinigrol (1), an unusual tricyclic diterpenoid, was isolated from a culture broth of the fungal strain identified as Virgaria nigra by Ando et al., who also described the antihypertensive and platelet aggregation inhibitory properties of 1.1 Furthermore, it was found that 1 is a tumor necrosis factor (TNF) antagonist.2 Therefore, 1 may be used to control conditions attributable to TNF such as endotoxic shock, inflammation, infection, cachexia, and the progression from the AIDS-related complex to AIDS. It features a complicated fused ring system involving an 8membered ring. Its interesting physiological activity and unique structure distinguish the molecule as a very interesting target for total synthesis.3 In conjunction with our program directed toward the total synthesis of 1, we synthesized the 8-6 fused ring system of this diterpene through the efficient cyclization of the 8-membered ring using an SmI₂-induced Barbier coupling. 4-6 Natural products possessing 8-membered ring substructural units have attracted much attention because of their potent biological activity and the synthetic challenge posed by the required formation of the carbocycle or heterocycle.

The stereoselective cross aldol reaction of 3-benzyloxypropionaldehyde (3) with the Li-enolate generated from (+)-chlorodihydrocarvone (2)⁸ provided the hydroxy ketone 4, which upon treatment with 2-fluoropyridinium tosylate gave the (E)- α -enone 5 (Scheme I). The 1,2-addition of CH₂=CHCH₂MgBr to 5 occurred in a stereoselective manner and the βalcohol 6 was obtained along with a small amount of its α -epimer (α : β = 5:1). The β-alcohol 6 was converted into the aldehyde 9 in the following sequence, (1) protection of the hydroxyl group of 6 as its MOM ether, (2) regioselective hydroboration-oxidation of the terminal olefin of the MOM ether 7 by thexylborane, and (3) Dess-Martin oxidation of the resulting alcohol 8.

Treatment of 9 with SmI₂ in the presence of HMPA cleanly allowed the 8-membered ring closing reaction to furnish the cyclooctanol 109 relevant to the 8-6 fused ring system of vinigrol (1) as the only detectable product. The quantitative yield of 10 is remarkable in view of the difficulties normally encountered during the cyclization of 8-membered rings. The addition of HMPA was essential for this SmI2-promoted intramolecular Barbier coupling. 10 The reaction in the absence of HMPA produced a substantial decrease in product yield (15%). The major isolated product was the reduced alcohol 8.

The structure of 10 was confirmed by 2D-COSY and 2D-NOESY experiments on the acetate 119 prepared from 10 through acetylation (Ac₂O, Py, rt, 100%). The stereochemical assignment to 11 definitely follows from inspection of the selected key coupling constants and NOE's as illustrated in Figure I.

The synthesis of the 8-6 fused ring skeleton of vinigrol (1) described here is noteworthy for the high efficiency with which the 8-membered ring is assembled. The protocol should be applicable to the construction of various 8-membered rings.

X=H, Y=CH=CH₂ X=CH₂OMe, Y=CH=CH₂ 8 X=CH₂OMe, Y=CH₂CH₂OH

(a) (1) LDA, THF, -78 °C. (2) 3, -78 °C. (b) FC₅H₅NMe•OTs, Et₃N, CH₂Cl₂, reflux. (c) CH₂=CHCH₂MgBr, ether, -78 °C. (d) MeOCH₂Cl, ⁱPr₂NEt, CH₂Cl₂, rt. (e) (1) ThexylBH₂, THF, 0 °C. (2) 30% H₂O₂, rt. (f) Dess-Martin Periodinane, Py, CH₂Cl₂, rt.

Scheme I

NOE H OMe

H₃C

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Figure I

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- Physical and ¹H-NMR spectral data for 10 and 11 are as follows. **10**: $[\alpha]_D^{25}$ -80.9° (c 0.915, CHCl₃); ¹H-NMR (CDCl₃) δ 0.81 (3H, d, J = 6.9 Hz, C_8 -Me), 1.31 (1H, brd, J = 12.3 Hz, C_{9eq} -H), 1.44 (1H, brdq, J = 6.5, 11.1 Hz, C_{5ax} -H), 1.74 (1H, brdd, J = 11.1, 14.5 Hz, C_{6ax} -H), 2.45 (1H, dq, J = 14.2, 7.1 Hz, C_{2} -H), 2.47 (1H, dd, $J = 4.0, 11.1 \text{ Hz}, C_{3eq}$ -H), 2.51 (1H, dq, $J = 14.2, 7.1 \text{ Hz}, C_{2}$ -H), 3.37 (3H, s, OMe), 3.42 (1H, brs, C₁-H), 3.59 (3H, m, C₄-H, C₃- H_2), 4.49 (1H, d, J = 7.5 Hz, CHOMe), 4.53 (2H, s, PhC H_2), 4.77 (1H, d, J = 7.5 Hz, CHOMe), 4.97, 4.99 (each 1H, brs, C₂=CH₂),5.98 (1H, t, J = 7.3 Hz, C_1 '-H), 7.25 - 7.38 (5H, m, Ph). 11: $[\alpha]_D^{25}$ -60.6° (c 0.930, CHCl₃); ¹H-NMR (CDCl₃) δ 0.81 (3H, d, J = 6.9 Hz, C_8 -Me), 1.31 (1H, brd, J = 12.5 Hz, C_{9eq} -H), 1.52 (1H, brdq, J = 6.5, 11.0 Hz, C_{5ax} -H), 1.58 – 1.72 (3H, m, C_{5eq} -H, C_{9ex} -H, C_{10} -H), 1.76 (1H, brdd, J = 11.0, 15.1 Hz, C_{6ax} -H), 1.90 (1H, brdd, J = 6.5, 15.1 Hz, C_{6eq} -H), 1.95 (1H, t, J = 11.0 Hz, C_{3ax} -H), 1.97 (3H, s, Ac), 2.02 (1H, m, C_{8} -H), 2.44 (1H, dq, J =14.2, 7.1 Hz, $C_{2'}$ -H), 2.48 (1H, dd, J = 4.2, 11.0 Hz, C_{3eq} -H), 2.54 (1H, dq, J = 14.2, 7.1 Hz, $C_{2'}$ -H), 3.36 (3H, s, OMe), 3.43 (1H, brs, C_1 -H), 3.58 (2H, m, C_3 -H₂), 4.47 (1H, d, J = 7.5 Hz, CHOMe), 4.55 (2H, s, PhC H_2), 4.71 (1H, tt, J = 4.2, 11.0 Hz, C_4 -H), 4.76 (1H, d, J = 7.5 Hz, CHOMe), 5.05, 5.10 (each 1H, brs, $C_2=CH_2$), 5.96 (1H, t, J = 7.1 Hz, C_1 -H), 7.25 – 7.38 (5H, m, Ph).
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