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STEREOSELECTIVE TOTAL SYNTHESIS OF THE BLOOD GROUP I-ACTIVE BIANTENNARY NEOLACTO-GLYCODECAOSYL CERAMIDE¹

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Abstract: A stereocontrolled total synthesis of biantennary *neolacto*-glycodecaosyl ceramide was achieved for the first time.

In 1981, Hanfland² et al. reported isolation and characterization of an I-active glycodecaosyl ceramide 3. Because of the biological significance³ as well as the structural diversity⁴ of I-active glycosyl ceramides, we started our project on their synthesis and in 1986 reported the first synthesis⁵ of the octasaccharide 1. Both chemical⁶ and enzymic⁷ synthesis of closely related I-active hexasaccharide were reported independently in 1986. Here we describe for the first time a total synthesis of both glycooctaosyl ceramide 2 and glycodecaosyl ceramide 3. Retrosynthetic analysis of 2 and 3 shown in scheme 1 led us to design two glycosyl donors 10⁸ and 11, and a glycosyl acceptor 12. The glycosyl donor 11 was synthesized from the coupling of either 13 or 14 with 15, while the key glycotetraosyl glycosyl acceptor 12 designed as a common intermediate for the synthesis of both 2 and 3 was synthesized via the regioselective glycosylation of the tetraol 17 with the imidate 16.

The fluoride 13^9 was readily obtainable from 18^{10} via 19^9 in three steps (1 BnBr, NaH in DMF, 2 CAN¹¹ in 4:1 CH₃CN-H₂O, 0°, 30 min, 3 DAST¹² in (ClCH₂)₂, 0°, 30 min, 64% overall). The glycotriosyl donors 24-26 were prepared as follows. Compound 21^{13} was converted into 15^9 via 22^9 in 4 steps (1 LiOH, 30% H₂O₂ in THF¹⁴, 2 Ag₂O, KI, BnBr in DMF, 3 [Ir(COD)(PMePh₂)₂]PF₆ in THF¹⁵, then I₂ in 4:1 THF-H₂O, 4 AcCl in Py at 0°, 43% overall). Glycosylation of 15 with fluoride 13 in the presence of SnCl₂¹⁶, AgClO₄ and molecular sieve 4A (MS4A) in Et₂O at -20° ~10° gave 80% of 23⁹ together with 12% of the β-anomer. Use of the thioglycoside 14¹⁷, however, improved





the stereoselectivity. Reaction of $14(\beta)$ with 15 in the presence of CuBr₂-nBu₄NBr-MS4A¹⁸ in 5:1 (ClCH₂)₂-DMF gave 95% of 23 with the concomitant formation of only 2% of the β -anomer. Compound 23 was transformed into trichloroacetimidate 24⁹ in two steps (*I* CAN in 25:19:12 CH₃CN-MePh-H₂O, 25°, 20

min, 2 CCl₃CN¹⁹ and DBU in (ClCH₂)₂, 0°, 50 min, overall 78%). Treatment of 24 with Bu₃SnSMe and BF₃•OEt₂ in (ClCH₂)₂ at -23° for 80 min gave 95% of thioglycoside 25⁹. Conversion of 23 into fluoride 26⁹ was carried out in 2 steps as described in the preparation of 13 in 77% overall. Now glycosyl donors designed in scheme 1 being available, synthesis of key glycosyl acceptor 12 was performed as follows. Partial acylation of 27²⁰ with ^tBuCOCl-DMAP-Et₃N in (ClCH₂)₂ at -5° afforded 51% of 28⁹ and 27% of 29, which were treated with TsOH in 1:1 dioxane-MeOH at 50° afforded 17⁹ (92%) and 30⁹ (94%), respectively. The trichloracetimidate 16⁹ was prepared from 22 via 31 in 3 steps (*l* Lev₂O, DMAP in 4:3 Py-(ClCH₂)₂, *2* CAN in 4:3:2 CH₃CN-MePh-H₂O, 20°, 1h, *3* Cl₃CCN, DBU in (ClCH₂)₂, 0°, 1h, 82% overall). TMSOTf promoted glycosylation of 17 with 16 in 2:1 PhMe-(ClCH₂)₂ at -23° afforded 72% yield of the $\beta 1 \rightarrow 3$ linked compound 32⁹ as a main product along with 13% yield of the minor regioisomers. Conversion of 32 into 12⁹ was carried out via 33 in 2 steps (*l* Ac₂O in Py, 2 NH₂NH₂•AcOH in EtOH, 20°, 50 min, 81% overall).



TMSOTf promoted glycosylation of 12 with trichloroacetimidate 10 at -23° proceeded to give 63% of the desired compound 34⁹ which was subsequently converted to the glycooctaosyl donor 35⁹ in 4 steps (1 Pd(OH)₂; H₂

in MeOH, 2 Ac₂O, DMAP in Py, 3 NH₂NH₂·AcOH in DMF, 50°, 20 min, 4 Cl₃CCN, DBU in (ClCH₂)₂, 71% overall). Coupling between 35 and 6^{21} in CHCl₃ in the presence of TMSOTf and MS4A afforded 59% of 36^9 which was converted into glycooctaosyl ceramide 2^9 in 3 steps (1 40% MeNH₂²² in MeOH, 20°, 40h, 2 Ac₂O, DMAP in Py, 50°, 16h, 3 MeONa in 1:1 MeOH-THF, 65°, 16h; 73% overall). ¹H-nmr data for synthetic 2 was in complete agreement with that for natural sample²³.



Next, we studied the coupling reactions of glycotriosyl donors 24, 25, and 26 with glycotetraosyl acceptor 12. TMSOTf promoted glycosylation of 12 with trichloroacetimidate 24 in (CICH₂)₂ at -23° gave only 23% of 37⁹. Similarly glycosylation of 12 with thioglycoside 25 in the presence of CuBr2-Bu4NBr-AgOTf afforded only 16% of 37. Best result was obtained by employing fluoride 26, which was reacted with 12 in the presence of Cp2HfCl2²⁴ and AgOTf in (C1CH₂)₂ at -23° to give 88% of 37. Conversion of 37 into 38⁹ was achieved as described for 35 in 4 steps in 58% overall. Crucial coupling between 38 and 6 was achieved in the presence of TMSOTf and MS4A in CHCl₃ at -23° to give 54% of 39^9 which was further converted into the target molecule 3 as described for 2 in 3 steps in 56% overall yield. ¹H-nmr data for synthetic 3 were in complete agreement with those for natural 3 reported by Hanfland et al^2 .



In conclusion, I-active glycooctaosyl and glycodecaosyl ceramides 2 and 3 were synthesized for the first time by employing glycotetraosyl acceptor 12 as a key intermediate and trichloroacetimidates 35 and 38 as the glycosyl donors for the crucial coupling with ceramide derivative 6.

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References and Notes

- 1 Part 84 in the series "Synthetic Studies on Cell-Surface Glycans". For part 83, see C. Murakata and T. Ogawa, Carbohydr. Res. submitted.
- 2 P. Hanfland, H. Egge, U. Dabrowski, S. Kuhn, D. Roelcke, and J. Dabrowski, Biochemistry, 20, 5310 (1981).
- T. Feizi and R.A. Childs, Biochem.J., 245,1(1987); D, Roelcke, Transfusion Med. Rev., 3, 140(1989).
- 4 S. Hakomori, K. Watanabe and R. A. Laine, Pure and Appl. Chem., 49, 1215(1977); M. E. Breimer, G. C. Hansson, K.-A. Karlsson, H. Leffler, W. Pimlott, and B. E. Samuelsson, FEBS Lett., 124,299 (1981); M. N. Fukuda and S. Hakomori, J. Biol. Chem., 257,446(1982); P. Hanfland, M. Kordowicz, H. Niermann, H. Egge, U. Dabrowski, J. Peter-Katalinic, and J. Dabrowski, Eur. J. Biochem., 145, 531(1984); H. Clausen, S. B. Levery, E. D. Nudelman, M. Boldwin, and S. Hakomori, Biochem. 25, 7075(1986); S. B. Levery, E. D. Nudelman, M. E. K. Salyan, and S. Hakomori, ibid., 28,7772(1989).
- Y. Ito and T. Ogawa, Agric. Biol. Chem., 50, 3231 (1986).
- A. Maranduba and A. Veyrières, Carbohydr. Res., 151, 105 (1986). 6
- 7
- C. Augé, C. Mathien, and C. Mérienne, Carbohydr. Res., 151, 145 (1986). R. R. Schmidt and G. Grundler, Angew. Chem. Int. Ed. Engl., 22, 776 (1986). 8
- Physical data for new compounds are given below, values of $[\alpha]_D$ and $\delta_{H,C}$ were measured at 25°±3° for solutions in CHCl3 and CDCl3, respectively, unless noted otherwise. 2: RF 0.34 in 6:4:1 CHCl₃-MeOH-H₂O; $[\alpha]_D$ +10.0° (c 0.1, py); δ_H (49:1:1 DMSOd₆-D₂O-CD₃OD, 60°) 1.828 (s, Ac), 1.846 (s, 2Ac), 4.180 (d, 7.7Hz, 1¹), 4.237 (d, 6.6Hz, 1⁶ and 1^{6'}), 4.285 (d, 7.0Hz, 1²), 4.318 (d, 7.0Hz, 14), 4.434 (d, 8.1Hz, 15'), 4.688 (d, 8.1Hz, 13 and 15), 5.380 (dd, 7.0 and 15.4Hz, 4Cer), 5.563 (td, 7.0 and 15.4Hz, 5^{Cer}). 3: R_F 0.32 in 6:4:1 CHCl₃-MeOH-H₂O; [α]_D +39.5° (c 0.1, py); δ_H (49:1:1 DMSOd₆- $D_2O, 60^\circ$) 0.855 (t, 6.8Hz, 2Me), 1.831 (s, Ac), 1.850 (s, 2Ac), 4.175 (d, 7.3Hz, 1¹), 4.282 (d, 6.2Hz, 1.850 (s, 2Ac)), 4.175 (d, 7.3Hz, 1.850 (s, 2Ac)), 4.175 (s, 2Ac)) 12), 4.308 (d, 7.3Hz, 14, 16, and 16'), 4.432 (d, 7.3Hz, 15'), 4.689 (d, 8.1Hz, 13 and 15), 4.855 (d, 3.3Hz, 17 and 17'), 5.374 (dd, 7.0 and 15.4Hz, 4^{Cer}), 5.557 (td, 7.0 and 15.4Hz, 5^{Cer}). 12: RF 0.39 in

1:1 toluene-EtOAc; [a]D +1.5° (c 0.1); &H 1.080, 1.190, and 1.217 (3s, 3Piv), 1.688, 1.900, and 2.091 (35, 3Ac), 4.173 (d. 8.1Hz, 1²), 5.121 (d. 8.4Hz, 1³), 5.338 (d. 3.7Hz, 4²). 13: 8H 5.166 (dd. 0.5H. 7.0 and 53.1Hz, 1 β), 5.588 (dd, 2.5 and 53.5Hz, 1 α), for the alternative synthesis see ref. 25. 15: R_F 0.26 in 3:2 hexane-EtOAc; $[\alpha]_D$ +41.5° (c 1.0); δ_H 2.004 (s, Ac), 3.696 (s, OMe), 4.468 (d, 7. Hz, 1²), 5.623 (d, 8.6Hz, 1^{1}), 16; R_F 0.43 in 2:1 toluene-EtOAc; $\delta_{\rm H}$ 2.147 and 2.184 (2s, 2Lev), 6.399 (d, 8.4Hz, 1¹), 8.533 (s, NH), 17; R_F 0.40 in 10:1 CHCl₃-MeOH; [α]_D -11.2° (c 1.0); mp 124-125° (iPr₂O); $\delta_{\rm H}$ 1.171, 1.184, and 1.240 (3s, 3Piv), 4.293 (d, 8.1Hz, 1²), 4.476 (d, 8.1Hz, 1¹), 4.920 (dd, 8.1 and 9.9Hz, 2¹), 19: RF 0.60 in 3:1 hexane-EtOAc; [α]₁₀ -17.6° (c 1.1); δH 3.738 (OMe). 22: RF 0.33 in 3:2 toluene-EtOAc, $[\alpha]_D$ +44.2° (c 1.3); δ_H 4.459 (d, 7.3Hz, 1²), 5.638 (d, 8.2Hz, 1¹). 23: R_F 0.33 in 4:1 toluene-EtOAc; $[\alpha]_D$ +50.7° (c 0.9); δ_H 1.955 (s, Ac), 3.684 (s, OMe), 4.465 (d, 7.7Hz, 1²), 5.217 (d, 3.3 Hz, 1³), 5.575 (d, 8.4 Hz, 1¹). 24: R_F 0.53 in 4:1 tolucne-EtOAc; [α]_D +56.4° (c 1.0); δ_H 1.962 (s, Ac). 5.202 (d, 3.3 Hz, 1^3), 6.363 (d, 7.8Hz, 1^1), 8.505 (s, NH). 25: RF 0.55 in 3:1 toluene-EtOAc; $[\alpha]_D$ +45.1° (c 1.1); $\delta_{\rm H}$ 1.862 (s, SMc), 1.940 (s, Ac), 5.089 (d, 9.9Hz, 1¹), 5.209 (d, 2.9Hz, 1³). 26: R_F 0.56 in 4:1 toluene-EtOAc; $\delta_{\rm H}$ 1.952 (s, Ac), 5.205 (d, 3.3Hz, 1³), 5.600 (dd, 0.08H, 3.5 and 54.0 Hz, 1¹ α), 5.800 (dd, 0.92H, 7.5 and 53.5 Hz, 1¹B). 28: R_F 0.42 in 2:1 toluene-EtOAc; $[\alpha]_D$ +5.6° (c 1.2); δ_H 1.175, 1.192, and 1.236 (3s, 3Piv), 1.335 and 1.510 (2s, CMe2), 4.229 (d, 8.1Hz, 1²), 4.451 (d, 8.1Hz, 1^{1} , 4.930 (dd, 8.1 and 9.5Hz, 2^{1}), 30; R_F 0.45 in 1:1 toluene-EtOAc; $[\alpha]_{T}$ -8.1° (c 0.9); δ_{H} 1.175, 1.201, 1.217 and 1.235 (4s. 4Piv), 4.907 and 4.990 (2dd, 8.1 and 9.5Hz, 2¹ and 2²), 31: RF 0.60 in 1:1 toluene-EtOAc; mp 112-113° (Et2O-iPr2O); [a]D +47.9° (c 0.9); bH 2.146 and 2.192 (2s, 2Lev), 3.692 (s, OMe), 4.499 (d, 7.7Hz, 1²), 4.854 (dd, 2.8 and 9.9Hz, 3²), 5.609 (d, 8.1Hz, 1¹). 32: R_F 0.40 in 1:1 toluene-EtOAc; [α]_D +20.2° (c 1.3); δ_H 1.132, 1.143, and 1.146 (3s, 3Piv), 2.157 and 2.184 (2s, 2Lev), 5.338 (d. 8.8Hz, 1³), 34: RF 0.41 in 1:1 toluene-EtOAc; [a]D -10.3° (c 1.1); bH 1.069, 1.171, and 1.191 (3s, 3Piv), 1.629, 1.860, 1.877, 1.901 1.918, 1.963, 1.981, 2.028, 2.029, 2.036, 2.051, 2.055, 2.128, 2.138, and 2.149 (15s, 15Ac), 5.449 (d, 8.4Hz, 1⁵), 5.652 and 5.703 (2dd, 8.8 and 10.6 Hz, 3⁵ and 3^{5'}). 35: $R_F 0.60$ in 1:3 toluene-EtOAc; $[\alpha]_D + 23.8^{\circ}$ (c 0.4); $\delta_H 1.089$, 1.192, and 1.211 (3s, 3Piv), 1.693, 1.756, 1.771, 1.869, 1.885, 1.926, 1.943, 1.970, 1.973, 2.036, 2.040, 2.064, 2.083, 2.092, 2.098, 2.105, 2.134, 2.134, and 2.159 (18s, 19Ac), 5.416 (d, 8.4Hz, 1⁵), 5.542, 5.636, and 5.653 (3dd, 8.5 and 10.0Hz, 3³, 3⁵, and 3^{5'}), 6.441 (d, 3.7Hz, 1¹), 7.65-7.90 (m, 3Phth), 8.591 (s, NH). 36: R_F 0.52 in 2:1 CCl₄-Me₂CO; [a]_D +9.8° (c 0.6); $\delta_{\rm H}$ 0.878 (t, 7.0Hz, 2Me), 1.097, 1.129, and 1.218 (3s, 3Piv), 1.686, 1.740, 1.756, 1.869, 1.884, 1.906, 1.943, 1.970, 1.973, 2.036, 2.039, 2.063, 2.069, 2.083, 2.096, 2.104, 2.133, 2.134, and 2.159 (19s, 19Ac), 5.833 (td, 7.3 and 15.0Hz, 5^{Cer}), 7.40-8.00 (m, Bz and 3Phth). 37: RF 0.40 in 3:1 toluene-EtOAc; [a]D +14.5° (c 1.4); $\delta_{\rm H}$ 1.069, 1.168, and 1.184 (3s, 3Piv), 1.631, 1.845, 1.890, 1.903, and 2.026 (5s, 5Ac), 5.180 (d, 3.3Hz, 4²), 5.217 (d, 3.3Hz, 1⁷ and 1⁷), 5.251 (d, 8.4Hz, 1³ or 5). 38: RF 0.47 in 1:3 toluene-EtOAc; $\delta_{\rm H}$ 1.088, 1.194, and 1.211 (3s, 3Lev), 5.540, 5.625 and 5.645 (3dd, 8.5 and 10.5Hz, 3³, 3⁵ and 3⁵), 6.441 (d, 3.7Hz, 1¹), 7.60-7.90 (m, 3Phth), 8.591 (s, NH). **39**: R_F 0.62 in 1:3 toluene-EtOAc; $[\alpha]_D$ +32.4° (c 0.6); δ_H 0.879 (t, 7.0Hz, 2Me), 1.098, 1.132, and 1.220 (3s, 3Piv), 5.834 (td. 7.0 and 15.0Hz, 5^{Cer}), 7.40-8.05 (m, Bz and 3Phth).

- 10 C. Murakata and T. Ogawa, Tetrahedron Lett., 31, 2439 (1990).
- 11 T. Fukuyama, A. A. Laird, and L. M. Hotchkiss, Tetrahedron Lett., 26, 6291 (1985).
- 12 Wm. Rosenbrook, Jr., D. A. Riley, and P. A. Lartey, Tetrahedron Lett., 26, 3 (1985); G. H. Posner and S. R. Haines, ibid., 26, 935 (1985).
- 13 T. Nakano, Y. Ito, and T. Ogawa, Tetrahedron Lett., 31, 1597 (1990). 14 E. J. Corey, S. Kim, S. Yoo, K. C. Nicolaou, L. S. Melvin, Jr., P. J. Brunelle, J. R. Flack, E. J. Trybulski, R. Lett, and R. W. Sheldrake, J. Am. Chem. Soc., 100, 4620 (1978).
- 15 L. M. Haines and E. Singleton, J. Chem. Soc. Dalton Trans., 1891 (1972); J. J. Oltvoort, C.A.A. van Boeckel, J. H. De Koning and J. H. van Boom, Synthesis, 305 (1981).
- 16 T. Mukaiyama, Y. Murai, and S. Shoda, Chem. Lett., 431 (1981); T. Mukaiyama, Y. Hashimoto, and S. Shoda, ibid., 935 (1985).
- 17 K. Koike, M. Sugimoto, S. Sato, Y. Ito, Y. Nakahara, and T. Ogawa, Carbohydr. Res., 163, 189(1987).
- 18 S. Sato, M. Mori, Y. Ito, and T. Ogawa, Carbohydr. Res., 155, C6 (1986).
- 19 R. R. Schmidt and J. Michel, Angew. Chem. Int. Ed. Engl., 19, 731 (1980).
- 20 T. Ogawa and M. Sugimoto, Carbohydr. Res., 135, C5 (1985).
- 21 K. Koike, Y. Nakahara, and T. Ogawa, Glycoconj. J. 1, 107 (1984); K. Koike, M. Numata, M. Sugimoto, Y. Nakahara, and T. Ogawa, Carbohydr. Res., 158, 113 (1986).
- 22 M. S. Motawia, J. Wengel, A. E. S. Abdel-Megid, and E. B. Pedersen, Synthesis, 384 (1989).
- 23 U. Dabrowski, P. Hanfland, H. Egge, S. Kuhn, and J. Dabrowski, J. Biol. Chem., 259, 7648 (1984).
- 24 T. Matsumoto, H. Maeta, K. Suzuki and G. Tsuchihashi, Tetrahedron Lett., 29, 3567 (1988).
- 25 H. Hayashi, S. Hashimoto, and R. Noyori, Chem. Lett., 1747 (1984).

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