

Available online at www.sciencedirect.com



CARBOHYDRATE RESEARCH

Carbohydrate Research 338 (2003) 117-121

www.elsevier.com/locate/carres

## Rapid Communication

# Synthesis of monomethylated dioscin derivatives and their antitumor activities

Ming Li,<sup>a</sup> Xiuwen Han,<sup>a,\*</sup> Biao Yu<sup>b,\*</sup>

<sup>a</sup>State Key Laboratory of Catalyst, Dalian Institute of Chemical Physics, Chinese Academy of Sciences, Dalian 116023, China <sup>b</sup>State Key Laboratory of Bio-organic and Natural Products Chemistry, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, 354 Fenglin Road, Shanghai 200032, China

Received 20 September 2002; accepted 29 October 2002

#### **Abstract**

All possible eight monomethylated dioscin derivatives (1-8) were synthesized. Their inhibitory activities against P388 and A-549 cells were determined, and the results indicate that six of the eight hydroxyls of dioscin are the 'key polar groupings' for tumor inhibitory activities. © 2002 Elsevier Science Ltd. All rights reserved.

Keywords: Dioscin; Methylation; Synthesis; Antitumor

Spirostan saponins are the largest group of steroidal saponins occurring in plants. While the spirostan aglycones provide a rigid bulky moiety, the glycoforms render a relatively mobile conformation for the molecule. A quite common feature of spirostan saponins is their inhibitory activities against tumor cells, with their IC50s being at the  $\mu$ M level. Dioscin, diosgenin-3-yl  $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 2)$ - $[\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 4)]$ - $\beta$ -D-glucopyranoside, represents a typical example of spirostan saponins. In fact, dioscin is one of the most common saponins occurring in plants

that has been isolated from some twenty genera. Many of these plants are vegetables or traditional medicinal plants, especially from the Orient. Besides antitumor activities, <sup>2,3</sup> antiviral, <sup>4</sup> antifungal, <sup>5</sup> antiinflammatory, <sup>6–8</sup> and immunostimulant activities <sup>9</sup> were also observed for this spirostan saponin. Recently, synthetic approaches toward dioscin and its congeners have been developed <sup>1b,10</sup> that give access to their derivatives for developing more potent compounds, as well as probing structure—activity relationships and deciphering biological mechanisms.

0008-6215/03/\$ - see front matter © 2002 Elsevier Science Ltd. All rights reserved. PII: S0008-6215(02)00443-3

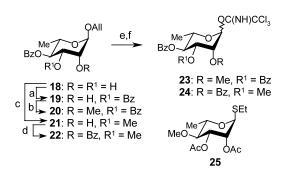
<sup>\*</sup> Corresponding author. Fax: +21-64166128 *E-mail address:* byu@pub.sioc.ac.cn (B. Yu).

Scheme 1. Reagents and conditions: (a)  $Bu_2SnO$ , PhMe, then  $CH_3I$ , t- $Bu_4N^+Br^-$ , 78%; (b) p-TsOH, MeOH $-CH_2Cl_2$ , 40% for 10; 46% for 11. (c) BzCl, pyridine, -40 °C, 81%; (d) 13 (4.0 equiv), TMSOTf (0.2-0.3 equiv),  $CH_2Cl_2$ , 81% for 14; 92% for 17. (e)  $CH_2Cl_2$ , 76% for 1; 78% for 2. (f)  $CH_2Cl_2$ , 76%. (g)  $CH_2Cl_2$ , 76%. (g)  $CH_2Cl_2$ , 76% then  $CH_3I$ ,  $CH_3Cl_2$ , 76%.

To find a starting point for derivatization, we managed to prepare all possible monomethylated dioscins (1–8). Replacement of a hydroxyl group on a sugar with a methoxy group has been an effective strategy to probe the involvement of a particular hydroxyl group in the recognition of the parent sugar on target proteins. For example, substitution of a 'key polar hydroxyl group' with a methoxy group on a disaccharide substrate of a glycosyltransferase completely blocked their recognition; while substitution of all the hydroxyl groups with methoxy groups on the heparin pentasaccharide did not affect its binding with antithrombin III, demonstrating no 'key polar hydroxyl groups' are involved in the interaction of heparin with antithrombin III. 12

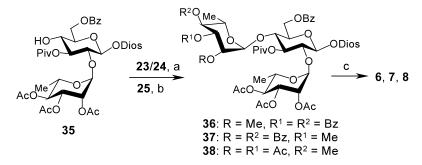
Along the versatile synthetic route toward dioscin and its congeners, <sup>10</sup> the monomethylated derivatives (1–8) were readily synthesized as shown in Schemes 1–4. Compounds 1 and 2 have the methoxy groups substituted on the core glucose residue and were prepared starting from diosgenyl glucopyranosides 9<sup>10</sup> and 15<sup>10</sup> (Scheme 1). Compounds 3–8 have the methoxy groups substituted on the peripheral rhamnose residues. Their preparation thus requires the glycosylation of the corresponding diosgenyl glucopyranoside monols (15 and 35<sup>10</sup>) with monomethylated rhamnopyranosyl donors (Schemes 3 and 4). Rhamnosyl donors 23 and 24 were readily prepared as shown in Scheme 2, and donor 25 was readily prepared from ethyl 4-*O*-methyl-1-thio-α-L-rhamnopyranoside<sup>14</sup> by acetylation.

Since the present syntheses adopted modifications of the well-documented synthetic route toward dioscin and its congeners, <sup>10</sup> the chemistry worth discussion here is the organotin-mediated regioselective methylation <sup>15</sup> and glycosylation with monomethylated rhamnopyranosyl donors. Regioselective benzoylation or methylation of the  $2_{\rm ax}$ ,  $3_{\rm eq}$ -diol 18 via a dibutylstannylene intermediate gave the  $3_{\rm eq}$ -substituted products (19 and 21) in excellent yields. Methylation of  $2_{\rm eq}$ ,  $3_{\rm eq}$ -diol 9 under similar conditions gave no regioselectivity, leading to, after cleavage of the 4,6-O-benzylidene group, 3-OMe and 2-OMe products 10 and 11 in 1:1.2 ratio. Attempts at selective methylation of the primary 6-OH of a triol to obtain 16 via tributylstannyl ether intermediates gave



Scheme 2. Reagents and conditions: (a) Bu<sub>2</sub>SnO, PhMe, 140 °C; then BzCl, pyridine, rt, 71%. (b) CH<sub>2</sub>N<sub>2</sub>, BF<sub>3</sub>·OEt<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, rt, 87%. (c) Bu<sub>2</sub>SnO, PhMe, 140 °C; then CH<sub>3</sub>I, *t*-Bu<sub>4</sub>N<sup>+</sup>Br<sup>-</sup>, rt; (d) BzCl, pyridine, rt, 80% (for 2 steps). (e) PdCl<sub>2</sub>, MeOH, rt; (f) CCl<sub>3</sub>CN, DBU, CH<sub>2</sub>Cl<sub>2</sub>, rt, 76% for **23**; 59% for **24** (2 steps).

Scheme 3. Reagents and conditions: (a) **23/24** (3.0 equiv), TMSOTf (0.05 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 80% for **26**; 72% for **27**. (b) **25** (3.0 equiv), NIS/AgOTf, CH<sub>2</sub>Cl<sub>2</sub>, 81% (for **28**). (c) *p*-TsOH, MeOH–CH<sub>2</sub>Cl<sub>2</sub>, 40 °C; (d) BzCl, pyridine, 0 °C, 78% for **29**; 76% for **30**; 66% for **31** (2 steps). (e) **13** (3.0 equiv), TMSOTf (0.05 equiv), CH<sub>2</sub>Cl<sub>2</sub>, rt, 66% for **32**; 76% for **33**; 79% for **34**. (f) NaOMe, MeOH–CH<sub>2</sub>Cl<sub>2</sub>, 72% for **3**; 80% for **4**; 89% for **5**.



Scheme 4. Reagents and conditions: (a) 23/24 (3.0 equiv), TMSOTf (0.05 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 85% for 36; 96% for 37. (b) 25 (3.0 equiv), NIS/AgOTf, CH<sub>2</sub>Cl<sub>2</sub>, 84% (for 38). (c) NaOMe, MeOH–CH<sub>2</sub>Cl<sub>2</sub>, 73% for 6; 80% for 7; 75% for 8.

Table 1 Growth inhibition rate (%) of monomethylated dioscin derivatives 1–8 on tumor cells (P388 and A-549)

Compound	P388			A-549		
	$10^{-5} \text{ M}$	$10^{-6} \text{ M}$	$10^{-7} \text{ M}$	$10^{-5} \text{ M}$	$10^{-6} \text{ M}$	10 <sup>-7</sup> M
1	16.8	0	0	36.5	1.9	0
2	100	90.6	4.4	99.5	69.4	0
3	100	16.1	0	99.3	15.7	0
4	54.6	0	0	99.3	34.3	8.9
5	94.8	0.9	0	97.4	7.4	7.9
6	91.4	12.1	2.2	72.3	0	0
7	97.7	22.3	5.6	99.6	10.4	0
8	99.4	74.9	0	99.6	63.5	0
Adriamycin	100	100	100		70.2	55.9

the desired 16 as a major product, albeit in only 29% yield. In addition, methylation of the monol 19 was not successful under many of the usual conditions, e.g., MeI/Ag<sub>2</sub>O/DMF, MeOTf/DTBMP, due to the neighboring acyl group migrations. Methylation was finally effected by means of CH<sub>2</sub>N<sub>2</sub> in the presence of BF<sub>3</sub>·OEt<sub>2</sub>, giving **20** in 87% yield. Glycosylation with monomethylated rhamnopyranosyl trichloroacetimidates 23 and 24 and thioglycoside 25 under the promotion of TMSOTf and NIS/AgOTf, respectively, afforded the expected products (26-28 and 36-38) in satisfactory yields (66–96%). Especially, glycosylation with donor 23, in the absence of neighboring group participation at the C-2 position, provided only the  $\alpha$ products (26 and 36), reflecting a strong anomeric effect in the glycosylation with L-rhamnopyranosyl donors.<sup>16</sup> In comparison, the monomethylated rhamnopyranosyl donors 23-25 demonstrated more reactivity than the peracetylated trichloroacetimidates 13, producing the corresponding glycosylation products, in glycosylation with the 4-OH of the disaccharides 29-31 and 35, in higher yields (cf., 66-79% yield for 32-34; 84-96% yield for 36-38).

The in vitro inhibitory activities of the monomethylated dioscins  $1-8^{\dagger}$  against the growth of P388 (mouse leukemia) and A-549 (human lung adenocarcinoma) were evaluated by the standard MTT assay.17 The results were listed in Table 1. In comparison with dioscin, which has an IC<sub>50</sub> of 0.46 μM against P388,<sup>2</sup> the activities of the monomethylated compounds 2 and 8 were largely retained, while the inhibitory activities of the other six compounds were considerably decreased. These results indicated that the six hydroxyl groups (OH-1 and OH-3-OH-7) on dioscin might be the 'key polar hydroxyl groupings'13 contributing to its antitumor activity. Thus the OH-2 and OH-8 could be the sites for labeling to provide derivatives for further mechanistic studies. In fact, the OH-2 and OH-8 of dioscin and its congeners have been demonstrated to be the most distant hydroxyl groups from the aglycone. Thus these could be regioselectively acylated by Novozyme 435. The resulting monoacetylated derivatives showed similar antitumor activities as the parent compounds.<sup>18</sup> Those findings are in agreement with the present results.

### Acknowledgements

This work is supported by the Ministry of Science and Technology of China (G1998051104), the Shanghai/Hong Kong/Anson Research Foundation, and the

National Natural Science Foundation of China (29925203).

#### References

- 1. (a) Hostettmann, K.; Marston, A. Saponins. Cambridge University Press: Cambridge, UK, 1995;
  - (b) Yu, B.; Hui, Y. Chemical Synthesis of Bioactive Steroidal Saponins. In *Glycochemistry: Principles, Synthesis, and Application*; Wang, P. G.; Bertozzi, C. R., Eds.; Marcel Dekker, Inc. New York, 2001; pp 163–174.
- 2. (a) Hu, K.; Dong, A. J.; Yao, X. S.; Kobayashi, H.; Iwasaki, S. *Planta Med.* **1996**, *62*, 573–575;
  - (b) Nakamura, T.; Komori, C.; Lee, Y.-y.; Hashimoto, F.; Yahara, S.; Nohara, T.; Ejima, A. *Biol. Pharm. Bull.* **1996**, *19*, 564–566.
- Chiang, H. C.; Tseng, T. H.; Wang, C. J.; Chen, C. F.; Kan, W. S. Anticancer Res. 1991, 11, 1911–1917.
- Ikeda, T.; Ando, J.; Miyazono, A.; Zhu, X.-H.; Tsumagari, H.; Nohara, T.; Yokomizo, K.; Uyeda, M. *Biol. Pharm. Bull.* 2000, 23, 363–364.
- (a) Takechi, M.; Shimada, S.; Tanaka, Y. *Phytochemistry* 1991, 30, 3943–3944;
   (b) Hufford, C. D.; Liu, S.; Clark, A. M. *J. Nat. Prod.*
- **1988**, *51*, 94–98.
  6. Kim, S. Y.; Son, K. H.; Chang, H. W.; Kang, S. S.; Kim,
- H. P. Arch. Pharm. Res. **1999**, 22, 313–316.
  7. Mimaki, Y.; Nakamura, O.; Sashida, Y.; Nikaido, T.;
- Ohmoto, T. *Phytochemistry* **1995**, *38*, 1279–1286. 8. Baek, S. H.; Kim, S. H.; Son, K. H.; Chung, K. C.;
- Chang, H. W. Arch. Pharm. Res. 1994, 17, 218–222.
  9. Chiang, H. C.; Wang, J. J.; Wu, R. T. Anticancer Res. 1992, 12, 1475–1478.
- 10. (a) Deng, S.; Yu, B.; Hui, Y.; Yu, H.; Han, X. Carbohydr. Res. 1999, 317, 53-62;
  - (b) Deng, S.; Yu, B.; Hui, Y. Tetrahedron Lett. 1998, 39, 6511–6514.
- 11. (a) Mukherjee, A.; Palcic, M. M.; Hindsgaul, O. *Carbohydr. Res.* **2000**, *326*, 1–21;
  - (b) Kanie, O.; Crawley, S. C.; Palcic, M. M.; Hindsgaul, O. *Carbohydr. Res.* **1993**, *243*, 139–164;
  - (c) Srivastava, O. P.; Hindsgaul, O. Carbohydr. Res. 1988, 179, 137–161.
- (a) Basten, J.; Jaurand, G.; Olde-Hanter, B.; Duchaussoy,
   P.; Petitou, M.; van Boeckel, C. A. A. Bioorg. Med. Chem. Lett. 1992, 2, 905–910;
  - (b) Basten, J.; Jaurand, G.; Olde-Hanter, B.; Petitou, M.; van Boeckel, C. A. A. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 901–904;
  - (c) Jaurand, G.; Basten, J.; Lederman, I.; van Boeckel, C. A. A.; Petitou, M. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 897–900.
- 13. Lemieux, R. U. Chem. Soc. Rev. 1989, 18, 347-374.
- Zuurmand, H. M.; Veeneman, G. H.; van der Marel, G. A.; van Boom, J. H. Carbohydr. Res. 1993, 241, 153–164.
- (a) David, S.; Hanessian, S. Tetrahedron 1985, 41, 643–663;
  - (b) Veyriéres, A. J. Chem. Soc., Perkin Trans. 1 1981, 1626–1629;
  - (c) Takeo, K.; Shibata, K. Carbohydr. Res. 1984, 133, 147–151.
- 16. (a) Yu, B.; Li, B.; Xing, G.; Hui, Y. *J. Comb. Chem.* **2001**, *3*, 404–406;

 $<sup>^{\</sup>dagger}$  The structures of 1–8 were unambiguously determined by extensive 2D NMR analysis and were further confirmed by ESIMS and elemental analysis.

- (b) Duynstee, H. I.; van Vliet, M. J.; van der Marel, G. A.; van Boom, J. H. *Eur. J. Org. Chem.* **1998**, 303–307;
- (c) Douglas, N. L.; Ley, S. V.; Lucking, U.; Warriner, S. L. *J. Chem. Soc.*, *Perkin Trans.* 1 **1998**, 51–66;
- (d) Ley, S. V.; Priepke, H. W. M. Angew. Chem., Int. Ed.
- Engl. 1994, 33, 2292-2294.
- Kuroda, M.; Mimaki, Y.; Sashida, Y.; Hirano, T.; Oka, K.; Dobashi, A.; Li, H.; Harada, N. *Tetrahedron* 1997, 53, 11549–11562.
- 18. Yu, B.; Xing, G.; Hui, Y.; Han, X. Tetrahedron Lett. **2001**, 42, 5513–5516.