Chemistry of Natural Compounds and Bioorganic Chemistry

Synthesis of modified ether phospholipids

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A number of modified ether phospholipids with additional substituents in the 2 position of the C(1)-alkyl chain, 1-O-[2'-(R,S)-hydroxyhexadecyl]-2-O-methyl-rac-glycero-3-phosphocholine, <math>1-O-[2'-(R,S)-acetoxyhexadecyl]-2-O-methyl-rac-glycero-3-phosphocholine, and <math>1-O-[2'-(R,S)-hydroxyhexadecyl]-2-chloro-2-deoxy-rac-glycero-3-phosphocholine, have been synthesized.

Key words: ether phospholipids; analogs of platelet activating factor; modified phospholipids.

Ether lipids possess a broad spectrum of biological activities. A representative of this class of substances, 1 - O-alky1 - 2 - O-acety1 - sn-glycero-3-phosphocholine (platelet activating factor, PAF), is known to be a potent wide-range biological regulator. PAF has been shown to be a mediator of a number of biological processes. It stimulates platelet degranulation and aggregation;¹ causes the contraction of smooth muscles, bronchoconstriction, and coronary vasoconstriction;¹ increases vascular permeability;^{1,2} and stimulates immune response.³⁻⁵ PAF also plays an important role in a number of physiological and pathological processes, such as allergy, hypotension, anaphylaxis, thrombosis, ischemia, acute infections in transplantation, nephritis, gastric ulcer, etc.⁶

A large number of PAF structural analogs were synthesized to study the mechanism of PAF interaction with target cells. Modifications of the structure of the PAF molecule made it possible to separate various types of biological activities. For example, substances with more potent platelet aggregating and hypotensive effects than that of PAF; specific PAF antagonists;⁷ and immunomodulators without platelet aggregating activity, which are hardly metabolizable as compared to PAF,⁸⁻¹⁰ have been obtained. Many PAF structural analogs have been found to possess antitumor activity.¹¹ Some ether phospholipids are shown to inhibit HIV-1 replication in infected T-cells.¹² Thus, ether phospholipids are of considerable interest as potential therapeutic preparations because of the variety of their biological effects.

At present the main directions of the modifications of phospholipid structure, which cause the appearance or the disappearance of various biological activities, are already known. For example, the substitution of the ether bond in the sn-1 position by a thioether, amide, or

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carbamoyl function or the replacement of the acetyl group in the sn-2 position with a small hardly metabolizable moiety (*e.g.*, a short chain alkoxy group or a halogen atom) cause a lack of platelet aggregating activity.⁷ Sometimes such modifications lead to the appearance of PAF antagonism⁷ and an antitumor effect.¹¹ The absence of platelet aggregating and hypotensive activities is indispensable in the synthesis of substances with possible antitumor and immunomodulating effects.

In confirming the studies on the structure—activity relations of ether phospholipids, we synthesized the compounds with additional substituents in the 2 position of the long alkyl chain. The additional moiety influences the steric surroundings in closest proximity to the glycerol backbone, which we expected to cause a decrease or disappearance of PAF-like activities. Such structures are of interest, because 1 - O - [2' - (R) - methoxy-hexadecy] - sn-glycerol, which was isolated from natural



R = OMe (a), Cl (b)

Reagents and conditions: a. BnCl, KOH, C₆H₆, 80 °C; b. TsOH, MeOH; c. TrCl, Py, 7 °C; d. MeI, KOH, C₆H₆, 80 °C; e. TsCl, Py, 0 °C; f. LiCl, DMF, 80 °C; g. Me₃SiCl, silica gel; h. POCl₃, Et₃N, CHCl₃; HOCH₂CH₂N⁺Me₃OTs⁻, Py; H₂O; i. H₂, Pd/C, MeOH; j. Ac₂O, HClO₄, CHCl₃.

sources, characterized, and then obtained by chemical synthesis, is known to inhibit tumor growth and to modulate the immune response in mice during oral administration.

In our previous communication¹⁵ we described the synthesis of modified phosphatidylcholines with 2-methoxy substituents in the long alkyl chain. Now we report the synthesis of modified ether phospholipids (**9a,b** and **10a,b**) with hydroxy and acetoxy groups in the 2 position of the long alkyl chain. These substances are of interest as possible PAF antagonists, immunomodulators, and antitumor agents.

Initially, 1-O-[2'-(R,S)-hydroxyhexadecyl]-2,3-di-Oisopropylidene-*rac*-glycerol (1)¹⁶ was refluxed with KOH powder in C₆H₆ with azeotropic distillation of water followed by alkylation with BnCl (Scheme 1) to give derivative 2. The isopropylidene protective group of 2 was removed by methanolysis in the presence of a catalytic amount of TsOH. 1-O-[2'-(R,S)-Benzyloxyhexadecyl]-*rac*-glycerol (3) was tritylated to protect the primary hydroxy group, and the trityl derivative 4 was alkylated with MeI to yield 1-O-[2'-(R,S)-benzyloxyhexadecyl]-2-O-methyl-3-O-trityl-*rac*-glycerol (6a).

In the synthesis of 2-chloro-2-deoxy derivatives compound 4 was treated with TsCl in the presence of Py in $CHCl_3$ to obtain the tosyl derivative 5. The tosyl group of 5 was then substituted by a chlorine atom by refluxing with LiCl in DMF via the Finkelstein reaction to give **6b**. Using DMF as solvent made it possible to avoid the partial detachment of the trityl group, which took place when the reaction was performed in MeC(O)Et. Then compounds **6a** and **6b** were treated with Me₃SiCl in the presence of silica gel to remove the trityl groups under mild conditions.¹⁷ The resulting 1 - O - [2' - (R, S) - (R, S)]benzyloxyhexadecyl]-2-O-methyl-rac-glycerol (7a) and 1-O-[2'-(R,S)-benzyloxyhexadecyl]-2-chloro-2-deoxyrac-glycerol (7b) were converted to phosphatidylcholines (8a and 8b) by subsequent treatment with POCl₃ in the presence of Et₃N and with HOCH₂CH₂N⁺Me₃OTs⁻ in the presence of Py in CHCl₃ according to Brockerhoff et al.¹⁸

The catalytic hydrogenolysis of **8a** and **8b** in the presence of Pd/C in MeOH to remove the benzyl protective groups afforded 1 - O - [2' - (R, S) - hydroxy-hexadecyl] - 2 - O - methyl - rac-glycero - 3 - phosphocholine (**9a**) and <math>1 - O - [2' - (R, S) - hydroxyhexadecyl] - 2 - chloro - 2 - deoxy-rac-glycero - 3 - phosphocholine (**9b**).

The 2'-hydroxyl functions of **9a** and **9b** were acetylated with Ac_2O in the presence of $HClO_4$ (see Ref. 19) to give 1-O-[2'-(R,S)-acetoxyhexadecyl]-2-O-methylrac-glycero-3-phosphocholine (**10a**) and <math>1-O-[2'-(R,S)-acetoxyhexadecyl]-2-chloro-2-deoxy-rac-glycero-3-phosphocholine (**10b**).

The identity and structure of the compounds obtained were confirmed by TLC, IR, ¹H and ³¹P NMR spectroscopy, and mass spectrometry data.

Experimental

IR spectra (films) were recorded on a Shimadzu IR-435 spectrometer. ¹H NMR spectra were registered on Bruker MSL-200 (200 MHz for 3, 5, 7a,b, and 8a) and WM-500 (500 MHz for 9a,b and 10a,b) spectrometers in CDCl₃ for neutral lipids and in a mixture of CDCl₃-CD₃OD-D₂O (1:1:0.15) for phosphocholines with SiMe₄ as internal standard. Broad-band ³¹P NMR spectra with proton decoupling were obtained on a Bruker MSL-200 instrument (81 MHz with H_3PO_4 as external standard) in a mixture of $CDCl_3$ - CD_3OD - D_2O (1 : 1 : 0.15). Mass spectra were taken on an MSBKh (Sumy, Ukraine) plasma desorption mass spectrometer with ionization by the products of californium-252 fission and at an accelerating voltage of ± 5 or ± 20 eV. TLC was performed on Silufol UV-254 (Kavalier, Czechoslovakia) plates using the following developing systems: $Et_2O(A)$; hexane $-Et_2O$, 4 : 1 (B); hexane $-Et_2O$, 1 : 1 (C); CHCl-MeOH-H₂O, 65 : 25 : 4 (D). The substances were detected by burning or treating with molybdenum blue (in the case of phospholipids). Column chromatographic purification was carried out on L 40/100 µm (Chemapol, Czechoslovakia) silica gel.

1-O-[2'-(R,S)-Benzyloxyhexadecyl]-2,3-di-O-isopropylidene-rac-glycerol (2). To a solution of 4.16 g (11.2 mmol) of 1-O-[2'-(R,S)-hydroxyhexadecyl]-2,3-di-Oisopropylidene-rac-glycerol (1) in 50 mL of C₆H₆ 5.0 g (99.8 mmol) of KOH powder was added with vigorous stirring. The mixture was refluxed for 6 h with stirring and azeotropic distillation of water. Then 4.18 g (33.0 mmol, 3.80 mL) of BnCl was added, and the mixture was refluxed for 4 h with stirring. After the reaction mixture was cooled, the solvent and the excess of BnCl were removed in vacuo. The residue was dissolved in 100 mL of Et₂O and washed with water (3×100 mL) up to pH 7. The organic phase was separated, dried over anhydrous Na₂SO₄, and concentrated in vacuo. The residue was purified by column chromatography on silica gel $(30 \times 300 \text{ mm}; \text{hexane}-\text{Et}_2\text{O}, 95 : 5, \text{ as the eluent})$ and dried in vacuo (130 Pa, 60 °C, 8 h) to yield 4.40 g (85 %) of 2, $R_{\rm f}$ 0.62 (system B). IR, v/cm⁻¹: 3100-3050 (w, C-H arom.); 2900, 2840 (s, C-H); 1460 (s, C-H); 1280, 1270, 1250, 1210 (m, C-H, C(CH₃)₂); 1100-1150 (s, C-O-C); 840, 715, 695 (m, π ,* C–H arom.); 720 (m, C–H).

1-O-[2'-(R,S)-Benzyloxyhexadecyl]-rac-glycerol (3). Isopropylidene derivative 2 (4.4 g) was dissolved in 50 mL of MeOH in the presence of a catalytic amount of TsOH, and the solvent was removed in vacuo. The operation was repeated 5 times, up to the absence of the initial compound. The resulting residue was chromatographed on a silica gel column $(30 \times 300 \text{ mm}; \text{Et}_2\text{O} \text{ as the eluent})$ and dried in vacuo to give 2.73 g (68 %) of 3, R_f 0.45 (system A). IR, v/cm⁻¹: 3400 (m, O-H); 3100-3050 (w, C-H arom.); 2900, 2840 (s, C-H); 1470, 1450 (m, C-H); 1120 (s, C-O-C); 1080 (s, C-O, C-OH); 715 (m, C-H); 700, 690 (m, π, C-H arom.). ¹H NMR, δ : 0.86 (t, 3 H, (CH₂)₁₂CH₃, J = 7 Hz); 1.22 (br.s, 24 H, $(CH_2)_{12}$); 1.50 (m, 2 H, $CH(OBn)CH_2$); 3.43-3.70 (m, 7 H, CH₂OH, CH₂OCH₂CHOBn); 3.82 (m, 1 H, CHOH); 4.57 (dd, 2 H, CH₂Ph, J = 11.5 and 13.0 Hz); 7.2–7.35 (m, 5 H, Ph). MS, m/\bar{z} (I_{rel} (%)): 424.0 [M+H]⁺ (100).

^{*} π is extraplanar vibration.

1-0-[2'-(R, S)-Benzyloxyhexadecyl]-3-0-trityl-rac-glycerol (4). A solution of 5.43 g (12.8 mmol) of 3 in 15 mL of anhydrous CHCl₃ and 5 mL of anhydrous Py was cooled to 7 °C, and 3.57 g (12.8 mmol) of TrCl was added with stirring. The mixture was stirred for 4 h at 7 °C, diluted with 35 mL of CHCl₃, and washed with 1 % HCl (50 mL) and then with water (2×50 mL) up to pH 7. The organic layer was separated, dried over anhydrous Na₂SO₄, and concentrated. The residue was dried *in vacuo* (130 Pa, 50 °C, 6 h) to give 9.10 g of crude 4, R_f 0.27 (system B). Compound 4 was used for the preparation of 5 and 6a without purification.

1-O-[2'-(R,S)-Benzyloxyhexadecyl]-2-O-tosyl-3-O-tritylrac-glycerol (5). To a solution of 2.21 g (3.31 mmol) of 4 in 15 mL of dry Py cooled to 0 °C 0.75 g (3.97 mmol) of TsCl was added with stirring. The mixture was stirred for 20 h at 20 °C and then concentrated in vacuo. The residue was dissolved in 5 mL of C_6H_6 . The solution was filtered to remove TsOH and Py · HCl and was evaporated in vacuo. The residue was dissolved in Et₂O and passed through Al₂O₃. After concentrating in vacuo the residue was chromatographed on a silica gel column (25×300 mm; hexane-Et₂O, 85 : 15, as the eluent) and dried in vacuo (130 Pa, 40 °C, 6 h). Yield 2.20 g (81 %), R_f 0.52 (system C). Found (%): C, 76.02; H, 8.03; S, 3.95. $C_{52}H_{66}O_6S$. Calculated (%): C, 76.25; H, 8.12; S, 3.91. IR, v/cm⁻¹: 3100-3050 (w, C-H arom.); 2900, 2840 (s, C-H); 1480, 1430 (m, C-H); 1350, 1160 (s, S=O); 1110 (s, C-O-C); 900 (s, S-O); 790, 740 (m, π , C-H arom.); 720 (m, C–H); 670 (s, π , C–H arom.). ¹H NMR, δ : 0.86 (t, 3 H, $(CH_2)_{12}CH_3$, J = 7 Hz); 1.24 (br.s, 24 H, $(CH_2)_{12}$); 1.32 (m, 2 H, CH(OBn)C \underline{H}_2); 2.37 (s, 3 H, C \underline{H}_3 (OTs)); 3.18-3.42 (m, 5 H, CH₂OCH₂CHOBn); 3.59, 3.65 (dd, 1 H, CH₂OTr, J = 5 and 11 Hz); 4.44 (dd, 13.0, 2 H, CH_2Ph , J = 11.5 Hz); 4.66 (m, 1 H, CHOTs); 7.14–7.39 (m, 22 H, 4Ph; m-H (OTs)); 7.77 (dt, 2 H, o-H (OTs), J =8 and 2 Hz).

1-O-[2'-(R,S)-Benzyloxyhexadecyl]-2-O-methyl-3-Otrityl-rac-glycerol (6a). To a solution of 1.90 g (2.85 mmol) of 4 in 25 mL of C₆H₆ 0.8 g (14.3 mmol) of KOH powder was added. The mixture was refluxed for 3 h with stirring and azeotropic distillation of water and then cooled to 30 °C. MeI (2.02 g, 14.3 mmol) was added, and the mixture was stirred for 6 h at 50 °C. The solvent and the excess MeI were removed in vacuo. The residue was dissolved in 100 mL of Et_2O and washed with water (3×100 mL) up to pH 7. The organic layer was separated, dried over anhydrous Na₂SO₄, and concentrated in vacuo. The residue was purified by column chromatography (25×200 mm; hexane-Et₂O, 9 : 1, as the eluent) and dried in vacuo (130 Pa, 50 °C, 6 h). Yield 1.63 g (84 %), R_f 0.54 (system B). Found (%): C, 81.28; H, 9.17. $C_{46}H_{62}O_4$. Calculated (%): C, 81.37; H, 9.20. IR, v/cm⁻¹: 3100-3050 (w, C-H arom.); 2900, 2840 (s, C-H); 1490 (m, C=C arom.); 1460, 1450 (m, C-H); 1110 (s, C-O-C); 1090 (s, C-O, C-OH); 760, 740, 630 (m, π, C-H arom.); 725 (m, C-H); 695 (s, π , C-H arom.).

1-O-[2'-(R, S)-Benzyloxyhexadecyl]-2-chloro-2-deoxy-3-O-trityl-rac-glycerol (6b). To a solution of 2.06 g (2.52 mmol) of tosyl derivative 5 in 25 mL of DMF 0.56 g (13.1 mmol) of LiCl was added, and the mixture was stirred for 7 days at 80 °C. The solvent was removed *in vacuo*. The residue was dissolved in 25 mL of Et₂O; the solution was washed with water (2×25 mL) and concentrated *in vacuo*. The residue was dried *in vacuo* (130 Pa, 40 °C, 6 h) to give 1.73 g of crude 6b, R_f 0.74 (system C). Compound 6b was then detritylated without purification.

1-O-[2'-(R,S)-Benzyloxyhexadecyl]-2-O-methyl-rac-glycerol (7a). To a solution of 1.43 g (2.10 mmol) of 6a in 20 mL of anhydrous CHCl₃ 5.0 g of activated (110 °C, 8 h) silica gel (L 40/100 μ m) was added with stirring. Then a solution of 1.14 g (10.5 mmol, 1.32 mL) of Me₃SiCl in 20 mL of anhydrous CHCl₃ was gradually poured in, and the reaction mixture was stirred for 1 h at 20 °C. Py-H₂O mixture (1 : 2, 60 mL) was added; the reaction mixture was stirred for 1 h, filtered and washed with water up to pH 7. The solvents were removed in vacuo. The resulting residue was crystallized from hexane, chromatographed on a silica gel column (25×200 mm; toluene-Et₂O, 7 : 3, as the eluent), and dried in vacuo (130 Pa, 50 °C, 6 h). Yield 0.842 g (91 %), R_f 0.16 (system C). IR, v/cm⁻¹: 3400 (m, O-H); 3100-3050 (w, C-H arom.); 2900, 2840 (s, C-H); 1460, 1450 (m, C-H); 1120 (s, C-O-C); 1080 (s, C-O, C-OH); 715 (m, C-H); 710, 690 (m, π , C-H arom.). ¹H NMR, δ : 0.86 (t, 3 H, (CH₂)₁₂CH₃, J = 7 Hz); 1.23 (br.s, 24 H, $(CH_2)_{12}$); 1.30 (m, 2 H, CH(OBn)CH₂); 3.31-3.65 (m, 8 H, CH₂OH, CHOMe, CH_2OCH_2CHOBn ; 3.35 (s, 3 H, OMe); 4.40 (dd, 2 H, CH₂Ph, J = 11 and 19 Hz); 7.0-7.25 (m, 5 H, Ph). MS, m/z (I_{rel} (%)): 437.7 [M+H]⁺ (100).

1-O-[2'-(R,S)-Benzyloxyhexadecyl]-2-chloro-2-deoxy-*rac***glycerol (7b)** was obtained by detritylation of **6b** under the conditions of the previous experiment. Yield 0.843 g (76 % from **5**), $R_{\rm f}$ 0.36 (system C). Found (%): C, 70.77; H, 10.18. C₂₆H₄₅O₃Cl. Calculated (%): C, 70.80; H, 10.28. ¹H NMR, δ : 0.86 (t, 3 H, (CH₂)₁₂CH₃, J = 7 Hz); 1.26 (br.s, 24 H, (CH₂)₁₂); 1.50 (m, 2 H, CH(OBn)CH₂); 3.45–3.92 (m, 7 H, CH₂OH, CH₂OCH₂CHOBn); 4.09 (m, 1 H, CHCl); 4.58 (dd, 2 H, CH₂Ph, J = 12 and 17 Hz); 7.21–7.36 (m, 5 H, Ph). MS, m/z ($I_{\rm rel}$ (%)): 441.7 [M+H]⁺ (100).

1-O-[2'-(R,S)-Benzyloxyhexadecyl]-2-O-methyl-racglycero-3-phosphocholine (8a). To a solution of 0.37 g (2.41 mmol, 0.22 mL) of POCl₃ in 5 mL of anhydrous CHCl₃ 0.842 g (1.93 mmol) of 7a and 0.244 g (2.41 mmol, 0.334 mL) of dry Et₃N in 10 mL of anhydrous CHCl₃ were added dropwise with stirring. After 30 min 1.13 mL of anhydrous Py was added, and the reaction mixture was stirred for 30 min. Then 1.06 g (3.86 mmol) of HOCH₂CH₂N⁺Me₃OTs⁻ was added, and the reaction mixture was stirred for an additional 4 h. Then it was diluted with 25 mL of CHCl₃ and washed with 3 % aqueous solution of Na2CO3 (20 mL), 5 % HCl (20 mL), and water (3×20 mL) up to pH 7. The water layer was extracted twice with CHCl₃. The combined organic extract was evaporated in vacuo. The residue was chromatographed on a silica gel column (20×250 mm; CHCl₃-MeOH-H₂O, 65 : 25 : 4, as the eluent) and dried in vacuo (130 Pa, 20 °C, 12 h) to yield 0.804 g (69 %) of **8a**, R_f 0.20 (system D). ¹H NMR, δ : 0.86 (t, 3 H, (CH₂)₁₂CH₃, J = 7 Hz); 1.21 (br. s, 24 H, (CH₂)₁₂); 1.47 (m, 2 H, CH(OBn)CH₂); 3.19 (s, 9 H, N⁺(CH₃)₃); 3.34–3.66 (m, 8 H, CH₂N, CHOMe, CH2OCH2CHOBn); 3.47 (s, 3 H, OCH3); 3.93 (m, 2 H, CH₂OP); 4.24 (m, 2 H, POCH₂); 4.61 (dd, 2 H, CH₂Ph, J = 11.5 and 24 Hz); 7.25-7.45 (m, 5 H, Ph). MS, m/z (I_{rel} (%)): 604.4 [M+H]⁺ (100).

1-O-[2'-(R,S)-Berzyloxyhexadecyl]-2-chloro-2-deoxy-racglycero-3-phosphocholine (8b) was obtained from 0.829 g (1.88 mmol) of 7b, 0.36 g (2.35 mmol) of POCl₃, and 1.03 g (3.76 mmol) of HOCH₂CH₂N⁺Me₃OTs⁻ in the presence of 0.238 g (2.35 mmol, 0.326 mL) of anhydrous Et₃N and 1.19 g (15.03 mmol, 1.21 mL) of anhydrous Py in 20 mL of anhydrous CHCl₃ under the conditions of the previous experiment. The product was chromatographed on a silica gel column (25×200 mm; CHCl₃-MeOH-H₂O, 65 : 25 : 4, as the eluent) and dried *in vacuo* (130 Pa, 20 °C, 12 h) to give 0.923 g (81 %) of 8b, $R_{\rm f}$ 0.20 (system D). MS, m/z ($I_{\rm rel}$ (%)): 607.4 [M+H]⁺ (100).

1-O-[2'-(R,S)-Hydroxyhexadecyl]-2-O-methyl-racglycero-3-phosphocholine (9a). To 0.10 g of 10 % Pd/C (previously activated in an H₂ atmosphere at 100 °C and cooled to 20 °C) a solution of 0.80 g (1.33 mmol) of 8a in 15 mL of anhydrous MeOH was added, and the mixture was stirred in the H₂ atmosphere for 8 h at 20 °C. The catalyst was removed by filtration, and the filtrate was evaporated in vacuo. The residue was chromatographed on a silica gel column $(25 \times 250 \text{ mm}; \text{ CHCl}_3 - \text{MeOH} - \text{H}_2\text{O}, 65 : 25 : 4, \text{ as the}$ eluent) and dried in vacuo (130 Pa, 20 °C, 12 h) to give 0.513 g (75 %) of **9a**, R_f 0.11 (system D). Found (%): C, 56.55; H, 10.59; N, 2.71. $C_{25}H_{54}O_7PN \cdot H_2O$. Calculated (%): C, 56.67; H, 10.66; N, 2.65. ¹H NMR, 8: 0.86 (t, 3 H, $(CH_2)_{12}CH_3$, J = 7 Hz); 1.26 (br.s, 24 H, $(CH_2)_{12}$); 1.42 (m, 2 H, CH(OH)CH₂); 3.21 (s, 9 H, N⁺(CH₃)₃); 3.29–3.66 (m, 7 H, CH_2N , CH_2OCH_2 , CHOMe); 3.45 (s, 3 H, OCH_3); 3.72 (m, 1 H, CHOH); 3.94 (m, 2 H, CH₂OP); 4.25 (m, 2 H, POCH₂). ³¹P NMR, δ: 1.13. MS, m/z (I_{rel} (%)): 512.8 [M+H]⁺ (100).

1-*O*-[2'-(*R*, *S*)-Hydroxyhexadecyl]-2-chloro-2-deoxy-racglycero-3-phosphocholine (9b) was obtained by catalytic hydrogenolysis of 0.854 g of 8b under the conditions of the previous experiment. Yield 0.431 g (59 %), $R_{\rm f}$ 0.12 (system *D*). Found (%): C, 53.99; H, 10.06; N, 2.70. C₂₄H₅₁O₆PNCl·H₂O. Calculated (%): C, 53.90; H, 10.19; N, 2.62. ¹H NMR, δ: 0.86 (t, 3 H, (CH₂)₁₂CH₃, J = 7 Hz); 1.26 (br.s, 24 H, (CH₂)₁₂); 1.42 (m, 2 H, CH(OH)CH₂); 3.22 (s, 9 H, N⁺(CH₃)₃); 3.34– 3.53 (m, 2 H, CH₂OCH₂); 3.61 (m, 2 H, CH₂N); 3.67–3.80 (m, 3 H, CHOH, CH₂OCH₂); 4.09 (m, 2 H, CH₂OP); 4.21 (m, 1 H, CHCl); 4.27 (m, 2 H, POCH₂). ³¹P NMR, δ: 0.65. MS, m/z ($I_{\rm rel}$ (%)): 517.1 [M+H]⁺ (100).

1-O-[2'-(R,S)-Acetoxyhexadecyl]-2-O-methyl-rac-glycero-3-phosphocholine (10a). To a solution of 0.273 g (0.534 mmol) of 9a in 5 mL of CHCl₃ 1.96 g (16.21 mmol, 1.81 mL) of Ac₂O was added. After stirring for 30 s the reaction mixture was cooled to 0 °C, 0.91 mL of 8 N HClO₄ was added dropwise, and the mixture was stirred for 10 s. Then 18 mL of ice-cold water, 19.5 mL of CHCl₃ and 20 mL of MeOH were added. The organic layer was separated and extracted with MeOH-H₂O, 10 : 9 (2×38 mL). The water layer was extracted twice with CHCl₃. The combined organic extract was evaporated in vacuo. The residue was chromatographed on a silica gel column (10×200 mm; CHCl₃-MeOH-H₂O, 65: 25: 4, as the eluent) and dried in vacuo (130 Pa, 20 °C, 12 h) to yield 0.177 g (60 %) of 10a, $R_{\rm f}$ 0.21 (system D). Found (%): C, 55.86; H, 10.13; N, 2.46. C₂₇H₅₆O₈PN · 1.5H₂O. Calculated (%): C, 55.82; H, 10.25; N, 2.41. ¹H NMR, δ : 0.86 (t, 3 H, (CH₂)₁₂CH₃, J = 7 Hz); 1.26 (br.s, 24 H, $(CH_2)_{12}$); 1.47 (m, 2 H, $CH(OAc)CH_2$); 2.07 (s, 3 H, $OC(O)CH_3$); 3.21 (s, 9 H, $N^+(CH_3)_3$); 3.41– 3.67 (m, 7 H, CH₂N, CH₂OCH₂, CHOMe); 3.45 (s, 3 H, OCH₃); 3.88 (m, 2 H, CH₂OP); 4.24 (m, 2 H, POCH₂); 5.0 (m, 1 H, CHOAc). ³¹P NMR, δ : 1.17. MS, m/z (I_{rel} (%)): 554.7 [M+H]⁺ (100).

1-O-[2'-(R,S)-Acetoxyhexadecyl]-2-chloro-2-deoxy-racglycero-3-phosphocholine (10b) was obtained by treating 9b (0.196 g, 0.379 mmol) with Ac₂O (1.39 g, 13.64 mmol) in the presence of HClO₄ (1 mL of 8 N solution, 0.777 mmol) under the conditions of the previous experiment. Yield 0.164 g (77 %), R_f 0.15 (system D). Found (%): C, 54.34; H, 9.62; N, 2.39. C₂₆H₅₃O₇PNCl·H₂O. Calculated (%): C, 54.23; H, 9.63; N, 2.43. ¹H NMR, δ : 0.86 (t, 3 H, (CH₂)₁₂CH₃, J = 7 Hz); 1.25 (br.s, 24 H, (CH₂)₁₂); 1.55 (m, 2 H, CH(OAc)CH₂); 2.08 (s, 3 H, OC(O)CH₃); 3.22 (s, 9 H,

N⁺(CH₃)₃); 3.51–3.83 (m, 4 H, CH₂OCH₂); 3.61 (m, 2 H, CH₂N); 4.04 (m, 2 H, CH₂OP); 4.18 (m, 1 H, CHCl); 4.27 (m, 2 H, POCH₂); 5.0 (m, 1 H, CHOAc). ³¹P NMR, δ : 0.64. MS, *m/z* (*I*_{rel} (%)): 559.2 [M+H]⁺ (100).

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References

- F. Snyder, T. Ch. Lee, and M. L. Blank, in Annals of New York Academy of Sciences, Ed. Z. S. Khachaturien et al., New York, 1989, 568, 35.
- 2. Sh. D. Shukia, Lipids, 1991, 26, 1128.
- 3. N. M. Ruls, J. K. Rose, and F. N. Valone, *Lipids*, 1991, **26**, 1160.
- P. Braquet, D. Hosford, P. Kolts, J. Gullbaud, and M. Paubert-Braquet, *Lipids*, 1991, 26, 1071.
- 5. H. Hayashi, I. Kudo, Sh. Nojima, and K. Inoue, *Lipids*, 1991, 26, 1193.
- 6. Platelet Activating Factor and Related Lipid Mediators, Ed. F. Snyder, Plenum Press, New York, 1987.
- 7. M. A. Sablina, I. P. Ushakova, and G. A. Serebrennikova, *Khim.-Farm. Zh.* [*Chem. Pharm. J.*], 1994, **28**, No. 6, 9 (in Russian).
- 8. R. Andreesen and P. G. Munder, in *Platelet-Activating* Factor and Cell Immunology. New Trends in Lipid Mediator Research, Ed. P. Braquet, Karger, Basel, 1988, 2, 16.
- 9. J. E. Talmadge, M. Schneider, B. Lenz, H. Phillips, and C. Long, Lipids, 1987, 22, 871.
- 10. R. Andreesen and V. Giese, Lipids, 1987, 22, 836.
- M. A. Sablina, I. P. Ushakova, and G. A. Serebrennikova, *Khim.-Farm. Zh.* [*Chem. Pharm. J.*], 1993, 27, No. 6, 3 (in Russian).
- L. S. Kucera, N. Iyer, E. Leake, A. Raben, E. J. Modest,
 L. W. Daniel, and C. Piantadosi, *AIDS Res. and Hum. Retroviruses*, 1990, 6, 491.
- J.-J. Gogfroid, G. Dive, J. Lammotte-Brasseur, J.-P. Batt, and F. Heymans, *Lipids*, 1991, 26, 1162.
- 14. G. A. M. Stallberg, Acta Chem. Scand., 1990, 44, 368.
- 15. M. A. Sablina, I. P. Ushakova, and G. A. Serebrennikova, Mendeleev Commun., 1995, 1, 6.
- 16. E. A. Parfenov, G. A. Serebrennikova, and N. A. Preobrazhenskii, *Zh. Org. Khim.*, 1966, **2**, 633 [*J. Org. Chem. USSR*, 1966, **2** (Engl. Transl.)].
- M. A. Sablina, I. P. Ushakova, E. V. Dement'eva, A. A. Dergousov and G. A. Serebrennikova, *Bioorg. Khim.*, 1993, 19, 354 (in Russian).
- H. Brockerhoff and N. K. N Ayengar, *Lipids*, 1979, 14, 88.
- 19. R. Kumar, S. T. Weintraub, L. M. McManus, R. N. Pinckard, and R. N. Hanahan, J. Lipid Res., 1984, 25, 198.

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