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FACILE ASYMMETRIC SYNTHESIS OF THE D-MYO-INOSITOL DERIVATIVE FROM DIETHYL 2,3-O-ISOPROPYLIDENE-D-TARTRATE

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Abstract: A facile asymmetric synthesis of the D-myo-inositol derivative was achieved from diethyl 2,3-O-isopropylidene-D-tartrate in 7 steps. The two carboxylate groups of diethyl tartrate were converted simultaneously into dialdehyde to afford a key intermediate (1) for the synthesis of various protected D-myo-inositols.

Polyphosphoinositides play significant roles in the cellular signal transduction systems¹. Phosphatidyl-D-myo-inositol 4,5-bisphosphate is hydrolyzed by phospholipase C, which is activated on stimulation with many growth factors, to give diacylglycerol and D-myo-inositol 1,4,5-trisphosphate. Then diacylglycerol activates protein kinase C and D-myo-inositol 1,4,5-trisphosphate promotes release of intracellular Ca²⁺. In addition, PI-3 kinase phosphorylates phosphatidyl-D-myo-inositol 4,5-bisphosphate to phosphatidyl-D-myoinositol 3,4,5-trisphosphate, which may act as a second messenger in cellular signal transduction².

Since the supply of polyphosphoinositides from natural sources is very limited, extensive efforts have been made to synthesize racemic and homochiral inositol derivatives¹. Recently we synthesized 1-O-alkyland 1-O-acyl-D-myo-inositol 3,4,5-trisphosphates as new analogues of phosphatidyl-D-myo-inositol 3,4,5trisphosphate³. Here we describe a highly efficient asymmetric synthesis of enantiomerically pure inositol derivatives from diethyltartrate.

In 1987, Ozaki and co-workers⁴⁴ reported the synthesis of D-*myo*-inositol derivatives through intramolecular pinacol coupling mediated by low-valent titanium reagent. In 1994, Chiara et al.^{4b} and Mioskowski et al.^{4c} independently reported an elegant stereoselective synthesis of inositol derivatives via pinacol coupling of dialdehyde (1) using samarium diiodide. However, preparation of 1 requires multistep reactions from D-mannitol^{4b} or L-sorbose^{4c}. We planned to synthesize 1 from readily available diethyl 2,3-O-isopropylidene-D-tartrate in a stereo-controlled manner (Scheme 1).

Scheme 1



Scheme 2



Reagents and conditions: a. DIBALH, toluene, -78°C, 2 h, then sodium trimethylphosphonoacetate, DME, -78°C to r.t., 12 h, 74%; b. DIBALH, CH₂Cl₂, -78°C to 0°C, 1 h, 78%; c. Ti(OiPr)₄, D-(-)-diethyltartrate, cumene hydroperoxide, MS-4A, CH₂Cl₂, -20°C, 2 days, 73%; d. triphenylphosphine, imidazole, iodine, THF, 0°C, 5 min, then, NH₄Cl, Zn, EtOH, r.t., 30 min, 97%; e. *t*-butylchlorodiphenylsilane, imidazole, DMF, 0°C to r.t., 18 h, 99%; f. O₃, MeOH, -78°C, 20 min, then dimethylsulfide, -78°C to r.t., 30 min, 94%; g. samarium diiodide, *t*-BuOH, THF, -78°C to r.t., 18 h, 87%

DIBALH reduction of diethyl-2,3-O-isopropylidene-D-tartrate (3) followed by Wittig-Horner reaction in one pot afforded an (E,E)-diester $(EE : EZ = 10 : 1)^5$, which was reduced with DIBALH to give the (E,E)-bisallyl alcohol (4)⁶. Sharpless asymmetric epoxidation of 4 proceeded diastereoselectively to give the bisepoxy alcohol 5⁷ as the sole product. Substitution of the hydroxy groups in 5 with iodine and successive reduction with Zn⁸ gave 6⁹ in 97% yield. The protection of both hydroxy groups with t-butylchlorodiphenylsilane and ozonolysis of olefins, followed by reductive workup, gave the known dialdehyde 7^{4b}, which was transformed to 3,6-di-O-t-butyldiphenylsilyl-4,5-O-isopropylidene-D-myo-inositol (8) according to the reported procedure ^{4b}. The overall yield of 8 from 3 through 7 steps was 33 %.

Our current synthetic method should provide easy access to a variety of chiral polyphosphoinositides and other *myo*-inositol polyphosphate derivatives.

References and notes

- (1) Potter, B.V.L.; Lampe, D. Angew. Chem. Int. Ed. Engl., 1995, 34, 1933.
- (2) Kapeller, R.; Cantley, L.C. BioEssays, 1994, 16, 565.
- (3) Sawada, T.; Shirai, R.; Matsuo, Y.; Kabuyama, Y.; Kimura, K.; Fukui, Y.; Hashimoto, Y.; Iwasaki, S. BioMed. Chem. Lett., 1995, 5, 2263.
- (4) a) Watanabe, Y.; Mitani, M.; Ozaki, S. Chem. Lett., 1987, 123. b) Chiara J.L.; Martin-Lomas, M. Tetrahedron Lett., 1994, 35, 2969. c) Guidot, J.P.; Le Gall, T.; Mioskowski, C. Tetrahedron Lett., 1994, 35, 6671.
- (5) Krief, A.; Dumont, W.; Pasau, P.; Ph. Lecomte, Tetrahedron, 1989, 45, 3039.
- (6) Barrett, A.G.M.; Kasdorf, K.; Williams, D.J. J. Chem. Soc., Chem. Commun., 1994, 1781.
- ⁽⁷⁾ ¹H NMR (500 MHz, CDCl₃): δ 1.41 (s, 6H), 2.37 (br, 2H), 3.15 (brm, 2H), 3.22 (m, 2H), 3.76 (dd, J 3.5, 13.0 Hz, 2H), 3.87 (dd, J 3.0 and 13.0 Hz, 2H), 3.90 (m, 2H). ¹³C NMR (125 MHz, CDCl₃): δ 26.6, 54.3, 55.3, 60.9, 77.3, 110.8. [α]_D +33.8 (c=1.07, CHCl₃).
- (8) Hayashi, N.; Mine, T.; Fujiwara, K.; Murai, A. Chem. Lett., 1994, 2143.
- (9) ¹H NMR (500 MHz, CDCl₃): δ 1.45 (s, 6H), 2.29 (d, J 7 Hz, 2H), 4.01 (dd, J 1.5, 3.0 Hz, 2H), 4.14 (m, 2H), 5.27 (td, J 1.5, 10.5 Hz, 2H), 5.38 (td, J 1.5, 17.5 Hz, 2H), 5.89 (ddd, J 6.0, 10.5, 17.5 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃): δ 27.1, 71.8, 79.3, 109.6, 116.9, 136.7. [α]_D -3.05 (c=1.44, CHCl₃).

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