Synthesis and Biological Activities of Lipid A Analogs: Modification of a Glycosidically Bound Group with Chemically Stable Polar Acidic Groups and Lipophilic Groups on the Disaccharide Backbone with Tetradecanoyl or N-Dodecanoylglycyl Groups

Tsuneo Kusama,* Tsunehiko Soga, Yoshiyuki Ono, Eiji Kumazawa, Emiko Shioya, Kiyoshi Nakayama, Kouichi Uoto, and Yasuaki Osada

Exploratory Research Laboratories I, Daiichi Pharmaceutical Co., Ltd., 16-13, Kitakasai 1-chome, Edogawa-ku, Tokyo 134, Japan. Received June 19, 1991

Six novel lipid A analogs were synthesized. The first two analogs, 4 and 5, have an α -glycosidically bound carboxymethyl or 1,3-dicarboxyisopropyl group on the disaccharide backbone with four tetradecanoyl groups. The next three analogs, 6, 7 and 8, have two or four N-dodecanoylglycyl groups on the 1- α -O-phosphonooxyethylated disaccharide backbone. Analog 6 bears N-dodecanoylglycyl groups on the hydroxyl functions at positions 3 and 3', and tetradecanoyl groups on the amino functions at positions 2 and 2'. Analog 7 is a 2, 3, 2' and 3'-tetrakis(N-dodecanoylglycyl) derivative, and analog 8 resembles compound 6, but the binding of the N-dodecanoylglycyl and tetradecanoyl groups at positions 2, 2' and 3, 3' are reversed. The third analog, 9, has the same acyl group configuration as compound 6, but has a 1,3-dicarboxyisopropyl group at position C-1.

Compounds 4 and 5 exhibited definite antitumor activity against Meth A fibrosarcoma, indicating that the phosphate group at the C-1 position in lipid A could be replaced by the carboxylic acid without reducing the antitumor activity. In rabbits, compounds 6 and 9 exhibited potent antitumor activity, but their toxicity was extremely low. On the other hand, compounds 7 and 8 showed no antitumor activity. The levels of antitumor activity of 6 and 9 were similar to those of the natural-type lipid A. The antitumor activities of analogs with a N-dodecanoylglycyl group on the disaccharide backbone depended on the connecting sites of the acyl groups.

Keywords lipid A analog; *N*-dodecanoylglycyl group; phosphate group; phosphonooxyethyl group; carboxymethyl group; 1,3-dicarboxyisopropyl group; antitumor activity; Meth A; toxicity; structure-activity; rabbit

The complete structure of natural Escherichia coli (E. coli) lipid A was deduced¹⁾ and unequivocally confirmed by total synthesis.²⁾ This chemically synthesized E. coli lipid A showed biological activities identical to natural type, which possesses not only undesirable lethal toxicity but also beneficial characteristics such as its antitumor activity.³⁾

However, we found that 1, a novel synthetic analog of lipid A with an α -glycosidically bound phosphonooxyethyl group instead of the α -glycosyl phosphate group of natural lipid A, exhibited antitumor activity at the same level as synthetic *E. coli* lipid A.⁴⁾ It was proven that the α -glycosyl phosphate group is not essential for antitumor activity, and is replaceable, without loss of the activity, with other acidic groups such as chemically stable polar acidic groups.

We furthermore reported that compound 2, which has four (R)-3-hydroxytetradecanoyl groups as acyl groups on

HO)₂OPO
$$\begin{array}{c} OPO(OH)_2 \\ O$$

the disaccharide backbone and an α -glycosidically bound phosphonooxyethyl group, exhibited distinctly less toxicity in rabbits than 1- α -O-phosphonooxyethylated compound 3, which has four tetradecanoyl groups, and that, nevertheless, the difference of antitumor potency between these two compounds was not remarkable⁵⁾ (Fig. 1).

The above results suggested the possibility of separating the antitumor activity from the toxicity of lipid A. Because the only structural difference between these two compounds was the presence or absence of hydroxyl groups in the fatty acid moiety, it was presumed that the effect of hydroxyl groups in decreasing the toxicity resulted from the capacity for hydrogen bond formation and an increase of polarity.

We therefore directed our effort to synthesize new antitumor compounds toward investigating the possibility of converting the phosphate group at the C-1 position into other chemically stable polar acidic groups, such as carboxylic acids. We also believed that the conversion of the hydroxyl group in (R)-3-hydroxytetradecanoyl groups into other functional groups, likely to have the same effect as the hydroxyl group, might lead to the separation of the beneficial antitumor activity and the lethal toxicity of lipid A.

As regards the acidic groups other than phosphoric

i) GIyC1 2OH, DCC, DMAP; ii) 90% AcOH; iii) Troc-Cl, pyridine; iv) $(PhO)_2POCI$, DMAP; v) $(COD)Ir[PCH_3(C_6H_5)_2]_2 + PF_6^-$; vi) $I_2 - H_2O$; vii) Ac_2O , pyridine; viii) HBr/AcOH

Chart 1

acid, we tried introducing carboxylic acid derivatives to the C-1 position. We chose glycolic acid as a monobasic acid and 3-hydroxyglutaric acid as a dibasic acid. Accordingly, we synthesized compounds 4 and 5 by glycosylation so that these carboxylic acid substituents were attached to the disaccharide backbone bearing four tetradecanoyl groups without a hydroxyl.

We were also interested in replacing the hydroxyl groups of the fatty acid residues of compound 2 with NH amides which can be expected to exert an effect similar to that of hydroxyl on the capacity for hydrogen bond formation and on the increase of polarity. We considered that the amide bond "CONH", which can confer properties similar to those described above without having an asymmetric carbon and without basicity, would be a suitable functional group for our purpose. Since the -CONH- sites of the acyl groups should be located at

nearly the same positions as the hydroxyl groups in the (R)-3-hydroxytetradecanoyl group, and since the chain length should be similar to the length of the hydroxytetradecanoyl group, the N-dodecanoylglycyl group was chosen for introduction to the disaccaride backbone (Fig. 2).

Thus, compounds 6, 7, and 8, substituted totally or partially with N-dodecanoylglycyl groups, were synthesized (Chart 5). The results of the biological activity evaluation of the synthesized compounds described above made it desirable to synthesize compound 9, which has the same composition of acyl groups as compound 6, as well as a 1,3-dicarboxyisopropyl group at the C-1 position.

This paper describes the synthesis and biological activities of compound 4—9.

Chemistry The glycosyl donor 16 was synthesized from 11 by a method similar to that used for synthesizing the

glycosyl bromide 17 in our previous study,⁵⁾ as shown in Chart 1.

After 3-O-acylation of acetonide 11 with N-dodecanoylglycine in the presence of dicyclohexylcarbodiimide (DCC) and 4-dimethylaminopyridine (DMAP), the isopropylidene group was removed to give 3-O-acylated 12. Selective O-trichloroethoxycarbonylation at position C-6, followed by phosphorylation with diphenyl chlorophosphate at the C-4 position, yielded the 4-O-diphenylphosphorylated compound 13. After deallylation and acetylation of the C-1 position, the resulting glycosyl acetate 15 was brominated with 25% HBr-acetic acid to give the glycosyl bromide 16.

Routes for the synthesis of glucosamine derivatives 33a, 33b and 33f, glycosyl acceptors, are shown in Charts 2 and 3.

The selective glycosylation of bromide 18 with methylglycolate was employed in a similar manner, using zinc bromide, as reported for the synthesis of 24, 6) to give α -glycoside 19. The 2,2,2-trichloroethoxycarbonyl (Troc) group of compounds 19 and 24 were cleaved by treatment with zinc powder in acetic acid (AcOH), and a tetradecanoyl group was introduced into the resulting amino group by the acid chloride method or the DCC method to give N-tetradecanoyl derivatives 20a and 25b. Thereafter, all acetyl and methyl groups of 20a were removed by

i) $(CH_3)_2C(OMe)_2$, H⁺; ii) $(PhO)_2POCI$, DMAP; iii) $C_{14}OCI$, DMAP, pyridine; iv) $GlyC_{12}OH$, DCC, DMAP; v) Zn/AcOH; vi) 90%AcOH

Troc =CCI3CH2OCO

Chart 4

treatment with a NaOH solution, and the resulting carboxyl group of 21a was protected by a benzyl group to give 22a. On the other hand, compound 25b was deprotected by treatment with a LiOH solution, and without isolation of the deprotected compound, was immediately esterified with benzyl bromide and sodium hydrogen carbonate to give 26b. Hydroxyl groups at the 4 and 6 positions of 22a and 26b were protected by isopropylidene fromation, and the residual hydroxyl groups of 23a and 27b were acylated with tetradecanoic acid or N-dodecanoylglycine (10) by the same method as described for 20a and 25b. Deprotection of isopropylidene groups with 90% AcOH afforded the glycosyl acceptors 33a, 33b and 33f.

The other glucosamine components, 33c-e, were prepared from α -glycoside 28 as shown in Chart 4.

After acetonide formation at the 4 and 6 hydroxyl groups of 28, the remaining primary hydroxyl group was phosphorylated by treatment with diphenyl chlorophosphate to give the phosphonate 30. Acylation of 30 with tetradecanoyl chloride or N-dodecanoylglycine in the presence of DCC and DMAP gave 31c or 31e. After removal of the Troc group of 31c and 31e with zinc powder in AcOH, the products were acylated as described for 31c and 31e to yield 32c and 32e, respectively. Subsequent hydrolysis with 90% AcOH produced the glycosyl acceptors 33c and 33e. Glycosyl acceptor 33d was prepared from 30. After removal of the Troc group of 30, introduction of N-dodecanoylglycine to both of the resulting 2-amino and 3-hydroxyl functions, followed by hydrolysis in 90% AcOH, gave 33d.

The basic strategy for the synthesis of compounds **4**—**9** was similar to that employed in our previous synthesis⁵⁾ of the 1-O-phosphonooxyethylated compound **3**, as shown in Chart 5.

Coupling of glycosyl bromides 16 and 17 with the acceptors 33a—f yielded the disaccharides 34a—f. After cleavage of the Troc groups, the resultant free amino groups were acylated with tetradecanoic acid by the 1-hydroxybenzotriazole (HOBt) active ester method to give compounds 35a—f. Finally, hydrogenolytic deprotection of 35a—f afforded the desired compounds 4—9.

Antitumor Activity The antitumor activity of synthetic compounds was tested in BALB/c mice as described earlier. Briefly, a group of 7 or 8 mice were inoculated intradermally with Meth A syngeneic fibrosarcoma cells (2×10^5) . The triethylamine (Et_3N) salt of each compound was dissolved in an aqueous solution containing 5% glucose and 0.1% Et_3N . The resulting solution was then administered to the mice at a dose of $100 \,\mu\text{g/mouse}$ through the tail vein on the 7th, 12th and 17th days after implantation. The percentage antitumor effect on the growth of Meth A was determined by dividing the average tumor weight of the treated group on the 21st day by the average tumor weight of the control group, then multiplying the quotient by 100. Tables I and II show the

Toxicity The Et₃N salts of each of the compounds were dissolved in 5% (v/v) glucose containing 0.1% Et₃N to prepare a 100 μ g/ml solution. This solution was administered through the ear vein to three NZW rabbits per group at a dose of 50 or 500 μ g/kg-body weight for 3

TABLE I. Antitumor Activity of Lipid A Analogs against Meth A Fibrosarcoma

Compound No.	Dose (µg/mouse)	T/C (%) ^{a)}	Cured mice/ treated mice ^{b)}		
4	100 × 3	45°)	1/7		
5	100×3	24 ^{d)}	1/8		
6	100×3	21 ^{d)}	1/8		
7	100×3	80	0/7		
8	100×3	77	0/7		
9	100×3	20^{d}	0/7		
Control		100	0/8		

a) (Mean tumor weight in tested group/that in control group) \times 100. Results given are at 21 d after tumor inoculation. b) Number of tumor-free mice/number of mice tested. c) p < 0.01 vs. control (Student's test). d) p < 0.001 vs. control (Student's test).

consecutive days. Toxicity was evaluated by mortality, clinical signs, decrease in body weight, decrease in platelets (%), blood chemistry and pathology (Table III).

Discussion

Compounds 4 and 5, which have the same acyl composition and an α -glycosidically bound carboxymethyl group or 1,3-dicarboxyisopropyl group, exhibited definite antitumor activity. It becomes more clear that the conversion of substituents possessing carboxylic acid as an acidic group at the C-1 position, as well as those having glycosidically bound phosphoric acid or a phosphonooxy-

TABLE II. Antitumor Activity of Lipid A and Compound 6 against Meth A Fibrosarcoma

Compound No.	Dose (μg/mouse)	T/C (%) ^{a)}	Cured mice/ treated mice ^{b)}	
6	100 × 3	21 ^{d)}		
Synthetic E. coli	100 × 3	18 ^{d)}	0/8	
Lipid A ^{c)} Control —		100	0/8	

a) (Mean tumor weight in tested group/that in control group) \times 100. Results given are at 21 d after tumor inoculation. b) Number of tumor-free mice/number of mice tested. c) Synthetic *E. coli* lipid A (No. 506) was purchased from Daiichi Pure Chemicals Co., Ltd., Tokyo. d) p < 0.001 vs. control (Student's test).

ethyl group, retained the antitumor activity (Table I). The acidic polar moiety at the C-1 position of the dibasic acid analog 5 (which has greater acidity) demonstrated stronger antitumor activity than did monobasic acid analog 4. On the other hand, concerning toxicity in rabbits, dicarboxylic acid analog 5 showed a tendency to be more toxic than monocarboxylic acid analog 4, whereas the conversion of a phosphoric acid type group to a carboxylic acid type group at the C-1 position did not cause a decrease in toxicity compared with the phosphate type.

The antitumor activity of compound 6, which has two N-dodecanoylglycyl groups at the C-3 and 3' positions, was comparable to that of synthetic natural-type lipid A (Table II). However, the antitumor activity of 8, which is

TABLE III. Toxicity of Lipid A Analogs in Rabbits

Observations	Compound								
	4 Dose: 50	5 50	6 50	7 50	8 50	9 50	6 500	9 500 μg/kg	
Mortality ^{a)}	0/3	1/3	0/3	0/3	0/3	0/3	0/3	0/3	
Clinical signs ^{b)}	HE, RD	HE, AD, PT, LY	HE	NO ⁱ⁾	НE	NO	HE	NO	
Decrease in body weight (g) ^{c)}	100—400	350	NO	NO	NO	NO	NO	NO	
Hematological examination ^d	GPT \uparrow , UN \uparrow , CRE \uparrow , CPK \uparrow , TG \uparrow	GPT↑, UN↑, CRE↑, TP↓, ALB↓	NO	NO	UN†, CRE†, CHO†	NO	NO	NO	
Pathology ^{a)}	1	-							
Thrombus	$3/3 (3+)^{h}$	1/3 (+)	NO	NO	NO	NO	NO	NO	
Change in liver ^{e)}	1/3(3+)	2/3 (+)	NO	NO	NO	NO	NO	NO	
Change in kidney ^{f)}	1/3 (+)	3/3 (+)	NO	NO	1/3 (+)	NO	NO	NO	
Change in heart ^{g)}	2/3(+,3+)	1/3(3+)	NO	NO	NÒ	NO	NO	NO	

a) Mortality and pathology: number of rabbits changed/number of rabbits tested. b) HE, hyperemia of eye; RD, respiratory depression; AD, decrease in locomotor activity; PT, ptosis; LY, lying on side. c) 24 h after final injection. d) GPT, glutamate pyruvate transaminase; UN, urea nitrogen; CRE, creatinine; CPK, creatine phosphokinase; TG, triglyceride; TP, total protein; ALB, albumin; CHO, total cholesterol. e) Liver change: degeneration and necrosis of liver cells. f) Kidney change: degeneration and necrosis of epithelium of uriniferous tubules. g) Heart change: degeneration and necrosis of muscle fiber. h) +, slight; 2+, moderate; 3+, severe. i) NO: no change.

a positional isomer of compound 6 or 7 having four N-dodecanoylglycyl groups, was extremely weak. On the other hand, compounds 6, 7 and 8 did not show any lipid A toxicity on treatment with $50 \mu g/kg$ -body weight dose for 3 consecutive days in rabbits.

Since compound 6 showed both strong antitumor activity and low toxicity, it was compared to the activities of compound 9 in which only the C-1 substituent was different. As a result, 9 was found to have the same strong activity and low toxicity as the original compound 6, thus suggesting that, with the same acyl composition on the disaccharide backbone, the biological effects of the 1,3-dicarboxyisopropyl and phosphonooxyethyl groups were nearly the same. These results indicate that the structural requirements for the antitumor activities of analogs with N-dodecanoylglycyl groups on a disaccharide backbone are very strict, and the binding position on the disaccharide backbone of this group is very important for the expression of activity. For example, compounds 6 and 9, having a high antitumor activity, showed almost no toxicity on treatment with a dose of 500 μ g/kg-body weight (Table III), and these compounds showed a clear separation between the antitumor activity and the undesired toxicity of lipid A. Since compound 3, which has four (R)-3-hydroxytetradecanoyl groups, exhibited a definite antitumor activity, as reported in previous paper,5) and although compound 8, with four N-dodecanoylglycyl groups, did not show any such activity, the influence of the -CONH- of N-dodecanoylglycyl groups on antitumor activity differed considerably from that of the hydroxyl of 3-hydroxytetradecanoyl groups. This difference suggested that the N-dodecanoylglycyl group has another relevant property in addition to its capacity for hydrogen bond formation and polarity, probably due to the restricted rotation of the amide bond in a N-dodecanoylglycyl group.

In conclusion, compounds 6 and 9 represent the first successful examples of separating toxicity in natural-type lipid A from its significant antitumor activity, while retaining this activity. We hope that these findings will open the way for the clinical application of lipid A derivatives, and that these compounds will be useful for the investigation of a toxicological mode of action of natural-type lipid A as an endotoxin.

Experimental

All melting points are uncorrected. 1H -Nuclear magnetic resonance (1H -NMR) spectra were determined on a Varian XL-200 (200 MHz) or JEOL-GSX 500 (500 MHz) spectrometer in deuteriochloroform solution unless otherwise noted. The chemical shifts are given in δ values with tetramethylsilane (TMS) as the internal standard. Optical rotations were measured with a Horiba SEDA-200 polarimeter at 25 $^{\circ}$ C. Mass spectra (MS) were obtained on a JMS-HX 110 or JMX-300 instrument. Precoated Silica gel 150 A PLK5F plates (1.0 mm thickness; Whatmann) were used for preparative thin layer chromatography (TLC). Organic solution were dried over sodium sulfate before concentration.

N-Dodecanoylglycine (10) Dodecanoyl chloride (10.9 g, 50 mmol) and an aqueous solution (30 ml) of NaOH (2.00 g, 50 mmol) were added to a mixture of glycine (4.13 g, 55 mmol) and an aqueous solution (30 ml) of NaOH (2.20 g, 55 mmol) under ice-cooling maintained at pH 9. The mixture was stirred for 30 min, and then neutralized with 35% HCl with ice-cooling to about at pH 1. After the mixture was extracted with ethyl acetate (EtOAc), the organic layer was washed with H_2O , saturated aqueous NaCl, dried and concentrated. The residue was crystallized from EtOAc-hexane to give $10 (11.3 \, g, 88\%)$, mp $118-119^{\circ}C (118-119^{\circ}C).^{7}$ NMR (CDCl₃+CD₃OD) δ : 0.88 (3H, t, J=6 Hz, CH₃), 1.27 (16H, s, CH₂), 1.60 (2H, br, CH₂CCO₂).

Allyl 2-Deoxy-3-O-(N-dodecanoylglycyl)-2-(2,2,2-trichloroethoxycarbonylamino)- α -D-glucopyranoside (12) N-Dodecanoylglycine (10) (1.42 g, 5.52 mmol), DMAP 0.28 g (5.52 mmol) and DCC (1.14 g, 5.53 mmol) were added to a solution of 11 (2.00 g, 4.60 mmol) in CH₂Cl₂ (60 ml) with ice-cooling, and the mixture was stirred for 15 h at room temperature. The precipitate was filtered off, and the filtrate was concentrated in vacuo. The residue in 90% AcOH (50 ml) was heated at 90 °C for 20 min. Evaporation of the solvent gave a viscous oil, which was purified by silica gel column chromatography (CHCl₃-MeOH, 19:1) to give 12 as a colorless oil (2.60 g, 89%), [α]_D +54.8° (c=1.0, CHCl₃). NMR δ : 0.89 (3H, t, t=7 Hz, CH₃), 1.28 (16H, s, CH₂), 1.64 (2H, br, CH₂CH₂CO), 2.26 (2H, t, t=8 Hz, CH₂CON), 4.69 and 4.84 (each 1H, AB type d, t=12 Hz, CH₂CCl₃), 4.97 (1H, d, t=4 Hz, H-1), 5.2—5.5 (4H, m, H-3, CH=CH₂ and NHTroc), 5.96 (1H, m, CH=CH₂), 6.24 (1H, m, CH₂CONH). MS m/z: 633 [(M+H)⁺].

Allyl 2-Deoxy-4-*O*-diphenylphosphono-3-*O*-(*N*-dodecanoylglycyl)-6-*O*-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)-

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α-D-glucopyranoside (13) Troc-Cl (0.77 ml, 5.62 mmol) was gradually added to a solution of 12 (2.55 g, 4.02 mmol) in pyridine (20 ml) with ice-cooling. After the mixture was stirred for 1.5 h, H₂O was added and the solvent was evaporated. The residue was purified by silica gel column chromatography (CHCl₃-acetone, 9:1) to give a colorless oil (2.44 g, 75%). Diphenyl chlorophosphate (1.20 g, 4.47 mmol), DMAP (0.55 g, 4.47 mmol) and pyridine (0.35 ml, 4.47 mmol) were added to a solution of the above oil (2.41 g, 2.98 mmol) in CH₂Cl₂ (30 ml). After stirring for 1 h at room temperature, the mixture was diluted with CHCl₃, and washed successively with 1 m HCl, H2O, 5% aqueous NaHCO3 and saturated aqueous NaCl, and dried. After the solvent was evaporated, hexane was added to the residual oil to give 13 as a white powder (2.71 g, 87%), mp 80—82 °C, $[\alpha]_D$ +46.4° (c = 1.1, CHCl₃). Anal. Calcd for C₄₁H₅₃Cl₆N₂O₁₄P: C, 47.28; H, 5.13; N, 2.69. Found: C, 47.34; H, 5.04; N, 2.74. NMR δ : 0.89 (3H, t, J=7 Hz, CH₃), 1.28 (16H, s, CH₂), 1.58 (2H, br, CH_2CH_2CO), 2.10 (2H, t, J=8 Hz, CH_2CON), 3.80 (1H, dd, J=18, 4Hz, NCH₂CO₂), 4.00 (1H, dd, J=18, 4Hz, NCH₂CO₂), 4.67 and 4.82 (each 1H, AB type d, J=12 Hz, CH_2CCl_3), 4.80 (3H, m, H-4 and CH_2CCl_3), 5.01 (1H, d, J=4Hz, H-1), 5.3—5.5 (4H, m, H-3, NHTroc, $CH = C\underline{H}_2$), 5.96 (1H, m, $C\underline{H} = CH_2$), 6.27 (1H, m, CH₂CONH), 7.2-7.4 (10H, m, arom. H).

2-Deoxy-4-O-diphenylphosphono-3-O-(N-dodecanoylglycyl)-6-O-(2,2,2trichloroethoxy carbonyl) - 2 - (2,2,2 - trichloroethoxy carbonylamino) - D - glucose(14) Compound 13 (2.64 g, 2.53 mmol) and 1,5-cyclooctadienebis(methyldiphenylphosphine)iridium hexafluorophosphate (50 mg) were dissolved in tetrahydrofuran (THF, 40 ml) in a nitrogen atmosphere. After activation of the catalyst with a hydrogen atmosphere for 1 min, the mixture was heated at 50 °C in the nitrogen atmosphere for 2h. After cooling, iodine (1.28 g, 5.06 mmol) and H₂O (4 ml) were added to the solution and the mixture was stirred for 30 min. The solution was neutralized with $5\% \text{ Na}_2\text{SO}_3$ and extracted with CHCl₃. The solution was washed with saturated NaCl solution, dried, and concentrated. The residue was purified by silica gel column chromatography (CHCl₃acetone, 9:1) to give 14 (2.34 g, 92%) as a yellow oil, $[\alpha]_D$ +31.0° $(c=1.0, \text{ CHCl}_3)$. NMR δ : 0.88 (3H, t, J=7 Hz, CH₃), 1.28 (16H, s, CH_2), 1.57 (2H, br, CH_2CH_2CO), 2.12 (2H, t, J=8Hz, CH_2CON), 3.80 (1H, dd, J=18, 4Hz, NCH_2CO_2), 3.98 (1H, dd, J=18, 4Hz, NCH₂CO₂), 4.50—4.90 (4H, m, CH₂CCl₃), 5.38 (1H, m, H-1), 5.58 (2H, m, H-3 and NHTroc), 6.30 (1H, m, CH₂CONH), 7.1-7.4 (10H, m, arom. H).

1-O-Acetyl-2-deoxy-4-O-diphenylphosphono-3-O-(N-dodecanoylglycyl)-6-O-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)-D-glucopyranose (15) Acetic anhydride (1.18 g, 11.6 mmol) and pyridine (0.93 ml, 11.6 mmol) were added to a solution of 14 (2.31 g, 2.31 mmol) in CH₂Cl₂ (40 ml). After being stirred for 19 h, the mixture was diluted with CHCl₃. The solution was washed with 1 M HCl, H₂O and 5% NaHCO₃, dried, and concentrated. The residue was purified by silica gel column chromatography (CHCl₃-acetone, 19:1) to give 15 (2.25 g, 93%) as a pale yellow oil, $[\alpha]_D + 43.8^\circ$ (c = 1.2, CHCl₃). NMR δ : 0.88 (3H, t, J=7 Hz, CH₃), 1.28 (16H, s, CH₂), 1.58 (2H, CH₂CH₂CO), 2.11 (2H, t, J=8 Hz, CH₂CON), 2.25 (3H, s, OAc), 3.77 (1H, dd, J=18, 4 Hz, $NC_{12}CO_{2}$, 4.07 (1H, dd, J=18, 4Hz, $NC_{12}CO_{2}$), 4.57 and 4.74 (each 1H, AB type d, J = 12 Hz, CH_2CCl_3), 4.86 and 4.98 (each 1H, AB type d, J=12 Hz, CH_2CCl_3), 4.92 (1H, q, J=8 Hz, H-4), 5.18 (1H, d, J=10 Hz, NHTroc), 5.50 (1H, t, J=10 Hz, H-3), 6.31 (2H, m, H-1 and CH₂CONH), 7.2—7.4 (10H, m, arom. H).

Methoxycarbonylmethyl 3,4,6-Tri-O-acetyl-2-deoxy-2-(2,2,2-trichloro-ethoxycarbonylamino)-α-D-glucopyranoside (19) Zinc bromide (140 mg, 0.62 mmol) was added to a suspension of 18^{71} (330 mg, 0.61 mmol), methyl glycolate (65 mg, 0.73 mmol) and CaSO₄ (100 mg) in CH₂Cl₂ (5 ml) at room temperature, and the mixture was refluxed for 5 h, then diluted with CHCl₃, and filtered. The filtrate was washed with 5% aqueous NaHCO₃, dried, and concentrated. The residue was purified by silica gel column chromatography (CHCl₃-acetone, 50:1) to give 19 (204 mg, 61%) as a powder, mp 110—111 °C, [α]_D +85.6° (c=0.9, CHCl₃). Anal. Calcd for C₁₈H₂₄Cl₃NO₁₂: C, 39.11; H, 4.38; N, 2.53. Found: C, 39.13; H, 4.37; N, 2.48. NMR δ: 2.01, 2.03 and 2.10 (each 3H, s, OAc), 3.78 (3H, s, OCH₃), 4.1—4.3 (6H, m), 4.63 and 4.86 (each 1H, AB type d, J=12 Hz, CH₂CCl₃), 4.97 (1H, d, J=4 Hz, H-1), 5.13 (1H, t, J=10 Hz, H-4), 5.31 (1H, t, J=10 Hz, H-3), 5.63 (1H, d, J=10 Hz). MS m/z: 552 [(M+H)⁺].

Methoxycarbonylmethyl 3,4,6-Tri-O-acetyl-2-deoxy-2-tetradecanoylamino-α-D-glucopyranoside (20a) Zinc powder (2.0 g) was added to a solution of 19 (1.60 g, 2.89 mmol) in AcOH (20 ml), and the mixture was vigorously stirred for 3 h at room temperature. The insoluble materials

were removed by filtration, and the filtrate was concentrated in vacuo. The resulting oily product was dissolved in EtOAc, and this solution was washed with 5% aqueous NaHCO₃, and dried. After evaporation of the solvent, tetradecanoyl chloride (1.07 g, 4.34 mmol) and Nmethylmorpholine (0.48 ml, 4.34 mmol) were added to a solution of the resulting oily product in CH₂Cl₂ (20 ml). The mixture was vigorously stirred for 1 h. MeOH (5 ml) was added to the reaction mixture. After 1 h of stirring, the mixture was diluted with CHCl₃, washed with 1 m HCl, saturated aqueous NaCl, dried, and concentrated. The residue was purified by silica gel column chromatography (CHCl₃-acetone, 50:1) to give **20a** (1.68 g, 99%) as a wax, $[\alpha]_D + 68.6^{\circ}$ (c = 1.2, CHCl₃). NMR δ : 0.89 (3H, t, J=7 Hz, CH₃), 1.26 (20H, s, CH₂), 1.63 (2H, br, CH₂CH₂CO), 2.02, 2.04 and 2.11 (each 3H, s, OAc), 2.20 (2H, m, CH₂CO), 3.78 (3H, s, CO₂CH₃), 4.1—4.5 (4H, m), 4.26 (2H, s, OCH₂CO₂), 4.90 (1H, d, J=4 Hz, H-1), 5.18 (1H, t, J=10 Hz, H-4), 5.32 (1H, t, J=10 Hz, H-3), 6.15 (1H, d, J = 10 Hz, NH). MS m/z: 588 [(M+H)⁺].

Carboxymethyl 2-Deoxy-2-tetradecanoylamino- α -D-glucopyranoside (21a) A solution of 1 M NaOH (12.2 ml) was added to a solution of 20a (1.63 g, 2.77 mmol) in THF (20 ml) with ice-cooling, and the mixture was stirred for 20 min at the same temperature, then neutralized with 10% citric acid. The insoluble material was collected by filtration, washed with H_2O and dried in vacuo to give 21a (1.16 g, 93%) as a white solid, mp 153-156 °C, $[\alpha]_D + 93.5$ ° (c=1.0, DMF). NMR (CD₃OD-CDCl₃, 1:1) δ : 0.88 (3H, t, J=7 Hz, CH₃), 1.26 (20H, s, CH₂), 1.60 (2H, br, CH₂CH₂CO), 2.26 (2H, t, J=7 Hz), 4.20 (2H, d, J=5.4 Hz, OCH₂CO₂). MS m/z: 448 $[(M+H)^+]$.

Benzyloxycarbonylmethyl 2-Deoxy-2-tetradecanoylamino-α-D-glucopyranoside (22a) Benzyl bromide (0.86 g, 5.00 mmol) and Et₃N (0.70 ml, 5.00 mmol) were added to a solution of 21a (1.12 g, 2.50 mmol) in dimethylformamide (DMF, 20 ml), and the mixture was heated at 60 °C for 2 h. The mixture was diluted with EtOAc, and washed with H₂O and 1 m HCl. The organic layer was concentrated *in vacuo*, and the residue was purified by silica gel column chromatography (CHCl₃-MeOH, 9:1) to give 22a (1.18 g, 88%) as a white powder, mp 268—269 °C, $[\alpha]_D$ +82.0° (c=1.1, DMF). Anal. Calcd for C₂₉H₄₇NO₈: C, 64.78; H, 8.81; N, 2.61. Found: C, 64.63; H, 8.78; N, 2.52. NMR δ: 0.87 (3H, t, J=7 Hz, CH₃), 1.24 (20H, s, CH₂), 1.50 (2H, br, CH₂CH₂CO), 2.10 (2H, t, J=7 Hz, CH₂CO), 4.26 (2H, s, OCH₂CO₂), 4.83 (1H, d, J=4 Hz, H-1), 5.18 (2H, s, CH₂C₆H₅), 7.42 (5H, s, arom. H), 7.64 (1H, d, J=8 Hz, NH). MS m/z: 538 [(M+H)⁺].

Benzyloxycarbonylmethyl 2-Deoxy-4,6-O-isopropylidene-2-tetradecanoylamino-α-D-glucopyranoside (23a) Dimethoxypropane (0.45 g, 4.28 mmol) was added to a solution of 22a (1.15 g, 2.14 mmol) in DMF (40 ml) in the presence of p-toluenesulfonic acid (p-TsOH·H₂O, 38 mg, 0.2 mmol), and the solution was stirred for 3.5 h. After evaporation of the solvent, the residue was purified by silica gel column chromatography (CHCl₃–acetone, 20:1) to give 23a (1.10 g, 89%) as a white powder, mp 72—73 °C, [α]_D +35.0° (c=1.1, CHCl₃). Anal. Calcd for C₃₂H₅₁NO₈: C, 66.52; H, 8.90; N, 2.42. Found: C, 66.42; H, 8.87; N, 2.51. NMR δ: 0.88 (3H, t, J=6 Hz), 1.26 (20H, s, CH₂), 1.45 and 1.56 (each 3H, s, CCH₃), 1.66 (2H, br, CH₂CH₂CO), 2.30 (2H, t, J=8 Hz, CH₂CO), 4.28 (2H, s, OCH₂CO₂), 4.82 (1H, d, J=4 Hz, H-1), 5.25 (2H, s, CH₂C₆H₅), 7.42 (5H, s, arom. H). MS m/z: 578 [(M+H)⁺].

Benzyloxycarbonylmethyl 2-Deoxy-3-O-tetradecanoyl-2-tetradecanoylamino-α-D-glucopyranoside (33a) As described for 20a, compound 23a (1.07 g, 1.85 mmol) was reacted with tetradecanoyl chloride (640 mg, 2.60 mmol) in CH₂Cl₂ (20 ml) in the presence of pyridine (0.73 ml, 2.60 mmol) and DMAP (40 mg) with ice-cooling. After the usual workup, the resulting oily residue was dissolved in 90% AcOH (40 ml), and the mixture was heated at 90 °C for 30 min. Evaporation of the solvent gave an oil, and the oily product was purified by silica gel column chromatography (CHCl₃-MeOH, 19:1) to give 33a (1.24g, 90%) as a colorless solid, mp 54—55 °C, $[\alpha]_D$ +46.9° (c=1.1, CHCl₃). Anal. Calcd for C₄₃H₇₃NO₉: C, 69.04; H, 9.84; N, 1.87. Found: C, 68.75; H, 9.67; N, 1.82. NMR δ : 0.89 (6H, t, J=7 Hz), 1.28 (40H, s, CH₂), 1.60 (4H, br, $CH_2CH_2CO \times 2$), 2.15 and 2.35 (each 2H, m, CH_2CO), 3.8—3.9 (4H, m), 4.27 (2H, s, OCH₂CO₂), 4.33 (1H, m, H-2), 4.89 (1H, d, J=4 Hz, H-1), 5.18 (1H, m, H-3), 5.22 (2H, s, $C_{12}C_{6}H_{5}$), 6.17 (1H, d, J=9 Hz, NH), 7.41 (5H, s, arom. H). MS m/z: 748 $\lceil (M+H)^+ \rceil$.

1,3-Di(methoxycarbonyl)isopropyl 2-Deoxy-3,4,6-tri-O-acetyl-2-tetra-decanoylamino-α-D-glucopyranoside (25b) Zinc powder (20 g) was added to a solution of 24⁶¹ (23.2 g, 0.04 mmol) in AcOH (100 ml), and the mixture was vigorously stirred for 1 h at room temperature. The insoluble materials were filtered off, and the filtrate was concentrated in vacuo. The resulting oily substance was dissolved in EtOAc, and the

solution was washed with 5% aqueous NaHCO3, and dried. The solvent was distilled off at reduced pressure, and the oily product was dissolved in CH₂Cl₂ (200 ml). Tetradecanoic acid (8.2 g, 0.04 mol) and DCC (7.50 g, 0.04 mol) were added to the solution with ice-cooling, and the mixture was stirred for 1 h at room temperature. The precipitate was filtered off, and the filtrate was concentrated in vacuo. The oily residue was purified by silica gel column chromatography (CHCl3-acetone, 20:1) to give 25b (15.6 g, 77%) as a colorless oil, $[\alpha]_D$ +49.3° (c=1.4, CHCl₃). NMR δ : 0.88 (3H, t, J=7 Hz), 1.25 (20H, s, CH₂), 1.58 (2H, m, CH₂CH₂CO), 1.99, 2.02 and 2.09 (each 3H, s, OAc), 2.56-2.67 (5H, m, CH_2CON , $OCH(CH_2CO_2Me)_2 \times 3$), 2.82 (1H, dd, J=16, 6 Hz, OCH(CH₂CO₂Me)₂), 3.70 and 3.72 (each 3H, s, CO₂CH₃), 4.04 (1H, m, H-5), 4.10 (1H, dd, J=12, 2Hz, H-6), 4.20 (1H, dd, J=12, 5 Hz, H-6), 4.36 (1H, td, J = 10, 4 Hz), 4.45 (1H, m, OCH(CH₂CO₂Me)₂), 4.96 (1H, d, J=4 Hz, H-1), 5.09 (1H, t, J=10 Hz, H-4), 5.14 (1H, t, J = 10 Hz, H-3), 6.28 (1H, d, J = 10 Hz, NH).

1,3-Di(benzyloxycarbonyl)isopropyl 2-Deoxy-2-tetradecanoylamino-α-Dglucopyranoside (26b) Compound 25b (500 mg, 0.74 mmol) was dissolved in MeOH-H₂O (3:1, 20 ml), and LiOH·H₂O (472 mg, 11 mmol) was added to the mixture with ice-cooling. The mixture was stirred for 12h at room temperature. 1 M HCl was added to the mixture until the pH of the solution became acidic, then 5% NaHCO3 was added until the neutral pH was achieved. After the solution was concentrated in vacuo, the residue was dissolved in DMF (14 ml). NaHCO₃ (624 mg, 7.4 mmol) and benzyl bromide (2.2 ml, 18.5 mmol) were added to this solution. After stirring for 24h at room temperature, the solvent was removed at reduced pressure. The residue was purified by silica gel column chromatography (CHCl₃-MeOH, 20:1) to give 26b (337 mg, 65%) as a colorless wax, $[\alpha]_D + 13.2^\circ$ (c=0.5, CHCl₃). Anal. Calcd for C₃₉H₅₇NO₁₀·H₂O: C, 65.25; H, 8.30; N, 1.95. Found: C, 65.20; H, 8.21; N, 2.12. NMR (500 MHz) δ : 0.88 (3H, t, J = 7.3 Hz), 1.24 (20H, s, CH₂), 1.64 (2H, m, CH₂CH₂CO), 2.25 (2H, m, CH₂CO), 2.59 (1H, dd, J=16.7, 5.5 Hz, $OCH(CH_2CO_2Bzl)_2$), 2.65 (1H, dd, J=16.7, 4.0 Hz, $OCH(C\underline{H}_2CO_2Bzl)_2)$, 2.72 (1H, d, J=16.7, 8.7 Hz, $OCH(C\underline{H}_2CO_2Bzl)_2)$, 2.82 (1H, dd, J=16.7, 6.4 Hz, OCH(CH₂CO₂Bzl)₂), 3.49 (1H, t, J= $8.7 \,\mathrm{Hz}$), $3.58 \,\mathrm{(1H,\ t,\ } J\!=\!10.3 \,\mathrm{Hz}$), $3.71 \,\mathrm{(2H,\ m)}$, $3.81 \,\mathrm{(1H,\ m)}$, $3.94 \,\mathrm{(1H,\ m)}$ m), 4.46 (1H, m, $OCH(CH_2CO_2BzI)_2$), 4.92 (1H, d, J=4.0Hz, H-1), 5.14 (2H, s, $CH_2C_6H_5$), 5.15 and 5.18 (each 1H, AB type d, J=11.9 Hz, $CH_2C_6H_5$), 7.12 (1H, m, NH), 7.3—7.4 (10H, m, arom. H). MS m/z:

1,3-Di(benzyloxycarbonyl)isopropyl 2-Deoxy-4,6-O-isopropylidene-2tetradecanoylamino-a-d-glucopyranoside (27b) In a manner similar to that described for 23a, compound 26b (640 mg, 0.91 mmol) was treated with 2,2-dimethoxypropane (0.5 ml) in the presence of p-TsOH·H₂O (30 mg) in acetone (10 ml) to give 27b (540 mg, 80%) as a colorless oil, $[\alpha]_D$ +3.3° (c=0.7, CHCl₃). Anal. Calcd for $C_{42}H_{61}NO_{10}\cdot 1/4H_2O$: C, 67.75; H, 8.34; N, 1.87. Found: C, 67.53; H, 8.09; N, 1.97. NMR (500 MHz) δ : 0.88 (3H, t, J = 7.3 Hz), 1.24 (20H, s, CH₂), 1.43 and 1.52 (each 3H, s, CCH₃), 1.64 (2H, m, C \underline{H}_2 CH₂CO), 2.25 (2H, td, J=7.3, 2.8 Hz, CH₂CO), 2.57 (1H, dd, J=15.6, 6.4 Hz, OCH(CH₂CO₂Bzl)₂), 2.69 (2H, m, OCH($CH_2CO_2Bzl)_2 \times 2$), 2.78 (1H, dd, J=15.6, 6.4 Hz, OCH(CH₂CO₂Bzl)₂), 3.57—3.70 (4H, m), 3.80 (1H, m), 4.05 (1H, m), 4.43 (1H, m, $OCH(CH_2CO_2BzI)_2$), 4.89 (1H, d, J=3.7Hz, H-1), 5.10 and 5.13 (each 1H, AB type d, $J=12.8\,\mathrm{Hz}$, $\mathrm{C}\underline{\mathrm{H}}_{2}\mathrm{C}_{6}\mathrm{H}_{5}$), 5.16 (2H, s, $CH_2C_6H_5$), 6.96 (1H, d, J=7.3 Hz, NH), 7.3—7.4 (10H, m, arom. H). MS m/z: 741 [(M+2)⁺].

1,3-Di(benzyloxycarbonyl)isopropyl 2-Deoxy-3-O-tetradecanoyl-2-tetradecanoylamino- α -D-glucopyranoside (33b) Compound 27b (480 mg, 0.65 mmol) and tetradecanoic acid (190 mg, 0.85 mmol) in CH₂Cl₂ (10 ml) were added to DMAP (100 mg, 0.85 mmol) and DCC (170 mg, 0.85 mmol) with ice-cooling, and the mixture was stirred for 2 h at room temperature. The insoluble material was filtered off, and the filtrate was concentrated at reduced pressure. The residue was dissolved in 90% AcOH (40 ml), and the mixture was heated at 90 °C for 20 min. After evaporation of the solvent, the residue was purified by silica gel column chromatography (CHCl₃-acetone, 10:1) to give 33b (530 mg, 89%) as a colorless wax, [α]_D +32.8° (c=0.9, MeOH). Anal. Calcd for C₅₃H₈₃NO₁₁: C, 69.94; H, 9.19; N, 1.54. Found: C, 69.73; H, 9.03; N, 1.60. NMR & 0.88 (6H, t, CH₃), 1.26 (40H, s, CH₂), 1.60 (4H, br, CH₂CH₂CO), 2.16 (2H, m, CH₂CO), 2.34 (2H, m, CH₂CO), 4.94 (1H, s, H-1), 5.20 (4H, s, CH₂C₆H₅×2), 7.40 (10H, s, arom. H). MS m/z: 911 [(M+2)+].

1,3-Di(benzyloxycarbonyl)isopropyl 2-Deoxy-3-O-(N-dodecanoylglycyl)-2-tetradecanoylamino-α-D-glucopyranoside (33f) As described for 33b, compound 27b (598 mg, 0.81 mmol) was treated with 10 (250 mg, 0.97 mmol) in the presence of DMAP (49 mg, 0.40 mmol) and DCC

(200 mg, 0.97 mmol), and the resulting oil was treated with 90% AcOH to give 33f (632 mg, 83%) as a wax, $[\alpha]_D$ +36.9° (c=1.3, CHCl₃). NMR (500 MHz) δ : 0.88 (6H, t, J=7.3 Hz, CH₃), 1.28 (36H, m, CH₂), 1.55—1.67 (4H, m, CH₂CH₂CO×2), 2.16 (2H, m, CH₂CO), 2.26 (2H, t, J=7.3 Hz, CH₂CO), 2.60 (1H, dd, J=16.5, 6.4 Hz, OCH(CH₂CO₂Bzl)₂), 2.72 (1H, dd, J=16.5, 8.2 Hz, OCH(CH₂CO₂Bzl)₂), 2.81 (1H, dd, J=16.5, 5.5 Hz, OCH(CH₂CO₂Bzl)₂), 3.66 (1H, t, J=9.2 Hz), 3.74—3.83 (4H, m), 4.12 (1H, dd, J=17.4, 5.5 Hz), 4.19 (1H, td, J=9.2, 3.7 Hz), 4.46 (1H, m, OCH(CH₂CO₂Bzl)₂), 4.92 (1H, d, J=4.6 Hz, H-1), 5.03 (1H, t, J=9.2 Hz, H-3), 5.10—5.18 (4H, m, CH₂C₆H₅), 6.43 (1H, br, NH), 6.65 (1H, d, J=9.2 Hz, NH), 7.32—7.40 (10H, m, arom. H). MS m/z: 939 [(M+H)⁺].

2-Hydroxyethyl 2-Deoxy-4,6-*O*-isopropylidene-2-(2,2,2-trichloroethoxy-carbonylamino)-α-D-glucopyranoside (29) As described for 23a, compound 28 (3.58 g, 8.98 mmol) was treated with 2,2-dimethoxypropane (3.70 ml, 30.2 mmol) in the presence of p-TsOH·H₂O (170 mg, 0.89 mmol) to give 29 (2.78 g, 71%) as a white powder, mp 190—192 °C. *Anal.* Calcd for C₁₄H₂₂Cl₃NO₈: C, 38.33; H, 5.06; Cl, 24.25; N, 3.19. Found: C, 38.40; H, 5.02; Cl, 24.46; N, 3.59. NMR δ: 1.45 and 1.54 (each 3H, s, CCH₃), 4.75 and 4.86 (each 1H, AB type d, J=12 Hz, CH₂CCl₃), 4.95 (1H, d, J=4 Hz, H-1), 5.80 (1H, d, NH). MS m/z: 438 [(M+H)⁺].

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-4,6-O-isopropylidene-2-(2,2,2trichloroethoxycarbonylamino)-\a-D-glucopyranoside (30) Diphenyl chlorophosphate (0.80 ml, 3.86 mmol), pyridine (0.4 ml) and DMAP (0.46 g, 3.77 mmol) were added to a solution of 29 (1.12 g, 2.55 mmol) in CH_2Cl_2 (20 ml) with ice cooling. After the mixture was stirred for 3 h, diphenyl chlorophosphate (0.30 ml, 1.45 mmol) and DMAP (0.19 mmol, 1.51 mmol) were added with ice cooling. After the addition of MeOH (1 ml), the mixture was stirred for 1h and diluted with CHCl3. The solution was washed with 1 M HCl, with aqueous 5% NaHCO₃, then with H₂O, and dried. After evaporation of the solvent, the residue was purified by silica gel column chromatography (CHCl₃-acetone, 30:1) to give 30 (1.23 g, 71.8%) as a white solid, mp 121—124 °C, $[\alpha]_D$ +46.4° (c=1.0, CHCl₃). Anal. Calcd for C₂₆H₃₁Cl₃NO₁₁P: C, 46.55; H, 4.66; N, 2.09. Found: C, 46.28; H, 4.55; N, 2.13. NMR δ : 1.45 and 1.52 (each 3H, s, CCH₃), 4.45 (2H, m, CH₂OP), 4.73 and 4.80 (each 1H, AB type d, J = 12 Hz, CH_2CCl_3), 4.85 (1H, d, H-1), 7.10—7.50 (10H, m, arom. H). MS m/z: 671 [(M+2)⁺].

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-4,6-*O*-isopropylidene-3-*O*-(*N*-dodecanoylglycyl)-2-(2,2,2-trichloroethoxycarbonylamino)- α -D-glucopyranoside (31c) Compound 30 (1.89 g, 2.82 mmol) was allowed to react with *N*-dodecanoylglycine (0.83 g, 3.22 mmol) in the presence of DCC (0.67 g, 3.23 mmol) and DMAP (0.17 g, 1.40 mmol), in a manner similar to that used for 33b, to give 31c (2.57 g, quant.) as a colorless oil, $[\alpha]_D$ + 32.2° (c=0.8, CHCl₃). NMR δ : 0.88 (3H, t, J=6 Hz), 1.28 (16H, br, CH₂), 1.38 and 1.48 (each 3H, s, CCH₃), 2.20—2.35 (2H, t, CH₂CON), 4.20 (2H, m, CH₂OP), 5.25 (1H, m, H-3), 7.2—7.5 (10H, m, arom. H).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-4,6-*O*-isopropylidene-3-*O*-tetradecanoyl-2-(2,2,2-trichloroethoxycarbonylamino)-α-D-glucopyranoside (31e) As described for 33a, compound 30 (0.50 g, 0.75 mmol) was treated with tetradecanoyl chloride (221 mg, 0.90 mmol) in the presence of pyridine (0.30 ml, 3.75 mmol) and DMAP (20 mg) in CH₂Cl₂ to give 31e (0.49 g, 74%) as a colorless oil, $[\alpha]_D$ +36.1° (c=1.0, CHCl₃). *Anal.* Calcd for C₄₀H₅₇Cl₃NO₁₂P: C, 54.52; H, 6.52; N, 1.59. Found: C, 54.41; H, 6.81; N, 1.72. NMR δ: 0.88 (3H, t, J=7Hz), 1.28 (16H, s, CH₂), 1.37 and 1.47 (each 3H, s, CCH₃), 1.60 (2H, br, CH₂CH₂CO), 2.30 (2H, m), 4.42 (2H, m, CH₂OP), 4.58 and 4.80 (each 1H, AB type d, J=12Hz, CH₂CCl₃), 4.86 (1H, d, J=4 Hz, H-1), 5.22 (1H, m, H-3), 5.65 (1H, d, J=10 Hz, NH), 7.2—7.5 (10H, m, arom. H). MS m/z: 881 [(M+2)⁺].

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-4,6-O-isopropylidene-3-O-(N-dodecanoylglycyl)-2-tetradecanoylamino-α-D-glucopyranoside (32c) As described for 20a, compound 31c (0.66 g, 0.73 mmol) was treated with zinc powder in AcOH (5 ml), and the resulting oil was treated with tetradecanoyl chloride (0.23 g, 0.93 mmol) in CH₂Cl₂ (5 ml) to give 32c (0.67 g, 97%) as a colorless oil, $[\alpha]_D$ +30.1° (c=1.7, CHCl₃). NMR δ: 0.88 (6H, t, J=6 Hz), 1.28 (40H, br, CH₂), 1.38 and 1.48 (each 3H, s, CCH₃), 1.64 (5H, br), 2.09 (2H, m), 2.26 (2H, m), 2.35 (2H, m), 3.6—4.2 (6H, m), 4.3—4.5 (3H, m), 4.78 (1H, d, J=4 Hz, H-1), 5.21 (1H, m, H-3), 6.25 (1H, br, NH), 6.95 (1H, m, NH), 7.2—7.5 (10H, m, arom. H).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-2-(N-dodecanoylglycylamino)-4,6-O-isopropylidene-3-O-tetradecanoyl-\(\alpha\)-Delucopyranoside (32e) As described for 25b, compound 31e (0.47 mg, 0.53 mmol) was treated with zinc

powder (0.5 g), and the resulting oil was allowed to react with 10 (0.21 g, 0.8 mmol) to give 32e (0.48 g, 94%) as a powder, mp 79—80 °C, $[\alpha]_D$ +28.1° (c=1.1, CHCl₃). NMR δ : 0.88 (6H, t, J=6 Hz), 1.28 (36H, s, CH₂), 1.37 and 1.63 (each 3H, s, CCH₃), 2.12 (2H, m, CH₂CO), 2.30 (2H, t, J=8 Hz, CH₂CO), 4.42 (2H, m, CH₂OP), 4.86 (1H, d, J=4 Hz, H-1), 5.20 (1H, t, J=10 Hz, H-3), 6.78 (1H, br, NH), 6.94 (1H, d, J=8 Hz, NH), 7.2—7.5 (10H, m, arom. H).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-3-*O*-(*N*-dodecanoylglycyl)-2-tetradecanoylamino-α-D-glucopyranoside (33c) A solution of 32c (0.63 g, 0.67 mmol) in 90% AcOH (20 ml) was heated at 90 °C for 30 min. After evaporation of the solvent, the residue was purified by silica gel column chromatography (CHCl₃-acetone, 10:1) to give 33c (0.45 g, 73%) as an oil, $[\alpha]_D$ +46.8° (c=0.9, CHCl₃). NMR δ: 0.88 (6H, t, J=6 Hz), 1.2—1.8 (40H, br, CH₂), 2.08 (2H, t), 2.27 (2H, t), 3.7—4.0 (8H, m), 4.05—4.50 (4H, m), 4.83 (1H, d, J=4 Hz, H-1), 5.18 (1H, m, H-3), 6.43 (1H, m, NH), 6.85 (1H, m, NH), 7.2—7.5 (10H, m, arom. H). MS m/z: 906 $[(M+2)^+]$ 1

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-2-(*N*-dodecanoylglycylamino)-3-*O*-tetradecanoyl-α-D-glucopyranoside (33e) As described for 33c, compound 32e (0.46 g, 0.48 mmol) was treated with 90% AcOH (20 ml) to give 33e (0.39 g, 90%) as a waxy solid, $[\alpha]_D + 36.1^\circ$ (c = 1.1, CHCl₃). NMR δ: 0.90 (6H, t, J = 6 Hz), 1.28 (36H, s, CH₂), 1.63 (4H, br, CH₂CH₂CO × 2), 2.13 (2H, m, CH₂CO), 2.36 (2H, t, J = 8 Hz, CH₂CO), 4.90 (1H, d, J = 4 Hz, H-1), 5.10 (1H, m, H-3), 6.74 (1H, br, NH), 6.96 (1H, d, J = 9 Hz, NH), 7.2—7.5 (10H, m, arom. H). MS m/z: 906 $[(M+2)^+]$.

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-3-O-(N-dodecanoylglycyl)-2-(N-dodecanoylglycylamino)- α -D-glucopyranoside (33d) Compound 30 (0.60 g, 0.89 mmol) was treated with zinc powder (0.6 g) in AcOH (10 ml). After stirring for 1 h, the mixture was filtered off, and the filtrate was concentrated in vacuo to give an oil, which was then allowed to react with 10 (0.69 g, 2.67 mmol). The resulting oil was treated with 90% AcOH, in the same manner as described for 33f, to give 33d as a colorless oil (0.57 g, 93%), $[\alpha]_D$ +16.6° (c=0.8, CHCl₃). NMR δ : 0.90 (6H, t, J=6 Hz), 1.28 (32H, s, CH₂), 1.60 (4H, br, CH₂CH₂CO×2), 2.0—2.3 (4H, m, CH₂CO×2), 4.85 (1H, d, J=4 Hz, H-1), 5.24 (1H, t, J=10 Hz, H-3), 6.88 (1H, d, J=10 Hz, NH), 7.2—7.5 (12H, m, arom. H and NH×2). MS m/z: 934 [(M+H)⁺].

 $Benzyloxy carbonyl methyl\ 2-Deoxy-6-\emph{O-}[2-deoxy-4-\emph{O-}diphenyl phosphono-diphenyl phosphon-diphenyl phosphono-diphenyl ph$ 3-O-tetradecanoyl-6-O-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)-\(\beta\)-D-glucopyranosyl]-3-O-tetradecanoyl-2-tetradecanoylamino-α-D-glucopyranoside (34a) Compounds 17 (515 mg, 0.51 mmol) and 33a (380 mg, 0.51 mmol) were dissolved in CH₂Cl₂ (8 ml), and mercuric cyanide (253 mg, 1.0 mmol) and CaSO₄ (600 mg) were added to the mixture, which was then refluxed for 16 h. After the reaction mixture was diluted with CHCl₃, the mixture was filtered through Cerite 545, and the filtrate was washed with 5% aqueous potassium iodide and saturated aqueous NaCl, then dried. After evaporation of the solvent, the residue was purified by silica gel column chromatography (CHCl₃-acetone, 9:1) to give 34a (717 mg, 83%) as a colorless oil, $[\alpha]_D$ +18.7° (c=1.1, CHCl₃). NMR δ : 0.89 (9H, t, J=6 Hz), 1.28 (60H, s, CH₂), 1.60 (6H, br, CH₂CH₂O × 3), 2.16 (4H, m, $CH_2CO \times 2$), 2.36 (2H, t, J=8 Hz, CH_2CO), 4.64—4.86 (4H, m, $CH_2CCl_3 \times 2$), 5.13 (1H, t, J = 10 Hz, H-3), 5.24 (2H, s, $C\underline{H}_2C_6H_5$), 5.56 (1H, t, J=10 Hz, H-3'), 7.2—7.4 (15H, m, arom. H). MS m/z: 1698 (M^+) , 1703 $[(M+5)^+]$.

1,3-Di(benzyloxycarbonyl)isopropyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenyl-phosphono-3-O-tetradecanoyl-6-O-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)- β -D-glucopyranosyl]-3-O-tetradecanoyl-2-tetradecanoylamino- α -D-glucopyranoside (34b) Similarly to the preparation of 34a, compound 17 (500 mg, 0.49 mmol) was treated with 33b (450 mg, 0.49 mmol) in CH₂Cl₂ (4 ml) in the presence of CaSO₄ (0.6 g) and mercuric cyanide (250 mg, 1.0 mmol) to give 34b (790 mg, 86%) as a colorless oil, $[\alpha]_D$ +14.0° (c=0.8, CHCl₃). Anal. Calcd for C₉₁H₁₃₁Cl₆N₂O₂₃P: C, 58.61; H, 7.08; N, 1.50. Found: C, 58.54; H, 7.08; N, 1.47. NMR δ : 0.88 (9H, t, J=6 Hz), 1.26 (60H, s, CH₂), 4.70 (4H, m, CH₂CCl₃ × 2), 5.18 (4H, d, CH₂C₆H₅ × 2), 7.16—7.46 (20H, m, arom. H). MS m/z: 1860 (M⁺), 1864 [(M+4)⁺].

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-3-O-(N-dodecanoylglycyl)-6-O-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)- β -D-glucopyranosyl]-3-O-(N-dodecanoylglycyl)-2-tetradecanoylamino- α -D-glucopyranoside (34c) HBr-AcOH (25%, 16 ml) was added to a solution of 15 (2.54 g, 2.43 mmol) in CH₂Cl₂ (18 ml), and the mixture was stirred for 3 h. The reaction mixture was diluted with CHCl₃ and the solution was washed with

ice-H₂O, 5% aqueous NaHCO₃ and saturated aqueous NaCl, and dried. After evaporation of the solvent, the residue (16) and 33c (2.51 g, 2.77 mmol) were dissolved in CH₂Cl₂ (18 ml), then CaSO₄ (3.0 g) and mercuric cyanide (1.23 g, 4.86 mmol) were added to the solution. After being refluxed for 2.5 h, the reaction mixture was filtered through Celite 545, and the filtrate was washed with 5% aqueous potassium iodide and saturated aqueous NaCl, dried, and concentrated. The residue was purified by silica gel chromatography (CHCl₃-MeOH, 19:1) to give 34c (4.06 g, 88%) as a colorless oil, $[\alpha]_D$ +24.0° (c=1.0, CHCl₃). NMR δ : 0.89 (9H, t, J=7 Hz), 1.26 (52H, s, CH₂), 1.60 (6H, br, CH₂CH₂CO × 3), 2.10 (4H, m, CH₂CO × 2), 2.27 (2H, t, J=8 Hz, CH₂CO), 5.00 (1H, d, J=8 Hz, H-1'), 5.11 (1H, t, J=10 Hz, H-3), 5.79 (1H, t, J=10 Hz, H-3'), 7.1—7.5 (20H, m, arom. H). MS m/z: 1884 (M⁺), 1889 [(M+5)⁺].

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-3-O-(N-dodecanoylglycyl)-6-O-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)- β -D-glucopyranosyl]-3-O-(N-dodecanoylglycyl)-2-(N-dodecanoylglycylamino)- α -D-glucopyranoside (34d) Compound 16, obtained from 15 (0.28 g, 0.27 mmol) was allowed to react with 33d (0.23 g, 0.24 mmol) in the presence of mercuric cyanide (0.14 g, 0.54 mmol), as described for 34c, to give 34d (0.41 g, 88%) as a colorless viscous oil, [α]_D +11.9° (c=1.0, CHCl₃). NMR δ : 0.90 (9H, t, J=6 Hz), 1.28 (48H, s, CH₂), 1.60 (6H, br, CH₂CH₂CO × 3), 2.0—2.3 (6H, m, CH₂CO × 2), 4.96 (1H, d, J=8 Hz, H-1'), 5.19 (1H, t, J=10 Hz, H-3'), 5.58 (1H, t, J=10 Hz, H-3'), 6.1, 6.2, and 6.8 (each 1H, br, NH), 7.1—7.5 (22H, m, arom. H and NH). MS m/z: 1913 (M⁺), 1916 [(M+3)⁺].

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-3-O-tetradecanoyl-6-O-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)- β -D-glucopyranosyl]-3-O-tetradecanoyl-2-(N-dodecanoylglycylamino)- α -D-glucopyranoside (34e) Compound 17 was allowed to react with 33e (0.32 g, 0.35 mmol) in the presence of mercuric cyanide (0.20 g, 0.78 mmol), as described for 34a, to give 34e (0.59 g, 90%) as a colorless viscous oil, [α]_D +14.8° (c=1.0, CHCl₃). NMR δ : 0.89 (9H, t, J=7 Hz), 1.28 (56H, s, CH₂), 1.60 (6H, br, CH₂CH₂CO × 3), 2.14 (4H, m, CH₂CO × 2), 2.35 (2H, t, J=8 Hz, CH₂CO), 5.04 (1H, d, J=10 Hz, H-3), 5.56 (2H, m, H-3' and NH), 6.66 and 6.84 (each 1H, br, NH), 7.1—7.5 (20H, m, arom. H). MS m/z: 1855 (M⁺), 1859 [(M+4)⁺].

1,3-Di(benzyloxycarbonyl)isopropyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-3-O-(N-dodecanoylglycyl)-6-O-(2,2,2-trichloroethoxycarbonyl)-2-(2,2,2-trichloroethoxycarbonylamino)-β-D-glucopyranosyl]-3-O-(Ndodecanoylglycyl)-2-tetradecanoylamino-\alpha-D-glucopyranoside (34f) As described for 34c, compound 16 obtained from 15 (416 mg, 0.40 mmol), was treated with 33f (376 mg, 0.40 mmol) to give 34f (531 mg, 69%) as a colorless viscous oil, $[\alpha]_D$ +19.4° (c=1.1, CHCl₃). NMR (500 MHz) δ : 0.88 (9H, t, J = 7.3 Hz, CH₃), 1.25 (52H, s, CH₂), 1.54 (2H, m, CH₂CH₂CO), 1.63 (2H, m, CH_2CH_2CO), 2.07 (2H, t, $J=7.3\,Hz$, CH_2CO), 2.25 (2H, m, CH_2CO), 2.26 (2H, t, J=7.3 Hz, CH_2CO), 2.64—2.74 (3H, m, $OCH(CH_2CO_2Bzl)_2 \times 3)$, 2.99 (1H, m, $OCH(CH_2CO_2Bzl)_2$), 3.29 (1H, m), 3.49 (1H, m), 3.62 (1H, m), 3.78—3.91 (m), 4.09 (1H, dd, *J*=17.4, 5.5 Hz), 4.17 (3H, m), 4.33 (1H, dd, *J*=11.9, 4.6 Hz), 4.46—4.56 (m), 4.68 (2H, m), 4.80 (1H, m), 4.89 (1H, d, J=3.4 Hz, H-1), 4.99 (2H, m), 5.09-5.18 (5H, m), 5.74 (1H, t, J=9.2 Hz, H-3'), 6.06 (1H, d, J=7.3 Hz, NH), 6.17 (1H, m, NH), 6.36 (1H, m, NH), 6.54 (1H, d, J=9.2 Hz, NH), 7.10—7.37 (20H, m, arom. H). MS m/z: 1918 (M⁺), 1923 [(M+5)⁺].

Benzyloxycarbonylmethyl 2-Deoxy-6-O-(2-deoxy-4-O-diphenylphosphono-3-O-tetradecanoyl-2-tetradecanoylamino- β -D-glucopyranosyl)-3-O-tetradecanoyl-2-tetradecanoylamino- α -D-glucopyranoside (35a) Zinc powder (0.7 g) was added to a solution of 34a (700 mg, 0.41 mmol), and the mixture was vigorously stirred for 1 h at room temperature. The insoluble materials were removed by filtration, and the filtrate was concentrated in vacuo. The residue was dissolved in benzene, and washed with 5% NaHCO₃ aqueous solution, and then with saturated aqueous NaCl, and dried. Evaporation of the solvent gave an oil.

Separately, DCC (134 mg, 0.65 mmol) was added to a solution of tetradecanoic acid (142 mg, 0.62 mmol) and HOBt (99 mg, 0.65 mmol) in THF (3 ml) with ice-cooling. The mixture was stirred for 3 h at room temperature, and the precipitate was filtered off to give an active ester solution. This solution was added to a solution of the above oil in CH₂Cl₂ (5 ml), then N-methylmorpholine (71 μ l, 0.65 mmol) was added to the mixture with ice cooling, and the final mixture was stirred for 18 h at room temperature. After evaporation of the solvent, the residue was purified by silica gel column chromatography (CHCl₃-MeOH, 50:1) to give 35a (400 mg, 62%) as a colorless oil, [α]_D +8.5° (c=0.9, CHCl₃). NMR δ : 0.90 (12H, t, J=6 Hz), 1.26 (80H, s, CH₂), 1.58 (8H, br,

C \underline{H}_2 CO \times 4), 2.1—2.4 (8H, m, C \underline{H}_2 CO \times 4), 4.75 (1H, m, H-4'), 4.84 (1H, d, J=4 Hz, H-1), 4.96 (1H, d, J=8 Hz, H-1'), 5.16 (1H, t, J=10 Hz, H-3), 5.22 (2H, s, C \underline{H}_2 C₆H₅), 5.51 (1H, t, J=10 Hz, H-3'), 7.2—7.4 (15H, m, arom. H). MS m/z: 1562 [(M+2)⁺].

1,3-Di(benzyloxycarbonyl)isopropyl 2-Deoxy-6-O-(2-deoxy-4-O-diphenylphosphono-3-O-tetradecanoyl-2-tetradecanoylamino- β -D-glucopyranosyl)-3-O-tetradecanoyl-2-tetradecanoylamino- α -D-glucopyranoside (35b) As described for 35a, compound 34b (780 mg, 0.42 mmol) was treated with zinc powder (1.2 g), and the resulting oil was allowed to react with the HOBt ester of tetradecanoic acid (140 mg, 0.63 mmol) to give 35b (550 mg, 76%) as a white powder, mp 78—80 °C, [α]_D +4.4° (c=1.3, CHCl₃). Anal. Calcd for C₉₉H₁₅₅N₂O₂₀P: C, 68.96; H, 9.06; N, 1.62. Found: C, 68.72; H, 8.95; N, 1.63. NMR δ: 0.90 (12H, t, J=6 Hz), 1.26 (80H, s, CH₂), 2.1—2.4 (8H, m, CH₂CO×4), 2.64—2.98 (4H, m, OCH(CH₂CO₂Bzl)₂), 4.75 (1H, m, H-4'), 4.94 (1H, d, J=4 Hz, H-1), 5.00 (1H, d, J=8 Hz, H-1'), 5.16 (4H, s, CH₂C₆H₅×2), 5.58 (1H, t, J=10 Hz, H-3').

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-3-O-(N-dodecanoylglycyl)-2-tetradecanoylamino- β -D-glucopyranosyl]-3-O-(N-dodecanoylglycyl)-2-tetradecanoylamino- α -D-glucopyranoside (35c) As described for 35a, compound 34c (4.06 g, 2.15 mmol) was treated with zinc powder, and the resulting oil was allowed to react with the HOBt ester of tetradecanoic acid (0.74 g, 3.23 mmol) in THF (15 ml) to give 34c (3.30 g, 88%) as a colorless oil, $[\alpha]_D + 20.9^\circ$ (c = 1.1, CHCl₃). NMR δ : 0.88 (12H, t, J = 6 Hz), 1.28 (76H, s, CH₂), 1.60 (8H, br, CH₂CH₂CO×4), 2.0—2.4 (8H, m, CH₂CON×4), 4.70 (1H, q, J = 10 Hz, H-4'), 4.90 (1H, d, J = 4 Hz, H-1), 5.16 (1H, t, J = 10 Hz, H-3, 5.28 (1H, d, J = 8 Hz, H-1), 5.66 (1H, t, J = 10 Hz, H-3'), 7.2—7.4 (20H, m, arom. H).

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-3-O-(N-dodecanoylglycyl)-2-(N-dodecanoylglycylamino)- β -D-glucopyranosyl]-3-O-(N-dodecanoylglycyl)-2-(N-dodecanoylglycylamino)- α -D-glucopyranoside (35d) As in the preparation of 35a, compound 34d (0.39 g, 0.20 mmol) was treated with zinc powder, and the resulting oil was allowed to react with the HOBt ester of 10 (0.10 g, 0.40 mmol) to give 34d (0.20 g, 54%) as a colorless oil, [α]_D +16.7° (c=1.0, CHCl₃). NMR δ : 0.9 (12H, t, J=6Hz), 1.28 (64H, s, CH₂), 1.64 (8H, br, CH₂CH₂CO × 4), 2.0—2.3 (8H, m, CH₂CO × 4), 4.74 (1H, m, H-4'), 4.84 (1H, d, J=4 Hz, H-1), 5.01 (1H, d, J=8 Hz, H-1'), 5.21 (1H, m, H-3), 5.60 (1H, t, J=10 Hz, H-3'), 7.2—7.5 (m, arom. H and NH). MS m/z: 1806 [(M+2)+].

2-(Diphenylphosphonooxy)ethyl 2-Deoxy-6-O-[2-deoxy-4-O-diphenylphosphono-2-(N-dodecanoylglycylamino)-3-O-tetradecanoyl- β -D-glucopyranosyl]-2-(N-dodecanoylglycylamino)-3-O-tetradecanoyl- α -D-glucopyranoside (35e) As described for 35a, compound 34e (0.58 g, 0.31 mmol) was treated with zinc powder, and the oily product was reacted with the HOBt ester of 10 to give 35e (0.49 g, 89%) as a colorless oil, [α]_D +16.4° (c=1.0, CHCl₃). NMR δ : 0.90 (12H, t, J=7 Hz), 1.26 (72H, s, CH₂), 2.1—2.3 (8H, m, CH₂CO×4), 4.73 (1H, m, H-4'), 4.88 (1H, d, J=4 Hz, H-1), 4.93 (1H, d, J=8 Hz, H-1'), 5.41 (1H, m, H-3), 5.65 (1H, t, J=10 Hz, H-3'), 6.42 (1H, br, NH), 6.66 (2H, m, NH × 2), 6.88 (1H, d, J=9 Hz, NH), 7.2—7.5 (20H, m, arom. H). MS m/z: 1748 [(M+2)+].

1,3-Di(benzyloxycarbonyl)isopropyl 2-Deoxy-6-O-[2-deoxy-3-O-(Ndodecanoylglycyl)-2-tetradecanoylamino-4-O-diphenylphosphono-β-Dglucopyranosyl]-3-O-(N-dodecanoylglycyl)-2-tetradecanoylamino-α-Dglucopyranoside (35f) As described for 35a, compound 34f (509 mg, 0.26 mmol) was treated with zinc dust, and the resulting oil was allowed to react with the HOBt ester to tetradecanoic acid (91 mg, 0.40 mmol) to give 35f (376 mg, 80%) as a colorless powder, mp 135—139 °C, $[\alpha]_D$ $+18.8^{\circ}$ (c=0.5, CHCl₃). Anal. Calcd for C₉₉H₁₅₃N₄O₂₂P: C, 66.62; H, 8.65; N, 3.14. Found: C, 66.81; H, 8.54; N, 3.11. NMR (500 MHz) δ : 0.89 (12H, t, $J = 7.3 \,\mathrm{Hz}$), 1.25 (72H, s, CH₂), 1.54 (2H, m, C $\mathrm{\underline{H}_2CH_2CO}$), 1.63 (2H, m, CH_2CH_2CO), 2.12 (4H, m, $CH_2CO \times 2$), 2.24 (2H, t, $J = 7.3 \text{ Hz}, \text{CH}_2\text{CO}, 2.62 (1\text{H}, \text{dd}, J = 16.5, 7.3 \text{ Hz}, \text{OCH}(\text{CH}_2\text{CO}_2\text{Bzl})_2),$ 2.67 (2H, d, J = 6.4 Hz, OCH(C \underline{H}_2 CO₂Bzl)₂ × 2), 2.90 (1H, dd, J = 16.5, 4.6 Hz, OCH(CH₂CO₂Bzl)₂), 3.28 (1H, m), 3.56—3.67 (4H, m), 3.74—3.90 (5H, m), 4.07 (2H, m), 4.36 (1H, m), 4.66 (1H, m, OCH(CH₂CO₂Bzl)₂), 4.67 (1H, q, J=9.2 Hz, H-4'), 4.90 (1H, d, J=3.7 Hz, H-1), 5.03 (1H, t, J = 10.1 Hz, H-3), 5.12 (4H, m, $C\underline{H}_2C_6H_5 \times 2$), 5.31 (1H, d, J = 8.3 Hz, H-1'), 5.67 (1H, t, J = 10.1 Hz, H-3'), 6.26 (1H, t, J = 5.5 Hz, NH), 6.29 (1H, t, J=5.5 Hz, NH), 6.37 (1H, d, J=6.4 Hz, NH), 6.50 (1H, d, J=9.2 Hz, NH), 7.14—7.36 (20H, m, arom. H). MS m/z: 1782 [(M+2)⁺].

Carboxymethyl 2-Deoxy-6-*O*-(2-deoxy-4-*O*-phosphono-3-*O*-tetradecanoyl-2-tetradecanoylamino-β-D-glucopyranosyl)-3-*O*-tetradecanoyl-2-tetradecanoylamino-α-D-glucopyranoside (4) Compound 35a (379 mg, 0.24 mmol)

was dissolved in THF (30 ml) and shaken with 5% palladium-carbon (400 mg) at room temperature for 1 h in H₂ at atmospheric pressure. Then, platinum dioxide (200 mg) was added to the mixture, and the mixture was further shaken at room temperature for 1.1 h in H, at atmospheric pressure. The catalyst was removed by filtration and washed with CHCl₃-MeOH-H₂O (8:3:1, lower layer). The combined filtrate washings were concentrated at reduced pressure. The residue was purified by preparative TLC (CHCl₃-MeOH-H₂O, 6:4:0.2). After extraction with CHCl₃-MeOH-H₂O-Et₃N (6:4:1:0.02), the solvent was evaporated in vacuo. The residue was dissolved in CHCl₃-MeOH-H₂O (6:4:0.5), and the solution was desalted with Dowex 50 (H⁺ type). A portion of the desalted solution was concentrated, and the residue was lyophilized from a dioxane suspension to give 4 (249 mg, 78%) as a white powder, mp 145—148 °C (dec.), $[\alpha]_D$ +14.2° (c=0.5, CHCl₃-MeOH, 3:1). IR (KBr): 3450, 2925, 2855, 1740, 1640 cm⁻¹. NMR (CDCl₃-CD₃OD) δ : 0.90 (12H, t, J = 6 Hz), 1.28 (80H, s, CH₂), 1.56 (8H, br, CH₂CH₂CO × 4), 2.1—2.4 (8H, m, $CH_2CO \times 4$), 4.80 (1H, d, J=4 Hz, H-1), 5.24 (2H, m, H-3, H-3').

The product (220 mg) in 0.1% aqueous Et_3N was lyophilized to give 250 mg of Et_3N salt as a white powder.

1,3-Dicarboxyisopropyl 2-Deoxy-6-O-(2-deoxy-4-O-phosphono-3-O-tetradecanoyl-2-tetradecanylamino- β -D-glucopyranosyl)-3-O-tetradecanoyl-2-tetradecanoylamino- α -D-glucopyranoside (5) In a manner similar to that described for 4, compound 35b (280 mg, 0.20 mmol) was hydrogenolyzed with palladium-black (200 mg) in dioxane (30 ml) for 2.5 h, and then with platinum dioxide (300 mg) for 18 h to give 5 (173 mg, 77%) as a white powder, mp 142—147 °C (dec.), $[\alpha]_D + 11.7^\circ$ (c=0.7, CHCl₃-MeOH, 3:1). NMR δ : 0.90 (12H, t, J=6 Hz), 1.30 (80H, s, CH₂), 2.20 (m), 2.36 (m), 2.70 (m), 4.98 (1H, d, H-1).

The Et₃N salt was prepared similarly to that of 4.

2-Phosphonooxyethyl 2-Deoxy-6-O-[2-deoxy-3-O-(N-dodecanoylglycyl)-4-O-phosphono-2-tetradecanoylamino-β-D-glucopyranosyl]-3-O-(N-dodecanoylglycyl)-2-tetradecanoylamino- α -D-glucopyranoside (6) Platinum dioxide (0.40 g) was added to a solution of 35c (0.42 g, 0.24 mmol) in THF (80 ml). The mixture was stirred in a hydrogen atmosphere for 25 h, then a mixture of CHCl₃-MeOH-H₂O (8:3:1, lower layer) was added. After the catalysts had been filtered off, the filtrate was concentrated, and the resulting residue was purified by preparative TLC (CHCl₃-MeOH-H₂O, 6:4:0.8). The extracted solution with CHCl₃-MeOH-H₂O-Et₃N (6:4:1:0.02) was concentrated by evaporation. After dissolution in CHCl₃-MeOH-H₂O (8:3:1, lower layer), the solution was desalted with Dowex 50 (H⁺). The desalting solution was concentrated and the dioxane suspension was freeze-dried to give 4 (95 mg, 27%) as a white powder, mp 140—145 °C (dec.), $[\alpha]_D = 13.3^\circ$ $(c=0.6, CHCl_3-MeOH, 3:1)$. IR (KBr): 3400, 2930, 2855, 1750, 1660, 1560 cm⁻¹. NMR (CDCl₃-CD₃OD, 1:1) δ : 0.90 (12H, t, J=7 Hz), 1.32 (72H, s, CH_2), 1.60 (8H, br, $CH_2CH_2CO \times 4$), 2.1—2.3 (8H, m, $CH_2CO \times 4$), 5.10 (1H, t, J = 10 Hz, H-3), 5.38 (1H, t, J = 10 Hz, H-3').

The Et₃N salt was prepared in a manner similar to that described for 4. **2-Phosphonooxyethyl 2-Deoxy-6-[2-deoxy-3-***O-(N-***dodecanoylglycyl)-2-**(*N-***dodecanoylglycylamino)-4-***O-***phosphono-**β-D-glucopyranosyl]-3-*O-(N-***dodecanoylglycyl)-2-**(*N-***dodecanoylglycyl)-2-**(*N-***dodecanoylglycyl)-3-**(*N-***dodecanoylglycyl)-3-**(*N-***dodecanoylglycyl)-3-**(*N-***dodecanoylglycyl)-3-**(*N-***dodecanoylglycyl)** was hydrogenolyzed in the presence of platinum dioxide, and the resulting powder was purified by preparative TLC (CHCl₃-MeOH-H₂O, 6:4:0.9) and desalted. The desalting solution was concentrated and the dioxane suspension was freeze-dried to give 7 (0.14 g, 39%) as a white powder, mp 145—150 °C (dec.), [α]_D + 7.6° (c=0.8, CHCl₃-MeOH, 3:1). IR (KBr): 3300, 1760, 1665, 1555 cm⁻¹. NMR (CDCl₃-CD₃OD, 1:1) δ: 0.90 (12H, t, J=6 Hz), 1.30 (64H, s, CH₂), 1.66 (8H, br, CH₂CO × 4), 2.30 (8H, m, CH₂CO × 4), 4.84 (1H, d, J=4 Hz, H-1), 5.19 (1H, t, J=10 Hz, H-3), 5.33 (1H, t, J=10 Hz, H-3').

The Et₃N salt was prepared similarly to that of 4.

2-Phosphonooxyethyl 2-Deoxy-6-O-[2-deoxy-2-(N-dodecanoylglycylamino)-4-O-phosphono-3-O-tetradecanoyl- β -D-glucopyranosyl]-2-(N-dodecanoylglycylamino)-3-O-tetradecanoyl- α -D-glucopyranoside (8) In the manner described for 6, compound 35e (0.46 g, 0.26 mmol) was hydrogenolyzed in the presence of platinum dioxide, and the resulting powder was purified by preparative TLC (CHCl₃-MeOH-H₂O, 6: 4: 0.8) and desalted. The desalting solution was concentrated, and the dioxane suspension was freeze-dried to give 8 (0.22 g, 59%) as a white powder, mp 148—153 °C (dec.), [α]_D +18.4° (c=0.9, CHCl₃-MeOH, 3:1). IR (KBr): 3300, 1745, 1645, 1555 cm⁻¹. NMR (CDCl₃-CD₃OD, 1:1) δ : 0.90 (12H, t, J=6Hz), 1.30 (72H, s, CH₂), 1.6 (8H, br, CH₂CH₂CO × 4), 2.3 (8H, m, CH₂CO × 4), 4.86 (1H, d, J=4 Hz, H-1), 5.16 (1H, t,

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J = 10 Hz, H-3), 5.34 (1H, t, J = 10 Hz, H-3).

The Et₃N salt was prepared in a manner similar to that for 4.

1,3-Dicarboxyisopropyl 2-Deoxy-6-O-[2-deoxy-3-O-(N-dodecanoylglycyl)-4-O-phosphono-2-tetradecanoylamino- β -D-glucopyranosyl]-3-O-(N-dodecanoylglycyl)-2-tetradecanoylamino-α-D-glucopyranoside (9) In a manner similar to that described for 4, compound 35f (361 mg, 0.20 mmol) was hydrogenolyzed with 5% palladium-carbon (400 mg) in 5% aqueous THF (32 ml) for 6 h, and then with platinum dioxide for 3 h. The resulting residue was purified by preparative TLC (CHCl₃-MeOH-H₂O, 6:4:0.5), then desalted with Dowex 50 (H⁺ type). A portion of the desalted solution was concentrated, and the residue was lyophilized from dioxane suspension to give 9 (116 mg, 39%) as a white powder, mp 158—165 °C (dec.), $[\alpha]_D$ +18.6° (c=0.6, CHCl₃-MeOH, 3:1). Anal. Calcd for $C_{73}H_{133}N_4O_{22}P \cdot 0.5H_2O$: C, 60.10; H, 9.26; N, 3.84. Found: C, 60.11; H, 9.55; N, 3.87. IR (KBr): 3448, 2924, 2856, 1748, 1646, 1550, 1470 cm⁻¹. NMR (500 MHz, CDCl₃-CD₃OD, 1:1) δ : 0.88 (12H, t, J = 7.3 Hz), 1.26 (72H, s, CH₂), 1.56 (4H, m, CH₂CH₂CO × 2), 1.63 (4H, m, $CH_2CH_2CO \times 2$), 2.16 (2H, t, J=7.1 Hz, CH_2CO), 2.21 (2H, t, $J=7.1 \text{ Hz}, \text{ CH}_2\text{CO}$, 2.26 (4H, t, $J=7.1 \text{ Hz}, \text{ CH}_2\text{CO} \times 2$), 2.67 (3H, m, $OCH(C_{12}CO_{2}H)_{2} \times 3)$, 2.83 (1H, dd, J = 16.5, 5.5 Hz, $OCH(C_{12}CO_{2}H)_{2}$), 3.52 (2H, m, H-4 and H-5'), 3.70 (1H, t, J=9.5 Hz, H-2'), 3.77 (1H, dd, J = 11.2, 5.6 Hz, H-6), 3.84 (1H, dd, J = 12.7, 4.8 Hz, COCH₂N), 3.9—4.0 $(7H, m, COCH_2N \times 3, H-5, H-6 \text{ and } H-6' \times 2), 4.12 (1H, m, H-2), 4.30$ (1H, q, J=9.5 Hz, H-4'), 4.40 (1H, m, OCH(CH₂CO₂H)₂), 4.84 (1H, d,J=7.9 Hz, H-1'), 4.95 (1H, d, J=4.0 Hz, H-1), 5.05 (1H, t, J=9.5 Hz, H-3), 5.40 (1H, t, J=9.5 Hz, H-3').

The Et₃N salt was prepared similarly to that of 4.

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