Acknowledgment. This research has been supported by the Division of Chemical Sciences, Office of Basic Energy Sciences, U.S. Department of Energy (Report DOE/ER/02968-137). We thank Dr. S. Steenken for stimulating our interest in aliphatic ether radical cations.

Supplementary Material Available: The solid-state ESR spectrum of (CH₃)₂O⁺ at 155 K (1 page). Ordering information is given on any current masthead page.

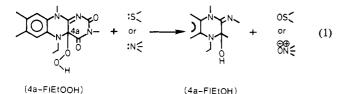
On the Nature of the Intermediate between 4a-Hydroperoxyflavin and 4a-Hydroxyflavin in the Hydroxylation Reaction of *p*-Hydroxybenzoate Hydroxylase. Synthesis of 6-Aminopyrimidine-2,4,5(3H)-triones and the Mechanism of Aromatic Hydroxylation by Flavin Monooxygenases

Albert Wessiak and Thomas C. Bruice*

Department of Chemistry University of California at Santa Barbara Santa Barbara, California 93106

Received August 6, 1981

There appears to be a consensus that the reactions of reduced flavoenzyme monooxygenases with molecular oxygen provide enzyme-bound 4a-hydroxyperoxyflavin (4a-FlHOOH).^{1-4a} In the N- and S-oxidation of amines by hepatic microsomal flavomonooxygenase,^{5,6} substrate oxidation is accompanied by the conversion 4a-FlHOOH \rightarrow 4a-FlHOH. The mechanism of the enzymatic reaction⁷ appears to be identical in essential features with the bimolecular N- and S-oxidations with authentic 4a-hydroperoxyflavins (reaction 1).^{2bf,i} No evidence for intermediates

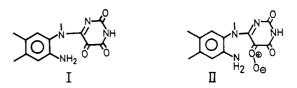


could be obtained, and the reactions are quantitative.^{2i,7} In contrast, the mechanisms for bacterial hydroxylases responsible for hydroxylation of electron-rich aromatic compounds are poorly understood. The enzyme p-hydroxybenzoate hydroxylase serves as the most useful example for this class of enzymes.⁸

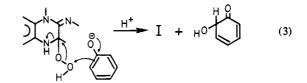
A strongly absorbing species (X) is formed from 4a-FlHOOH during the hydroxylation of a number of alternate substrates by p-hydroxybenzoate hydroxylase.⁸ The disappearance of X is accompanied by the appearance of a species to which a 4ahydroxyflavin structure (4a-FlHOH) was assigned (reaction 2).

4a-Fihooh — 4a-Fihoh		
λ _{max} 380-390 nm e ~9000 M ⁻¹ cm ⁻¹	λ _{max} 390-420 nm ε ~15 000 M ⁻¹ cm ⁻¹	$\lambda_{max} 380-385 \text{ nm} \\ e \sim 9000 \text{ M}^{-1} \text{ cm}^{-1}$
		(2)

The aromatic hydroxylation reaction differs, therefore, from the $N \in$ and $S \in$ oxidation reactions in which an intermediate (X) is not seen. The spectral observation of a well-defined intermediate occurring in time between 4a-FIHOOH and 4a-FIHOH is most important. Structure I has been assigned to X.⁸ Massey and



co-workers have proposed that I arises in concert with oxygen-atom transfer from 4a-FIHOOH to substrate (reaction 3).8 Species



I was predicted by Hamilton⁹ in 1971 as the immediate product of monooxygen transfer from his hypothetic carbonyl oxide II. Structures I and II have not only received a great deal of attention with respect to mechanism in flavomonooxygenase reactions, but structures completely analogous to I and II have been considered by Bailey and Ayling¹⁰ to arise along the reaction paths for pteridine monooxygenases (responsible for initiation of the biosynthesis of neuroactive amines through hydroxylation of phenylalanine, tyrosine, and tryptophan). An assessment of the plausibility that X (reaction 3) = I, which has been disputed,¹¹ is most easily accomplished by independent synthesis of I and comparison of the spectral properties of X and I. The objective of this study has been to synthesize N,N-dimethyl-I (i.e., IX) and compare its spectral properties to those of X (reaction 2). 6-Amino-5-oxouracils (or 6-aminopyrimidine-2,4,5-triones) have not been isolated.^{12,13} We describe herein our efforts to synthesize substituted 6-aminopyrimidine-2,4,5(3H)-triones from 5,6-diaminouracils and the first successful synthesis via 4a-5 ring opening of a suitably substituted flavin.

(9) Hamilton, G. A. Prog. Bioorg. Chem. 1971, 1, 83.

(10) Bailey, S. W.; Ayling, J. E. J. Biol. Chem. 1980, 255, 7774.

^{(1) (}a) Massey, V.; Hemmerich, P. Enzymes, 3rd. Ed. 1975, 12, 191. (b) Flashner, M. S.; Massey, V. "Molecular Mechanisms of Oxygen Activation";

<sup>Flashner, M. S.; Massey, V. "Molecular Mechanisms of Oxygen Activation";
O. Hayaishi, Ed.; Academic Press: New York, 1974; p 245.
(2) (a) Kemal, C.; Bruice, T. C.</sup> *Proc. Natl. Acad. Sci. U.S.A.* 1976, 73, 995.
(b) Kemal, C.; Chan, T. W.; Bruice, T. C. *Ibid.* 1977, 74, 405.
(c) Kemal, C.; Bruice, T. C. J. Am. Chem. Soc. 1977, 99, 7064.
(d) Kemal, C.; Bruice, T. C. *J. Am. Chem. Soc.* 1977, 99, 7064.
(d) Kemal, C.; Bruice, T. C. *Jid.* 1977, 99, 7272.
(e) Kemal, C.; Bruice, T. C. *Ibid.* 1977, 99, 7272.
(e) Kemal, C.; Bruice, T. C. *Joid.* 1977, 99, 7272.
(f) Ball, S.; Bruice, T. C. *Ibid.* 1979, 101, 4017.
(g) Iwata, M.; Bruice, T. C.; Carrell, H. L.; Glusker, J. P. Bid. 1980, 102, 5036.
(h) Muto, S.; Bruice, T. C. *Ibid.* 1980, 102, 4472.
(i) Ball, S.; Bruice, T. C. *Ibid.* 1980, 102, 7559.
(k) Shepherd, P. T.; Bruice, T. C. *Ibid.* 1980, 102, 7759.

 ^{1.} C. 1012. 1930, 102, 6498. (1) Muto, S.; Bruice, I. C. 1012. 1930, 102, 7559.
 (k) Shepherd, P. T.; Bruice, T. C. 1bid. 1980, 102, 7774.
 (3) (a) Spector, T.; Massey, V. J. Biol. Chem. 1972, 247, 5632. (b) Strickland, S.; Massey, V. 1bid. 1973, 248, 2953. Hastings, J. W.; Balny, C.; Le Peuch, C.; Douzou, P. Proc. Natl. Acad. Sci. U.S.A 1973, 70, 3468. (d) Poulsen, L. L.; Ziegler, D. M. J. Biol. Chem. 1979, 254, 6449.
 (4) (a) Ghisla, S.; Hastings, J. W.; Favandon, V.; Lhoste, J.-M. Proc. Natl. Acad. Sci. U.S.A. 1978, 75, 5860. (b) Moad, G.; Luthy, C. L.; Benkovic, P. A.; Benkovic, S. 1970, 104, 6065.

A.; Benkovic, S. J. J. Am. Chem. Soc. 1979, 101, 6068.
 (5) Poulsen, L. L.; Kadlubar, F. F.; Ziegler, D. M. Arch. Biochem. Biophys. 1974, 164, 774. Ziegler, D. M.; Mitchell, C. H. Ibid. 1972, 150, 116. (6) Hajjar, N. P.; Hodgson, E. Science (Washington, D.C.) 1980, 209, 1134.

⁽⁷⁾ Beaty, N. B.; Ballou, D. P. J. Biol. Chem. 1980, 255, 3817; 1981, 256, 4619.

^{(8) (}a) Entsch, B.; Ballou, D. P.; Massey, V. J. Biol. Chem. 1976, 251, 2550.
(b) Massey, V. "Oxygen: Biochemical and Clinical Aspects"; Caughey, W. S., Ed.; Academic Press: New York, 1979; p 477.

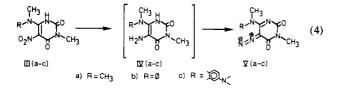
^{(11) (}a) Hemmerich, P.; Wessiak, A. "Flavins and Flavoproteins"; Singer,

T. P., Ed.; Elsevier: Amsterdam, 1976; p 9. (b) Hemmerich, P.; Wessiak, A. "Oxygen: Biochemical and Clinical Aspects"; Caughey, W. S., Ed.; Academic Press: New York, 1979; p 491. (c) Bruice, T. C. Prog. Bioorg. Chem. 1976, 4, 1.

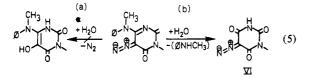
^{12) (}a) Brown, D. J. Chem. Heterocycl. Compd. 1962, 16. (b) Brown, D. J. Chem. Heterocycl. Compd. Suppl. I 1970, 16.

⁽¹³⁾ Bien, S.; Salemnik, G.; Zamir, L.; Rosenblum, M. J. Chem. Soc., Chem. Commun. 1968, 496. Bien, S.; Amith, D.; Ber, M. J. Chem. Soc., Perkin Trans. 1 1973, 1089.

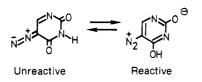
The three 5-diazo-6(R)-(methylamino)-3-methyluracils¹⁴ Va-c were obtained by the sequence of reaction 4. In contrast to the



easy hydrolysis reported for 5-diazouracil,²¹ compounds Va-c proved to be rather stable in acid solution. Hydrolysis of Vb yielded VI (IR 2180, 1745, 1720 and 1670 cm⁻¹; ¹H NMR 3.26 ppm) showing the extraordinary stability of the diazo group. The



difference in reactivity of V and 5-diazouracil may be related to the ability of the 5-diazouracil to tautomerize to an aromatic diazonium compound, a feature not allowed to V due to N(3) substitution. As already described by Sakuma and Yoneda,¹⁶



(14) The 5-nitrouracils IIIa-c¹⁵ were hydrogenated in aqueous solution over Pd on charcoal. After addition of 6 M sulfuric acid to make the solutions about 1 M, the catalyst was filtered off, and an excess of sodium nitrite was added. After 1 h and partial neutralization, the diazouracils Va-c were extracted with chloroform and precipitated with diethyl ether. Pure compounds were obtained by column chromatography on silica gel with chloroform points were obtained by column informatography of since get which following the formation of the sectate as eluants. 5-Diazo-6-(dimethylamino)-3-methyluracil (Va): $C_7H_9N_5O_2$ (M_r 195.18); mp 136-140 °C dec; IR (KBr) 2130 (N₂), 1675 (C(4)-O), 1625 (C(2)-O), 1560 cm⁻¹; ¹H NMR (Me₂SO-d₆) δ 3.21 (s, 6 H, C(6)N(CH₃)₂); 3.10 (s, 3 H, N(3)CH₃). 5-Diazo-3-methyl-6-methyl-H, C(6)N(CH₃)₂); 3.10 (s, 3 H, N(3)CH₃). 5-Diazo-3-methyl-6-methyl-phenylaminouracil (Vb) has already been synthesized by Sakuma and Yo-neda:¹⁶ C₁₂H₁₁N₅O₂ (M, 257.25); mp 178–180 °C dec (lit. 185 °C dec); IR (KBr) 2200 (w), 2140, 2110 (N₂), 1690 (C(4)=O), 1640 (C(2)=O), 1540, 1520 cm⁻¹; ¹H NMR (CDCl₃) δ 7.57–7.12 (m, 5 H, ArH₅), 3.48 (s, 3 H, N(6)CH₃), 3.24 (s, 3 H, N(3)CH₃). 5-Diazo-6-[[2-(dimethylamino)-4,5-dimethylphenyl]methylamino]-3-methyluracil (Vc): C₁₆H₂₀N₆O₂ (M, 328.37); mp 180–182 °C dec; IR (KBr) 2130 (N₂), 1695 (C(4)=O), 1646 (s, 1 H, C(3,6)H₂), 3.50 (s, 3 H, C(6)NCH₃), 3.32 (s, 3 H, N-(3)CH₃), 2.72 (s,6 H), C(2)N(CH₃)₂), 2.30 (s, 3 H) and 2.19 (s, 3 H, C(4,5)(CH₃)₂). Anal. Calcd: C, 58.52; H, 6.14; N, 25.60. Found: C 58.34; H, 6.29; N, 25.30. (15) 6-(Dimethylamino)-3-methyl-5-nitrouracil (IIIa)¹⁷⁶ and 3-methyl-6-(methylphenylamino)-5-nitrouracil (IIIb)¹⁸ were prepared as described in

6-(methylphenylamino)-5-nitrouracil (IIIb)¹⁸ were prepared as described in the literature by condensation of 6-chloro-3-methyl-5-nitrouracil¹⁷ with dimethylamine or phenylmethylamine. 6-[[2-(Dimethylamino)-4,5-dimethylphenyl]methylamino]-3-methyl-5-nitrouracil (IIIc) was prepared by analogous condensation with N, N, N, 4, 5-pentamethylphenylenediamine:¹⁹ C₁₆H₂₁ O₄H₂O (*M*, 365.38). Anal. Calcd: C, 52.59; H, 6.34; N, 19.17. Found: C 52.75%; H 6.04%; N 19.08%.

(16) Sakuma, Y.; Yoneda, F. Heterocycles 1977, 6, 1911.

 (17) (a) Daves, B. D.; Robins, R. K.; Cheng, C. C. J. Am. Chem. Soc.
 1962, 84, 1724. (b) Pfleiderer, W.; Walter, H. Liebigs Ann. Chem. 1964, 677, 113.

(18) Yoneda, F.; Sakuma, Y.; Shinomura, K. J. Chem. Soc., Perkin Trans. 1 1978, 348.

(19) Prepared by hydrolysis of N-formyl-N,N',N',4,5-pentamethyl-phenylenediamine²⁰ in 3 M sulfuric acid and extracted with chloroform after neutralization: yellow oil, ¹H NMR (CDCl₃) δ 6.83 (s, 1 H) and 6.43 (s, 1 H, C(3,6)H₂), 2.84 (s, 3 H, (C(1)NCH₃), 2.63 (s, 6 H, C(2)N(CH₃)₂, 2.23 (s, 3 H) and 2.17 (s, 3 H, $C(4,5)(CH_3)_2$).

(20) Synthesized analogous to N-formyl-N,N',N'-trimethylphenylenediamine from 4,5-dimethylphenylenediamine by following the procedure of: Sekiya, M.; Tomie, M.; Leonard, N. J. J. Org. Chem. 1968, 33, 318. The compound has already been prepared via a different route by: Saito, I.; Abe,
S.; Takahashi, Y.; Matsuura, T. Tetrahedron Lett. 1974, 4001.
(21) Chang, S. H.; Kim, I. K.; Hahn, B.-S. Taehan Hwahakhoe Chi 1965,

9, 75; Chem. Abstr. 1965, 64, 17588g.

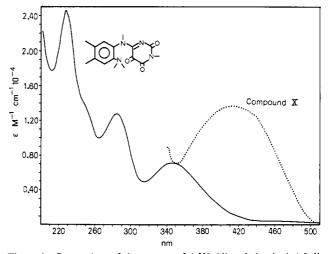
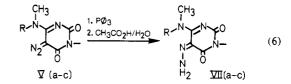


Figure 1. Comparison of the spectra of 6-[[2-(dimethylamino)-4,5-dimethylphenyl]methylamino]-3-methylpyrimidine-2,4,5(3H)-trione (IX) -) and the intermediate X (...) observed with p-hydroxybenzoate hydroxylase (spectra of X computer digitized from: Ballou, D. P. Flavoprotein Monooxygenases, 7th International Symposium on Flavins and Flavoproteins, Ann Arbor, MI. Also see ref 8 of text).

the diazouracil Vb is converted to 3,9-dimethylpyrimido[4,5-b]indole-2,3(1H,3H)-dione by both photochemical and thermal reactions. In addition, after heating with Cu(I) in 1 M aqueous HCl solutions, we obtained, again, the pyrimido[4,5-b]indole-2,3(1H,3H)-dione, while Cu(II) had no catalytical effect. Bestmann et al.²² have described the synthesis of α,β -diketones by reacting α -diazoketones with triphenylphosphine followed by acid hydrolysis and reaction of the resulting hydrazone with sodium nitrite. Using a method analogous to Bestmann et al.²² the three hydrazones $VIIa-c^{23}$ could be obtained. However, hydrolysis of



the hydrazones (VII) to the triones could not be effected. No reaction occurred on treatment of VIIa with NO2⁻ in dioxane/1 M H₂SO₄. Similar treatment of VIIb provided 3,10-dimethylisoalloxazine N⁵ oxide and 3,10-dimethylisoalloxazine, as established by comparison with authentic materials. As already known in the case of α -ketohydrazones,²² compounds VII show great stability in aqueous acids so that treatment with acid alone was also unsuccessful.

^{(22) (}a) Bestmann, H.-J.; Buckschewski, H.; Leube, H. Chem. Ber. 1959, 92, 1348. (b) Bestmann, H.-J.; Klein, O.; Goethlich, L.; Buckschewski, H. Chem. Ber. 1963, 96, 2259.

⁽²³⁾ The 5-diazouracils Va-c were dissolved in dry dioxane, and an excess of triphenylphosphine was added. After stirring for 1 h, the reaction mixture was poured into 0.3 M acetic acid. Following stirring for 30 min, the products were extracted with chloroform and purified via column chromatography on silica gel using ethyl acetta eas eluant: 6-(Dimethylamino)-3-methyl-pyrimidine-2,4,5(3H)-trione-5-hydrazone (VIIa): $C_7H_{11}N_5O_2$ -1/gH₂O (M_r 199.45); mp 214 °C dec; IR (KBr) 3250 (NH); 1650 (C(4)=O); nf 20 (C(2)=O) cm⁻¹; ¹H NMR (CDCl₃ + 10% CF₃CO₂H) δ 3.68 (s, 3 H), 3.54 (s, 3 H), 3.36 (s, 3 H). Anal. Calcd: C, 42.15; H, 5.68; N, 35.11. Found: C, 42.00; H, 5.55; N, 34.44. 3-Methyl-6-(methylphenylamino)pyrimidine-2,45(3H)-trione-5-hydrazone (VIIb): $C_{12}H_{13}N_5O_2$ -1/ $_4H_2O$ (M_r 261.75) mp 141–146 °C dec; IR (KBr) 3350 (NH), 1680 (C(4)=O), 1640 (C(2)=O) cm⁻¹; ¹H NMR (CDCl₃) δ 7.50–7.00 (m, 5 H, Ph), 3.57 (s, 3 H), C(6)NCH₃), 3.30 (s, (N(3)CH₃). Anal. Calcd: C, 54.60; H, 5.15; N, 26.55. Found: C, 54.73; H, 5.15; N, 25.86. 6-[[2-(Dimethylamino)-4,5-dimethylphenyl]-methylamino]-3-methylpyrimidine-2,4,5(3H)-trione-5-hydrazone (VIIc): $C_{16}H_{22}N_6O_2$ (M_r , 330.38) mp 162–164 °C dec; IR (KBr) 3300, 3350 (NH), 1680 (C(4)=O), 1640 (C(2)=O), 1530 cm⁻¹; ¹H NMR (CDCl₃) δ 6.70 (s, 2 H), (C(3,6)H₂), 3.44 (s, 3 H, C(6)NCH₃), 3.28 (s, 3 H, N(3)CH₃), 2.63 (s, 6 H), C(2)N(CH₃)₂), 2.23 (s, 1 H), and 2.15 (s, 3 H, C(4,5)(CH₃)₂). Anal. Calcd: C, 58.16; H, 6.71; N, 25.44. Found: C, 57.96; H, 6.81; N, 25.19. were extracted with chloroform and purified via column chromatography on 25.19.

A suitable procedure for the synthesis of the desired trione (IX) was found in the reaction of 3,5,5-trimethyl-1,5-dihydrolumiflavin² (VIII) with a stoichiometric amount of m-chloroperbenzoic acid in CHCl₃ (followed by washing with aqueous NaHCO₃ solution and column chromatography on silica with CHCl₃ as eluant). By this means, IX was obtained in moderate yield ($\sim 20\%$). The

$$\underbrace{\operatorname{VIII}}_{XIII}^{h} \underbrace{\operatorname{V}}_{N}^{0} \xrightarrow{} \left[\underbrace{\operatorname{VII}}_{N}^{h} \underbrace{\operatorname{VII}}_{N}^{0} \xrightarrow{} \left[\underbrace{\operatorname{VII}}_{N} \xrightarrow{} \left[\underbrace{\operatorname{VII}}_{N} \underbrace{\operatorname{VII}}_{N} \xrightarrow{} \left[\operatorname{VII}_{N} \xrightarrow{} \left[\operatorname{V$$

structure of IX is proven by elemental analysis and IR, ¹H NMR, and finally ¹³C NMR spectroscopy.²⁵ It represents the first isolated 6-aminopyrimidine-2,4,5(3H)-trione and thereby ends speculations¹¹ concerning its properties. Elemental analysis shows that C(5)=O is not hydrated, which is confirmed by four ¹³C signals between 163 and 154 ppm. An sp³-hybridized $\dot{C}(5)$ would absorb at much higher field (\sim 70 ppm; cf. Ghisla et al.^{4a} and Benkovic et al.^{4b}). When compound IX is put into aqueous solution, spectral changes show fast hydration and, at higher pH values, probably hydrolysis. These reactions will be subject to further investigation.

When 5-acetyl-3-methyl-1,5-dihydrolumiflavin²⁶ was treated with *m*-chloroperbenzoic acid, as in the case of VIII, again a new compound could be isolated.²⁷ In this case, the 13 C NMR spectrum shows a signal at 71 ppm, clearly indicating an sp³hybridized C(4a). We ascribe the 5-acetyl-4a-hydroxy-4a,5-dihydroflavin structure to the new product, although we cannot exclude a ring-opened hydrated form.

All attempts to convert the ketone IX with hydrazine to the hydrazone VIIc failed, which is not totally unexpected. Thus, alloxan (pyrimidine-2,4,5,6(1H,3H)-trione) oxidizes phenylhydrazine giving dialuric acid, nitrogen, and benzene.²⁸ We have not been successful in obtaining VII via condensation of IX with hydrazine salts, as described for alloxan.²⁹ The redox properties of IX, and its reactivity toward different carbonyl reagents and nucleophiles (amines and alcohols), are presently under investigation.

From our (preliminary) results, we can draw some conclusions of biological relevance. As our model (IX) is stable and does not "self-destruct" by intramolecular redox reactions, those arguments¹¹ against a 4a,5 ring opening during enzymatic catalysis are invalid. On the other hand, the UV spectrum of the model compound IX (λ_{max} 342 nm, ϵ 7120 M⁻¹ cm⁻¹) clearly shows no resemblance to the spectrum of the enzyme-bound X (λ_{max} 390-420 nm, ϵ 15000 M⁻¹ cm⁻¹)³⁰ (Figure 1). This finding does

(26) Hemmerich, P.; Prijs, B.; Erlenmeyer, H. Helv. Chim. Acta 1960, 43, 372

not support the proposal that the pyrimidine-2,4,5(3H)-trione structure I represents X, and reaction 3 appears to have no experimental basis. There remains the possibility of X being the p- or o-quinoid tautomer of I: a 4-hydroxypyrimidine-2,4-dione or a 2-hydroxypyrimidine-4,5-dione. These two tautomers are expected to be less stable than I and, therefore, less probable. Nonetheless, efforts are under way in this laboratory to synthesize these tautomers. Finally, the fluorescence properties³¹ of IX, unless drastically altered by the apoprotein, do not support its role as the emitter for bacterial luciferase.

Acknowledgment. This work was supported by grants from the National Science Foundation and the National Institutes of Health. We are very grateful to Professors J. J. Villafranca and S. J. Benkovic for the natural abundance ¹³C NMR spectra.

Intercalation of Potentially Reactive Transition-Metal Complexes in the Lamellar MnPS₃ Host Lattice

René Clement

Laboratoire de Physicochimie Minérale and ERA 672 Université de Paris Sud, 91405 Orsay Cedex, France

Received July 8, 1981 . Revised Manuscript Received September 9, 1981

The increasing interest in intercalated layer systems has remained largely centered, during the past years, on those systems exhibiting electrical conductivity.¹ Molecular reactions in the interlayer space are an attractive topic for research; recently, several novel organic reactions have been carried out in the in-terlayer space of layered silicates.²⁻⁵ The catalytic^{6,7} and photocatalytic^{8,9} properties of organometallic intercalated silicates are also of great potential interest. This communication describes the synthesis of new intercalation compounds based on MnPS₃ host layers, containing large cationic species potentially suitable for further chemistry or photochemical experiments.

MnPS₃ belongs to a class of lamellar semiconducting materials¹⁰ known, as the structurally analogous transition-metal dichalcogenides, to intercalate electron-donor species, such as amines¹¹ or cobaltocene.¹² It has been recently shown that $MnPS_3$

- (2) Adams, J. M.; Davies, S. E.; Graham, S. H.; Thomas, J. M. J. Chem. Soc., Chem. Commun. 1978, 930-931; 1979, 527-528.
 (3) Adams, J. M.; Ballantine, J. A.; Graham, S. H.; Laub, R. J.; Purnell, J. H.; Reid, P. I.; Shaman, W. Y. M.; Thomas, J. M. Angew. Chem., Int. Ed. Engl. 1978, 17, 282-283.
 (4) Bellasting A. Device M. Directly M. D.
- (4) Ballantine, J. A.; Davies, M.; Purnell, H.; Rayanakorn, M.; Thomas, J. M.; Williams, K. J. J. Chem. Soc., Chem. Commun. 1981, 8-9.
 (5) Ballantine, J. A.; Purnell, H.; Rayanakorn, M.; Thomas, J. M.; Wil-

- (c) Definition, S. A., Fullet, A., Reganatorin, M., Hollas, J. M., Williams, K. J. J. Chem. Soc., Chem. Commun. 1981, 9–10.
 (d) Welty, P. K.; Pinnavaia, T. J. J. Am. Chem. Soc. 1975, 97, 3819–3820.
 (7) Pinnavaia, T. J.; Raythatha, R.; Guo Shuh Lee, J.; Halloran, L. J.; Hoffman, J. F. J. Am. Chem. Soc. 1979, 101, 6891–6897.
 (8) Krenske, D.; Abdo, S.; Van Damme, H.; Cruz, M.; Fripiat, J. J. J. Phys. Chem. 1980, 84, 2447-2457
- (9) Abdo, S.; Canesson, P. Cruz, M.; Fripiat, J. J.; Van Damme, H. J. Phys. Chem. 1981, 85, 797-809.
- (10) Klingen, W.; Ott, R.; Hahn, H. Z. Anorg. Allg. Chem. 1973, 396, 271-278.

⁽²⁴⁾ Ghisla, S.; Hartmann, U.; Hemmerich, P.; Mueller, F. Liebigs Ann. Chem. 1973, 1388.

^{(25) 6-[[2-(}Dimethylamino)-4,5-dimethylphenyl]methylamino]-3-(25) 6-[[2-(Dimethylamino)-4,5-dimethylphenyl]methylamino]-3-methylpyrimidine-2,4,5(3*H*)-trione (IX): $C_{16}H_{20}N_{4}O_{3}$ (*M*, 316.35); mp 230-235 °C dec; IR (KBr) 1720 (C(5)=O), 1700 (C(4)=O), 1665 (C(2)-=O) cm⁻¹; UV (acetonitrile) λ_{max} (c, M^{-1} cm⁻¹) 229 (24 600), 245 (sh), 281 (12 900), 342 (7120) nm; ¹H NMR (CDCl₃) δ 7.10 (s, 1 H) and 6.87 (s, 1 H, C(3',6')H₂), 3.60 (s, 3 H, C(6)NCH₃), 3.39 (s, 3 H, N(3)CH₃), 2.51 (s, 6 H, C(2')N(CH₃)₂), 2.24 (s, 6 H, C(4',5')(CH₃)₂); ¹³C NMR (CDCl₃) δ 162.7, 160.5, 159.7, 154.5 (C(2,4,5,6)), 139.2, 136.4, 134.7, 124.9, 123.0 (C(1',2',3',4',5',6')), 43.0 (C(2')N(CH₃)₂), 38.2 (C(6)NCH₃), 28.5 (N(3)CH₃), 19.6, 19.3 (C(4',5')(CH₃)₂). Anal. Calcd: C, 60.74; H, 6.37; N, 17.71. Found: C, 60.56; H, 6.43; N, 17.53. (26) Hemmerich, P: Priis, B: Erlenwever, H. Helv. Chim. Acta **1960**, 43.

^{(27) 5-}Acetyl-4a,5-dihydro-4a-hydroxy-3-methyllumiflavin or 6-[(2-(27) 5-Acetyl-4a,5-dihydro-4a-hydroxy-3-methyllumiflavin or 6-[(2-(Acetylamino)-4,5-dimethylphenyl)methylamino]-3-methylpyrimidine-2,4,5-(3*H*)-trione hydrate: C₁₆H₁₈N₄O₄* 4 ₃H₂O (*M*, 354.36); mp 185–190 °C dec (to 3-methyllumiflavin); IR (KBr) 3400, 3250 (OH), 1735 (C(4)=O), 1690 and 1660 (C(2)=O and N(5)C=O), 1630 cm⁻¹; ¹H NMR (CDCl₃) δ 7.20 (s, 1 H) and 7.01 (s, 1 H, C(6,9)H₂), 3.63 (s, 3 H, N(10)CH₃), 3.36 (s, 3 H, N(3)CH₃), 2.36 (s, 6 H, C(7,8)(CH₃)₂), 2.10 (s, 3 H, C(5)NCOCH₃); ¹³C NMR (CDCl₃) δ 173.0 (N(5)COCH₃), 165.9, 162.1, 155.7 (C(2,4,10a)), 136.5, 133.1, 132.5, 127.5, 124.9, 117.5 (C(5a,6,7,8,9,9a)), 71.0 (C(4a)), 31.9 (N(10)CH₃), 2.7.6 (N(3)CH₃), 22.5 (N(5)COCH₃), 19.6, 19.2 (C(7,8)(C-H₃), 2.4 (S) (C(3,2) (C(3, $(H_3)_2$). Anal. Calcd: C, 54.23; H, 5.88; N, 15.81. Found: C, 54.13; H, 5.58; N, 15.58.

^{(28) (}a) Pellizzari, G. G. Orosi 1887, 10 (7,8); Gazz. Chim. Ital. 1887, 17, (b) Kuehling, O. Ber. Deutsch. Chem. Ges. 1891, 24, 4140.
 (29) Kuehling, O. Ber. Deutsch. Chem. Ges. 1898, 31, 1972.

⁽³⁰⁾ In fact, the UV data of IX resemble more those of 4a-hydroxy-4a,5-dihydroflavins (cf. Ghisla, S.; Entsch, B.; Massey, V.; Husein, M. Eur. J. Biochem. 1977, 76, 139), which should lead to caution in ascribing 4a-FIOH structures (cf. reaction 2) to enzyme-bound intermediates solely on the basis of UV data.

⁽³¹⁾ Compound IX is nonfluorescent in solution (solvent acetonitrile) and shows only very weak fluorescence ($\lambda_{max} \sim 410-430$ nm; λ_{max} (excitation) 330-340 nm) in DMF/ethylene glycol dimethyl ether glass at 77 K. Whether an excited state of IX can transfer its energy to another fluorescer (as proposed for a model reaction^{2k}) will be subject to further investigation.

⁽¹⁾ Whittingham, M. S.; Dines, M. B. Surv. Prog. Chem. Vol. 9; 1980, 9, 55-87.