



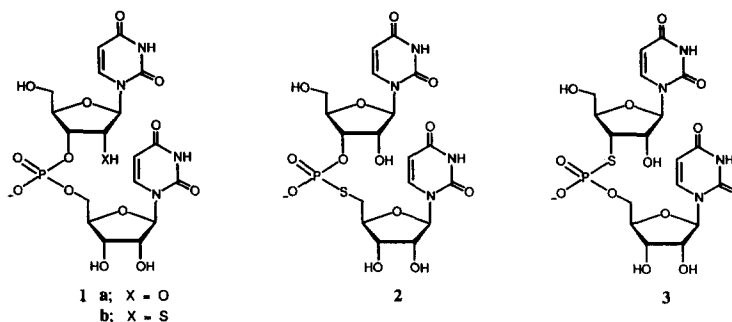
3' - Thiouridylyl - (3' → 5') - uridine

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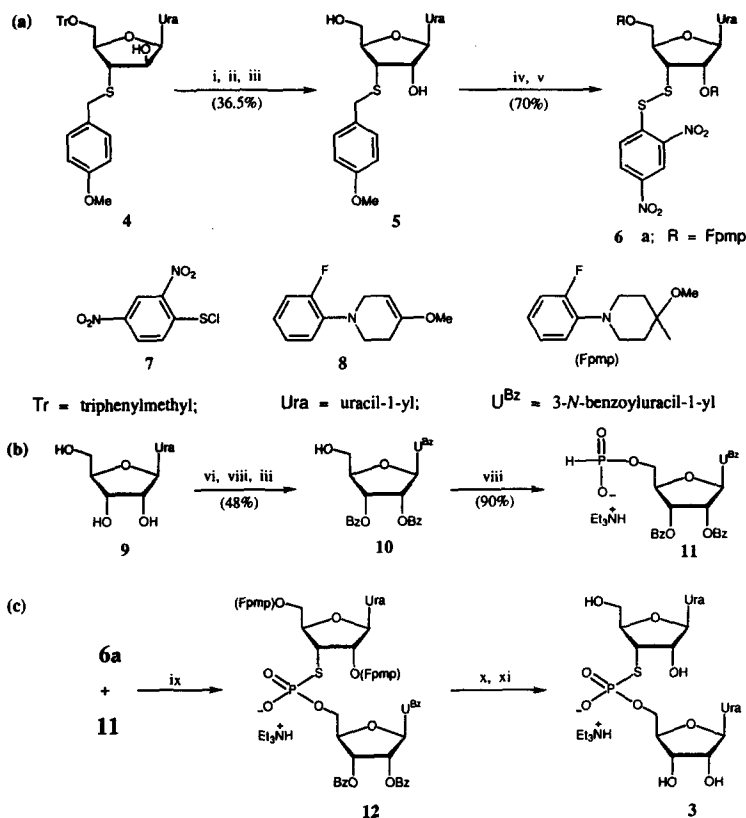
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Abstract: 3'-Thiouridylyl-(3'→5')-uridine **3** undergoes base-catalysed hydrolysis more rapidly than UpU **1a**; it also undergoes cleavage more rapidly than UpU in glacial acetic acid solution, but shows much less (if any) tendency to isomerize.

The possible development of antisense chemotherapy¹ has encouraged organic chemists to undertake the synthesis of oligodeoxyribo- and oligoribo-nucleotide analogues, and particularly analogues in which the sugar residues and internucleotide linkages are modified. Studies directed towards the elucidation of the mechanism of ribozyme action² have also led to the synthesis of modified oligoribonucleotides. Our own studies in the synthesis of oligonucleotide analogues have been further stimulated by a long-standing intrinsic interest³ in the chemistry of ribonucleic acids (RNA). As part of these studies, we have examined the effects of replacing the 2'-hydroxy function of uridylyl-(3'→5')-uridine (UpU) **1a** by a thiol function⁴ (as in **1b**) and the 5'-bridging oxygen atom in the internucleotide linkage of UpU by a sulfur atom⁵ (as in **2**). In order to complete what we consider to be a fundamental study in RNA chemistry, we have now⁶ synthesized 3'-thiouridylyl-(3'→5')-uridine **3** and have thereby examined the effect of replacing the 3'-bridging oxygen atom in the internucleotide linkage of UpU by a sulfur atom.



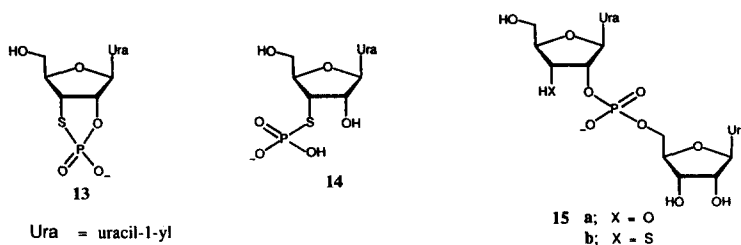
3'-Thiouridylyl-(3'→5')-uridine **3** was prepared (Scheme 1) from the building blocks **6a** and **11**. The thioether **4** was subjected (Scheme 1) to the Mitsunobu reaction⁸ in the presence of 4-nitrobenzoic acid. Saponification of the resulting 4-nitrobenzoate ester and removal of the 5'-*O*-trityl group gave the *S*-(4-methoxybenzyl) derivative⁹ **5** of 3'-thiouridine in 36.5% overall yield. Treatment of the latter compound with 2,4-dinitrophenylsulfanyl chloride¹¹ **7** in the presence of trifluoroacetic acid, followed by reaction of the product with the enol ether¹² **8**, also in the presence of acid, gave the fully-protected disulfide building block **6a** in 70% overall yield. The *H*-phosphonate building block¹³ **11** was prepared (Scheme 1b) in the usual way⁵ from 2,3'-*di-O*-benzoyl-3-*N*-benzoyluridine¹⁴ **10** in 90% yield. The three benzoyl



Scheme 1 Reagents and conditions: i, 4-(O_2N) $\text{C}_6\text{H}_4\cdot\text{CO}_2\text{H}$, Ph_3P , $\text{EtO}_2\text{C}-\text{N}=\text{N}-\text{CO}_2\text{Et}$, MeCN , 0°C to RT; ii, MeNH_2 , EtOH , RT; iii, $\text{AcOH}-\text{H}_2\text{O}$ (4:1 v/v), reflux; iv, 7, $\text{CF}_3\text{CO}_2\text{H}$, CH_2Cl_2 , 0°C ; v, 8, $\text{CF}_3\text{CO}_2\text{H}$, CH_2Cl_2 , RT; vi, TiCl_4 , $\text{C}_2\text{H}_5\text{N}$, 100°C ; vii, BzCl , $\text{C}_2\text{H}_5\text{N}$, RT; viii, a, reagent prepared from PCl_3 , Et_3N , 1,2,4-1*H*-triazole, THF , -35°C , 15 min, b, $\text{Et}_3\text{N}-\text{H}_2\text{O}$ (1:1 v/v), -35°C to RT; ix, Me_3SiCl , Et_3N , CH_2Cl_2 , RT, 17 hr; x, NH_3 , MeOH , RT, 17 hr; xi, $\text{AcOH}-\text{H}_2\text{O}$ (2:98 v/v), RT, 17 hr.

protecting groups were introduced to facilitate the isolation both of the *H*-phosphonate **11** and the intermediate dinucleoside phosphorothioate **12**. The two building blocks **6a** and **11** were coupled together (Scheme 1c) by essentially the same modification of the procedure of Li *et al*¹⁵ that was previously used in the preparation of uridylyl-(3'→5')-(5'-thiouridine)⁵ **2**. Following the removal of the benzoyl and Fmpmp protecting groups (steps x and xi) and chromatography of the products on DEAE-Sephadex A-25, 3'-thiouridylyl-(3'→5')-uridine **3** was isolated as its pure triethylammonium salt¹⁶ (δ_p [D_2O] 18.69).

When 3'-thiouridylyl-(3'→5')-uridine **3** was heated at 50°C in 0.05 mol dm^{-3} sodium glycinate buffer solution (pH 10.06), it was converted into uridine **9**, 3'-thiouridine 2',3'-cyclic phosphorothioate **13** and 3'-thiouridine 3'-phosphorothioate¹⁷ **14**. The reaction displayed pseudo first order kinetics with $t_{1/2} = 25 \text{ min}$ ¹⁹. Under the same conditions, $t_{1/2}$ for the hydrolysis of UpU **1a** was found to be 80 - 90 hr. Thus at pH 10.06 and 50°C , the 3'-thio-analogue **3** underwent base-catalyzed hydrolysis at a rate *ca.* 200 times faster than that of UpU **1a**. This result is perhaps surprising in that both reactions involve the formation and the cleavage of a phosphorus-oxygen bond. A possible contributing factor to the increased base lability of **3** relative to that of **1a** is the presence of a larger 3'-sulfur atom in **3** which presumably leads to a lower energy transition state in the hydrolysis reaction and to a less strained five-membered cyclic ester product **13**. The latter cyclic



phosphorothioate **13** was most probably the only primary nucleotide product in the above base-catalysed hydrolysis of **3**; in a separate study carried out in 0.05 mol dm⁻³ sodium glycinate buffer solution (pH 9.87) at 30°C, 3'-thiouridine 2',3'-cyclic phosphorothioate **13** was found to undergo hydrolysis ($t_{1/2}$ = 165 min) to give 3'-thiouridine 3'-phosphorothioate **14** as the sole product.

3'-Thiouridylyl-(3'→5')-uridine **3** was rapidly converted mainly into uridine **9** and 3'-thiouridine 2',3'-cyclic phosphorothioate **13** in glacial acetic acid solution at 30°C; after 6 min, only *ca.* 25% of substrate **3** remained and after 30 min, virtually none was left. Small quantities of 3'-thiouridine 3'-phosphorothioate²⁰ **14** and another product [t_R = 7.51 min (programme A)¹⁹, < 5% of the total absorbance at 260 nm] that may have been 3'-thiouridylyl-(2'→5')-uridine **15b** or the corresponding dimeric disulfide⁴ were observed after 30 min. UpU **1a** was found to be somewhat more stable in glacial acetic acid at 30°C, but it showed more tendency to isomerize to give uridylyl-(2'→5')-uridine **15a**; after 60 min, the products²¹ consisted of substrate **1a** (49.8%), its 2'→5'-isomer **15a** (15.7%), uridine 2'- and 3'-phosphates (4.6%), uridine 2',3'-cyclic phosphate (12.3%) and uridine (17.6%).

Finally, the action of various hydrolytic enzymes on 3'-thiouridylyl-(3'→5')-uridine **3** was investigated. The latter dinucleoside phosphorothioate **3** was found to be a good substrate for *Crotalus adamanteus* snake venom phosphodiesterase, but it did not undergo digestion in the presence of bovine spleen phosphodiesterase. 3'-Thiouridylyl-(3'→5')-uridine **3** also proved to be a good substrate for ribonuclease A: in the presence of the latter enzyme, it underwent digestion to give first a mixture of uridine **9**, 3'-thiouridine 2',3'-cyclic phosphorothioate **13** and 3'-thiouridine 3'-phosphorothioate **14**, and finally a 1:1 mixture of uridine and 3'-thiouridine 3'-phosphorothioate. In confirmation of the latter observation, 3'-thiouridine 2',3'-cyclic phosphorothioate **13** was independently found to be a good substrate for ribonuclease A; in the presence of the latter enzyme it underwent quantitative digestion to give 3'-thiouridine 3'-phosphorothioate **14** as the sole product.

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9. Found : C, 53.51, H, 5.30, N, 7.29. C₁₇H₂₀N₂O₆S requires : C, 53.67; H, 5.30; N, 7.36%, m.p. 144-146°C; δ_{H} [(CD₃)₂SO] 3.19 (1 H, dd, *J* 4.9 and 9.3), 3.63 (1 H, m), 3.72 (3 H, s), 3.73-3.83 (3 H, m), 4.01 (2 H, m), 5.27 (1 H, t, *J* 4.7), 5.58 (1 H, d, *J* 8.1), 5.65 (1 H, d, *J* 1.7), 5.92 (1 H, d, *J* 5.2), 6.84 (2 H, d, *J* 8.6), 7.24 (2 H, d, *J* 8.6), 8.02 (1 H, d, *J* 8.1), 11.29 (1 H, br s). The identical compound **5** was obtained, albeit in low yield, by the acidic hydrolysis of the products of the reaction between the conjugate base of 4-methoxybenzyl mercaptan and 3'-*O*-mesyl derivative of 1-(2,5-di-*O*-trityl- β -D-xylofuranosyl)uracil¹⁰.
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16. In an experiment starting from disulfide **6a** (0.1 mmol) and *H*-phosphonate **11** (0.2 mmol), the isolated yield of pure dinucleoside phosphorothioate **3** was 387 A₂₆₀ units.
17. 3'-Thiouridine 3'-phosphorothioate **14** was prepared by a modification of the procedure of Müller and Roth¹⁸: 3'-deoxy-3'-(2,4-dinitrophenyldisulfanyl)uridine **6**, R = H (0.1 mmol) was treated first with Me₃SiCl (0.5 mmol) and Et₃N (0.55 mmol) and then with P(OSiMe₃)₃ (0.15 mmol) in CH₂Cl₂ solution. Aqueous work-up and fractionation of the products on DEAE-Sephadex A-25 gave the triethylammonium salt of 3'-thiouridine 3'-phosphorothioate **14** (507 A₂₆₀ units), δ_{P} [D₂O] 16.1. Treatment of a vigorously stirred solution of the latter material (210 A₂₆₀ units) in water (0.5 ml) with *n*-Bu₃N (0.5 mmol) and ClCO₂Et (0.2 mmol), and fractionation of the products on DEAE-Sephadex A-25 gave the triethylammonium salt of 3'-thiouridine 2',3'-cyclic phosphorothioate **13** (168 A₂₆₀ units), δ_{P} [D₂O] 37.1.
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19. The progress of the reaction was monitored by HPLC. Reverse phase HPLC was carried out on a 5 μ Jones APEX ODS column, following programme A [linear gradient of CH₃CN - 0.1 mol dm⁻³ aqueous triethylammonium acetate (pH 7.0) (3:97 to 20:80 v/v) over 10 min, followed by a linear gradient (20:80 to 30:70 v/v) over a further 5 min]. Retention times for uridine **9**, 3'-thiouridine 2',3'-cyclic phosphorothioate **13**, 3'-thiouridine 3'-phosphorothioate **14** and 3'-thiouridylyl-(3'→5')-uridine **3** were 3.37, 4.83, 6.27 and 7.76 min, respectively.
20. The formation of 3'-thiouridine 3'-phosphorothioate **14** was presumably due to the presence of traces of moisture in the acetic acid.
21. This reaction was monitored by reverse phase HPLC, following programme B [linear gradient of CH₃CN - 0.1 mol dm⁻³ aqueous triethylammonium acetate (pH 7.0) (3:97 to 7:93 v/v) over 10 min]. The numbers in parentheses represent percentages of the total absorbance at 260 nm.

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