Glycosylation of 4,7,8,9-Tetra-O-acetyl-2-deoxy- 2β ,3 β -epoxy-N-acetylneuraminic Acid Methyl Ester¹⁾

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Reaction of the 2β , 3β -epoxyneuraminic acid derivative prepared from 2,3-dehydroneuraminic acid derivative with primary or secondary alcohol gave the corresponding glycoside(s) or ortho ester.

In the course of our glycosylation study on N-acetylneuraminic acid we have prepared some new glycosyl donors such as 2,3-dibromo-, 2,3-epoxy-, and 2-halo-3-hydroxy-N-acetylneuraminic acid derivatives from tetra-O-acetyl-2-deoxy-2,3-dehydro-N-acetylneuraminic acid methyl ester (1). We have already reported that the 2,3-dibromo compound is well-suited for the stereoselective β -glycosylation since the bromo group prevents the elimination reaction. We report here glycosylation of the 2,3-epoxy compound $2^{(2)}$ whose epoxy ring is β -oriented (Scheme 1). If the glycosylation reaction occurs in S_N2 type, 2 is suited to give α -glycosides.

Treatment of 2 with methanol in the presence of d-camphor-10-sulfonic acid or Amberlyst 15 catalyst (catalytic amount) gave only methyl α -glycoside 3 in

†Present address: Institute of Bio-Active Science (IBAS), Nippon Zoki Pharmaceutical Co., Ltd., Kinashi, Yashirocho, Kato-gun, Hyogo 673-14. nearly quantitative yield. To determine the anomeric configuration the glycoside 3 was treated with phenyl chlorocarbonothioate⁴⁾ in the presence of 4-dimethylaminopyridine (DMAP) to give 5, which was reduced with tributylstannane to the known α -glycoside $6^{5)}$ quantitatively. Antimony pentachloride (1.0 equiv) in methanol was also effective catalyst for this glycosylation. By use of 2 equivalents of methanol in the presence of antimony pentachloride or trimethylsilyl triflate (TMSOTf) catalyst, however, an inseparable mixture of α - and β -glycosides was obtained.

Treatment of **2** with methyl 2,3,4-tri-O-benzyl- α -D-glucoside (**8**)⁶⁾ in the presence of boron trifluoride-ether complex or titanium(IV) chloride gave methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-2,5-dideoxy-2-fluoro- β -D-erythro-L-gluco-2-nonulopyranosonate (**12**)²⁾ or methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-2-chloro-2,5-dideoxy- β -D-erythro-L-gluco-2-nonulopyranosonate (**7**)²⁾ without any glycosides. The hydroxy- β -glycoside **9** was

Table 1. Reaction of	ne Epoxide 2 and Methanol
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Catalyst(equiv)	MeOH	Solvent	Rea	ction	Yield/% of Produc		
Catalyst(equiv)	(equiv)	Solvent	Temp	Time	3	4	
CSA (cat.)		Methanol	rt	15 min	99ª)		
Amberlyst 15 (cat.)		Methanol	rt	15 min	98ª)		
$SbCl_5$ (1.0)		Methanol	rt	15 min	89ª)		
$SbCl_5$ (1.0)	2.0	ClCH ₂ CH ₂ Cl	$0 \circ C$	15 min	41 b)	20 ^{b,c)}	
TMSOTf (1.0)	2.0	ClCH ₂ CH ₂ Cl	$0 { m c}$	15 min	62 ^{b)}	29 ^{b)}	

a) Isolated yield. b) Determined by ¹H NMR. c) Methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-2-chloro-2,5-dideoxy-β-D-*erythro*-L-*gluco*-2-nonulopyranosonate (**7**) was also obtained in 37% yield.

Table 2. Reaction of the Epoxide 2 and Glucose Derivative 8^{a)}

Catalyst(equiv)	Solvent	Rea	ction	Yield/% ^{b)} of 9
Catalyst(equiv)	Solvent	Temp	Time/h	11elu/70 Ol 9
CSA (cat.)	ClCH ₂ CH ₂ Cl	rt	0.5	no reaction
$BF_3 \cdot OEt_2 (1.0)$	THF	rt	0.2	c)
TiCl ₄ (1.0)	ClCH ₂ CH ₂ Cl	rt	0.2	d)
SbCl ₅ (1.0)	ClCH ₂ CH ₂ Cl	rt	0.2	49
$TMSOTf^{e)}$ (1.0)	ClCH ₂ CH ₂ Cl	$0 ^{\circ} \mathrm{C}$	0.2	56

a) Two equivalents of **8** were used. b) Isolated yield. c) Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-2,5-dideoxy-2-fluoro- β -D-erythro-L-gluco-2-nonulopyranosonate (**12**) was obtained in 80% yield. d) Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-2-chloro-2,5-dideoxy- β -D-erythro-L-gluco-2-nonulopyranosonate (**7**) was obtained in 80% yield. e) Na₂HPO₄ was used as a buffer.

obtained in a moderate yield by the use of either antimony pentachloride or trimethylsilyl triflate. In the ^1H NMR spectrum of **9**, the coupling constant $J_{7.8}$ was 1.5 Hz and the $\Delta\delta|\text{H-9'-H-9}|$ value was 0.94 ppm. In cases of the carbonothioate **10** and the glycoside **11**, the $J_{7.8}$ values were 1.5 and 2.2 Hz and the $\Delta\delta|\text{H-9'-H-9}|$ values were 0.94 and 0.94 ppm, respectively. As **11** was determined to be a β -glycoside by comparing its physical data with those of the authentic sample, $^{3)}$ it is safe to deduce the anomeric configuration of **9** from the $J_{7.8}$ and $\Delta\delta|\text{H-9'-H-9}|$ values in ^{1}H NMR spec-

trum.^{7,8)} Similarly, reaction of **2** with methyl 2,4,6-tri-O-benzyl- β -D-galactopyranoside (**13**)⁹⁾ afforded in 64% yield only β -glycoside **14**, whose $J_{7,8}$ was 1.8 Hz and $\Delta\delta$ |H-9'-H-9| value was 1.02 ppm. To make sure that the anomeric position was β configuration, the hydroxy-glycoside **14** was carbonothioated and reduced in the same manner as above to give the protected 3-O-(2 β -neuraminyl)galactoside **16**, which was identical with the authentic sample.³⁾ Although the reaction of **2** with the acceptor **13** in the presence of mercury(II) chloride catalyst gave no glycosides, treatment of **2**

with a less sterically hindered acceptor 17 afforded an unstable product 18 in 75% yield. In its 1H NMR spectrum, the coupling constant $J_{7.8}$ was 4.2 Hz and the $\Delta\delta|H-9'-H-9|$ value was 0.89 ppm, which were fit for neither α - nor β -glycoside. The product 18 was treated with phenyl chlorocarbonothioate to determine the structure. In spite of the presence of a carbonothioate group in 19 as evident from its 1H NMR spectrum, the signal corresponding to H-3 of 19 was not shifted toward lower field. The carbonothioated 19 was reduced with tributylstannane to give 20, which exhi-

bited $J_{2ax,3eq}$ coupling constant as 2.4 Hz. From these data a series of compounds obtained by the reaction of 2 with 17 was revealed to be the ortho esters as illustrated in Scheme 5. Treatment of 20 with dilute trifluoroacetic acid in aqueous methanol gave almost instantaneously two degradation products, which were treated with acetic anhydride in pyridine to afford the acetylated compounds 21 and 22. These compounds were characterized by IR, 1H NMR, and mass spectra.

In summary, the epoxide 2 gave α -glycosides by the use of highly reactive acceptors such as methanol, and

 β -glycosides by sugar derivatives such as a 6-unprotected glucoside and a 3-unprotected galactoside in the presence of antimony pentachloride or trimethylsilyl triflate. On the other hand, the ortho ester was yielded in the presence of mercury(II) chloride catalyst. The anomeric configuration of neuraminic acid derivatives was able to be determined from the $J_{7,8}$ coupling constant and the $\Delta\delta|\text{H-9'-H-9}|$ value in ¹H NMR spectrum. For example, 2β -glycosides showed ca. 2 Hz in $J_{7,8}$ and ca. 1 ppm of $\Delta\delta|\text{H-9'-H-9}|$ value, whereas 2α -glycosides showed more than 7 Hz in $J_{7,8}$ coupling constant and less than 0.5 ppm of $\Delta\delta|\text{H-9'-H-9}|$ value. These values in glycosides remain unaltered even if 3 position of NeuAc unit is substituted by any group.

Experimental

General. Melting points were taken on a Mitamura Riken flat-bulb thermometer with a heating metal block and

uncorrected. Elemental analyses were done on a Perkin-Elmer 240C elemental analyzer. Nuclear magnetic resonance spectra (NMR) were obtained with a JEOL GX-500 instrument in the FT mode. Chemical shifts were expressed in parts per million from internal tetramethylsilane (δ). Coupling constants are in hertz(Hz) and splitting pattern abbreviations are: s, singlet; d, doublet; dd, doublet of doublets; ddd, doublet of double doublets; m, mutiplet; br, broad. Mass spectra (MS) were obtained on a JEOL DX-300 spectrometer. Infrared spectra (IR) were recorded on a JASCO A-3 spectrophotometer. Optical rotations [α]_D were recorded on a JASCO DIP-181 digital polarimeter.

Analytical thin-layer chromatography (TLC) was conducted on precoated TLC glass sheets (silica gel 60F-254, layer thickness 0.25 mm) manufactured by E. Merck. Detection was effected with 2% concentrated sulfuric acid in ethanol.

¹H NMR data were summarized in Tables 3, 4, and 5, and MS, elemental analyses, mp, R_f , $[\alpha]_D$, and IR data in Table 6. Reaction of the Epoxide 2 and Methanol. Procedure A

Table 3. ¹H NMR Data of NeuAc Unit at 500 MHz in Chloroform-d

								Che	mical	shifts	, δ (multi	plicit	y)				
Com- pound	H-2ax (d)	H-3eq (d)	H-3ax (d)	H-4 (dd)	H-5 (ddd)	H-6 (dd)	H-7 (dd)	H-8 (ddd)	H-9 (dd)	H-9' (dd)	Me ester (s)	NH (d)	OH-2 (br.s)	OH-3 (d)	O-Ac, N-Ac (s)	Phenyl (m)	Aglycon (s)
3			3.82ª)	5.15	4.22	4.54	5.28	5.37	4.08	4.32	3.84	5.63		3.02	1.89, 2.03, 2.08 2.09, 2.14		3.46
5			5.87	5.50	4.42	4.63	5.31	5.36	4.10	4.32	3.82	5.52			1.92, 2.04, 2.06 2.09, 2.16	7.04—7.44	3.51
6		2.57	1.94	4.86	4.07	4.13	5.33	5.43	4.11	4.32	3.82	5.14			1.89, 2.03, 2.04 2.14, 2.15		3.32
9			3.85 ^{a)}	5.05	4.21	4.30	5.35	5.24	4.04	4.98	3.80	5.39		2.53	1.87, 1.97, 2.04 2.06, 2.14		
10			5.87	5.43	4.41	4.44	5.41	5.28	4.08	5.02	3.68	5.63			1.88, 1.98, 2.05 2.06, 2.17	6.85—7.50	
11		2.47	1.88	5.16	4.11	4.27	5.38	5.27	4.13	5.07	3.72	5.49			1.86, 1.97, 2.01 2.05, 2.13		
14			3.90 ^{a)}		4.08						3.54	4.52		3.68	1.72, 1.93, 2.01 2.02, 2.08		
15			5.82		4.25						3.51	3.95			1.65, 1.99, 2.00 2.03, 2.09	6.90—7.65	
16		2.70	1.80		3.99						3.54	3.86			1.64, 1.98, 2.00 2.03, 2.06		
18		4.60			3.87						3.68	5.84	5.48		1.69, 1.93, 2.03 2.07, 2.13		
19		4.72			4.18						3.71	6.64			1.72, 1.92, 1.93 2.06, 2.13	7.10—7.50	
20		4.74 ^{a)}			3.39						3.66	5.69			1.65, 1.95, 2.03 2.06, 2.12		
21	4.37	5.71 a)		5.22	4.25	3.88	5.39	5.30	4.20	4.73	3.75	5.87			1.90, 1.99, 2.05 2.07, 2.13, 2.17		

Compound-		First-order coupling constants, Hz														
Compound	$J_{2ax,3ex}$	$J_{3 m eq,3ax}$	$J_{3 m eq,4}$	$J_{3ax,4}$	$J_{4,5}$	$J_{5,6}$	$J_{5, m NH}$	$J_{6,7}$	$J_{7,8}$	$J_{8,9}$	$J_{8,9}$	$J_{9,9}$	J _{OH,3ax}			
3				9.8	10.1	11.0	10.1	2.1	7.9	6.1	2.6	-12.5	6.1			
5				8.6	10.4	11.0	9.8	2.1	8.0	6.3	2.5	-12.5				
6		-12.8	4.6	12.5	10.1	10.2	9.8	2.1	8.5	5.5	2.7	-12.5				
9				9.5	10.4	10.7	10.1	2.1	1.5	9.2	2.3	-12.5	7.3			
10				9.8	9.8	10.1	9.5	2.0	1.5	9.2	2.1	-12.5				
11		-12.9	5.1	10.8	11.2	10.4	10.4	2.3	2.2	9.3	2.2	-12.5				
14				9.5	9.8	10.0	10.1	2.0	1.8	9.2	2.4	-12.5	7.0			
15				9.8	10.4	10.5	10.5	2.1	2.1	8.9	2.2	-12.5				
16		-13.4	4.6	11.6	11.2	10.7	10.4	1.8	b)	8.5	2.1	-12.5				
18			5.2		8.2	10.7	9.8	2.1	4.2	7.9	2.1	-12.5				
19			5.5		7.6	10.0	9.2	2.7	2.7	7.6	2.1	-12.5				
20	2.4		5.8		7.6	11.3	8.2	1.8	5.5	6.7	2.4	-12.5				
21	1.2		3.5		10.7	9.4	9.8	2.1	4.9	7.3	2.4	-12.5				

a) Multiplicity: dd. b) Not assigned owing to the complexity of the spectrum.

Table 4. ¹H NMR Data of Glc or Gal Unit at 500 MHz in Chloroform-d

Com-			Chem	ical sł	nifts, δ	(mul	tiplici	ty)		First-order coupling constants, Hz							
pound	H-1 (d)	H-2 (dd)	H-3 (dd)	H-4 (dd)	H-5 (ddd)	H-6 (dd)	H-6' (dd)	OMe (s)	OH-4 (d)	$J_{1,2}$	$J_{2,3}$	$J_{3,4}$	$J_{4,5}$	$J_{5,6}$	$J_{5,6}$	$J_{6,6}$	<i>Ј</i> он,4
9	4.90	3.64	3.99	3.72	3.79	3.88	4.09	3.36		3.7	9.5	9.2	8.9	2.0	2.3	-10.9	
10	4.93	3.67	3.99	3.84	3.82	3.87	4.19	3.36		3.7	9.5	8.9	9.0	1.0	2.0	-10.3	
11	4.89	3.61	3.97	3.70	3.74	3.80	3.81	3.35		3.7	9.5	9.2	10.1	2.5	2.0	-10.8	
14	4.35	3.87	4.39	4.10	3.82	3.69	3.73	3.49		7.0	9.2	3.1	1.0	6.7	6.3	-10.0	
15	4.38	3.84	4.42	4.08 ^{a)}	3.88	3.77	3.79	3.47		7.3	9.8	3.1	0	7.2	7.1	-10.2	
16	4.34	3.75	4.09	3.94 ^{a)}	3.77	3.68	3.71	3.46		7.3	9.8	2.8	0	6.1	6.1	-9.8	
18	4.28	3.51	3.67	4.01	3.57	3.72	3.78	3.55	2.98	7.6	9.5	3.7	0	6.1	4.6	-10.1	1.5
19	4.29	3.53	3.70	3.90^{b}	3.58	3.75	3.81	3.56	2.68	7.9	9.8	3.4	0.5	5.4	5.8	-10.1	2.5
20	4.27	3.50	3.69	3.95	3.56	3.72	3.79	3.55	2.64	7.9	9.5	3.4	0	5.8	5.5	-9.8	2.5
22	4.38	3.58	4.98	5.42	3.83	3.49	3.57	3.59		7.6	10.1	3.7	1.0	6.7	6.3	-9.8	

a) Multiplicity: d. b) Multiplicity: ddd.

Table 5. ¹H NMR Data of Other Groups at 500 MHz in Chloroform-d

Com		Chemical s	shifts, δ (multipl	licity)		First-order coupling constants, Hz
Com- pound		PhC <u>H2</u> (AB quartet)		Ph (m)	O-Ac (s)	$J_{f g}$ em
9	4.74 and 4.89,	4.76 and 4.82,	4.85 and 4.97	7.23—7.45		-11.0, -11.9, -11.0
10	4.78 and 4.85,	4.88 and 4.96,	4.92 and 5.00	6.85 - 7.50		-11.9, -11.0, -11.0
11	4.70 and 4.87,	4.75 and 4.82,	4.84 and 4.97	7.20 - 7.50		-10.8, -12.0, -11.0
14	4.54 and 4.56,	4.74 and 4.87,	4.88 and 4.90	7.20 - 7.50		-11.9, -11.3, -13.7
15	4.60, a)	4.82 and 5.02,	4.95 and 5.03	6.90 - 7.65		-11.9, -13.7
16	4.56, ^{a)}	4.68 and 4.88,	4.79 and 4.80	7.20 - 7.50		-11.0, -12.5
18	4.57 and 4.61,	4.63 and 4.81,		7.25 - 7.50		-11.9, -11.0
19	4.58 and 4.60,	4.62 and 4.85,		7.10 - 7.50		-10.0, -10.7
20	4.57 and 4.61,	4.61 and 4.85,		7.23 - 7.45		-12.2, -11.0
22	4.44 and 4.55,	4.61 and 4.85,			1.94, 2.01	-11.9, -11.9

a) Multiplicity: s.

Table 6. MS, Elemental Analyses, Mp, R_f , $[\alpha]_D$, and IR Data of 3—22

Com-	Formula	MS ^{a)}		Ar	nal.		Мр	$R_{ m f}$	$[\alpha]_{D}(c)^{b}$			$ u_{ m max}^{ m KBr}$	
pound		(M+H)		C (%)	H(%)	N(%)	$\theta_{\rm m}/^{\circ}{ m C}$	·	(Temp)	NH, OH	ester	amide I	amide II
3	C ₂₁ H ₃₁ NO ₁₄	522	Calcd Found	48.37 48.35	5.99 5.89	2.69 2.75	c)	0.27 ^{d)}	-34.3°(1.1) (15°C)	3430	1743	1660	1540
5	$C_{28}H_{35}NO_{15}S$	658	Calcd Found	51.14 50.85	5.36 5.43	2.13 2.32	— c)	0.56 ^{d)}	-18.4°(1.2) (15°C)	3430	1748	1660	1540
6	$C_{21}H_{31}NO_{13}$	506	Calcd Found	49.90	6.18 6.01	2.77 2.92	87—89 ^{e)}	0.43 ^{d)}	-23.7°(1.1) (21°C)	3420	1742	1660	1540
9	$C_{48}H_{59}NO_{19}$	955	Calcd Found	60.43 60.71	6.23 6.45	1.47 1.39	c)	0.46 ^{d)}	+11.5°(1.3) (21°C)	3430	1743	1665	1540
10	$C_{55}H_{63}NO_{20}S$		Calcd Found	60.60 60.23	5.82 5.78	1.29 1.44	^{c)}	0.60 ^{f)}	-5.6°(2.6) (20°C)	3400	1748	1665	1540
11	C ₄₈ H ₅₉ NO ₁₈	938	Calcd Found	61.46 61.59	$6.34 \\ 6.34$	1.49 1.31	— ^{c)}	0.52 ^{f)}	+23.3°(1.0) (29°C)	3400	1748	1665	1540
14	C ₄₈ H ₅₉ NO ₁₉	955	Calcd Found	60.43 60.56	6.23 6.57	1.47 1.59	— ^{c)}	0.48 ^{f)}	-15.7°(1.0) (11°C)	3380	1743	1670	1540
15	$C_{55}H_{63}NO_{20}S$		Calcd Found	60.60 60.91	5.82 5.75	1.29 1.46	— ^{c)}	0.64 ^{f)}	+9.1°(0.7) (12°C)	3420	1752	1660	1540
16	$C_{48}H_{59}NO_{18}$	938	Calcd Found	61.46 61.23	$6.34 \\ 6.27$	1.49 1.08	— c)	0.58 ^{f)}	+5.3°(1.0) (29°C)	3400	1743	1660	1540
18	C ₄₈ H ₅₃ NO ₁₉		Calcd Found	57.00 56.72	6.18 6.08	1.62 1.87	c)	0.40 ^{d)}	+31.8°(1.2) (11°C)	3400	1745	1660	1540
19	C ₄₈ H ₅₇ NO ₂₀ S		Calcd Found	57.65 57.83	5.75 5.41	1.40 1.42	— c)	0.54 ^{f)}	-9.5°(1.0) (12°C)	3400	1748	1660	1530
20	$C_{41}H_{53}NO_{18}$		Calcd Found	58.08 57.82	6.30 6.07	1.65 1.36	— c)	0.40 ^{f)}	+12.3°(0.9) (11°C)	3420	1742	1660	1530
21	$C_{22}H_{31}NO_{14}$	534	Calcd Found	49.53 49.50	5.86 5.79	2.63 2.38	c)	0.26 ^{f)}	+49.8°(1.6) (12°C)	3370	1748	1660	1540
22	C ₂₅ H ₃₀ O ₈	459	Calcd Found	65.49 65.44	6.60 6.73		c)	0.54 ^{g)}	+6.5°(2.0) (12°C)		1740		

a) Fast atom bombardment method. b) Measured in chloroform. c) Viscous syrup. d) Solvent system is benzene-acetone (3:2). e) Recrystallized from hexane-ethyl acetate. f) Solvent system is benzene-acetone (2:1). g) Solvent system is benzene-ethyl acetate (6:1).

(d-camphor-10-sulfonic acid or SbCl $_5$ catalyst in the presence of large excess methanol). To a solution of **2** (100 mg, 0.20 mmol) in absolute methanol (2.5 ml) was added d-camphor-10-sulfonic acid (cat. amount) or antimony pentachloride (26 μ l, 0.20 mmol) at room temperature under argon atmosphere. After stirring for 15 min, the mixture was evaporated to dryness, and the residue was dissolved in ethyl acetate. The solution was washed with 5% NaHCO $_3$ and brine, dried over anhydrous Na $_2$ SO $_4$. The solvent was removed in vacuo to give a syrupy residue, which was chromatographed on a silica-gel column (benzene-acetone, 1:1) to give methyl glycoside of methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-5-deoxy- α -D-erythro-L-gluco-2-nonulopyranosonate (**3**) as a syrup.

Procedure B (Amberlyst 15 catalyst in the presence of large excess methanol). A mixture of 2 (100 mg, 0.20 mmol) and Amberlyst 15 (cat. amount) in absolute methanol (2.5 ml) was stirred for 15 min under argon atmosphere. It was filtered and the resin was washed well with methanol. The combined filtrate and washings were evaporated to a syrup, which was purified in the same manner as procedure A.

Procedure C (SbCl₅ catalyst in the presence of 2 equiv of methanol). To a solution of 2 (100 mg, 0.20 mmol) in 1,2-dichloroethane (2.0 ml) containing 0.2 M (1M=1 mol dm⁻³) methanol (2.0 ml) was added antimony pentachloride (26 μ l, 0.20 mmol) at 0 °C under argon atmosphere. The mixture was stirred for 15 min, diluted with chloroform, washed with 5% NaHCO₃ and brine, dried over anhydrous Na₂SO₄, and evaporated to dryness. The residue was purified in the same manner as procedure A. The fast migrating zone was methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-2-chloro-2,5-dideoxy-*β*-Derythro-L-gluco-2-nonulopyranosonate (7)²⁰ (40 mg, 37%) and the slow migrating zone was an inseparable mixture of 3 (41%) and 4 (20%).

Procedure D (TMSOTf catalyst in the presence of 2 equiv of methanol). To a solution of **2** (100 mg, 0.20 mmol) in 1,2-dichloroethane (2.0 ml) containing 0.2 M methanol was added trimethylsilyl triflate (40 μ l, 0.20 mmol) at 0 °C under argon atmosphere. The mixture was worked up in the same manner as above to give **3** (62%) and **4** (29%).

Methyl (Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-5-deoxy-3-O-[phenoxy(thiocarbonyl)]- α -D-erythro-L-gluco-2-nonulopyranosid)onate (5). A solution of 3 (50 mg, 0.096 mmol), 4-dimethylaminopyridine (50 mg, 0.41 mmol), and phenyl chlorocarbonothioate (27 μ l, 0.20 mmol) in anhydrous acetonitrile (1.0 ml) was stirred for 1.5 h at room temperature under argon atmosphere. The mixture was evaporated to dryness and the residue, which was partitioned between ethyl acetate and water. The organic layer was separated, washed with water and brine, dried over anhydrous Na₂SO₄, and evaporated in vacuo to give a crude material, which was chromatographed on a silica-gel column (benzene-acetone, 5:2) to give 5 (64 mg, 98%) as a colorless syrup.

Methyl (Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy- α -p-glycero-p-galacto-2-nonulopyranosid)onate (6). To a solution of 5 (20 mg, 0.030 mmol) in toluene (1.0 ml) were added tributylstannane (20 μ l, 0.074 mmol) and AIBN (cat. amount) under argon atmosphere. The mixture was heated at 110 °C for 10 min, evaporated to dryness and the residue was chromatographed on a silica-gel column (benzene-acetone, 2:1) to give crystals. Recrystallization from hexane-ethyl acetate gave 6 (15 mg, 98%) as white needles.

Reaction of the Epoxide 2 and 8. Procedure A (BF₃·OEt₂)

catalyst). To a stirred solution of **2** (20 mg, 0.041 mmol) and **8**⁶⁾ (40 mg, 0.086 mmol) in tetrahydrofuran (0.6 ml) was added boron trifluoride-ether complex (5.2 μ 1, 0.041 mmol) at room temperature under argon atmosphere. After stirring for 0.2 h, the solvent was removed and the residue was dissolved in ethyl acetate. The solution was washed with 5% NaHCO₃ and brine, dried over anhydrous Na₂SO₄, and evaporated in vacuo to give a crude syrup, which was chromatographed on a silica-gel column (benzene-acetone, 1:1). Unreacted **8** (35 mg) was recovered and then methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-2,5-dideoxy-2-fluoro- β -Derythro-L-gluco-2-nonulopyranosonate (12)²⁾ (16.7 mg, 80%) was obtained.

Procedure B (TiCl₄ catalyst). To a solution of 2 (20 mg, 0.041 mmol) and 8 (40 mg, 0.086 mmol) in 1,2-dichloroethane (0.6 ml) was added titanium(IV) chloride (4.5 μ l, 0.041 mmol) at room temperature under argon atmosphere. The mixture was stirred for 0.2 h, diluted with ethyl acetate, washed with saturated Na₂SO₄ and brine, dried over anhydrous Na₂SO₄, and evaporated in vacuo to give a residue, which was treated in the same manner as procedure A to give 8 (30 mg) and 7 (17.3 mg, 80%).

Procedure C (SbCl₅ catalyst). To a solution of **2** (20 mg, 0.041 mmol) and **8** (40 mg, 0.086 mmol) in 1,2-di chloroethane (0.6 ml) was added antimony pentachloride (5.2 μl, 0.041 mmol) at room temperature under argon atmosphere. The mixture was stirred for 0.2 h, diluted with ethyl acetate, washed with 5% NaHCO₃ and brine, dried over anhydrous Na₂SO₄, and evaporated in vacuo to give a crude material, which was chromatographed on a silica-gel column (benzene-acetone, 2:1) to give methyl 2,3,4-tri-O-benzyl-6-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-5-deoxy-β-D-erythro-L-gluco-2-nonulopyranosyl)onate]-α-D-glucopyranoside (**9**) (19 mg, 49%) as a syrup.

Procedure D (TMSOTf catalyst). To a mixture of 2 (20 mg, 0.041 mmol), 8 (40 mg, 0.086 mmol), anhydrous disodium hydrogenphosphate (20 mg), and 1,2-dichloroethane (0.6 ml) was added trimethylsilyl triflate (8 μl, 0.041 mmol) at 0 °C under argon atmosphere. It was stirred for 0.2 h at the same temperature and worked up in the same manner as above to give 9 (22 mg, 56%).

Methyl 2,3,4-Tri-O-benzyl-6-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-5-deoxy-3-O-[phenoxy(thiocarbonyl)]- β -D-erythro-L-gluco-2-nonulopyranosyl)onate]- α -D-glucopyranoside (10). A solution of 9 (22 mg, 0.023 mmol), 4-dimethylaminopyridine (6 mg, 0.049 mmol), and phenyl chlorocarbonothioate (7 μ l, 0.051 mmol) in anhydrous acetonitrile (0.3 ml) was stirred for 24 h at room temperature under argon. The mixture was evaporated to dryness, and the residue was dissolved in ethyl acetate, washed with water and brine, dried over anhydrous Na₂SO₄, and evaporated in vacuo to give a crude material. This was chromatographed on a silica-gel column (benzene-acetone, 5:2), triturated with hexane-ethyl acetate, and washed with hexane to give 10 (24 mg, 95%).

Methyl 2,3,4-Tri-O-benzyl-6-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy- β -D-glycero-D-galacto-2-nonulo-pyranosyl)onate]- α -D-glucopyranoside (11). To a solution of 10 (20 mg, 0.018 mmol) in toluene (1 ml) were added tributylstannane (10 μl, 0.037 mmol) and AIBN (cat. amout) under argon atmosphere. The mixture was heated at 100 °C for 15 min and the solvent was removed in vacuo to give a crude material, which was chromatographed on a silica-gel

column (benzene-acetone, 2:1) to give 11 (16.5 mg, 96%) as a syrup.

Methyl 2,4,6-Tri-O-benzyl-3-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-5-deoxy- β -D-erythro-L-gluco-2-nonulopyranosyl)-onate]- β -D-galactopyranoside (14). To a mixture of 2 (20 mg, 0.041 mmol), 13¹⁰⁾ (40 mg, 0.086 mmol), anhydrous disodium hydrogenphosphate (20 mg), and 1,2-dichloroethane (0.3 ml) was added trimethylsilyl triflate (8 μ l, 0.041 mmol) at -20 °C under argon atmosphere. It was stirred for 10 min, diluted with ethyl acetate, washed with 5% NaHCO₃ and brine, dried over anhydrous Na₂SO₄, and evaporated to dryness. The residue was chromatographed on a silica-gel column (benzene-acetone, 2:1) to give 14 (25 mg, 64%) as a syrup.

Methyl 2,4,6-Tri-O-benzyl-3-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-5-deoxy-3-O-[phenoxy(thiocarbonyl)]- β -D-erythro-L-gluco-2-nonulopyranosyl)onate]- β -D-galactopyranoside (15). A solution of 14 (18 mg, 0.019 mmol), 4-dimethylaminopyridine (6 mg, 0.049 mmol), and phenyl chlorocarbonothioate (5 μ l, 0.036 mmol) in anhydrous acetonitrile (0.3 ml) was stirred for 24 h at room temperature under argon atmosphere. After evaporation, the residue was partitioned between ethyl acetate and water. The organic layer was separated, washed with water and brine, dried over anhydrous Na₂SO₄, and evaporated in vacuo to give a crude material, which was chromatographed on a silica-gel column (benzene-acetone, 4:1) to give 15 as a syrup.

Methyl 2,4,6-Tri-O-benzyl-3-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy- β -p-glycero-p-glacto-2-nonulopyranosyl)onate]- β -p-galactopyranoside (16). To a solution of 15 (15 mg, 0.014 mmol) in toluene (1 ml) were added tributylstannane (7 μl, 0.026 mmol) and AIBN (cat. amount) under argon atmosphere. The mixture was heated at 110 °C for 10 min and evaporated to dryness. The residue was chromatographed on a silica-gel column (benzene-acetone, 4:1) to give a syrup. This was triturated with hexane-ethyl acetate and washed with hexane to give 16 (12 mg, 93%).

Reaction of the Epoxide 2 with 17. A mixture of 2 (100 mg, 0.20 mmol), 17^{10} (160 mg, 0.43 mmol), mercury(II) chloride (60 mg, 0.22 mmol), powdered molecular sieves 4A (200 mg), and 1,2-dichloroethane (3 ml) was stirred for 40 h. The reaction mixture was filtered and the solid was washed with chloroform. The combined filtrate and washings were washed with water and brine, dried over anhydrous Na_2SO_4 , and evaporated in vacuo to give a residue, which was chromatographed on a silica-gel column (benzene-acetone, 3:2) to give the ortho ester 18 (132 mg, 75%).

To a solution of **18** (100 mg, 0.12 mmol) and 4-dimethylaminopyridine (60 mg, 0.49 mmol) in anhydrous acetonitrile (1.5 ml) was added phenyl chlorocarbonothioate (18 μ 1, 0.13

mmol) under argon atmosphere. The mixture was stirred for 1 h at room temperature, evaporated to dryness, and the residue was partitioned between ethyl acetate and water. The ethyl acetate layer was separated, washed with water and brine, dried over anhydrous Na₂SO₄, and evaporated in vacuo to give a syrup, which was chromatographed on a silica-gel column (benzene-acetone, 3:2) to give 19 (100 mg, 86%).

To a mixture of **19** (80 mg, 0.08 mmol), anhydrous sodium acetate (40 mg), and toluene (3 ml) was added tributylstannane (43 μ 1, 0.16 mmol) under argon atmosphere. The mixture was heated at 110 °C for 10 min, diluted with ethyl acetate, washed with water and brine, dried over anhydrous Na₂SO₄, and evaporated to dryness. The residue was chromatographed on a silica-gel cloumn (benzene-acetone, 3:2), triturated with hexane-ethyl acetate, and washed with hexane to give **20** (50 mg, 74%).

To a solution of **20** (100 mg, 0.12 mmol) in 60% aq methanol (3 ml) was added trifluoroacetic acid (1 drop) at room temperature. After stirring for 10 min, the mixture was evaporated to dryness and the residue was treated with acetic anhydride (0.4 ml) and pyridine (0.8 ml) for 10 h at room temperature under argon atmosphere. The resulting mixture was evaporated to a syrup, which was chromatographed on a silica-gel column (benzene-ethyl acetate, 6:1, followed by benzene-acetone, 1:1) to give methyl 3,4-di-O-acetyl-2,6-di-O-benzyl- β -D-galactopyranoside (**22**) (50 mg, 92%) and methyl 5-acetamido-3,4,7,8,9-penta-O-acetyl-2,5-dideoxy-D-erythro-L-manno-2-nonulopyranosonate (**21**) (51 mg, 91%) as a syrup.

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