gel permeation on Sephadex LH-20. Sugar [3,4] and methylation [5] analyses showed 3 and 4 to be glucopyranosides. The structure of 3 is evident from its elemental composition, spectral properties and the fact that it gave *p*-cresyl- β -D-glucopyranoside upon catalytic hydrogenation. Parishin (4) is a neutral substance which on catalytic hydrogenation followed by treatment with diazomethane yielded *p*-cresyl- β -D-glucopyranoside and trimethyl citrate in the molar ratio 3:1. The fission must be due to hydrogenolysis of benzyl esters, as no signals for aromatic methyl groups were observed in the NMR spectrum of 4. These results demonstrate that 4 is tris[4-(β -D-glucopyranosyloxy)benzyl] citrate, a structure also consistent with the NMR spectrum.

EXPERIMENTAL

General conditions were the same as in an earlier communication [2].

Plant material. Vanda parishii. Rchb.f. was delivered from Mr N. Prakash, Chandra Orchid and Bulb Nurseries, 8 1/2 miles P.O. Kalimpong, West Bengal, India.

Isolation of 3 and 4. Fresh plants of V. parishii (3 kg) were extracted with MeOH (101), and the solution was concentrated to 0.651. A part (100 ml) of this extract was diluted to 300 ml with water and washed with $CHCl_3$ (4 × 50 ml). The aqueous layer was saturated with butanol and extracted with butanol saturated with water (7 \times 50 ml). The butanolic phase was washed with water (25 ml) and evaporated to dryness. A part (2 g) of the residue (6.4 g) was chromatographed on a silica gel column (5×8.5 cm) using CHCl₃-MeOH-H₂O (13:7:2, lower phase) as eluent. The fraction containing 3 (300 mg) was filtered through a column of Sephadex LH-20 $(5 \times 70 \text{ cm})$ using EtOH-H₂O (1:1) as eluent giving crude 3 (96 mg), which was crystallised from iso-PrOH-H2O. Recrystallisation from EtOAc-EtOH gave 3 (56 mg). A part (220 mg) of the fraction containing 4 (450 mg) was chromatographed on silica gel $(2.6 \times 11 \text{ cm})$ using the same eluent as above. The fraction containing 4 (150 mg) was filtered through a column of Sephadex LH-20 (2.5 × 83 cm) using EtOH-H₂O (1:1) as eluent, giving 4 as a colourless amorphous solid (78 mg).

Glucoside 3. Needles (EtOAc-EtOH), mp 154-157°C; $[\alpha]_{278}^{27}$ - 63° (c 0.77, MeOH). (Found: C 54.6; H 6.3; O 39.1. C₁₃H₁₈O₇ requires: C 54.5; H 6.3; O 39.1). IR: $\nu_{\text{max}}^{\text{Kar}}$ 3700-3000(s), 1615(m), 1590(m), 1510(s) cm⁻¹. UV: $\lambda_{\text{max}}^{\text{MOH}}$ (log ϵ) 277.5 (2.96), 271 (3.04) nm. ¹H NMR (D₂O): δ 3.36-4.12 7.19 and 7.42 (4 H, A₂B₂ system, J 9 Hz). Parishin (4). Amorphous solid, $[x]_{27_8}^2 - 59^\circ$ (c 0.80, MeOH). IR: ν_{max}^{KB7} 3700-3000(s), 1735(s), 1615(m), 1590(m), 1515(s) cm⁻¹. ¹H NMR (D₂O): δ 2.76 and 2.94 (4 H, two AB systems, J 15 Hz), 3.4-4.0 (18 H), 4.4-5.1 (the benzylic and the anomeric protons; the HOD signal partially overlapping), 6.9-7.4 (12 H). ¹H NMR (Pyridine-d₃): δ 3.32 and 3.37 (4 H, two AB systems, J 15 Hz), 3.80-4.66 (18 H), 5.09 (s, 4 H), 5.28 (s, 2 H), 5.58 (d, 3 H, J 6 Hz), 7.1-7.5 (12 H).

Hydrogenation of 3. A soln of 3 (51 mg) in MeOH (9 ml) was hydrogenated over Pd (20 mg, 10% on carbon) at room temp. and atm. pres. After 7 hr the catalyst was filtered off and the soln was evaporated to dryness giving p-cresyl- β -D-glucopyranoside, mp 180–182°C (*iso*-PrOH-H₂O); $[\alpha]_D^{22} - 67^\circ$ (c 0.76, H₂O) (lit. [6] mp 178–179.5°C; $[\alpha]_D^{20} - 67.7^\circ$ (H₂O)), further identified by NMR.

Hydrogenation of 4. Parishin (92 mg) was hydrogenated as described for 3. Catalyst was filtered off and the soln was treated with an excess of CH₂N₂ in Et₂O and evaporated to dryness. Residue was washed with CHCl₃ (5 × 1 ml) and the CHCl₃ phase was evaporated to dryness. Residue was erystallised from Et₂O-hexane at -20° C giving trimethyl citrate (14 mg), mp 76-78°C (lit. [7] mp 78.5-79°C). The total amount of trimethyl citrate was found by GLC to be 17.5 mg. The residue insoluble in CHCl₃ above was dissolved in H₂O and the soln washed with CHCl₃-MeOH (1:1). The aq phase was evaporated to dryness giving *p*-cresyl- β -D-glucopyranoside (68 mg), mp 181-182.5°C; $[\alpha]_{D^2}^{2^2} - 66^{\circ}$ (c 0.27, H₂O).

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NEOCAPILLEN, A NEW ACETYLENIC HYDROCARBON FROM ARTEMISIA CAPILLARIS

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Key Word Index-Artemisia capillaris; Compositae; acetylenic hydrocarbon; neocapillen.

Abstract—During an investigation of Artemisia capillaris, a new acetylenic hydrocarbon, neocapillen, was isolated as a minor component and its structure determined.

The structures of the new acetylenic compounds, 1-(2'-methoxyphenyl)-2,4-hexadiyne (o-methoxy capillen), capillanol and norcapillen, in the essential oil of the stalks and leaves of *Artemisia capillaris* was described previously [1-3]. As a continuation of our study on the components of *A. capillaris* extracts, which have been recognized to make an important contribution to the characteristic odour, we wish to report the identification of a new polyacetylenic hydrocarbon neocapillen (1), which seemed to be derived from capillen (2) [4].

Fresh roots of *A. capillaris* were chopped finely and extracted with ether. The extract was chromatographed on alumina and divided into 3 fractions from which compound 1, which had a Shiitake-like odour, was isolated by PLC.

By elementary analysis, the molecular formula of 1 was determined to be $C_{12}H_{10}$ (Found: C, 93.44; H, 6.56; Calcd.: 94.46; H, 6.54%). The spectral data of 1 are given in the Experimental. The IR showed absorptions at 2210, 2200 and 2120 cm⁻¹(-C=C-), at 1595 and 1490 cm⁻¹ (aromatic), and at 755 and 690 cm⁻¹ (aromatic H). These data indicated that the compound was an aromatic mono-substituted hydrocarbon with a C_6H_5 unit. The IR absorptions at 1210 and 1120 cm⁻¹ and the intense UV absorption at 222 nm in *n*-hexane indicated the presence of a 1-phenyl-1,3-dyne function [5]. The NMR spectrum showed no olefinic proton, a signal at δ 2.16(2H) was the two protons of a methylene attached to a conjugated diacetylenic bond. The five protons in the benzene ring appeared as a broad singlet from δ 7.05 to 7.55. Consequently, the splitting pattern of the signals in the NMR spectrum were consistent with structure (1) for neocapillen. It had a UV spectrum almost super-imposable with that of norcapillen [3].

Besides the molecular ion peak at m/e 154 the other significant MS peaks were at m/e 153(M⁺-H, 82%), 152(M⁺-H₂, 47%), 139(Ph-C=C-C=C-CH₂⁺, 36%), 113 (Ph-C=C-C⁺, 7%).

For the confirmation of the identity, compound 1 was prepared from capillen. Good agreement of the IR, NMR and MS spectra was observed between neocapillen (1) and the synthetic material. 1 may be an artefact produced from 2 by UV irradiation, although no acetylenic substance has been detected in the upper parts of the plants. 1 was catalytically hydrogenated over PtO₂ in ethanol to give octahydroneocapillen, which was found to be identical with *n*-hexylbenzene in all respects (IR, NMR, MS spectrum). Thus, the structure of neocapillen was confirmed to be 1-phenyl-1,3-hexadiyne. The synthesis of this phenylacetylene had been reported [6]. Many phenyl diacetylene (Ph-C=C-C=C-R) derivatives which have C₇ or C₁₁-side chains have been detected in the genus *Bidens, Coreopsis* and *Dahlia*.

The first isolation of neocapillen from a natural source is significant from biogenetic considerations as all the phytoacetylenes so far isolated, have a phenyl diacetylene skeleton or a modification thereof.

EXPERIMENTAL

UV spectra were measured in EtOH. NMR spectra were determined at 60 MHz, in CCl₄ and with TMS as internal standard.

Plant material and oil removal. Fresh roots (5 kg) of A. capillaris, collected in October at the suburbs of Osaka prefecture, were chopped finely and extracted with Et_2O for 5 weeks at room temp. Neocapillen was also obtained after only several hr extraction.

Isolation of neocapillen (1). The extract (15.8 g) was chromatographed on deactivated alumina with *n*-hexane, C_6H_6 and E1.0. and divided into 3 fractions. Elution with C_6H_6 gave neocapillen (1) which was then isolated by PLC (R_f 0.60) developed with *n*-hexane-EtOAc (6:1). MS *m/e* (rel. int.): 154(100°₄) M⁺, 153(82), 45(63), 152(47), 139(36), 73(29); 89(24), 88(22), 43(20), 151(18). IR v_{max} cm⁻¹: 3025, 3005, 2960, 2900, 2855, 2825, 2210, 2200, 2120, 1943, 1880, 1595, 1570, 1490, 1450, 1440, 1425, 1375, 1330, 1290, 1170, 1150, 1065, 1050, 1020, 950, 785, 755, 690. UV $\lambda_{\text{ErOH}}^{\text{max}}$ nm: (log ϵ) 211 nm (3.42), 222(3.55), 244(2.63), 257(3.13), 272(3.32), 289(3.20). NMR: δ 1.00(3H, *t*, *J* = 6.2 Hz), 2.16(2H, *q*, *J* = 6.2 Hz), 7.05 ~ 7.55(5H, *m*).

Catalytic hydrogenation of 1. Catalytic hydrogenation of 1 (50 mg) in EtOH (5 ml) over PtO₂ (1.4 mg) was carried out at room temp for 3 hr. The product was purified by GLC (Carbowax-20 M 5%, 80 ~ 100 mesh, 4 mm × 3 m, 1.5 Kg/cm²), to give a colourless oil, NMR: δ 0.88(3H, t, $-CH_2-CH_3$), 1.26(8H, m, $-(CH_2)_4-$), 2.68(2H, t, $\phi-CH_2-$), 7.23(5H, m, $\phi-$). (Found: C, 88.80; H, 11.20; Calcd for C₁₂H₁₈:C, 88.81; H, 11.1×1

Isolation of capillen (2) The essential oil from stalk and leaves of a *A. capillaris* was chromatographed on activated alumina (60 g, 300 mesh) with *n*-hexane, C_6H_6 , Et_2O , EtOAc and EtOH, and divided into 5 fractions. Subsequent elution with C_6H_6 gave capillen (2) which was then isolated by prep GLC, and identified (UV, IR, NMR) by comparison with an authentic specimen [4,7].

Catalytic hydrogenation of 2. Catalytic hydrogenation of 2 (55 mg) in EtOH (5 ml) over PtO₂ (1.5 mg) was carried out at room temp for 3.5 hr. The product was purified by prep GLC. NMR: δ 0.88 (3H, t, -CH₂-CH₃), 1.26(8H, m, -(CH₂)₄-), 2.68(2H, t, ϕ -CH₂-), 7.23(5H, m, ϕ -). MS m/e: 162(M⁺, 26%).

Irradiation of capillen (2). 2 (50 mg) dissolved in a mixture of 70 ml MeOH and a trace of KOH were irradiated for about 24 hr in a Pyrex tube with light of wavelength 3500 Å. This soln was acidified and extracted with Et_2O ; the extract was washed, dried and evaporated to leave a viscous reddish oil. After separation by prep GLC 10 mg of neocapillen (1) were obtained as a pure compound.

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