

## Synthesis of Alkylphosphon(othio)ate Analogues of DNA

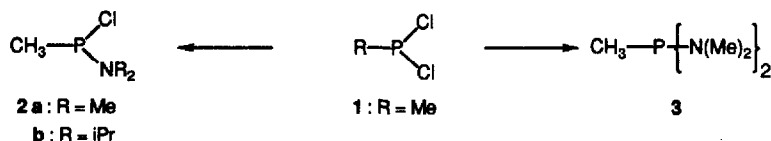
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**Abstract:** Easily accessible bis(diisopropylamino)alkylphosphines (alkyl is methyl, *n*-butyl or *n*-octyl) are convenient reagents for the synthesis of DNA fragments having an alkylphosphonate or alkylphosphonothioate linkage at a predetermined site in solution and on solid support.

Oligodeoxynucleotides in which one or more of the natural internucleosidic (3'-5') phosphodiester bonds have been replaced by non-charged methylphosphonate linkages are valuable tools in biophysical and biological (*e.g.* protein-DNA interaction, antisense) studies<sup>1</sup>. The assembly of oligodeoxynucleotide methylphosphonates has been accomplished using reagents **2a**<sup>2</sup>, **2b**<sup>3</sup> or **3**<sup>4</sup> which, in turn, are accessible starting with commercially available methyldichlorophosphine<sup>5</sup> (**1**, R = Me). However, the introduction of other alkylphosphonate bonds by a similar route is less convenient due to the poor accessibility of the requisite alkylidichlorophosphines (*i.e.* **1**; R = *n*-butyl or *n*-octyl).

In order to study<sup>6</sup> the effect of increasing chain length on the affinity of d-oligonucleotide alkylphosphon(othio)ates for complementary d-oligonucleotides, we report here that the conversion of bis(diisopropylamino)chlorophosphine (**4**)<sup>7</sup> into the bis(diisopropylamino)alkylphosphines **6a-c** opened the way to a solid-phase synthesis of oligodeoxynucleotides containing alkylphosphonate or alkylphosphonothioate linkages at preselected positions (*e.g.* hexamers **19a-c** and **20**, respectively).

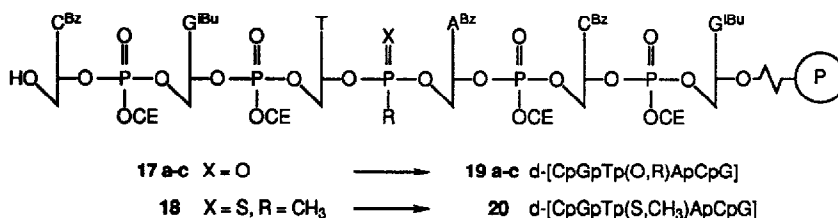


Earlier studies from our laboratory revealed that crystalline and readily accessible bis(diisopropylamino)chlorophosphine (**4**)<sup>7</sup> could not only be applied to the formation of internucleosidic 1*H*-phosphonate linkages<sup>8</sup>, but also to the preparation of the interesting phosphitylating reagents **5a**<sup>9</sup> and **5b**<sup>10</sup>. Apart from this, it was to be expected<sup>11</sup> that reagent **4** would also be a convenient starting compound for the synthesis of the bis(diisopropylamino)alkylphosphines **6a-c**. Indeed, replacement of the chlorine atom in **4** by an alkyl group using *n*-butyl lithium or methyl(*n*-octyl)magnesium halides, following a reported<sup>11</sup> procedure, gave the three reagents **6a-c** as homogeneous liquids (see Table 1).



In the second stage, the individual alkylphosphonamidites **8a-c** ( $B^1 = T$ ; 0.55 mmol each) in dry dichloromethane (5 mL) were condensed with 2-*N*-diphenylacetyl-3'-*O*-benzoyl-2'-deoxyguanosine<sup>13</sup> (**9**; 0.50 mmol) in the presence of 1-*H*-tetrazole (1.38 mmol). <sup>31</sup>P NMR analysis of the reaction mixture indicated in each case rapid formation of the dinucleoside phosphonites **10a-c**<sup>14</sup>. *In situ* oxidation of **10a-c** with *t*-butylhydroperoxide for 5 min at 20°C resulted, after work-up and purification by silica gel column chromatography, in the isolation of the fully protected modified dimers **12a-c**<sup>15</sup>. Removal of the diphenylacetyl and benzoyl groups from **12a-c** by ammonolysis ( $\text{NH}_3/\text{MeOH}$ ) for 48 h at 50°C and subsequent acidic hydrolysis of the DMT group afforded, after purification (Sephadex LH-20), the modified TG dimers **14a-c**<sup>16</sup>, the identity and homogeneity of which was unambiguously ascertained by <sup>31</sup>P- and <sup>1</sup>H-NMR spectroscopy. Apart from the successful preparation of the TG dimers **14a-c**, it was also demonstrated that the intermediate phosphonites **10a-c** could be converted smoothly into the corresponding alkylphosphonothioates **15a-c**. Thus, thiolation of **10a-c** (0.50 mmol) in dichloromethane (5 mL) with phenylacetyl disulfide (**11**)<sup>10,17</sup> (1.0 mmol) for 5 min at 20°C gave, after isolation and purification, the fully protected TG dimers **13a-c**<sup>18</sup>. Deblocking of the latter compounds, under the same conditions as mentioned for the conversion of **12a-c** into **14a-c**, yielded the homogeneous dimers **15a-c**<sup>16</sup>.

The ease of preparing intermediate dimers **10a-c** and their high propensity toward oxidation and thiolation, urged us to apply the alkylphosphonamidites **8a-c** in a solid-phase synthesis of the modified hexamers **19a-c** and **20**, using an automated Gene Assembler and controlled pore glass as the solid support.



To this end, the immobilized fully protected hexamers **17a-c** were assembled by stepwise elongation of immobilized (3'-*O*-succinyl linkage) 2-*N*-isobutyryl-2'-deoxyguanosine via 1-*H*-tetrazole-mediated coupling of the appropriate phosphoramidites **16** ( $B = T$ ,  $C^{\text{Bz}}$ ,  $A^{\text{Bz}}$  or  $G^{\text{IBu}}$ ), prepared by phosphorylation of **7** with **5a**<sup>19</sup>, and **8a-c** ( $B^1 = T$ ) followed by oxidation ( $\text{I}_2/\text{H}_2\text{O}$ ) and subsequent acidolysis of the DMT group. The efficiency of each elongation step was, as gauged spectrophotometrically by the released DMT cation, higher than 97%. Cleavage of the succinyl linkage and removal of all protective groups by ammonolysis ( $\text{NH}_3/\text{MeOH}$ , 50°C, 24 h) gave, after purification (Sephadex G-50), the target hexamers **19a-c**, the identity and homogeneity of which was firmly established by <sup>31</sup>P NMR spectroscopy and FPLC analysis. In the same way, the immobilized methylphosphonothioate-containing hexamer **18** was prepared by thiolation of the intermediate methylphosphonite linkage between the thymidine and 6-*N*-benzoyl-2'-deoxyadenosine unit of the growing chain. Ammonolysis of **18** resulted in the isolation of **20**, the methylphosphonothioate linkage of which was *inter alia* confirmed by <sup>31</sup>P NMR and FPLC analysis<sup>20</sup>.

In conclusion, the new and readily available bis(diisopropylamino)alkylphosphines **6a-c** promise to be convenient reagents for the future preparation of alkylphosphon(othio)ate analogues of nucleic acids and other biopolymers.

## References and notes

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- <sup>31</sup>P NMR (CH<sub>2</sub>Cl<sub>2</sub>) data of compounds **8a-c** (B<sup>1</sup> = T): **8a**; 120.91, 120.55. **8b**; 127.62, 127.38. **8c**; 127.59, 127.41. R<sub>f</sub> values (ethyl acetate/triethylamine, 95/5, v/v) of compounds **8a-c**: **8a**; 0.51. **8b**; 0.53. **8c**; 0.58. In a similar way, compounds **8a-c** (B<sup>1</sup> = C<sup>Bz</sup>, A<sup>Bz</sup> or G<sup>Bu</sup>) were obtained.
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- <sup>31</sup>P NMR (CH<sub>2</sub>Cl<sub>2</sub>) data of compounds **10a-c**: **10a**; 184.86, 184.16. **10b**; 189.12, 188.76. **10c**; 189.30, 188.48.
- Yields of compounds **12a-c**: **12a**; 87%. **12b**; 86%. **12c**; 89%. <sup>31</sup>P NMR (CH<sub>2</sub>Cl<sub>2</sub>) data of compounds **12a-c**: **12a**; 33.85, 32.40. **12b**; 35.66, 34.36. **12c**; 36.63, 34.39. R<sub>f</sub> values (dichloromethane/methanol, 92/8, v/v) of compounds **12a-c**: **12a**; 0.53, 0.51. **12b**; 0.56. **12c**; 0.60. Mass spectrometric [M+Na]<sup>+</sup> data of compounds **12a-c**: **12a**; 1193.4. **12b**; 1234.6. **12c**; 1290.8.
- All deprotected compounds were characterized by <sup>31</sup>P NMR spectroscopy as well as mass spectrometry.
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- Yields of compounds **13a-c**: **13a**; 84%. **13b**; 82%. **13c**; 85%. <sup>31</sup>P NMR (CH<sub>2</sub>Cl<sub>2</sub>) data of compounds **13a-c**: **13a**; 99.73, 98.64. **13b**; 104.35, 103.26. **13c**; 104.32, 103.32. R<sub>f</sub> values (dichloromethane/methanol, 92/8, v/v) of compounds **13a-c**: **13a**; 0.56, 0.53. **13b**; 0.58. **13c**; 0.63. Mass spectrometric [M+Na]<sup>+</sup> data of compounds **13a-c**: **13a**; 1209.2. **13b**; 1253.9. **13c**; 1307.4.
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