Stereochemical Determination of Roflamycoin: ¹³C Acetonide Analysis and Synthetic Correlation

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Abstract: The relative configuration of natural roflamycoin (1) was determined by ¹³C acetonide analysis, combined with other supporting spectroscopic data. A previous proposal for the configuration of roflamycoin was shown to be incorrect. The absolute configuration was determined by the advanced Mosher's method. Our proposed structure of natural roflamycoin was confirmed by the synthesis of a degradation fragment that incorporates all of the 11 independent stereogenic centers of roflamycoin. The complete structure of roflamycoin is illustrated in Figure 1.

Complex polyol segments make the stereochemical elucidation of polyene macrolide antibiotics a serious challenge. The most convenient method for determining the structure of a complex organic molecule is X-ray crystallography. Unfortunately, the polyene macrolide antibiotics exhibit poor crystallographic properties. Of the over 200 known polyene macrolide antibiotics, only the structures of amphotericin B and roxaticin have been determined by X-ray crystallography.^{2,3} Without crystalline compounds suitable for X-ray analysis, one must rely on spectroscopic analysis, chemical degradation, and synthetic studies to determine the structures of polyene macrolide antibiotics. Schreiber and Goulet determined the stereochemical pattern for mycoticin by synthesizing a compound that matched a degradation product obtained from the natural material.⁴ The stereochemical assignments of nystatin A₁⁵ and pentamycin,⁶ as well as the partial stereochemical assignment of lienomycin,⁷ have been determined by NMR analysis of degradation products complemented by synthetic studies. Lancelin and Beau used sophisticated NMR analysis to predict the stereochemical structure of pimaricin,8 and the structure was later confirmed by the synthesis of pimaricin aglycon.⁹ Most recently, the

stereochemical structure of candidin was determined by a combination of NMR methods. 10

Although several approaches have been suggested for determining the configuration of alternating polyol chains, ¹¹ we desired a more convenient method to determine the stereochemical structure of polyene macrolide antibiotics. One powerful tool which may be employed to determine the relative configuration of compounds containing 1,3-polyols is that of ¹³C NMR acetonide analysis. ¹² In general, it has been observed that syn 1,3-diol acetonides have acetal methyl shifts at 19 and 30 ppm, while anti 1,3-diol acetonides have methyl shifts at approximately 25 ppm. The ¹³C NMR acetonide analysis has proven to be a very reliable indicator of stereochemistry in that syn 1,3-diol acetonides are easily distinguished from anti 1,3-diol acetonides. ¹³ We report the stereochemical determination of roflamycoin using the ¹³C NMR analysis of acetonide derivatives along with other spectroscopic investigations.

Roflamycoin (1) was isolated over 20 years ago from the mycelium of *Streptomyces roseoflavus* ARIA 1951 var. *jenensis* nov. var. JA 5068 and was originally called flavomycoin (Figure 1).¹⁴ It is a polyene macrolide antibiotic with antifungal activity and has been shown to form sterol dependent ion-channels.¹⁵ The flat structure was determined by careful degradative studies, in addition to NMR and mass spectral analysis.¹⁶ The relative stereochemistry, however remained a mystery. Roflamycoin has 11 independent stereogenic centers leading to 2048 possible

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⁽¹⁾ Camille and Henry Dreyfus Teacher-Scholar, 1990-1995. Alfred P. Sloan Research Fellow 1992-1994.

⁽²⁾ The structure of amphotericin B was determined from its N-iodoacetyl derivative: Ganis, P.; Avitabile, G.; Mechinski, W.; Schaffner, C. P. J. Am. Chem. Soc. 1971, 93, 4560-4564.

⁽³⁾ The structure of roxaticin was determined from its heptaacetate derivative: Maehr, H.; Yang, R.; Hong, L.-N.; Liu, C.-M.; Hatada, M. H.; Todaro, L. I. J. Org. Chem. 1989, 54, 3816-3819

Todaro, L. J. J. Org. Chem. 1989, 54, 3816-3819.
(4) (a) Schreiber, S. L.; Goulet, M. T. J. Am. Chem. Soc. 1987, 109, 4718-4720. (b) Schreiber, S. L.; Goulet, M. T. J. Am. Chem. Soc. 1987, 109, 8120-8122.

^{(5) (}a) Lancelin, J.-M.; Beau, J.-M. Tetrahedron Lett. 1989, 30, 4521–4524.
(b) Prandi, J.; Beau, J. M. Tetrahedron Lett. 1989, 30, 4517–4520.
(c) Nicolaou, K. C.; Ahn, K. H. Tetrahedron Lett. 1989, 30, 1217–1220.

^{(6) (}a) Oishi, T. Pure Appl. Chem. 1989, 61, 427-430. (b) Tadashi Nakata, personal communication.

⁽⁷⁾ Pawlak, J.; Nakanishi, K.; Iwashita, T.; Borowski, E. J. Org. Chem. 1987, 52, 2896-2901.

⁽⁸⁾ Lancelin, J.-M.; Beau, J.-M. J. Am. Chem. Soc. 1990, 112, 4060-4061.

⁽⁹⁾ Duplantier, A. J.; Masamune, S. J. Am. Chem. Soc. 1990, 112, 7079-7081.

⁽¹⁰⁾ Pawlak, J.; Sowinski, P.; Borowski, E.; Gariboldi, P. J. Antibiotics 1993, 46, 1598-1603.

^{(11) (}a) Nakata, T.; Noriaki, H.; Nakanishi, K.; Oishi, T. Chem. Pharm. Bull. 1987, 35, 4355-4358. (b) Mori, Y.; Kohchi, Y.; Suzuki, M.; Furukawa, H.; J. Am. Chem. Soc. 1992, 114, 3557-3559. (c) Gonnella, N. C.; Nakanishi, K.; Martin, V. S.; Sharpless, K. B. J. Am. Chem. Soc. 1982, 104, 3775-3776. (d) Zhou, P.; Zhoa, N.; Rele, D. N.; Berova, N.; Nakanishi, K. J. Am. Chem. Soc. 1993, 115, 9313-9314.

⁽¹²⁾ Rychnovsky, S. D.; Skalitzky, D. J. Tetrahedron Lett. 1990, 31, 945-948.

⁽¹³⁾ Rychnovsky, S. D.; Rogers, B.; Yang, G. J. Org. Chem. 1993, 58, 3511-3515.

^{(14) (}a) Schlegel, R.; Fugner, R.; Bradler, G.; Thrum, H. Z. Allg. Mikrobiol. 1971, 11, 661-670. (b) Schlegel, R.; Thrum, H. J. Antibiot. 1971, 24, 360-367. (c) Schlegel, R.; Thrum, H. J. Antibiot. 1971, 24, 368-374.

^{(15) (}a) Schlegel, R.; Grigorjev, P. A.; Thrum, H. Studia Biophysica 1982, 92, 135–140. (b) Grigorjev, P.; Schlegel, R.; Thrum, H.; Ermishkin, L. Biochim. Biophys. Acta 1985, 821, 297–304.

⁽¹⁶⁾ Schlegel, R.; Thrum, H.; Zielinsky, J.; Borowski, E. J. Antibiot. 1981, 34, 122-123.

Figure 1. Structure and absolute configuration of natural roflamycoin.

Figure 2. Polyene macrolide antibiotics related to roflamycoin.

stereoisomers.¹⁷ In the structure determination of roxaticin, Maehr's group found that roxaticin and mycoticin shared the same relative and absolute stereochemical configurations and proposed that roflamycoin had the same stereochemical pattern in the analogous C19—C35 region of the chain (Figure 2).³ Their proposal was based on a presumed common biogenesis, but without access to natural roflamycoin, Maehr's group was unable to test it. We have synthesized a roflamycoin analog based on this stereochemical proposal, but found that the particular isomer we prepared did not have the same stereochemistry as natural roflamycoin.^{18,19} When an authentic sample of roflamycoin became available, we set out to investigate its stereochemistry.²⁰

The Relative Configuration of Roflamycoin. We set out to prepare suitable derivatives of roflamycoin that would shed light on its configuration. When roflamycoin was treated with Dowex 50W-X1 acid resin in MeOH, it formed spiro acetal 2 in good yield (Scheme 1). Treatment of 2 with Ac_2O and DMAP in THF gave the peracetate 4. The ¹H NMR and COSY analysis of 4 revealed several proton signals on the spiro acetal portion of the molecule (Figure 3). Of special note were the axial protons at C14 (1.18 ppm, q, J = 11.6 Hz), C18 (1.11 ppm, t, J = 12.2 Hz), and C20 (1.41 ppm, q, J = 11.7 Hz). Each of these protons showed large coupling constants to the adjacent protons on the six membered spiro acetal rings. To

account for these observed signals, the protons at C13, C15, C19, and C21 must all be axial. Thus the relative (acyclic) configurations at C13/C15 and C19/C21 must be anti.

If spiro acetal 2 was first subjected to acetonide-forming conditions, followed by acetylation, a mixture of compounds containing two acetonides and three acetates was obtained (Scheme 1). Careful separation by reverse-phase HPLC gave a single, pure compound that was identified as 3. The ¹H NMR and COSY information allowed us to unambiguously assign nearly every proton signal. This allowed us to perform ¹H NMR and NOE experiments to determine the relative configuration of the spiro acetal portion. There are only two possible combinations for which both C13/C15 and C19/C21 are anti (Figure 4). The first possibility, A, has a local meso-type symmetry (i.e. a mirror plane) through the keto center at C17. The second possibility, **B**, has a local C_2 -type symmetry through the keto center. If **B** was to form a spiro acetal, it should still exhibit C_2 symmetric properties; however when A forms a spiro acetal, the symmetry is lost. The chemical shifts for the protons at C13 and C21 of compound 3 are 3.40 and 4.32 ppm, respectively. This large difference in chemical shift suggests that the spiro acetal formed is the one derived from A and that the relative configuration at C15/C19 is syn. This was confirmed by the NOESY spectrum of 3, which showed cross peaks between the equatorial C18 proton and the axial protons at C13 and C15.

The ^{13}C NMR spectrum of 3 showed acetonide methyl signals in only the 30 and 19 ppm range. The actual positions of the methyl signals were confirmed by removing the acetonides and then reforming them with $[1,3^{-13}\text{C}_2]$ acetone. The ^{13}C -enriched compound showed methyl doublets at 30.65, 30.48, 19.54, and 19.47 ppm. 21 Thus the relative configurations at C23/C25 and C29/C31 must be syn.

The hemiacetal of roflamycoin could be reduced by treating with NaBH₄ in EtOH to give 17-dihydroroflamycoin as a 5:3 mixture of epimers at C17 as determined by HPLC integration (Scheme 2). The structures were confirmed by treating the epimeric mixture with Ac₂O and DMAP to form two peracetates that were easily separated by HPLC. Mass spectral and ¹H NMR analysis were consistent for both epimers having the expected 10 acetates.²² The dihydroroflamycoin epimers were then separated by reverse-phase HPLC. Each was treated with acetone, 2,2-dimethoxypropane (2,2-DMP), and CSA to give pentaacetonides 5 and 6 (Scheme 2). The ¹³C NMR spectra showed that 5 had two syn acetonides and three anti acetonides. The same analysis of 6 showed the presence of four anti acetonides and only one syn acetonide. The acetonide formed from C17/C19 must be syn in 5 and anti in 6. We know that the C29/C31 relationship is syn from compound 3; therefore the relationships at C13/C15, C21/C23, and C25/C27 must each be anti.

The remaining unknown stereochemical relationships were between C27/C29, C31/C34, and C34/C35. Roflamycoin spiro acetal 2 was silylated and then subjected to ozonolysis followed by treatment with NaBH₄ (Scheme 3). This effectively removed the polyene portion. The resulting diol was silylated, and the lactate ester at C35 was cleaved by treatment with LAH. This gave a compound where the only unprotected hydroxyl group was at C35. The alcohol was mesylated to give 7. Treatment with tetrabutylammonium fluoride (TBAF) removed the silyl protecting groups and was basic enough to promote cyclization. This cyclization should take place with inversion at C35. The

⁽¹⁷⁾ The configuration at C17 is dependent upon the configuration at C13.

⁽¹⁸⁾ Rychnovsky, S. D.; Griesgraber, G.; Kim, J. J. Am. Chem. Soc. 1994, 116, 2621-2622.

⁽¹⁹⁾ For synthetic work directed toward the hypothetical all syn roflamycoin, see: (a) Lipshutz, B. H.; Moretti, R.; Crow, R. *Tetrahedron Lett.* 1989, 30, 15–18. (b) Lipshutz, B. H.; Kotsuki, H.; Lew, W. *Tetrahedron Lett.* 1986, 27, 4825–4828.

⁽²⁰⁾ For a preliminary account, see: Rychnovsky, S. D.; Griesgraber, G.; Schlegel, R. J. Am. Chem. Soc. 1994, 116, 2623-2624.

⁽²¹⁾ The signals are doublets ($J \cong 5$ Hz) due to $^{13}\mathrm{C} - ^{13}\mathrm{C}$ coupling in the isotopically enriched acetonides.

⁽²²⁾ Major isomer: HRMS (FAB) 1161.5768 (M + H), calcd for $C_{69}H_{89}O_{22}$ 1161.5820. Minor isomer: HRMS (FAB) 1161.5854 (M + H), calcd for $C_{60}H_{89}O_{22}$ 1161.5820.

C or O
$$\frac{15}{4}$$
 $\frac{13}{15}$ $\frac{13}{15}$

 C_{20} H_{ax}: 1.41 ppm, q, J = 11.7 Hz 19/21 anti Figure 3. Coupling constants and relative configurations of the spiro acetal portion of 4.

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OAC

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Figure 4. Relative configuration of the spiro acetal portion of 3.

free alcohols were then acetylated to give 8. The ¹H NMR and COSY spectra of 8 revealed that the proton at C35 was a doublet of doublets at 3.18 ppm with J = 3.9 and 8.0 Hz. Decoupling revealed that $J_{34-35} = 8.0$ Hz, and $J_{35-36} = 3.9$ Hz.²³

There are four possible diastereomeric relationships for the tetrahydropyran portion of compound 8. Knowing the coupling constants for the C35 proton, we turned to computer modeling to help us determine the configuration of the tetrahydropyran ring. A complete conformational search²⁴ was carried out on each of the four possible tetrahydropyran diastereomers (10ad), and the predicted coupling constants for each conformer within 10 kJ/mol of the minimum were weighted according to a Boltzmann distribution at 25 °C (Figure 5). Only the predicted coupling constants for stereoisomer 10d are consistent with the observed coupling found in 8. Because the cyclization occurred with inversion at C35, the relative configurations at C31/C34 and C34/C35 are both syn.

To find the final stereochemical relationship, C27/C29, we prepared compound 9 from mesylate 7 (Scheme 3). The tetrahydropyran ring was formed as before by treating 7 with TBAF. Subsequent treatment with acetone, 2,2-DMP, and PPTS introduced two acetonides. The crude reaction mixture was then benzoylated to give 9 as a UV active compound that was purified by reverse-phase HPLC. The ¹H NMR showed that we had formed a tribenzoate with two acetonides, but due to limited material, we did not have enough sample for the important ¹³C NMR analysis. This problem was solved by removing the acetonides (Dowex 50W-X1, MeOH) and reforming them with acetone that was isotopically enriched with 33% [1,3-13C₂]acetone. The ¹³C NMR of isotopically enriched 9 was easily obtained and revealed methyl signals at 30.74 (overlap), 20.22, and 20.04 ppm.²¹ These signals indicate that both acetonides are syn and that the final unassigned relationship, C27/C29, is syn.

We now have all the information necessary to determine the relative stereochemistry of roflamycoin. We know from spiro acetals 3 and 4 that the relationships C13/C15 and C19/C21 are anti and C15/C19 is syn. In addition, the acetonide information obtained from 3 tells us that the relationships C23/ C25 and C29/C31 are syn. The pentaacetonides 5 and 6 revealed that the relationships C21/C23 and C25/C27 are both anti. The relative stereochemistry for C31, C34, and C35 was determined by ¹H NMR analysis of the tetrahydropyran portion of 8, and the final relationship, C27/C29, was shown to be syn by ¹³C NMR analysis of 9. The relative configuration of natural roflamycoin is illustrated in Figure 6. It is epimeric with the structure proposed by Maehr at C21, C25, and C27.

To strengthen this conclusion, pentaacetonide 12 was prepared from 5 as described in Scheme 4. Compound 5 was subjected to ozonolysis, silylation, LAH reduction, and mesylation to give the expected C35 mesylate 11. The acetonides were removed by treatment with acidic Dowex in MeOH, and cyclization was induced by treatment with TBAF. Now that the C31 hydroxyl is tied up in the tetrahydropyran ring, reprotection directs the

⁽²³⁾ The protons of interest were identified from the COSY spectrum. The C36-H (δ 1.88 ppm) has COSY off-diagonal peaks with the two methyl groups (δ 1.15 ppm, d, J=6.9 Hz; δ 1.00 ppm, d, J=6.9 Hz) attached to C36. The C34-H (δ 1.42 ppm) has an off-diagonal peak with the methyl group (δ 0.76 ppm, d, J = 6.7 Hz) attached to C34. The C35-H (δ 3.19 ppm, dd, J = 3.9, 8.0 Hz) has an off-diagonal peak with C34-H, but a weak off-diagonal peak with C36-H. Our assignments were confirmed by the decoupling experiment. When we decoupled at 1.88 ppm, the signals at 1.15 and 1.00 ppm collapsed to singlets, and the signal at 3.19 ppm collapsed to a doublet (J = 8.0 Hz). When we decoupled at 1.42 ppm, the signal at 0.76 ppm collapsed to a singlet, and the signal at 3.19 ppm collapsed to a doublet (J = 3.9 Hz).

⁽²⁴⁾ A Monte Carlo search with 500 initial structures using the MM2 force field was carried out using Macromodel 3.5 (Mohamadi, F.; Richards, N. G. J.; Guida, W. C.; Liskamp, R.; Lipton, M.; Caufield, C.; Chang, G.; Hendrickson, T.; Still, W. C. J. Comput. Chem. 1990, 11, 440-467).

Scheme 3

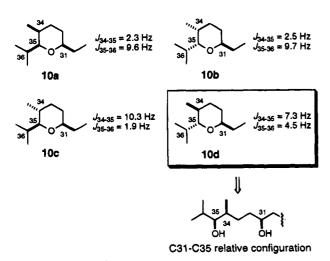


Figure 5. Predicted ¹H NMR coupling constants based on molecular modeling.

Figure 6. Relative configuration of natural roflamycoin.

acetonide protecting groups one oxygen to the right with respect to their position in 5 and thus shifts the "reading frame" for the ¹³C acetonide analysis. ¹² The crude polyol was reacted with [1,3-¹³C₂]acetone and CSA to give pentaacetonide 12 that showed the presence of one anti and four syn acetonides by ¹³C NMR analysis. ²¹ This result was consistent with the configurational assignment in Figure 6.

Determination of the Absolute Configuration of Roflamycoin. Only the absolute stereochemistry of roflamycoin remained to be determined. We used the advanced Mosher ester technique refined by Kakisawa.²⁵ The secondary alcohol at C35 is best suited for the Mosher analysis because the three methyl groups and the protons at C34 and C36 are easily identified by 1 H NMR. To liberate the alcohol at C35, pentaacetonide 6 was ozonized and treated with NaBH₄ to give a diol (Scheme 5). Both of the (R)- and (S)-MTPA esters, 13R and 13S, were prepared. The protons of interest were identified by 1 H NMR and COSY analysis, and the $\Delta\delta_{\rm H}$ values were measured and are listed in Table 1. The configuration at C35 is S based on this analysis.

The magnitude of the $\Delta\delta_{\rm H}$ values were quite small, which led us to question the reliability of the advanced Mosher's method with this type of hindered secondary alcohol. To test its validity, we repeated the advanced Mosher's analysis on synthetic material of known configuration. Compound 14 was an intermediate in our synthesis of a roflamycoin analogue, 18 and the stereochemistry at C35 was known to be S. We prepared both MTPA esters of compound 14, followed by removal of the TBS protecting group, which obscured important proton signals (Scheme 5). The measured $\Delta\delta_{\rm H}$ values for 15R and 15S were in very close agreement to those found for the material derived from natural roflamycoin, lending support to the prediction that C35 has an S configuration. The measured

^{(25) (}a) Kusumi, T.; Ohtani, I.; Inouye, Y.; Kakisawa, H. Tetrahedron Lett. 1988, 29, 4731-4734. (b) Ohtani, I.; Kusumi, T.; Ishitsuka, O. M.; Kakisawa, H. Tetrahedron Lett. 1989, 30, 3147-3150. (c) Ohtani, I.; Kusumi, T.; Kashman, Y.; Kakisawa, H. J. Am. Chem. Soc. 1991, 113, 4092-4096.

Scheme 5

Table 1. Comparison of the $\Delta\delta_{\rm H}$ (= $\delta_{\rm S}$ - $\delta_{\rm R}$) Values for Compounds 13 and 15

| - | 13(R) δ (ppm) | 13(S) δ (ppm) | $\Delta \delta_{\rm H}$ (Hz) | 15(R) δ (ppm) | 15(S) δ (ppm) | $\Delta \delta_{\rm H} \ ({ m Hz})$ |
|-----------|------------------|------------------|------------------------------|------------------|------------------|-------------------------------------|
| C34-H | 1.742 | 1.770 | +14 | 1.726 | 1.743 | +8.5 |
| C34-Me | 0.794 | 0.831 | +18.5 | 0.791 | 0.824 | +16.5 |
| C35-H | 4.865 | 4.871 | +3 | 4.863 | 4.868 | +2.5 |
| C36-H | 1.945 | 1.928 | -8.5 | 1.948 | 1.926 | -11 |
| C36-Me(1) | 0.867 | 0.852 | -7.5 | 0.868 | 0.849 | -9.5 |
| C36-Me(2) | 0.819 | 0.787 | -16 | 0.826 | 0.786 | -20 |

 $\Delta\delta_H$ values for the protons of interest are summarized in Table 1. Taken together, the data and analysis presented here are only consistent with roflamycoin having the structure shown in Figure 1.

Synthesis and Correlation of Degradation Fragment 12. We were interested in pursuing a synthesis of natural roflamycoin, but before attempting a revised total synthesis, we considered it prudent to confirm the predicted stereochemical assignment by preparing a less complicated degradation piece. Degradation product 12, vide infra, was chosen as a synthetic target because it contains all of the independent stereogenic centers of natural roflamycoin. Our methods for the rapid construction of polyol chains allowed us to prepare this molecule in an efficient manner from optically active precursors that we had on hand

Our synthetic approach to 12 was based on the cyanohydrin acetonide approach for the convergent synthesis of polyol chains.²⁶ It appeared that 12 could be constructed from four relatively simple optically active precursors (Figure 7). Our strategy involved the coupling of these pieces using standard

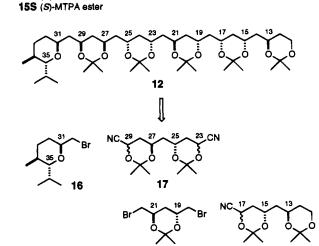


Figure 7. Synthetic analysis of the degradation fragment 12.

alkylation chemistry followed by a stereoselective reductive decyanation to give pentaacetonide 12. We have already shown that dibromide 18 is an effective alkylating agent for the construction of polyol chains and its synthesis has been described.²⁷ This left us with the task of synthesizing bromide 16, cyanohydrin acetonide 19, and the C_2 symmetric dinitrile 17. The preparation of these optically active synthons and the synthesis of the roflamycoin degradation product 12 are described below.

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Synthesis of the Pieces. Our strategy for the synthesis of 16 involved the coupling of two optically active fragments, 21 and 22, followed by an intramolecular cyclization to form the tetrahydropyran ring (Scheme 6). Starting with (2R,3R)-2,4-dimethyl-1,3-pentanediol,²⁸ 20, we could selectively convert the primary alcohol to a phenyl thioether by treatment with tributylphosphine and diphenyl disulfide.²⁹ The resulting alcohol was protected as its benzyl ether by treatment with KH

^{(26) (}a) Rychnovsky, S. D.; Zeller, S.; Skalitzky, D. J.; Griesgraber, G. J. Org. Chem. 1990, 55, 5550-5551. (b) Rychnovsky, S. D.; Griesgraber, G. J. Org. Chem. 1992, 57, 1559-1563.

⁽²⁷⁾ Rychnovsky, S. D.; Griesgraber, G.; Zeller, S.; Skalitzky, D. J. J. Org. Chem. 1991, 56, 5161-5169.

⁽²⁸⁾ Hoye, R. C.; Rychnovsky, S. D. J. Am. Chem. Soc. **1994**, 116, 1753–1765.

⁽²⁹⁾ Nakagawa, I.; Hata, T. Tetrahedron Lett. 1975, 1409–1412.

Scheme 7

and benzyl bromide, and the thiol was then oxidized with Oxone to give sulfone 21 in 80% overall yield. Deprotonation of the sulfone with butyllithium in THF at 0 °C followed by the addition of DMPU and (R)-(2,2-dimethyl-1,3-dioxolan-4-yl)methyl bromide (22)³⁰ gave the coupled product 23 in 53% yield. The sulfone was removed by reduction with sodium amalgam, followed by removal of the benzyl protecting group by hydrogenolysis. Mesylation of the resulting C35 alcohol gave the mesylate 24 in 80% yield from 23. Removal of the acetonide was accomplished by treatment with Dowex 50W-X1 acid resin in MeOH to give a diol which was treated with NaH in THF to give the cyclized product, tetrahydropyranylmethanol 25, in 79% yield for the two steps. The free alcohol was converted to its mesylate, followed by displacement with tetrabutylammonium bromide, to give 16 in 80% yield for the two steps. The synthesis of 16 was completed in a straightforward manner from diol 20 in 22% overall yield.

The C_2 symmetric dinitrile 17 appeared to be a useful synthon for the synthesis of alternating polyol chains. Sequential alkylation of 17 with different electrophiles, followed by reductive decyanation, would introduce four new stereogenic centers into a chain. Dinitrile 17 was prepared from the optically active diepoxide 26 (Scheme 7).26a The inexpensive alkyne 1-hexyne was chosen as the nucleophile because it could be easily converted to an aldehyde by the two-step process of hydrogenation and ozonolysis. Deprotonation of 1-hexyne with butyllithium, followed by addition to the diepoxide 26 with BF₃·OEt₂ catalysis, gave the expected diol in excellent yield.³¹ The diol was protected as an acetonide to give the C_2 symmetric dialkyne 27 in 80% overall yield. Acetonide 27 was then hydrogenated with 5% Pd/BaSO₄ and quinoline to give the Z,Zdiene, which was cleaved by treatment with ozone to give the unstable dialdehyde 28 in 88% yield over the two steps.

Scheme 8

Dialdehyde 28 was immediately treated with 2 equiv of TMSCN and catalytic KCN/18-crown-6 complex followed by acetone, 2,2-dimethoxypropane and CSA to give nitriles 17 as a 1:2:1 mixture of isomers in 91% yield. This mixture of isomers was of no consequence as the cyanohydrin stereogenic centers are set during a subsequent reductive decyanation step. The transformation of diepoxide 26 into diacetonide 17 in 64% overall yield is one example of the potentially important conversion of an epoxide into a chain extended cyanohydrin acetonide.

The final fragment needed for the preparation of pentaacetonide 12 was prepared from hydroxy ester 29 by a procedure similar to that used to synthesize a piece of roxaticin.²⁸ Hydroxy ester 29 was prepared in optically active form by a Noyori enantioselective hydrogenation of the corresponding β -keto ester.32,33 Reduction of 29 with LAH followed by treatment with acetone, 2,2-dimethoxypropane, and catalytic CSA gave acetonide 30 in 81% yield (Scheme 8). The next steps were carried out carefully to avoid possible epimerization that could occur by migration of the acetonide group. Hydrogenolysis of the benzyl ether was accomplished with Pearlman's catalyst in the presence of H₂. The crude alcohol was immediately oxidized with Dess-Martin reagent to give an aldehyde which was treated with Brown's (+)-Ipc₂B(allyl) reagent³⁴ to give 31 as a single homoallylic alcohol in 44% overall yield for the three-step sequence. Silvlation of the free hydroxyl group and stepwise oxidation of the terminal alkene (OsO₄, NMO, NaIO₄) gave the intermediate aldehyde. The aldehyde was treated with TMSCN and catalytic KCN/18-crown-6 followed by addition of acetone, 2,2-dimethoxypropane and CSA to give cyanohydrin acetonide 19 in 77% yield from alcohol 31.

Preparation of Pentaacetonide 12. The four fragments were assembled to give pentaacetonide **12** using our alkylation and reductive decyanation protocol (Scheme 9). Dinitrile **17** has effective C_2 symmetry, so only one monoalkylation product is possible in the coupling to give the left-hand fragment **32**. Overalkylation could have been a problem, but it was avoided by using the monoanion of **17** in excess. Dinitrile **17** (1.9 equiv) was deprotonated with LiNEt₂ (1.1 equiv) in THF at -78 °C followed by addition of 1.0 equiv of bromide **16**. The reaction vessel was then transferred to a -25 °C MeOH/ice bath, and the reaction mixture was allowed to slowly warm to 15 °C overnight. The monoalkylated product was isolated in 40% yield, and 28% of the unreacted bromide was also recovered. The alkylation of **17** proceeds in only moderate yield, but it is the linchpin for this convergent synthesis and dramatically

⁽³⁰⁾ Prepared from 1,2:5,6-di-O-isopropylidene-D-mannitol by the following sequence: (i) Pb(OAc)₄; (ii) NaBH₄; (iii) TsCl, Pyr; (iv) Bu₄NBr, Δ. Kawakami, Y.; Asai, T.; Umeyama, K.; Yamashita, Y. J. Org. Chem. 1982, 47, 3585-3587.

⁽³¹⁾ Yamaguchi, M.; Hirao, I. Tetrahedron Lett. 1983, 24, 391-394.

⁽³²⁾ We thank Dr. Daniel E. Mickus for preparing hydroxy ester 29. (33) (a) Kitamura, M.; Ohkuma, T.; Inoue, S.; Sayo, N.; Kumobayashi, H.; Akutagawa, S.; Ohta, T.; Takaya, H.; Noyori, R. J. Am. Chem. Soc. 1988, 110, 629-631. (b) Noyori, R.; Ohkuma, T.; Kitamura, M.; Takaya, H.; Sayo, N.; Kumobayashi, H.; Akutagawa, S. J. Am. Chem. Soc. 1987, 109, 5856-5858. (c) Noyori, R.; Takaya, H. Acc. Chem. Res. 1990, 23, 345-350.

^{(34) (}a) Brown, H. C.; Jadhav, P. K. J. Am. Chem. Soc. 1983, 105, 2092—2093. (b) Jadhav, P. K.; Bhat, K. S.; Perumal, P. T.; Brown, H. C. J. Org. Chem. 1986, 51, 432—439.

simplifies the route. We will continue to investigate dinitrile 17 as a polyol synthon.

For the preparation of 33, dibromide 18 is now the C_2 symmetric piece, and again only one monoalkylation product is possible. To avoid overalkylation, an excess of 18 was used in the alkylation. Cyanohydrin acetonide 19 (1.0 equiv) was deprotonated with LiNEt₂ (1.2 equiv) in THF at -78 °C followed by addition of 2.5 equiv of dibromide 18. The reaction vessel was then transferred to a -23 °C MeOH/ice bath, and the reaction mixture was allowed to slowly warm to 15 °C overnight. The monoalkylated product was isolated in 62% yield, and 68% of the unreacted dibromide was recovered.

Fragments 32 and 33 were then assembled to give the complete carbon skeleton of our target molecule. Deprotonation of 32 (1.0 equiv) and alkylation with 33 (1.3 equiv) was carried out as described for the first two coupling reactions to give adduct 34 in 81% yield. Reductive decvanation of 34 with lithium in ammonia cleanly gave pentaacetonide 12 in 81% yield.

Comparison of Synthetically and Naturally Derived Pentaacetonide 12. The synthetic pentaacetonide 12 was identical to the compound obtained by the degradation of natural roflamycoin by ¹H NMR. ¹³C NMR. HRMS. and TLC.³⁵ Especially convincing was the comparison of ¹H NMR spectra shown in Figure 8. The two spectra were identical except for a few small peaks observed at approximately 1.7 ppm in the spectrum of the material derived from natural roflamycoin. These peaks are the result of using 20% enriched [1,3-13C₂]acetone in the preparation of pentaacetonide 12. The extra signals in the NMR are due to ¹³C-¹H coupling. comparison of our synthetic material with that obtained from the natural product confirms the configurational assignment of roflamycoin (1) based on our ¹³C acetonide analysis. Direct comparison of natural and synthetic samples can provide a valuable check on NMR based structural assignments.

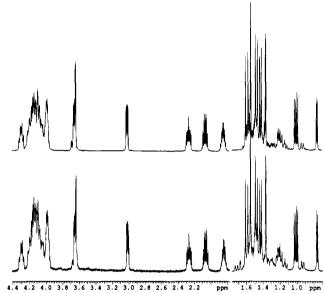


Figure 8. The ¹H NMR spectrum of synthetic 12 (top) and the roflamycoin degradation product 12 (bottom). Note: the 4.40-1.77 ppm region is shown at $5 \times$ the intensity of the 1.77-0.70 ppm region.

Experimental Section

General Experimental Details. Liquid chromatography was performed using forced flow (flash chromatography) of the indicated solvent system on EM reagent silica gel 60 (230-400 mesh).36 HPLC purifications were performed on a reverse-phase column (Spherisorb S5 ODS2, 25-cm \times 10-mm) eluting with H₂O/MeOH mixtures. Commercial CH2Cl2 was distilled from CaH2 under N2. Air and/or moisture sensitive reactions were carried out under N2 or Ar using flame-dried glassware and standard syringe/septa techniques. NMR data for ¹³C DEPT experiments are reported as quaternary (C), tertiary (CH), secondary (CH₂), and primary (CH₃) carbon atoms. For overlapping signals, the number of carbon atoms are given in

Degradation and NMR Analysis of Roflamycoin. Roflamycoin (1); ¹H NMR (500 MHz, CD₃OD) δ 7.175 (dd, J = 1.2, 11.1 Hz, 1

⁽³⁵⁾ The optical rotation of synthetic 12 is $[\alpha]^{27}_D = +20.5^{\circ}$ (c 0.85, CHCl₃). The optical rotation of naturally derived 12 was not measured due to insufficient material.

⁽³⁶⁾ Still, W. C.; Kahn, M.; Mitra, A. J. Org. Chem. 1978, 43, 2923-2925.

H), 6.577 (dd, J=10.3, 14.6 Hz, 1 H), 6.509 (dd, J=11.3, 14.6 Hz, 1 H), 6.403 (dd, J=10.6, 14.7 Hz, 1 H), 6.324 (dd, J=10.3, 14.7 Hz, 1 H), 6.284 (dd, J=10.6, 14.7 Hz, 1 H), 6.173 (dd, J=10.6, 14.7 Hz, 1 H), 5.746 (ddd, J=5.5, 8.7, 15.0 Hz, 1 H), 4.728 (dd, J=2.3, 9.9 Hz, 1 H), 4.219 (m, 1 H), 4.00-3.85 (m, 7 H), 3.528 (m, 1 H), 2.311 (dt, J=2.2, 15.2 Hz, 1 H), 2.130 (dt, J=9.3, 15.1 Hz, 1 H), 1.896 (s, 3 H), 1.88-1.81 (m, 3 H), 1.72 (m, 1 H), 1.704 (dd, J=7.8, 14.0 Hz, 1 H), 1.63-1.57 (m, 2 H), 1.50-1.24 (m, 13 H), 1.150 (t, J=11.7 Hz, 1 H), 1.143 (m, 1 H), 1.048 (q, J=11.8 Hz, 1 H), 1.020 (m, 1 H), 0.881 (d, J=6.7 Hz, 3 H), 0.849 (d, J=6.7 Hz, 3 H), 0.795 (d, J=6.6 Hz, 3 H); HRMS (FAB) 739.4611 (M + H), calcd for C₄₀H₆₇O₁₂ 739.4634.

Roflamycoin Spiro Acetal (2). Roflamycoin (100 mg) was dissolved in 20 mL of MeOH and treated with Dowex 50W-X1 acid resin. The reaction was kept in the dark and stirred for 20 h under Ar. The reaction was then filtered and concentrated under reduced pressure. Chromatography (SiO₂, 15%-20% MeOH/ethyl acetate) gave the product (58 mg) as a yellow solid: ¹H NMR (500 MHz, CD₃OD) δ 7.17 (dd, J = 1.3, 10.9 Hz, 1 H), 6.58 (dd, J = 9.4, 14.7 Hz, 1 H), 6.52 (dd, J = 10.9, 14.4 Hz, 1 H), 6.41 - 6.38 (m, 2 H), 6.29 (dd, J =10.0, 14.7 Hz, 1 H), 6.22 (dd, J = 9.4, 14.7 Hz, 1 H), 6.16 (dd, J =10.0, 14.7 Hz, 1 H), 5.68 (ddd, J = 5.9, 9.1, 14.7 Hz, 1 H), 4.70 (dd, J = 2.2, 9.7 Hz, 1 H, 3.98 (m, 1 H), 3.91 - 3.78 (m, 4 H), 3.74 (m, 1 H)H), 3.63 (m, 1 H), 3.61 (m, 1 H), 3.47 (m, 1 H), 2.35 (m, 1 H), 2.31 (dd, J = 4.0, 13.1 Hz, 1 H), 2.10 (dt, J = 14.1, 9.0 Hz, 1 H), 1.90 (d, J = 14.1, 9.0 Hz, 1 H)J = 1.3 Hz, 3 H), 1.88 (m, 1 H), 1.87 (dd, J = 3.4, 13.1 Hz, 1 H), 1.82 (m, 1 H), 1.79 - 1.75 (m, 2 H), 1.67 (ddd, J = 5.9, 7.8, 13.4 Hz, 1 H),1.54 (ddd, J = 5.3, 8.1, 13.4 Hz, 1 H), 1.47 (m, 1 H), 1.38–1.17 (m, 11 H), 1.12-0.98 (m, 4 H), 0.90 (d, J = 6.9 Hz, 3 H), 0.86 (d, J = 6.6Hz, 3 H), 0.82 (d, J = 6.6 Hz, 3 H); HRMS (FAB) 721.4540 (M + H), calcd for $C_{40}H_{65}O_{11}$ 721.4528.

15,19,23,25,27,29,31-Hepta-O-acetylroflamycoin Spiro Acetal (4). A suspension of roflamycoin spiro acetal 2 (2 mg, 0.3 μ mol, 1 equiv) in 1.5 mL of THF was treated with DMAP (24 mg, 0.20 mmol, 67 equiv) and Ac₂O (18 μL, 0.19 mmol, 63 equiv). The reaction was kept in the dark, under Ar. After 18 h, the reaction was quenched by adding a drop of MeOH. The reaction mixture was then diluted in 20 mL of ethyl acetate and washed with 0.05 M H₂SO₄ (2 × 5 mL) and brine (5 mL). The organic layer was dried with Na₂SO₄ and concentrated under reduced pressure. The crude yellow solid was passed through a SiO₂ column eluting with 30-50% ethyl acetate/ hexanes and then purified by reverse-phase HPLC (12% H₂O/MeOH) to give 4 (1.5 mg) as a yellow solid: ^{1}H NMR (500 MHz, $C_{6}D_{6}$) δ 7.60 (d, J = 9.4 Hz, 1 H), 6.52 (dd, J = 11.2, 14.6 Hz, 1 H), 6.50-6.40 (m, 2 H), 6.33-6.25 (m, 3 H), 6.23 (dd, J = 10.9, 14.9 Hz, 1 H),5.85 (ddd, J = 5.7, 9.5, 14.6 Hz, 1 H), 5.21 - 5.13 (m, 4 H), 5.05 (m,1 H), 5.02 (m, 1 H), 4.97 (m, 1 H), 4.95 (m, 1 H), 4.24 (m, 1 H), 3.21 (m, 1 H), 2.35 (dd, J = 3.4, 12.3 Hz, 1 H), 2.09 (dd, J = 4.9, 12.3 Hz, 1 H), 2.042 (s, 3 H), 1.904 (s, 3 H), 1.846 (s, 3 H), 1.833 (s, 3 H), 1.785 (s, 3 H), 1.744 (s, 3 H), 1.689 (s, 3 H), 1.645 (s, 3 H), 2.02-1.52 (m, 18 H), 1.41 (q, J = 11.7 Hz, 1 H), 1.35 - 1.25 (m, 3 H), 1.18(q, J = 11.7 Hz, 1 H), 1.11 (t, J = 12.2 Hz, 1 H), 0.94 (d, J = 6.6 Hz, 1 Hz)3 H), 0.84 (d, J = 6.9 Hz, 3 H), 0.73 (d, J = 6.9 Hz, 3 H); HRMS (FAB) 1015.5259 (M + H), calcd for $C_{54}H_{79}O_{18}$ 1015.5267.

15,19,27-Tri-O-acetyl-23,25:29,31-bis-O-(1-methylethylidene)-roflamycoin Spiro Acetal (3). A solution of roflamycoin spiro acetal 2 (22.0 mg) dissolved in 4 mL of a 4:1 mixture of acetone and 2,2-DMP was treated with 2 mg of CSA. The reaction was stirred under N_2 , in the dark, for 1 h. The reaction was then quenched by addition of 5 μ L of Et₃N and concentrated under a stream of N_2 . Chromatography (SiO₂, 50% acetone/hexanes) gave 16.7 mg of a yellow solid.

The resulting yellow solid was dissolved in 5 mL of THF and treated with DMAP (30.2 mg, 0.248 mmol) and Ac₂O (21 μ L, 0.22 mmol). The reaction was kept under N₂, in the dark, for 20 h. The reaction was then quenched by addition of 3 drops of MeOH. The reaction mixture was then diluted in 20 mL of ethyl acetate and washed with H₂O (5 mL), 0.05 M H₂SO₄ (2 × 5 mL), and brine (5 mL). The organic layer was dried with Na₂SO₄ and concentrated under reduced pressure. The crude yellow solid was passed through a SiO₂ column eluting with 40% ethyl acetate/hexanes and then purified by reverse-phase HPLC (4% H₂O/MeOH) to give 3 (5.4 mg) as a yellow glass: ¹H NMR (500 MHz, C₆D₆) δ 7.51 (d, J = 11.7 Hz, 1 H), 6.29 (dd, J = 12.0, 14.4 Hz, 1 H), 6.24 (m, 1 H), 6.17–6.06 (m, 5 H), 5.79 (ddd, J = 5.0, 10.0,

14.9 Hz, 1 H), 5.67 (m, 1 H), 5.07 (m, 1 H), 5.06 (dd, J=2.9, 9.5 Hz, 1 H), 5.00 (m, 1 H), 4.32 (m, 1 H), 3.95 (m, 1 H), 3.88 (m, 1 H), 3.79 (m, 1 H), 3.59 (m, 1 H), 3.40 (m, 1 H), 2.51 (dd, J=2.9, 12.9 Hz, 1 H), 2.11 (dd, J=4.4, 12.9 Hz, 1 H), 2.10 (m, 1 H), 2.05 (m, 1 H), 1.99 (m, 1 H), 1.946 (s, 3 H), 1.926 (s, 3 H), 1.749 (s, 3 H), 1.638 (s, 3 H), 1.522 (s, 3 H), 1.514 (s, 3 H), 1.467 (s, 3 H), 1.443 (s, 3 H), 1.96-1.74 (m, 6 H), 1.70 (t, J=11.9 Hz, 1 H), 1.66-1.41 (m, 5 H), 1.31 (q, J=11.8 Hz, 1 H), 1.21-0.98 (m, 6 H), 1.16 (q, J=11.6 Hz, 1 H), 1.13 (t, J=12.2 Hz, 1 H), 1.02 (d, J=6.4 Hz, 3 H), 0.91 (d, J=6.7 Hz, 3 H), 0.72 (d, J=6.7 Hz, 3 H); HRMS (FAB) 926.5369 calcd for $C_{52}H_{78}O_{14}$ 926.5393.

A portion of 3 (4 mg) was treated with Dowex 50W-X1 resin in methanol to remove the acetonides. The resulting material was treated with $[1,3^{-13}C_2]$ acetone (10% in toluene) and CSA. After stirring for 18 h in the dark, the reaction mixture was filtered through a SiO₂ plug eluting with 40% ethyl acetate/hexanes. Purification by reverse-phase HPLC (5% $\rm H_2O/MeOH$) gave 1.5 mg of 3 with isotopically enriched acetonides: ^{13}C NMR (75 MHz, $\rm C_6D_6$) δ 30.66 (d, J=4.7 Hz), 30.48 (d, J=4.8 Hz), 19.54 (d, J=5.9 Hz), 19.47 (d, J=5.0 Hz).

16,17-Dihydro-13,15:17,19:21,23:25,27:29,31-pentakis-O-(1-methylethylidene)roflamycoin (5). A solution of roflamycoin (50 mg, 0.068 mmol) dissolved in 3 mL of 50% aqueous EtOH was treated with NaBH₄ (10 mg, 0.26 mmol, 4 equiv) and stirred under N₂, in the dark, for 16 h. The reaction was then quenched with 15 mL of pH 7 phosphate buffer solution. The reaction mixture was then extracted with ethyl acetate (3 \times 15 mL). The combined organic layers were washed with brine, dried (Na₂SO₄), and concentrated under reduced pressure. Chromatography (SiO₂, 30% MeOH/ethyl acetate) gave 42 mg of an epimeric mixture of reduced roflamycoin. Separation of these epimers was achieved by reverse-phase HPLC (25% H₂O/MeOH) to give 18.4 mg of the major isomer and 12.9 mg of the minor isomer, both as yellow solids.

The major isomer (14.7 mg) was dissolved in 4 mL of acetone and 0.5 mL of 2,2-DMP and treated with 3 mg of CSA. The reaction was heated to 50 °C and stirred in the dark, under Ar, for 16 h. The reaction appeared to reach an equilibrium at this point as indicated by TLC. The reaction was quenched with 10 µL of Et₃N and concentrated under a stream of Ar. Chromatography (SiO2, 50-100% ethyl acetate/ hexanes) gave 3.6 mg of the desired pentaacetonide and 12.7 mg of more polar compounds. The more polar material was resubjected to the reaction conditions to give another 10.4 mg of desired material and 4 mg of polar material. Again, the more polar material was resubjected to the reaction conditions. The desired material from all of the reactions was combined for a final purification by flash chromatography (SiO₂, 25% ethyl acetate/hexanes) to give 5 (17.0 mg) as a yellow solid: ¹H NMR (C₆D₆, 300 MHz) δ 7.64 (d, J = 11.1 Hz, 1 H), 6.37-6.04 (m, 7 H), 5.91 (m, 1 H), 5.10 (dd, J = 2.1, 9.3 Hz, 1 H), 4.32-3.82 (m, 9 H), 3.69 (m, 1 H), 2.50 (m, 1 H), 2.15-2.00 (m, 3 H), 1.998 (s, 3 H), 1.90-1.10 (m, 22 H), 1.556 (s, 3 H), 1.535 (s, 3 H), 1.420 (s, 12 H), 1.398 (s, 3 H), 1.384 (s, 3 H), 1.350 (s, 3 H), 1.342 (s, 3 H), 0.99 (d, J = 6.1 Hz, 3 H), 0.97 (d, J = 6.0 Hz, 3 H), 0.74 (d, J = 6.7 Hz, 3 H); ¹³C NMR (C₆D₆, 75 MHz, DEPT) C 168.0, 100.3 (2), 100.1, 98.6, 98.5; CH 140.1, 139.2, 136.8, 135.5, 134.1, 132.3, 131.6, 131.3, 127.6, 80.3, 69.4, 66.3, 65.9, 65.7, 65.6, 63.9, 63.5, 63.4, 63.3, 62.9, 34.9, 30.1; CH₂ 43.2, 43.1, 42.8, 42.4, 39.2, 39.1, 38.4, 37.9, 37.5, 36.1, 34.6, 29.8; CH₃ 30.7, 30.6, 26.3, 25.4, 25.1 (3), 24.9, 19.9, 19.9, 19.8, 19.0, 14.2, 13.1; HRMS (FAB) 925.6064 (M -CH₃), calcd for C₅₄H₈₅O₁₂ 925.6044.

16,17-Dihydro-13,15:17,19:21,23:25,27:29,31-pentakis-*O*-(1-methylethylidene)roflamycoin (6). Compound 6 was prepared from the minor isomer of reduced roflamycoin (12.9 mg) using the procedure described above for the preparation of **5**. We obtained 10.6 mg of 6 as a yellow solid: 1 H NMR (C_6D_6 , 300 MHz) δ 7.64 (d, J = 11.7 Hz, 1 H), 6.36 (dd, J = 11.3, 14.3 Hz, 1 H), 6.27–5.88 (m, 7 H), 5.12 (dd, J = 2.1, 9.2 Hz, 1 H), 4.25–3.90 (m, 9 H), 3.77 (m, 1 H), 2.56 (m, 1 H), 2.13–1.87 (m, 8 H), 2.022 (s, 3 H), 1.76–1.20 (m, 17 H), 1.570 (s, 3 H), 1.465 (s, 3 H), 1.415 (s, 6 H), 1.406 (s, 6 H), 1.380 (s, 6 H), 1.359 (s, 6 H), 0.98 (d, J = 6.4 Hz, 3 H), 0.97 (d, J = 6.6 Hz, 3 H), 0.72 (d, J = 6.6 Hz, 3 H); 13 C NMR (C_6D_6 , 75 MHz, signals for acetonide methyls) 30.66, 26.43, 25.36, 25.30, 25.14 (2), 25.08 (2), 25.00, 19.96; HRMS (FAB) 925.5988 (M – CH₃), calcd for $C_{54}H_{85}O_{12}$ 925.6044.

[2S,2(2S,4S,6S,8S,10S,13S,14S),4R,6R,8R,10S]-4,10-Bis((1,1-dimethylethyl)dimethylsiloxy)-8-(2-((1,1-dimethylethyl)dimethylsiloxy)-ethyl)-2-(13,15-dimethyl-2,4,6,8,10,14-hexakis((1,1-dimethylethyl)dimethylsiloxy)-14-hydroxyhexadecanyl)-1,7-dioxaspiro-[5.5]undecane (7 [Ms = H]). A solution of roflamycoin spiro acetal 2 (38.2 mg, 0.053 mmol) was dissolved in 5 mL of CH₂Cl₂ and treated with 2,6-lutidine (109 μ L, 0.532 mmol, 10 equiv) and TBSOTf (62 μ L, 0.475 mmol, 9 equiv) at 0 °C, under N₂. The reaction was allowed to warm to room temperature and stirred in the dark for 16 h. The reaction was quenched by addition of 3 mL of saturated NaHCO₃ solution. The reaction mixture was extracted into CH₂Cl₂ (20 mL) and the organic layer was washed with H₂O, followed by brine. The organic layer was then dried with Na₂SO₄ and concentrated under reduced pressure. Chromatography (SiO₂, 3% ethyl acetate/hexanes) gave 58.4 mg of a yellow solid.

The yellow solid was dissolved in 9 mL of a 2:1 mixture of MeOH and CH_2Cl_2 and cooled to -78 °C. Ozone was bubbled through the solution until a blue color persisted. Nitrogen was then bubbled through the solution until it was colorless, then NaBH₄ (32 mg) was added and the reaction mixture was allowed to slowly warm to room temperature. After 1 h, the reaction was diluted with Et₂O (30 mL) and quenched with 3 mL of saturated NaHCO₃ solution. The organic portion was decanted and the aqueous portion was washed with Et₂O (2 × 10 mL). The combined organic portions were then washed with brine, dried (MgSO₄), and concentrated under reduced pressure to give 56.2 mg of a colorless syrup.

The syrup was dissolved in 5 mL of CH_2Cl_2 and treated with 2,6-lutidine (25 μ L) and TBSOTf (42 μ L) at 0 °C, under N_2 . The reaction was allowed to warm to room temperature. After stirring overnight, the reaction was quenched by addition of 3 mL of saturated NaHCO₃ solution. The reaction mixture was extracted into CH_2Cl_2 (20 mL) and the organic layer was washed with H_2O , followed by brine. The organic layer was then dried with Na_2SO_4 and concentrated under reduced pressure to give 45 mg of a colorless oil.

This colorless oil (40 mg) was dissolved in 4 mL of THF and treated with LAH (10 mg). After stirring for 1 h, the reaction was quenched by addition of Na₂SO₄·10H₂O. The reaction mixture was filtered through a small column of Na₂SO₄, eluting with Et₂O, and concentrated under reduced pressure. Chromatography (3% ethyl acetate/hexanes) gave alcohol 7 [Ms = H] (21.7 mg) as a colorless syrup: ¹H NMR $(C_6D_6, 500 \text{ MHz}) \delta 4.51 \text{ (t, } J = 10.9 \text{ Hz, } 1 \text{ H)}, 4.46 \text{ (dd, } J = 8.7, 9.9)$ Hz, 1 H), 4.18-4.13 (m, 3 H), 3.99-3.90 (m, 3 H), 3.89 (dt, J = 10.0, 6.6 Hz, 1 H), 3.79 (dt, J = 10.0, 6.6 Hz, 1 H), 3.14 (dd, J = 6.0, 11.2 Hz, 1 H), 2.49 (dd, J = 3.0, 12.7 Hz, 1 H), 2.11 (dd, J = 3.3, 12.7 Hz, 1 H), 2.08-1.53 (m, 22 H), 1.52 (q, J = 11.9 Hz, 1 H), 1.41 (q, J = 1.41 (q, J = 1.41 H), 1.41 (q, J = 111.1 Hz, 1 H), 1.38 (t, J = 11.9 Hz, 1 H), 1.118 (s, 9 H), 1.103 (s, 9 H), 1.065 (s, 9 H), 1.034 (s, 9 H), 1.027 (s, 9 H), 1.019 (s, 18 H), 1.01 (d, J = 6.5 Hz, 3 H), 1.00 (d, J = 6.4 Hz, 3 H), 0.979 (s, 9 H), 0.86(d, J = 6.9 Hz, 3 H), 0.402 (s, 3 H), 0.352 (s, 3 H), 0.293 (s, 3 H),0.279 (s, 3 H), 0.249 (s, 3 H), 0.238 (s, 3 H), 0.213 (s, 3 H), 0.201 (s, 3 H), 0.190 (s, 3 H), 0.175 (s, 3 H), 0.170 (s, 3 H), 0.158 (s, 3 H), 0.140 (s, 3 H), 0.135 (s, 3 H), 0.067 (s, 3 H), 0.056 (s, 3 H).

[2S,2(2S,4S,6S,8S,10S,13S,14S),4R,6R,8R,10S]-4,10-Bis((1,1-dimethylethyl)dimethylsiloxy)-8-(2-((1,1-dimethylethyl)dimethylsiloxy)ethyl)-2-(13,15-dimethyl-2,4,6,8,10-hexakis((1,1-dimethylethyl)dimethylsiloxy)-14-O-(methylsulfonyl)hexadecanyl)-1,7-dioxaspiro-[5.5] undecane (7). Alcohol 7 [Ms = H] (21.7 mg) was dissolved in 4 mL of CH₂Cl₂ and treated with DMAP (40 mg, 0.328 mmol), Et₃N (45 μ L, 0.323 mmol), and MsCl (25 μ L, 0.323 mmol). After stirring under N₂ for 18 h, the reaction was quenched with 3 mL of saturated NaHCO₃ solution. The reaction mixture was extracted into Et₂O (20 mL), and the organic layer was washed with NaHCO₃, H₂O, and brine. The organic layer was then dried (MgSO₄) and concentrated under reduced pressure. Chromatography (3% ethyl acetate/hexanes) gave mesylate 7 as a colorless syrup: ¹H NMR (C₆D₆, 500 MHz) δ 4.63 (br t, J = 5.3 Hz, 1 H, 4.52 (m, 1 H), 4.46 (m, 1 H), 4.19-4.12 (m, 3 H),3.98-3.91 (m, 3 H), 3.89 (dt, J = 9.8, 6.8 Hz, 1 H), 3.79 (dt, J = 9.8, 6.5 Hz, 1 H), 3.64 (m, 1 H), 2.49 (dd, J = 3.6, 13.0 Hz, 1 H), 2.326 (s, 3 H), 2.11 (dd, J = 3.9, 13.0 Hz, 1 H), 2.09-1.35 (m, 24 H), 1.119 (s, 9 H), 1.107 (s, 9 H), 1.09 (d, J = 6.5 Hz, 3 H), 1.069 (s, 9 H), 1.045 (s, 9 H), 1.037 (s, 9 H), 1.028 (s, 9 H), 1.019 (s, 9 H), 1.00 (d, J = 7.1 Hz, 3 H, 0.980 (s, 9 H), 0.90 (d, J = 6.8 Hz, 3 H), 0.406 (s, J = 6.8 Hz, 3 H)3 H), 0.354 (s, 3 H), 0.308 (s, 3 H), 0.283 (s, 3 H), 0.274 (s, 3 H),

0.259 (s, 3 H), 0.241 (s, 3 H), 0.205 (s, 3 H), 0.191 (s, 3 H), 0.175 (s, 9 H), 0.140 (s, 3 H), 0.135 (s, 3 H), 0.066 (s, 3 H), 0.054 (s, 3 H).

[2S,2(2S,4S,6S,8S,10(2R,3S,6S)),4R,6S,8R,10S]-4,10-Di-O-acetyl-8-(2-O-acetyl-2-hydroxyethyl)-2-(2,4,6,8-tetra-O-acetyl-2,4,6,8-tetrahydroxy-9-(3-methyl-2-(1-methylethyl)tetrahydropyran-6-yl)nonanyl)-4,10-dihydroxy-1,7-dioxaspiro[5.5]undecane (8). Mesylate 7 (1.7 mg) was dissolved in 1 mL of THF and treated with 50 μ L of a 1.0 M solution of TBAF in THF. After stirring at 50 °C for 3 h, the reaction mixture was concentrated under a stream of N2 and purified by column chromatography (SiO₂, 20% MeOH/ethyl acetate) to give 1.0 mg of a white solid. The white solid was dissolved in 1.5 mL of THF and treated with DMAP (12 mg) and Ac₂O (9 µL). After stirring overnight, the reaction was quenched with a drop of MeOH. The reaction mixture was then diluted in 10 mL of ethyl acetate and washed with $0.05 \text{ M H}_2\text{SO}_4$ (2 × 5 mL) and brine (2 × 5 mL). The organic layer was dried with Na₂SO₄ and concentrated under reduced pressure. Chromatography (SiO₂, 50% ethyl acetate/hexanes) gave the product (0.7 mg) as a colorless solid: ^{1}H NMR (C₆D₆, 500 MHz) δ 5.41 (m, 1 H), 5.36-5.20 (m, 3 H), 4.97 (m, 1 H), 4.89 (m, 1 H), 4.32 (m, 1 H), 4.19-4.12 (m, 2 H), 3.87 (m, 1 H), 3.43 (m, 1 H), 3.19 (dd, J =3.9, 8.0 Hz, 1 H), 2.43 (dd, J = 4.1, 12.7 Hz, 1 H), 2.25 (m, 1 H), 2.08 (dd, J = 4.7, 12.4 Hz, 1 H), 2.05-1.35 (m, 17 H), 1.956 (s, 3 H),1.912 (s, 3 H), 1.880 (s, 3 H), 1.835 (s, 3 H), 1.778 (s, 3 H), 1.681 (s, 3 H), 1.637 (s, 3 H), 1.29–1.19 (m, 3 H), 1.15 (d, J = 6.9 Hz, 3 H), 1.12-0.94 (m, 3 H), 1.00 (d, J = 6.9 Hz, 3 H), 0.76 (d, J = 6.7 Hz, 3 H); HRMS (FAB) 857.4534 (M + H), calcd for $C_{43}H_{69}O_{17}$ 857.4536.

[2S,2(2R,4S,6S,8S,10(2R,3S,6S)),4R,6S,8R,10S]-4,10-Di-O-benzoyl-8-(2-O-benzoyl-2-hydroxyethyl)-2-(2,4:6,8-bis-O-(1-methylethyl)tetrahydroxy-9-(3-methyl-2-(1-methylethyl)tetrahydroxy-pyran-6-yl)nonanyl)-4,10-dihydroxy-1,7-dioxaspiro[5.5]undecane (9). Mesylate 7 (22.3 mg) was dissolved in 4 mL of THF and treated with 300 μ L of a 1.0 M solution of TBAF in THF. After stirring at 50 °C for 3 h, the reaction mixture was concentrated under a stream of N₂ and purified by column chromatography (SiO₂, 20% MeOH/ethyl acetate) to give 10.0 mg of a white solid.

The white solid was then dissolved in 4 mL of acetone and 0.4 mL of 2,2-DMP and treated with 5 mg of PPTS. After stirring at 50 °C for 8 h, the reaction was cooled and the acid quenched with a few drops of Et_3N . The reaction was concentrated under a stream of N_2 and purified by column chromatography (SiO₂, 75% acetone/hexanes) to give 3.5 mg of a colorless syrup.

The syrup was then dissolved in 1.5 mL of THF and treated with DMAP (17 mg) and Bz₂O (30 mg). After stirring overnight, the reaction was quenched with 2 mL of saturated NaHCO₃ solution. The reaction mixture was extracted into CH₂Cl₂ (10 mL) and the organic layer was then washed with saturated NaHCO3, H2O, and brine. The organic portion was dried with Na2SO4 and concentrated under reduced pressure. Chromatography (SiO₂, 20% ethyl acetate/hexanes) gave 3.0 mg of crude 9 as a colorless oil. The product was further purified by reverse-phase HPLC (MeOH) to give 2.9 mg of 9: ¹H NMR (C₆D₆, 500 MHz) δ 8.36-8.12 (m, 6 H), 7.20-7.05 (m, 9 H), 5.38 (m, 1 H), 5.19 (m, 1 H), 4.62 (m, 1 H), 4.57-4.45 (m, 3 H), 4.22-4.10 (m, 2 H), 4.05-3.96 (m, 2 H), 3.60 (m, 1 H), 3.01 (dd, J = 4.1, 7.9 Hz, 1 H), 2.55 (dd, J = 4.4, 12.6 Hz, 1 H), 2.28–2.22 (m, 2 H), 1.93–1.14 (m, 23 H), 1.616 (s, 3 H), 1.578 (s, 3 H), 1.510 (s, 3 H), 1.488 (s, 3 H), 1.015 (d, J = 6.8 Hz, 3 H), 0.99 (d, J = 6.5 Hz, 3 H), 0.76 (d, J= 6.8 Hz, 3 H); HRMS (FAB) 955.5207 (M + H), calcd for $C_{56}H_{75}O_{13}$ 955,5209.

In order to get better 13 C NMR data in the acetonide region, 9 (2.9 mg) was dissolved in 2 mL of MeOH and treated with a small amount of Dowex 50W-X1 acid resin to remove the acetonides. After 1.5 h, the reaction was filtered and concentrated under reduced pressure to give 2.1 mg of a white solid. This was dissolved in 200 μ L of acetone which was 33% enriched with $[1,3^{-13}C_2]$ acetone and treated with approximately 0.1 mg of PPTS. After stirring overnight, the acid was quenched with $10~\mu$ L of Et₃N and the reaction mixture was subjected to column chromatography (20% ethyl acetate/hexanes) to give 2.0 mg of product which had acetonides 33% 13 C-enriched at the methyl positions: 13 C NMR (signals for $1,3^{-13}$ C-acetone enriched acetonides, C_6D_6 , 125~MHz) $\delta~$ 30.746 (2) (d, J=4.8~Hz), 20.217 (d, J=3.2~Hz), 20.045 (d, J=4.8~Hz).

(3R,5S,7R,9S,11S,13S,15R,17R,19S,21S,24S,25S)-1-O-((1,1-Dimethylethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1-Dimethyl)dimethylsilyl

methylethylidene)-24,26-dimethylheptacosane-1,3,5,7,9,11,13,15,-17,19,21,25-dodecol (11 [Ms = H]). Pentaacetonide 5 (17.0 mg) was dissolved in 7.5 mL of a 2:1 mixture of MeOH and CH_2Cl_2 and cooled to -78 °C. Solid NaHCO₃ (10 mg) was added and then ozone was bubbled through the solution until a blue color persisted. Nitrogen was then bubbled through the solution until it was colorless, followed by addition of NaBH₄ (15 mg). The reaction mixture was allowed to slowly warm to room temperature. After 1 h, the reaction was diluted with Et₂O (30 mL) and quenched with 3 mL of saturated NaHCO₃ solution. The organic portion was decanted and the aqueous portion was washed with Et₂O (2 × 10 mL). The combined organic portions were then washed with brine, dried (MgSO₄), and concentrated under reduced pressure to give 15.2 mg of a colorless syrup.

This syrup was dissolved in 3 mL of CH_2Cl_2 and treated with 2,6-lutidine (15 μ L) and TBSOTf (25 μ L) at 0 °C, under N_2 . The reaction was allowed to warm to room temperature. After stirring overnight, the reaction was quenched by addition of 3 mL of saturated NaHCO₃ solution. The reaction mixture was extracted into CH_2Cl_2 (10 mL) and the organic layer was washed with H_2O and brine. The organic layer was then dried with Na_2SO_4 and concentrated under reduced pressure to give 17.2 mg of a colorless syrup.

The syrup was dissolved in 3 mL of THF and treated with LAH (10 mg). After stirring for 1 h, the reaction was quenched by addition of Na₂SO₄·10H₂O. The reaction mixture was filtered through a small column of Na₂SO₄, eluting with Et₂O, and concentrated under reduced pressure. Chromatography (30% ethyl acetate/hexanes) gave alcohol 11 [Ms = H] (8.6 mg) as a colorless syrup: ¹H NMR (300 MHz, C₆D₆) δ 4.35–3.98 (m, 9 H), 3.83–3.64 (m, 3 H), 2.98 (m, 1 H), 2.13–2.00 (m, 3 H), 1.84–1.20 (m, 27 H), 1.555 (s, 3 H), 1.546 (s, 3 H), 1.463 (s, 3 H), 1.444 (s, 6 H), 1.412 (s, 6 H), 1.370 (s, 6 H), 1.356 (s, 3 H), 0.95 (d, J = 6.6 Hz, 3 H), 0.88 (d, J = 6.5 Hz, 3 H), 0.77 (d, J = 6.7 Hz, 3 H), 0.079 (s, 3 H), 0.072 (s, 3 H); ¹³C NMR (C₆D₆, 75 MHz, DEPT) C 100.6, 100.4 (2), 98.7, 98.7, 18.6; CH 79.7, 69.4, 66.2, 66.1, 65.5, 63.7, 63.5 (2), 63.3 (2), 62.7, 35.4; CH₂ 59.7, 43.3 (2), 43.2, 42.7, 39.8, 39.3, 39.2, 39.0, 37.9, 37.5, 34.6, 29.8; CH₃ 31.3, 30.8, 26.3 (3), 25.3 (3), 25.2 (2), 25.1, 20.1, 20.0, 19.7, 18.8, 13.5, -5.1 (2).

(3R,5S,7R,9S,11S,13S,15R,17R,19S,21S,24S,25S)-1-O-((1,1-Dimethylethyl)dimethylsilyl)-3,5:7,9:11,13:15,17:19,21-pentakis-O-(1methylethylidene)-25-O-(methylsulfonyl)-24,26-dimethylheptacosane-1,3,5,7,9,11,13,15,17,19,21,25-dodecol (11). Alcohol 11 [Ms = H] (8.6 mg) was dissolved in 3 mL of CH₂Cl₂ and treated with DMAP (8 mg), Et₃N (9 μ L), and MsCl (5 μ L). After stirring under N₂ for 18 h, the reaction was quenched with 2 mL of saturated NaHCO₃ solution. The reaction mixture was extracted into Et₂O (20 mL), and the organic layer was washed with saturated NaHCO3, H2O, and brine. The organic layer was then dried (MgSO₄) and concentrated under reduced pressure. Chromatography (30% ethyl acetate/hexanes) gave mesylate 11 (8.5 mg) as a colorless syrup: ¹H NMR (C₆D₆, 500 MHz) δ 4.52 (dd, J =3.1, 7.5 Hz, 1 H), 4.31 (m, 1 H), 4.21-4.10 (m, 6 H), 4.10-4.00 (m, 2 H), 3.78 (m, 1 H), 3.71 (m, 1 H), 3.68 (m, 1 H), 2.229 (s, 3 H), 2.12-2.03 (m, 3 H), 1.81-1.20 (m, 23 H), 1.534 (s, 3 H), 1.525 (s, 3 H), 1.454 (s, 3 H), 1.433 (s, 6 H), 1.427 (s, 3 H), 1.402 (s, 3 H), 1.366 (s, 3 H), 1.359 (s, 3 H), 1.325 (s, 3 H), 0.985 (s, 9 H), 0.91 (d, J = 6.5)Hz, 3 H), 0.90 (d, J = 6.5 Hz, 3 H), 0.72 (d, J = 6.9 Hz, 3 H), 0.069(s, 3 H), 0.062 (s, 3 H).

[3R,5S,7S,9R,11R,13R,15S,17S,19S,20(2R,3S,6S)]-20-(3-Methyl-2-(1-methylethyl)tetrahydropyran-6-yl)-1,3:5,7:9,11:13,15:17,19-pentakis-O-(1-methylethylidene)eicosane-1,3,5,7,9,11,13,15,17,19-decol (12). Mesylate 11 (8.5 mg) was dissolved in 4 mL of MeOH and treated with Dowex 50W-X1 acid resin. After stirring for 2.5 h, the reaction was filtered and concentrated under reduced pressure. The resulting white solid was resubjected to the reaction conditions to ensure complete removal of the acetonide protecting groups.

The resulting polyol was then suspended in 3 mL of THF and treated with $100 \,\mu$ L of a 1.0 M solution of TBAF in THF. After stirring at 50 °C for 3 h, the reaction was concentrated under reduced pressure. Chromatography (SiO₂, 25% MeOH/ethyl acetate) gave 3.7 mg of a white solid

The white solid was dissolved in 250 μ L of acetone which was 20% enriched with [1,3-¹³C₂]acetone and treated with approximately 1 mg of CSA. After stirring overnight, the acid was quenched with 10 μ L of Et₃N and the reaction mixture was subjected to column chromatography (SiO₂, 50% ethyl acetate/hexanes) to give 1.0 mg of the desired

product which had acetonides 20% 13 C-enriched at the methyl positions: 1 H NMR ($^{\circ}$ C₆D₆, 500 MHz) δ 4.30 (m, 1 H), 4.22–3.95 (m, 9 H), 3.66–3.64 (m, 2 H), 3.01 (dd, J = 4.2, 7.8 Hz, 1 H), 2.27 (ddd, J = 5.3, 9.2, 13.8 Hz, 1 H), 2.07 (quintet, J = 7.1 Hz, 1 H), 1.84 (m, 1 H), 1.67–1.12 (m, 22 H), 1.609 (s, 3 H), 1.580 (s, 3 H), 1.549 (s, 6 H), 1.491 (s, 3 H), 1.466 (s, 3 H), 1.439 (s, 3 H), 1.420 (s, 3 H), 1.369 (s, 6 H), 1.02 (d, J = 6.7 Hz, 3 H), 1.00 (d, J = 6.7 Hz, 3 H), 0.93 (m, 1 H), 0.76 (d, J = 6.4 Hz, 3 H); 13 C NMR (signals for 1,3- 13 C-acetone enriched acetonides, $^{\circ}$ C₆D₆, 125 MHz) δ 30.729 (2) (d, J = 4.8 Hz), 30.673 (d, J = 4.8 Hz), 30.448 (d, J = 4.6 Hz), 24.963 (2) (d, J = 4.8 Hz), 20.107 (d, J = 4.8 Hz), 20.041 (d, J = 4.8 Hz), 19.488 (d, J = 4.6 Hz); HRMS (FAB) 767.5366 (M - CH₃), calcd for $^{\circ}$ C₄3H₇₅O₁₁ 767.5311. The 13 C-enriched compounds were also observed in the FABMS as (M - CH₃) 767 (30%), 769 (40%), 771 (20%).

(3R,5S,7S,9S,11S,13S,15R,17R,19S,21S,24S,25S)-1,25-Bis-O-((R)-2-methoxy-2-phenyl-2-(trifluoromethyl)acetyl)-3,5:7,9:11,13:15,17: 19,21-pentakis-O-(1-methylethylidene)-24,26-dimethylheptacosane-1,3,5,7,9,11,13,15,17,19,21,25-dodecol (13R). Pentaacetonide 6 (10.6 mg) was dissolved in 6 mL of a 2:1 mixture of MeOH and CH₂Cl₂ and cooled to -78 °C. Ozone was bubbled through the solution until a blue color persisted. Nitrogen was then bubbled through the solution until it was colorless, followed by addition of NaBH₄ (10 mg). The reaction mixture was allowed to slowly warm to room temperature. After 1 h, the reaction was diluted with Et₂O (20 mL) and quenched with 2 mL of saturated NaHCO₃ solution. The organic portion was decanted and the aqueous portion was washed with Et₂O (2 × 10 mL). The combined organic portions were then washed with brine, dried (MgSO₄), and concentrated under reduced pressure to give 10.3 mg of a colorless syrup.

The syrup was dissolved in 3 mL of THF and treated with LAH (10 mg). After stirring for 1 h, the reaction was quenched by addition of Na_2SO_4 +10H₂O. The reaction mixture was filtered through a small column of Na_2SO_4 , eluting with Et₂O, and concentrated under reduced pressure to give 9.3 mg of a diol as a colorless syrup.

A 2.3 mg portion of the diol was dissolved in 2 mL of CH₂Cl₂ and treated with DMAP (14 mg) and (S)-MTPA-Cl (17 μ L). ¹⁹ After stirring overnight, the reaction was quenched with saturated NaHCO₃ solution and extracted into CH₂Cl₂. The organic layer was washed with H₂O and brine, dried (NaSO₄), and concentrated under reduced pressure. Chromatography (SiO₂, 20% ethyl acetate/hexanes) gave the bis ester **13R** (3.5 mg) as a colorless oil: ¹H NMR (CDCl₃, 500 MHz) δ 7.58–7.34 (m, 10 H), 4.86 (dd, J = 3.9, 7.5 Hz, 1 H), 4.38–4.36 (m, 2 H), 3.99–3.88 (m, 8 H), 3.80 (m, 1 H), 3.65 (m, 1 H), 3.52 (s, 6 H), 1.95 (m, 1 H), 1.87–1.78 (m, 3 H), 1.74 (m, 1 H), 1.62–1.07 (m, 21 H), 1.359 (s, 3 H), 1.325 (s, 3 H), 1.309 (s, 6 H), 1.300 (s, 6 H), 1.294 (s, 6 H), 1.289 (s, 3 H), 1.275 (s, 3 H), 0.87 (d, J = 6.7 Hz, 3 H), 0.82 (d, J = 6.4 Hz, 3 H), 0.79 (d, J = 6.7 Hz, 3 H); HRMS (FAB) 1217.6188 (M – CH₃), calcd for C₆₃H₉₁O₁₆F₆ 1217.6213.

(3R,5S,7S,9S,11S,13S,15R,17R,19S,21S,24S,25S)-1,25-Bis-O-((S)-2-methoxy-2-phenyl-2-(trifluoromethyl)acetyl)-3,5:7,9:11,13:15,17: 19,21-pentakis-O-(1-methylethylidene)-24,26-dimethylheptacosane-1,3,5,7,9,11,13,15,17,19,21,25-dodecol (13S). The (S)-MTPA ester was prepared from the intermediate diol (4.2 mg) following the same procedure used to prepare the (R)-MPTA ester 13R described above. Chromatography (SiO₂, 20% ethyl acetate/hexanes) gave the bis ester 13S (3.5 mg) as a colorless oil: 1 H NMR (CDCl₃, 500 MHz) δ 7.58– 7.35 (m, 10 H), 4.87 (dd, J = 3.8, 8.0 Hz, 1 H), 4.42 (m, 1 H), 4.34 (dt, J = 10.7, 7.2 Hz, 1 H), 3.99-3.82 (m, 8 H), 3.80 (m, 1 H), 3.66 (m, 1 H), 3.52 (s, 3 H), 3.51 (s, 3 H), 1.93 (m, 1 H), 1.85-1.74 (m, 4 H), 1.61–1.07 (m, 21 H), 1.367 (s, 3 H), 1.332 (s, 3 H), 1.305 (s, 6 H), 1.300 (s, 6 H), 1.292 (s, 6 H), 1.281 (s, 3 H), 1.236 (s, 3 H), 0.85 (d, J = 6.9 Hz, 3 H), 0.83 (d, J = 6.9 Hz, 3 H), 0.79 (d, J = 6.6 Hz, 3 H); HRMS (FAB) 1217.6224 (M - CH₃), calcd for $C_{63}H_{91}O_{16}F_{6}$ 1217.6213.

[2R,4R,4(2S,4S,6S,8S,10S,12S,15S,16S),6S,8R,10S]-4-(2,4,6,8,10,-12,16-Heptahydroxy-16-O-((R)-2-methoxy-2-phenyl-2-(trifluoromethyl)acetyl)-15,17-dimethyl-2,4:6,8:10,12-tris-O-(1-methylethylidene)-octadecanyl)-8-(2-O-((R)-2-methoxy-2-phenyl-2-(trifluoromethyl)-acetyl)-2-hydroxyethyl)-2-methyl-10-hydroxy-1,3,7-trioxaspiro[5.5]-undecane (15R). A 3.9 mg sample of diol 14 was dissolved in 2 mL of CH₂Cl₂ and treated with DMAP (14 mg) and (S)-MTPA-Cl (14 μ L). ¹⁹ After stirring overnight, the reaction was concentrated and then

 $C_{59}H_{83}O_{16}F_6$ 1161.5587.

dissolved in THF. This was treated with 200 μ L of a 1.0 M solution of TBAF in THF. After stirring for 3 h, the reaction was quenched with saturated NaHCO₃ solution and extracted into CH₂Cl₂. The organic layer was washed with H₂O and brine, dried (NaSO₄), and concentrated under reduced pressure. Chromatography (SiO₂, 50% ethyl acetate/hexanes) gave the bis ester **15R** (4.7 mg) as a colorless oil: ¹H NMR (CDCl₃, 500 MHz) δ 7.59–7.35 (m, 10 H), 4.86 (dd, J = 4.1, 7.9 Hz, 1 H), 4.76 (q, J = 5.1 Hz, 1 H), 4.50 (dt, J = 10.9, 5.8 Hz, 1 H), 4.41 (dt, J = 10.9, 7.2 Hz, 1 H), 4.12–3.96 (m, 7 H), 3.73–3.64 (m, 2 H), 3.52 (s, 3 H), 3.51 (s, 3 H), 2.37 (br t, J = 7.5 Hz, 1 H), 2.01 (m, 1 H), 1.95 (m, 1 H), 1.90–1.86 (m, 4 H), 1.80–1.71 (m, 2 H), 1.58–1.04 (m, 18 H), 1.394 (s, 3 H), 1.357 (s, 3 H), 1.339 (s, 3 H), 1.328 (s, 3 H), 1.306 (s, 3 H), 1.302 (s, 3 H), 1.15 (d, J = 5.1 Hz, 3 H), 0.87 (d, J = 6.8 Hz, 3 H), 0.83 (d, J = 6.5 Hz, 3 H), 0.79 (d, J = 6.8 Hz, 3 H); HRMS (FAB) 1161.5554, (M – CH₃) calcd for

[2R,4R,4(2S,4S,6S,8S,10S,12S,15S,16S),6S,8R,10S]-4-(2,4,6,8,10,-12,16-Heptahydroxy-16-O-((S)-2-methoxy-2-phenyl-2-(trifluorom $ethyl) acetyl) - 15,17-dimethyl - 2,4:6,8:10,12-tris-{\it O-}(1-methylethylidene) - 10,12-tris-{\it O-}(1-methylethylidene) - 10$ octadecanyl)-8-(2-O-((S)-2-methoxy-2-phenyl-2-(trifluoromethyl)acetyl)-2-hydroxyethyl)-2-methyl-10-hydroxy-1,3,7-trioxaspiro[5.5]undecane (15S). The (S)-MTPA ester was prepared from diol 14 (4.0 mg) by following the same procedure used to prepare the (R)-MTPA ester described above. Chromatography (SiO2, 20% ethyl acetate/ hexanes) gave the bis ester 15S (4.7 mg) as a colorless oil: ¹H NMR (CDCl₃, 500 MHz) δ 7.58-7.36 (m, 10 H), 4.87 (dd, J = 3.7, 7.7 Hz, 1 H), 4.77 (q, J = 5.1 Hz, 1 H), 4.57 (dt, J = 5.6, 10.6 Hz, 1 H), 4.36(dt, J = 10.6, 7.0 Hz, 1 H), 4.07 - 3.96 (m, 7 H), 3.68 (m, 1 H), 3.61(m, 1 H), 3.53 (s, 3 H), 3.51 (s, 3 H), 2.38 (m, 1 H), 2.00 (m, 1 H), 1.93 (m, 1 H), 1.88-1.83 (m, 4 H), 1.80-1.73 (m, 2 H), 1.53-1.04 (m, 18 H), 1.393 (s, 3 H), 1.365 (s, 3 H), 1.335 (s, 6 H), 1.300 (s, 6 H), 1.131 (d, J = 5.1 Hz, 3 H), 0.85 (d, J = 6.7 Hz, 3 H), 0.82 (d, J = 6.7 Hz, 3 Hz, 3 Hz, 4 Hz, = 6.7 Hz, 3 H, 0.79 (d, J = 6.7 Hz, 3 H); HRMS (FAB) 1161.5564 $(M - CH_3)$, calcd for $C_{59}H_{83}O_{16}F_6$ 1161.5587.

Synthesis of Degradation Fragment 12. (2R,3S)-2,4-Dimethyl-1-(phenylthio)-3-pentanol. A solution of (2R,3R)-2,4-dimethyl-1,3pentanediol²⁸ (20) (584 mg, 4.42 mmol, 1.00 equiv) in 20 mL of dry acetonitrile was treated with tributylphosphine (1.10 mL, 4.42 mmol, 1.00 equiv) followed by the addition of diphenyl disulfide (1.00 g, 4.59 mmol, 1.04 equiv). After stirring under N2 for 20 h, a 40 mL portion of 1 N NaOH was added. The reaction mixture was extracted with Et₂O (3 \times 50 mL), and the combined organic layers were washed with 1 N NaOH and H₂O. The organic layer was dried with MgSO₄ and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 10% ethyl acetate/hexanes) gave 815 mg of the product (3.84 mmol, 87%) as a colorless oil: IR (neat) 3425, 3074, 3058, 2963, 2930, 2873, 1584, 1480, 1438, 1382, 1246, 1091, 1065, 1026, 980, 952, 737, 690 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.35-7.13 (m, 10 H), 3.37 (m, 1 H), 3.05 (dd, J = 7.4, 12.7 Hz, 1 H), 2.89 (dd, J = 6.6, 12.7 Hz, 1 H), 1.93 (m, 1 H), 1.68 (m, 1 H), 1.51 (d, J = 4.8 Hz, 1 H), 0.98 (d, J = 6.7 Hz, 6 H), 0.80 (d, J = 6.7 Hz, 3 H); ¹³C NMR (75 MHz, CDCl₃, DEPT) C 137.1; CH 129.3, 129.1, 126.0, 78.6, 34.9, 31.5; CH₂ 38.8; CH₃ 19.4, 19.3, 12.8. Anal. Calcd for C₁₃H₂₀OS: C, 69.59; H, 8.98. Found: C, 69.62; H, 8.83.

(2R,3S)-2,4-Dimethyl-3-O-(phenylmethyl)-1-(phenylthio)-3-pentanol. A solution of (2R,3S)-2,4-dimethyl-1-(phenylthio)-3-pentanol (815 mg, 3.84 mmol, 1.0 equiv) in 8 mL of THF was added via cannula to a suspension of KH (300 mg, 7.50 mmol, 2.0 equiv) in 10 mL of THF. After stirring for 15 min, the reaction mixture was cooled to 0 °C and benzyl bromide (500 µL, 4.20 mmol, 1.1 equiv) was added dropwise. After stirring for 18 h, the reaction was quenched with 20 mL of saturated NaHCO₃ solution and extracted with CH₂Cl₂ (3×). The combined organic layers were washed with brine $(2\times)$, dried with Na₂SO₄, and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 1% ethyl acetate/hexanes) gave 1.210 g (3.84 mmol, 100%) of the product as a colorless liquid: IR (neat) 3062, 3028, 2962, 2930, 2872, 1716, 1584, 1496, 1479, 1454, 1438, 1383, 1270, 1092, 1068, 1026, 966, 736, 696 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.38-7.14 (m, 5 H), 4.67 (d, J = 11.3 Hz, 1 H), 4.57 (d, J = 11.3 Hz, 1 H), 3.28 (dd, J = 3.0, 7.8 Hz, 1 H), 3.01 (dd, J = 7.3, 12.6 Hz, 1 H), 2.86 (dd, J = 6.8, 12.6 Hz, 1 H), 2.00 (m, 1 H), 1.89 (m, 1 H), 1.05(d, J = 7.0 Hz, 3 H), 1.03 (d, J = 6.9 Hz, 3 H), 0.85 (d, J = 6.8 Hz, 3 H)3 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) C 139.2, 136.9; CH 129.0, 128.8, 128.2, 127.4, 127.3, 125.7, 86.6, 35.2, 31.2; CH_2 75.1, 38.7; CH_3 19.7, 19.4, 14.0. Anal. Calcd for $C_{20}H_{26}OS$: C, 76.39; H, 8.33. Found: C, 76.50; H, 8.38.

(2R,3S)-2,4-Dimethyl-3-O-(phenylmethyl)-1-(phenylsulfonyl)-3pentanol (21). (2R,3S)-2,4-Dimethyl-3-O-(phenylmethyl)-1-(phenylthio)-3-pentanol (1.210 g, 3.85 mmol, 1.0 equiv) was dissolved in 20 mL of MeOH and treated with a solution of Oxone (3.22 g, 5.24 mmol, 1.4 equiv) dissolved in 20 mL of H₂O. After stirring for 1 h, the reaction mixture was diluted with H_2O and extracted with $CHCl_3$ (4 × 30 mL). The combined organic layers were washed with H₂O and brine. The organic layer was dried with Na2SO4 and concentrated under reduced pressure. Purification by flash chromatography (SiO2, 16% ethyl acetate/hexanes) gave 1.232 g (3.56 mmol, 92%) of the product as a slightly yellow oil: $[\alpha]^{27}_D = +9.0^{\circ}$ (c 1.10, CHCl₃); IR (neat) 3062, 3030, 2964, 2873, 1497, 1447, 1403, 1385, 1349, 1305, 1244, 1206, 1147, 1086, 1070, 1027, 999, 963, 927, 869, 856, 839, 785, 739, 722, 690 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.86-7.19 (m, 10 H), 4.61 (d, J = 11.7 Hz, 1 H), 4.49 (d, J = 11.7 Hz, 1 H), 3.25-3.19 (m, 2)H), 2.90 (dd, J = 6.9, 14.2 Hz, 1 H), 2.45 (dq, J = 3.1, 6.9 Hz, 1 H), 1.85 (m, 1 H), 1.06 (d, J = 6.9 Hz, 3 H), 0.98 (d, J = 6.7 Hz, 3 H), 0.89 (d, J = 6.8 Hz, 3 H); ¹³C NMR (75 MHz, CDCl₃, DEPT) C 140.0, 138.7; CH 133.5, 129.2, 128.2, 127.7, 127.4, 127.3, 86.7, 30.8, 30.7; CH₂ 74.4, 60.2; CH₃ 19.9, 19.3, 14.4. Anal. Calcd for C₂₀H₂₆O₃S: C, 69.33; H, 7.56. Found: C, 69.49; H, 7.31.

(2S,4R,5R,6S)- and (2S,4S,5R,6S)-5,7-Dimethyl-1,2-O-(1-methylethylidene)-6-O-(phenylmethyl)-4-(phenylsulfonyl)-1,2,6-octanetriol (23). A solution of sulfone 21 (2.137 g, 6.18 mmol, 1.00 equiv) dissolved in 30 mL of THF was cooled to 0 °C under N2 and treated with 2.50 mL of butyllithium (2.47 M in hexanes, 6.18 mmol, 1.00 equiv). After 1.5 h, a solution of (R)-(2,2-dimethyl-1,3-dioxolan-4yl)methyl bromide³⁰ (22) (1.279 g, 6.56 mmol, 1.06 equiv) and DMPU (3.75 mL, 5 equiv) dissolved in 5 mL of THF was added dropwise via cannula. The reaction mixture was allowed to warm slowly to room temperature over 24 h followed by quenching with saturated NaHCO₃ solution. The reaction mixture was extracted with CH_2Cl_2 (3 × 75 mL), and the combined organic layers were washed with H₂O and brine. The organic layer was dried with Na2SO4 and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 15-25% ethyl acetate/hexanes) gave 916 mg of unreacted sulfone (43%) and 1.506 g (3.27 mmol, 53%) of the product as a colorless oil: IR (neat) 3088, 3062, 3028, 2963, 2937, 1605, 1585, 1496, 1447, 1380, 1370, 1345, 1305, 1251, 1216, 1147, 1084, 1066, 1029, 998, 966, 924, 881, 866, 832, 731, 695, 606 cm⁻¹; ¹H NMR (300 MHz, C_6D_6) δ 7.83– 7.76 (m, 2 H), 7.33-6.91 (m, 8 H), 4.62-4.46 (m, 2.3 H), 4.34 (d, J = 11.3 Hz, 0.7 H), 4.17 (dd, J = 3.4, 6.8 Hz, 0.3 H), 3.80 (dd, J = 6.3, 8.1 Hz, 1.0 H), 3.56-3.48 (m, 1.0 H), 3.27 (dd, J = 6.7, 8.1 Hz, 0.7 H), 3.09 (dd, J = 5.3, 8.2 Hz, 0.3 H), 2.93 (dd, J = 3.6, 7.4 Hz, 0.7 H), 2.50 (m, 0.3 H), 2.38–2.22 (m, 1.7 H), 1.89–1.76 (m, 2.0 H), 1.46 (d, J = 7.0 Hz, 0.9 H), 1.33 (s, 2.1 H), 1.23 (d, J = 6.7 Hz, 2.1 H), 1.22 (s, 2.1 H), 1.13 (dd, J = 6.8 Hz, 0.9 H), 1.07 (d, J = 6.8 Hz, 0.9 H), 1.06 (s, 0.9 H), 0.81 (s, 0.9 H), 0.80 (d, J = 7.0 Hz, 2.1 H), 0.50 (d, J = 6.7 Hz, 2.1 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) C 140.5, 139.8, 139.7, 139.3, 109.1; CH 133.3, 133.1, 129.3, 129.1, 128.9, 128.7, 128.5, 127.6, 127.5, 86.9, 85.2, 74.6, 72.5, 63.5, 63.2, 35.9, 35.3, 31.0, 30.3; CH₂ 75.4, 74.7, 69.9, 69.4, 32.1, 29.4; CH₃ 27.4, 26.9, 25.6, 25.5, 20.6, 20.4, 16.4, 15.7, 13.7, 10.4. Anal. Calcd for C₂₆H₃₆O₅S: C, 67.80; H, 7.88. Found: C, 68.00; H, 8.08.

(2S,5S,6S)-5,7-Dimethyl-1,2-O-(1-methylethylidene)-6-O-(phenylmethyl)-1,2,6-octanetriol. A solution of sulfone 23 (794 mg, 1.73 mmol, 1.0 equiv) dissolved in 20 mL of MeOH was treated with Na₂-HPO₄ (1.56 g, 10.1 mmol, 6.4 equiv) and sodium amalgam (6% sodium content, 4.21 g, 10.5 mmol, 6.1 equiv). After stirring for 2.5 h, the reaction mixture was filtered through a Celite pad and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 10% ethyl acetate/hexanes) gave 511 mg (1.60 mmol, 93%) of the product as a colorless oil: $[\alpha]^{27}_D = +18.1^{\circ} (c \ 0.93, CHCl_3); IR (neat)$ 3062, 3028, 2965, 2939, 2874, 1718, 1602, 1496, 1454, 1378, 1344, 1273, 1216, 1160, 1096, 1067, 1028, 976, 854, 806, 791, 753, 714, 699 cm⁻¹; ¹H NMR (300 MHz, C₆D₆) δ 7.35-7.08 (m, 5 H), 4.48-4.42 (m, 2 H), 3.87 (m, 1 H), 3.79 (dd, J = 5.9, 7.5 Hz, 1 H), 3.35 (t,J = 7.5 Hz, 1 H), 2.80 (dd, J = 4.0, 6.5 Hz, 1 H), 1.85 (m, 1 H), 1.66-1.54 (m, 2 H), 1.45 (s, 3 H), 1.44-1.27 (m, 3 H), 1.35 (s, 3 H), 1.00 (d, J = 6.7 Hz, 3 H), 0.96 (d, J = 6.7 Hz, 3 H), 0.85 (d, J = 6.7

Hz, 3 H); 13 C NMR (75 MHz, C_6D_6 , DEPT) C 140.0, 108.9; CH 128.6, 127.6, 127.5, 76.5, 36.0, 31.4; CH₂ 75.3, 69.8, 31.9, 31.0; CH₃ 27.5, 26.1, 20.4, 19.0, 14.8. Anal. Calcd for $C_{20}H_{32}O_3$: C, 74.96; H, 10.06. Found: C, 75.17; H, 9.88.

(2S,5S,6S)-5,7-Dimethyl-1,2-O-(1-methylethylidene)-1,2,6-octanetriol. A suspension of 675 mg (2.11 mmol) of (2S,5S,6S)-5,7dimethyl-1,2-O-(1-methylethylidene)-6-O-(phenylmethyl)-1,2,6-octanetriol and 60 mg of 20% Pd(OH)₂/C in 10 mL of MeOH was flushed with H₂ and then stirred vigorously under balloon pressure. After 4 h the mixture was filtered through a Celite pad which had been moistened with MeOH and Et₃N. The resulting filtrate was concentrated under reduced pressure and purified by flash chromatography (SiO₂, 20-25% ethyl acetate/hexanes) to give 436 mg (1.90 mmol, 90%) of the product as a colorless oil: IR (neat) 3498, 2959, 2871, 1457, 1370, 1218, 1160, 1095, 1060, 981, 851, 792 cm⁻¹; ¹H NMR (300 MHz, C_6D_6) δ 3.87 (m, 1 H), 3.81 (m, 1 H), 3.37 (t, J = 7.3 Hz, 1 H), 2.93 (m, 1 H), 1.66–1.42 (m, 3 H), 1.43 (s, 3 H), 1.40–1.26 (m, 3 H), 1.34 (s, 3 H), 0.94 (d, J = 6.6 Hz, 3 H), 0.82 (d, J = 6.6 Hz, 3 H), 0.76 (d, $J = 6.7 \text{ Hz}, 3 \text{ H}); {}^{13}\text{C NMR} (75 \text{ MHz}, C_6D_6, \text{ DEPT}) C 108.7; CH$ 79.3, 76.5, 35.3, 31.2; CH₂ 69.6, 31.4, 30.4; CH₃ 27.2, 25.9, 19.5, 18.8, 13.2. Anal. Calcd for C₁₃H₂₆O₃: C, 67.79; H, 11.38. Found: C, 68.02; H, 11.48.

(2S,5S,6S)-5,7-Dimethyl-6-O-(methylsulfonyl)-1,2-O-(1-methylethylidene)-1,2,6-octanetriol (24). A solution of (2S,5S,6S)-5,7dimethyl-1,2-O-(1-methylethylidene)-1,2,6-octanetriol (460 mg, 2.00 mmol, 1.0 equiv) and DMAP (244 mg, 2.00 mmol, 1.0 equiv) dissolved in 20 mL of CH₂Cl₂ was cooled to 0 °C under N₂ and treated with Et₃N (668 μ L, 4.80 mmol, 2.4 equiv) and mesyl chloride (185 μ L, 2.39 mmol, 1.2 equiv). After stirring for 15 h, the reaction was quenched with saturated NaHCO3 solution and extracted with CH2Cl2 $(3\times)$. The combined organic layers were washed with H₂O and brine. The organic layer was dried with Na₂SO₄ and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 25% ethyl acetate/hexanes) gave 594 mg (1.93 mmol, 96%) of the product as a colorless oil: IR (neat) 2938, 2876, 1464, 1418, 1369, 1337, 1215, 1172, 1059, 973, 906, 848, 810, 764, 668 cm⁻¹; ¹H NMR (300 MHz, C_6D_6) δ 4.47 (dd, J = 3.5, 7.4 Hz, 1 H), 3.87 (m, 1 H), 3.80 (dd, J =6.0, 7.5 Hz, 1 H), 3.41 (t, J = 7.5 Hz, 1 H), 2.30 (s, 3 H), 1.75 (m, 1 H), 1.59 (m, 1 H), 1.51–1.31 (m, 4 H), 1.43 (s, 3 H), 1.34 (s, 3 H), 0.92 (d, J = 6.7 Hz, 3 H), 0.84 (d, J = 6.7 Hz, 3 H), 0.72 (d, J = 6.8 Hz, 3 H)Hz, 3 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) C 108.8; CH 90.8, 76.3, 35.1, 30.4; CH₂ 69.6, 31.4, 30.5; CH₃ 38.2, 27.2, 26.0, 19.7, 19.0, 14.4. Anal. Calcd for C₁₄H₂₈O₅S: C, 54.52; H, 9.15. Found: C, 54.32; H, 9.27.

(2R,3S,6S)-(3-Methyl-2-(1-methylethyl)tetrahydropyran-6-yl)methanol (25). Mesylate 24 (584 mg, 1.90 mmol, 1.0 equiv) was dissolved in 20 mL of MeOH and treated with Dowex 50W-X1 acid resin. After stirring for 2 h, the reaction was filtered and concentrated under reduced pressure to give 508 mg of a yellow oil which was dissolved in 15 mL of THF and treated with 250 mg of NaH (10.4 mmol, 5.5 equiv). After stirring for 2 h, the reaction was quenched with saturated NaHCO₃ solution and extracted with Et₂O (2 × 20 mL). The combined organic layers were washed with H2O and brine. The organic layer was dried with MgSO4 and concentrated under reduced pressure. Purification by flash chromatography (SiO2, 25% ethyl acetate/hexanes) gave 257 mg (1.50 mmol, 79%) of the product as a slightly yellow oil: IR (neat) 3428, 2957, 2931, 2872, 1461, 1384, 1367, 1238, 1207, 1173, 1156, 1141, 1107, 1054, 985, 905, 881, 856, 819 cm⁻¹; 1 H NMR (300 MHz, $C_{6}D_{6}$) δ 3.63–3.58 (m, 2 H), 3.34 (m, 1 H), 2.89 (t, J = 5.9 Hz, 1 H), 2.28 (m, 1 H), 1.80 (m, 1 H), 1.51-1.23 (m, 4 H), 0.94 (d, J = 6.7 Hz, 3 H), 0.84 (d, J = 6.8 Hz, 3 H), 0.81 (d, J = 6.7 Hz, 3 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) CH 81.7, 71.5, 29.9, 27.8; CH₂ 63.4, 26.6, 23.7; CH₃ 23.7, 20.2, 18.2.

(2R,3S,6S)-(3-Methyl-2-(1-methylethyl)tetrahydropyran-6-yl)-O-(methylsulfonyl)methanol. A solution of alcohol 25 (250 mg, 1.45 mmol, 1.0 equiv) and DMAP (177 mg, 1.45 mmol, 1.0 equiv) dissolved in 15 mL of CH₂Cl₂ was cooled to 0 °C under N₂ and treated with Et₃N (484 μ L, 3.84 mmol, 2.4 equiv) and mesyl chloride (135 μ L, 1.74 mmol, 1.2 equiv). After stirring for 17 h, the reaction was quenched with saturated NaHCO₃ solution and extracted with CH₂Cl₂ (3×). The combined organic layers were washed with H₂O and brine. The organic layer was dried with Na₂SO₄ and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 25%

ethyl acetate/hexanes) gave 340 mg (1.36 mmol, 94%) of the product as a colorless oil: IR (neat) 2959, 2934, 2873, 1462, 1355, 1176, 1119, 1093, 1059, 1031, 961, 886, 823, 747 cm $^{-1}$; 1 H NMR (300 MHz, C_6D_6) δ 4.06 (dd, J=7.7, 10.6 Hz, 1 H), 3.78 (dd, J=4.3, 10.6 Hz, 1 H), 3.64 (m, 1 H), 2.82 (t, J=6.0 Hz, 1 H), 2.27 (s, 3 H), 1.74 (m, 1 H), 1.40 (m, 1 H), 1.26 (m, 1 H), 1.15-1.08 (m, 2 H), 0.94 (m, 1 H), 0.91 (d, J=6.7 Hz, 3 H), 0.86 (d, J=6.7 Hz, 3 H), 0.74 (d, J=6.8 Hz, 3 H); 13 C NMR (75 MHz, C_6D_6 , DEPT) CH 82.0, 68.7, 29.7, 27.7; CH₂ 69.9, 26.2, 23.5; CH₃ 37.0, 20.1, 18.0, 16.7. Anal. Calcd for $C_{11}H_{22}O_4S$: C, 52.77; H, 8.85. Found: C, 52.75; H, 8.95.

(2R,3S,6S)-6-(Bromomethyl)-3-methyl-2-(1-methylethyl)tetrahy**dropyran (16).** A solution of (2R,3S,6S)-(3-methyl-2-(1-methylethyl)tetrahydropyran-6-yl)-O-(methylsulfonyl)methanol (325 mg, 1.30 mmol, 1.00 equiv) and nBu₄NBr (425 mg, 1.32 mmol, 1.02 equiv) in 10 mL of dry toluene was heated to reflux under N2. After 24 h, an additional 100 mg of nBu₄NBr was added and the reaction was refluxed for an additional 2 h. The reaction mixture was then cooled, diluted with 100 mL of Et₂O, and washed with H₂O (2×) and brine. The organic layer was dried with MgSO₄ and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 3% ethyl acetate/hexanes) gave 259 mg (1.10 mmol, 85%) of the product as a colorless liquid: IR (neat) 2959, 2931, 2872, 1460, 1384, 1222, 1155, 1114, 1078, 1062, 1027, 969 cm⁻¹; 1 H NMR (300 MHz, $C_{6}D_{6}$) δ 3.74 (m, 1 H), 3.22 (dd, J = 7.5, 10.3 Hz, 1 H), 2.97 (dd, J = 6.1, 10.3 Hz, 1 H), 2.78 (dd, J= 5.3, 6.7 Hz, 1 H), 1.74 (m, 1 H), 1.44-1.31 (m, 3 H), 1.26 (m, 1 H), 0.96 (d, J = 6.8 Hz, 3 H), 0.95 (d, J = 6.7 Hz, 3 H), 0.91 (m, 1 H), 0.71 (d, J = 6.7 Hz, 3 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) CH 80.9, 71.1, 30.3, 28.0; CH₂ 33.6, 26.5, 25.8; CH₃ 20.3, 17.7, 16.0. Anal. Calcd for C₁₀H₁₉BrO: C, 51.08; H, 8.14. Found: C, 51.16; H, 8.24.

(8S,10S)-5,12-Heptadecadiyne-8,10-diol. A solution of 1-hexyne (2.85 mL, 24.8 mmol, 4.0 equiv) in 50 mL of THF under N₂, cooled to -78 °C, was treated with 9.7 mL (23.7 mmol, 3.8 equiv) of a 2.44 M solution of butyllithium. After 20 min, (2R,4R)-1,2:4,5-dianhydro-3-deoxypentitol^{26a} (26) (622 mg, 6.22 mmol, 1.0 equiv), dissolved in 10 mL of THF, was added via cannula followed by dropwise addition of BF₃*OEt₂ (3.00 mL, 24.4 mmol, 3.9 equiv). After 30 min, the reaction was quenched with 40 mL of saturated NaHCO3, and the solution was warmed to room temperature. The reaction mixture was diluted with Et₂O (75 mL), and the layers were separated. The aqueous layer was treated with 40 mL of 1 N NaOH solution and extracted with Et₂O (3 \times 30 mL). The combined organic layers were washed with 1 N NaOH, H₂O, and brine. The organic portion was then dried (MgSO₄) and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 20-40% ethyl acetate/hexanes) gave the product (1.541 g, 94%) as a colorless syrup: $[\alpha]^{27}_D = +17.7^{\circ}$ (c = 1.18, CH₂-Cl₂); IR (neat) 3285, 2930, 2872, 1599, 1466, 1428, 1343, 1252, 1232, 1192, 1087, 1057, 1022 cm⁻¹; 1 H NMR (300 MHz, CDCl₃) δ 4.00 (m, 2 H), 2.60 (br s, 2 H), 2.35 (m, 4 H), 2.13 (m, 4 H), 1.72 (t, J = 5.8Hz, 2 H), 1.49-1.32 (m, 8 H), 0.87 (t, J = 7.1 Hz, 6 H); 13 C NMR (75 MHz, CDCl₃, DEPT) C 83.5, 76.1; CH 67.9; CH₂ 41.0, 31.3, 28.0, 22.2, 18.6; CH₃ 13.8. Anal. Calcd for C₁₇H₂₈O₂: C, 77.22; H, 10.67. Found: C, 77.13; H, 10.56.

(8S,10S)-8,10-O-(1-Methylethylidene)-5,12-heptadecadiyne-8,10diol (27). (85,105)-5,12-Heptadecadiyne-8,10-diol (1.541 g, 5.83 mmol) was dissolved in 40 mL of a mixture of acetone and 2,2dimethoxypropane (3:1) and treated with 12 mg of CSA. After stirring for 18 h, 0.25 mL of Et₃N was added and the reaction mixture was concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 4% ethyl acetate/hexanes) gave 1.503 g (4.94 mmol, 85%) of the product as a colorless liquid: IR (neat) 2987, 2957, 2932, 2872, 1468, 1434, 1379, 1329, 1303, 1225, 1170, 1111, 1094, 1022, 992, 940, 904, 842, 815, 728 cm $^{-1}$; 1 H NMR (300 MHz, C₆D₆) δ 3.98 (m, 2 H), 2.47 (ddt, J = 5.6, 16.3, 2.7 Hz, 2 H), 2.32 (ddt, J = 7.3, 16.3, 2.3 Hz, 2 H), 2.07 (m, 4 H), 1.85 (t, J = 7.5 Hz, 2 H), 1.43-1.32 (m, 8 H), 1.34 (s, 6 H), 0.81 (t, J = 7.0 Hz, 6 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) C 100.5, 81.9, 76.8; CH 66.2; CH₂ 37.2, 31.5, 26.6, 22.2, 18.8; CH₃ 25.3, 13.8. Anal. Calcd for C₂₀H₃₂O₂: C, 78.90; H, 10.59. Found: C, 78.83; H, 10.50.

(8S,10S)-8,10-O-(1-Methylethylidene)-5,12-heptadecadiene-8,10-diol. A suspension of 93 mg of 5% Pd on BaSO₄ and 930 μ L of quinoline in 80 mL of ethyl acetate was stirred under H₂ (balloon pressure) for 1 h followed by the addition of acetonide 27 (1.460 g, 4.80 mmol) dissolved in 10 mL of ethyl acetate. After stirring for 1 h,

the reaction mixture was filtered through Celite and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 2% ethyl acetate/hexanes) gave 1.430 g (4.64 mmol, 97%) of the product as a colorless liquid: IR (neat) 2986, 2957, 2930, 2873, 2859, 1657, 1548, 1512, 1459, 1404, 1378, 1224, 1169, 1107, 1022, 992, 942, 904, 836, 724 cm⁻¹; ¹H NMR (300 MHz, C_6D_6) δ 5.60–5.47 (m, 4 H), 3.88 (m, 2 H), 2.38 (dt, J = 13.8, 6.6 Hz, 2 H), 2.25 (dt, J = 13.8, 5.7 Hz, 2 H), 2.06-2.00 (m, 4 H), 1.56 (t, J = 7.6 Hz, 2 H), 1.48 (s, 6 H), 1.36-1.21 (m, 8 H), 0.87 (t, J = 7.0 Hz, 6 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) C 100.2; CH 131.9, 125.7, 66.8; CH₂ 38.0, 34.2, 32.1, 27.5, 27.5, 22.6; CH_3 25.1, 14.1. Anal. Calcd for $C_{20}H_{36}O_2$: C, 77.87; H, 11.76. Found: C, 78.00; H, 11.83.

(3R,5R)-3,5-Dihydroxy-3,5-O-(1-methylethylidene)-1,7-heptanedialdehyde (28). (85,105)-8,10-O-(1-Methylethylidene)-5,12-heptadecadiene-8,10-diol (1.40 g, 4.55 mmol, 1.00 equiv) was dissolved in 50 mL of dry CH₂Cl₂ and cooled to -78 °C. Ozone was then bubbled through the solution until a slight blue color persisted. Nitrogen was then bubbled through the solution until it was colorless. Triphenylphosphine (2.45 g, 9.35 mmol, 2.06 equiv) was then added and the reaction was warmed to room temperature. After stirring for 24 h, the reaction was concentrated under a stream of N2. Purification by flash chromatography (SiO₂, 33% ethyl acetate/hexanes) gave 828 mg (4.14 mmol, 91%) of the unstable dialdehyde as a colorless liquid: IR (neat) 2988, 2938, 2834, 2732, 1726, 1447, 1382, 1224, 1167, 1128, 1031, 1003, 899, 735 cm⁻¹; ¹H NMR (300 MHz, C_6D_6) δ 9.37 (dd, J= 1.0, 2.5 Hz, 2 H), 4.00 (ddt, J = 4.6, 8.3, 7.8 Hz, 2 H), 2.18 (ddd, J = 4.6, 8.3, 7.8 Hz, 2 H)J = 2.5, 8.3, 16.6 Hz, 2 H), 1.89 (ddd, J = 1.0, 4.6, 16.6 Hz, 2 H), 1.21 (s, 6 H), 1.18 (t, J = 7.8 Hz, 2 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) C 100.7; CH 199.2, 61.9; CH₂ 49.0, 37.3; CH₃ 24.5.

(2R,4R,6R,8R)-, (2R,4R,6R,8S)-, and (2S,4R,6R,8S)-2,4:6,8-Bis-O-(1-methylethylidene)-2,4,6,8-tetrahydroxynonane-1,9-dinitrile (17). A solution of dialdehyde 28 (818 mg, 4.09 mmol, 1.00 equiv) dissolved in 2 mL of CH₂Cl₂ was cooled to 0 °C and treated with TMSCN (1.12 mL, 8.40 mmol, 2.05 equiv) and a catalytic amount of KCN/18-crown-6 complex (caution: highly exothermic). After stirring for 90 min, the CH₂Cl₂ was evaporated under a stream of N₂, and the resulting oil was treated with 50 mg of CSA and 50 mL of a mixture of acetone and 2,2-dimethoxypropane (4:1). After stirring for 3 h, an additional 50 mg of CSA was added and the reaction was stirred overnight. The reaction was quenched with 0.5 mL of Et₃N and the reaction was concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 25% ethyl acetate/hexanes) gave 1.091 g (3.71 mmol, 91%) of the product as a colorless syrup which was a 1:2:1 mixture of the three possible diastereomers: ¹H NMR (300 MHz, C₆D₆) δ 4.22-4.09 (m, 2 H), 4.04-3.98 (m, 1 H), 3.56-3.45 (m, 1 H), 1.41-0.85 (m, 6 H), 1.53 (s, 1.5 H), 1.52 (s, 1.5 H), 1.32 (s, 1.5 H), 1.29 (s, 1.5 H), 1.27 (s, 1.5 H), 1.24 (s, 1.5 H), 0.97 (s, 3 H); ¹³C NMR (75 MHz, $C_6D_6,\, DEPT)\,\, C\,\, 119.9,\, 118.0,\, 118.0,\, 100.8,\, 100.6,\, 99.8,\, 99.7;\, CH\,\, 63.4,\, 100.6$ 63.1, 61.2, 59.1, 58.5; CH₂ 41.2, 41.1, 34.6, 34.4, 33.3, 33.2; CH₃ 29.5, 29.5, 21.7, 21.7, 18.9, 18.9. Anal. Calcd for C₁₅H₂₂N₂O₄: C, 61.21; H, 7.53. Found: C, 61.37; H, 7.59.

(3S)-1,3-O-(1-Methylethylidene)-5-O-(phenylmethyl)-1,3,5-pentanetriol (30). A solution of ethyl (3S)-3-hydroxy-5-(phenylmethoxy)pentanoate^{32,33} (29) (5.20 g, 20.6 mmol, 1 equiv) dissolved in 50 mL of Et₂O was added dropwise via cannula (over a 30 min period) to a suspension of LAH in 100 mL of Et₂O at 0 °C. After stirring for 60 min, the reaction mixture was warmed to room temperature and stirred an additional 60 min. The reaction was then cooled to 0 °C and quenched by slow addition of H₂O (2.9 mL), 15% NaOH (2.9 mL), and H₂O (12 mL). The aluminum salts were removed by filtration, and the filtrate was washed with brine, dried with MgSO4, and concentrated under reduced pressure. The resulting colorless oil was treated with 30 mg of CSA and 80 mL of a mixture of acetone and 2,2-dimethoxypropane (3:1). After stirring for 2 h, the reaction was quenched with 1 mL of Et₃N and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 12% ethyl acetate/hexanes) gave 4.19 g (16.7 mmol, 81%) of the product as a colorless syrup: IR (neat) 3063, 3029, 2991, 2946, 2866, 1496, 1454, 1380, 1369, 1272, 1238, 1202, 1168, 1135, 1100, 1057, 1028, 997, 969, 870, 860, 797, 738, 698 cm⁻¹; ¹H NMR (300 MHz, C_6D_6) δ 7.30-7.07 (m, 5 H), 4.33 (d, J = 12.2 Hz, 1 H), 4.31 (d, J = 12.2 Hz, 1 H), 3.91 (dddd, J= 2.6, 4.5, 7.4, 11.9 Hz, 1 H), 3.65 - 3.61 (m, 2 H), 3.52 (ddd, J =5.6, 8.0, 9.0 Hz, 1 H), 3.40 (ddd, J = 5.7, 5.8, 9.1 Hz, 1 H), 1.831.60 (m, 2 H), 1.49 (s, 3 H), 1.48-1.38 (m, 1 H), 1.30 (s, 3 H), 0.97 $(ddd, J = 2.3, 4.6, 12.9 \text{ Hz}, 1 \text{ H}); {}^{13}\text{C NMR} (75 \text{ MHz}, C_6D_6, \text{DEPT})$ C 139.4, 98.2; CH 128.4, 127.5, 66.1; CH₂ 73.0, 66.4, 59.8, 37.2, 31.7; CH_3 30.3, 19.3. Anal. Calcd for $C_{15}H_{22}O_3$: C, 71.97; H, 8.86. Found: C, 71.76; H, 8.60.

(3R,5R)-1,3-O-(1-Methylethylidene)-7-octene-1,3,5-triol (31). A suspension of 1.03 g (3.96 mmol) of benzyl ether 30 and 20 mg of 20% Pd(OH)₂/C in 10 mL of MeOH was flushed with H₂ and then stirred vigorously under balloon pressure. After 24 h the mixture was filtered through a Celite pad which had been moistened with MeOH and Et₃N. The resulting filtrate was concentrated under reduced pressure and purified by flash chromatography (SiO₂, 50% ethyl acetate/ hexanes) to give 580 mg of a colorless oil which was immediately dissolved in 40 mL of CH₂Cl₂ and treated with NaHCO₃ (4.53 g, 53.9 mmol) and Dess-Martin reagent (2.03 g, 4.79 mmol). After stirring for 40 min, the reaction mixture was diluted with 200 mL of Et₂O followed by the addition of 80 mL of saturated NaHCO3 solution and 80 mL of 0.5 M NaS₂O₃ solution. The layers were separated, and the organic portion was washed sequentially with NaHCO₃, H₂O and brine. The organic layer was dried with MgSO₄ and concentrated under reduced pressure to give the crude aldehyde as a light brown liquid.

The aldehyde was dissolved in 5 mL of Et₂O and added via cannula to a cooled (-78 °C), stirred solution of (+)-Ipc₂B(allyl)³⁴ (2.5 mmol) in 12 mL of Et₂O. The reaction mixture was allowed to warm to room temperature overnight followed by the addition of 2 mL of 15% NaOH and 4 mL of 30% H₂O₂. The reaction mixture was refluxed for 2 h and then cooled and diluted with Et₂O. The reaction mixture was then washed sequentially with H₂O (2×), NaHCO₃, and brine. The organic layer was dried with MgSO₄, concentrated under reduced pressure, and purified by flash chromatography (20-30% ethyl acetate/hexanes) to give the product (348 mg, 1.74 mmol, 44%) as a colorless oil: IR (neat) 3454, 3075, 2993, 2944, 2918, 2870, 1641, 1460, 1430, 1381, 1272, 1240, 1201, 1164, 1138, 1100, 1051, 998, 968, 936, 914, 884, 870, 813, 747 cm⁻¹; 1 H NMR (300 MHz, C₆D₆) δ 5.80 (m, 1 H), 5.05– 5.00 (m, 2 H), 4.00 (m, 1 H), 3.92 (m, 1 H), 3.68-3.53 (m, 2 H), 2.30 (d, J = 3.6 Hz, 1 H), 2.16 (dd, J = 6.8, 13.7 Hz, 1 H), 2.09 (dd, J = 6.8, 13.7 Hz)5.7, 13.7 Hz, 1 H), 1.59-1.32 (m, 3 H), 1.43 (s, 3 H), 1.29 (s, 3 H), 0.88 (m, 1 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) C 98.2; CH 135.4, 67.0, 66.4; CH₂ 117.0, 59.7, 42.9, 42.7, 31.3; CH₃ 30.1, 19.1. Anal. Calcd for $C_{11}H_{20}O_3$: C, 65.97; H, 10.07. Found: C, 66.07; H, 10.25.

(2R,4S,6R)- and (2S,4S,6R)-2,4:6,8-Bis-O-(1-methylethylidene)-2,4,6,8-tetrahydroxyoctanenitrile (19). A solution of alcohol 31 (336 mg, 1.68 mmol, 1.00 equiv) and TMSNMe₂ (330 μL, 2.06 mmol, 1.23 equiv) in 10 mL of CH₃CN was heated to reflux for 3 h. The reaction was then cooled and concentrated under reduced pressure to give 440 mg of a colorless liquid which was dissolved in 9 mL of acetone and 3 mL of H₂O. To this solution were added N-methylmorpholine N-oxide (270 mg, 2.31 mmol, 1.4 equiv) and 0.5 mL of OsO₄ solution (2.5% in tert-butyl alcohol). After stirring for 90 min, TLC analysis showed complete consumption of the alkene. A solution of NaIO₄ (740 mg, 3.45 mmol, 2.1 equiv) in 3 mL of H₂O was added all at once. After stirring for 60 min, the reaction mixture was diluted with H₂O, extracted (2 × Et₂O), washed (Na₂SO₃, brine), and concentrated under reduced pressure. The resulting residue was dissolved in CH2Cl2 and washed with saturated NH₄Cl solution. The organic layer was then dried with Na2SO4 and concentrated under reduced pressure to give 396 mg of the aldehyde as a colorless oil.

The aldehyde was dissolved in 3 mL of CH₂Cl₂, cooled to 0 °C, and treated with TMSCN (200 μ L, 1.50 mmol, 1.04 equiv) and a eatalytic amount of KCN/18-crown-6 complex. After stirring for 15 min, the CH₂Cl₂ was evaporated under a stream of N₂, and the resulting oil was treated with 15 mg of CSA and 10 mL of a mixture of acetone and 2,2-dimethoxypropane (3:1). After stirring overnight, the reaction was quenched with 0.2 mL of Et₃N and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 20% ethyl acetate/hexanes) gave 346 mg (1.28 mmol, 77% overall yield from alcohol 31) of the product as a colorless syrup which was a 1:1 epimeric mixture at the cyanohydrin center: IR (neat) 2992, 2943, 2870, 1462, 1431, 1382, 1270, 1240, 1199, 1160, 1105, 1047, 1002, 970, 946, 912, 871, 819, 805 cm⁻¹; ¹H NMR (300 MHz, C_6D_6) δ 4.29 (m, 0.5 H), 4.19 (m, 0.5 H), 4.01 (m, 0.5 H), 3.93-3.81 (m, 1 H), 3.66-3.57 (m, 2.5 H), 1.59 (s, 1.5 H), 1.45 (s, 1.5 H), 1.43 (s, 1.5 H), 1.34 (s, 1.5 H), 1.29 (s, 1.5 H), 1.26 (s, 3 H), 1.03 (s, 1.5 H), 1.53-1.13 (m, 4.5 H),

0.97-0.79 (m, 1.5 H); 13 C NMR (75 MHz, C_6D_6 , DEPT) C 120.0, 118.1, 100.8, 99.8, 98.3, 98.2; CH 64.5, 64.2, 63.8, 61.8, 59.3, 58.8; CH₂ 59.8, 42.6, 42.5, 34.9, 33.7, 31.9, 31.8; CH₃ 30.2, 30.2, 29.7, 21.9, 19.4, 19.3, 19.1. Anal. Calcd for $C_{14}H_{23}NO_4$: C, 62.43; H, 8.61. Found: C, 62.13; H, 8.79.

[2R,4S,6R,8S,9(2R,3S,6S)]-8-Cyano-2,4:6,8-bis-O-(1-methylethylidene)-9-(3-methyl-2-(1-methylethyl)tetrahydropyran-6-yl)-2,4,6,8tetrahydroxynonanenitrile (32). A solution of nitriles 17 (370 mg, 1.26 mmol, 1.88 equiv) in 8 mL of THF was added dropwise via cannula to a solution of LiNEt₂ (0.74 mmol, 1.1 equiv) in 8 mL of THF at -78 °C under argon. After 1 h, DMPU (660 μ L, 8 equiv) was added followed by the dropwise addition of a solution of bromide 16 (158 mg, 0.672 mmol, 1.00 equiv) in 2 mL of THF. The reaction mixture was then transferred to a -25 °C ice/MeOH bath. The system was allowed to slowly warm to 15 °C over 15 h. The reaction mixture was then quenched with saturated NH4Cl solution and extracted into CH₂Cl₂ (3×). The combined organic layers were washed sequentially with H₂O and brine (2×), dried with Na₂SO₄, and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 10-20% ethyl acetate/hexanes) gave 45 mg of the unreacted bromide (28%) and 121 mg (0.270 mmol, 40%) of the desired product as a colorless syrup: IR (neat) 2993, 2932, 2873, 1460, 1433, 1384, 1330, 1262, 1206, 1163, 1124, 1060, 1024, 1002, 976, 944, 920, 877, 814, 672 cm⁻¹; ¹H NMR (500 MHz, C₆D₆) δ 4.35 (dddd, J = 2.3, 4.8, 6.9, 12.1)Hz, 1 H), 4.24 (m, 1 H), 3.89 (dd, J = 2.8, 11.9 Hz, 1 H), 3.55 (dddd, J = 2.5, 5.0, 6.8, 11.8 Hz, 1 H), 2.82 (dd, J = 4.9, 7.0 Hz, 1 H), 2.10 (dd, J = 7.5, 14.7 Hz, 1 H), 1.92 - 1.80 (m, 2 H), 1.72 (dd, J = 3.3,14.7 Hz, 1 H), 1.70 (s, 3 H), 1.50 (m, 1 H), 1.43-1.28 (m, 5 H), 1.37 (s, 3 H), 1.30 (s, 3 H), 1.19 (m, 1 H), 1.09 (ddd, J = 2.6, 9.5, 13.7 Hz,1 H), 0.99 (d, J = 6.7 Hz, 3 H), 0.98 (m, 1 H), 0.94 (s, 3 H), 0.93 (d, J = 6.7 Hz, 3 H, 0.80 (dt, J = 12.9, 2.5 Hz, 1 H, 0.77 (d, J = 6.7 Hz, 1 Hz)3 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) C 122.5, 117.9, 100.9, 99.9, 69.1; CH 80.8, 66.6, 63.6, 62.4, 59.1, 30.8, 28.1; CH₂ 44.3, 41.5, 38.1, 34.7, 29.6, 26.8; CH₃ 30.7, 29.5, 21.5, 20.5, 18.9, 18.0, 16.5; HRMS (FAB) 449.3037 (M + H). Anal. Calcd for $C_{25}H_{40}N_2O_5$: C, 66.94; H, 8.99. Found: C, 67.04; H, 8.86.

(3R,5S,7R,9R,11R)-12-Bromo-7-cyano-1,3:5,7:9,11-tris-O-(1-methylethylidene)dodecane-1,3,5,7,9,11-hexol (33). A solution of nitriles 19 (283 mg, 1.05 mmol, 1.00 equiv) in 3 mL of THF was cooled to -78 °C and added dropwise via cannula to a solution of LiNEt₂ (1.20 mmol, 1.2 equiv) in 8 mL of THF at -78 °C under argon. After 1 h, a solution of dibromide 18 (800 mg, 2.65 mmol, 2.52 equiv) in 2 mL of THF, cooled to -78 °C, was added dropwise via cannula. The reaction mixture was then transferred to a -23 °C ice/MeOH bath. The system was allowed to slowly warm to 15 °C over 15 h. The reaction mixture was then quenched with saturated NH₄Cl solution and extracted into CH2Cl2 (3×), dried with Na2SO4, and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 10-20% ethyl acetate/hexanes) gave 540 mg of the unreacted dibromide (68%) and 319 mg (0.651 mmol, 62%) of the desired product as a colorless syrup: IR (neat) 2990, 2942, 2870, 1496, 1461, 1432, 1382, 1262, 1224, 1197, 1160, 1124, 1049, 988, 971, 955, 942, 909, 870, 850, 808, 733, 696, 651 cm⁻¹; ¹H NMR (300 MHz, C_6D_6) δ 4.44 (m, 1 H), 4.05 (m, 1 H), 3.92 (m, 1 H), 3.72 (m, 1 H), 3.63-3.60 (m, 2 H), 3.01 (dd, J = 6.5, 10.5 Hz, 1 H), 2.93 (dd, J = 5.0, 10.5 Hz, 1 H), 1.88 (dd, J = 8.3, 14.6 Hz, 1 H), 1.74–1.63 (m, 3 H), 1.72 (s, 3 H), 1.49-1.20 (m, 5 H), 1.42 (s, 3 H), 1.36 (s, 3 H), 1.31 (s, 3 H), 1.28 (s, 3 H), 1.24 (s, 3 H), 0.90 (m, 1 H); 13C NMR (75 MHz, C₆D₆, DEPT) C 122.3, 101.1, 101.0, 98.3, 68.5; CH 66.8, 64.6, 62.6, 62.0; CH₂ 59.8, 46.8, 42.8, 38.6, 37.5, 25.2, 32.0; CH₃ 31.1, 30.2, 24.7, 24.6, 21.7, 19.4; HRMS (FAB) 490.1795 (M + H). Anal. Calcd for C₂₂H₃₆-BrNO₆: C, 53.88; H, 7.40. Found: C, 54.04; H, 7.24.

[3R,5S,7R,9R,11R,13S,15R,17R,19S,20(2R,3S,6S)]-7,13,19-Tricy-ano-20-(3-methyl-2-(1-methylethyl)tetrahydropyran-6-yl)-1,3:5,7:9,-11:13,15:17,19-pentakis-O-(1-methylethylidene)eicosane-1,3,5,7,9,-11,13,15,17,19-decol (34). A solution of nitrile 32 (120 mg, 0.268 mmol, 1.00 equiv) in 1.7 mL of THF was added dropwise via cannula to a solution of LiNEt₂ (0.322 mmol, 1.2 equiv) in 2 mL of THF at

-78 °C under argon. After 1 h, DMPU (160 µL, 4 equiv) was added followed by the dropwise addition of a solution of bromide 33 (174 mg, 0.355 mmol, 1.33 equiv) in 1.3 mL of THF. The reaction mixture was then transferred to a -22 °C ice/MeOH bath. The system was allowed to slowly warm to 10 °C over 14 h. The reaction mixture was then quenched with saturated NH₄Cl solution and extracted into CH_2Cl_2 (3×). The combined organic layers were washed with brine, dried with Na₂SO₄, and concentrated under reduced pressure. Purification by flash chromatography (SiO₂, 15 - 20 - 25% ethyl acetate/ hexanes) gave 187 mg (0.218 mmol, 81%) of the desired product as a colorless syrup: $[\alpha]^{27}_D = +13.8^{\circ}$ (c 0.70, CHCl₃); IR (neat) 2990, 2939, 2871, 1462, 1434, 1383, 1262, 1208, 1161, 1026, 955, 871, 807, 680 cm⁻¹; ¹H NMR (500 MHz, C_6D_6) δ 4.58-4.50 (m, 3 H), 4.33 (m, 1 H), 4.26 (m, 1 H), 3.95 (m, 2 H), 3.63-3.61 (m, 2 H), 2.82 (dd, J =4.9, 7.1 Hz, 1 H), 2.13 (dd, J = 7.3, 14.6 Hz, 1 H), 1.97–1.92 (m, 2 H), 1.84-1.79 (m, 2 H), 1.78 (s, 3 H), 1.77 (s, 3 H), 1.76-1.64 (m, 5 H), 1.75 (s, 3 H), 1.54-1.28 (m, 15 H), 1.48 (s, 3 H), 1.44 (s, 3 H), 1.40 (s, 3 H), 1.39 (s, 3 H), 1.36 (s, 3 H), 1.31 (s, 3 H), 0.99 (m, 1 H), 0.99 (d, J = 6.7 Hz, 3 H), 0.88 (d, J = 6.7 Hz, 3 H), 0.86 (m, 1 H), 0.76 (d, J = 6.7 Hz, 3 H); ¹³C NMR (75 MHz, C₆D₆, DEPT) C 122.7, 122.5, 121.6, 101.3, 101.3, 101.2, 100.9, 69.3, 68.9, 68.7; CH 80.9, 66.9, 64.7, 63.0, 62.7, 62.4, 62.0, 30.9, 28.3; CH₂ 59.9, 48.1, 47.1, 44.4, 42.9, 41.7, 41.6, 39.5, 38.4, 32.0, 29.9, 27.1; CH₃ 31.2, 31.0, 30.3, 24.6, 24.6, 21.8, 21.7, 21.7, 20.7, 19.4, 18.1, 16.6; HRMS (FAB) 858.5540 (M + H). Anal. Calcd for $C_{47}H_{75}N_3O_{11}$: C, 65.79; H, 8.81. Found: C, 65.68; H, 8.66.

[3R,5S,7S,9R,11R,13R,15S,17S,19S,20(2R,3S,6S)]-20-(3-Methyl-2-(1-methylethyl)tetrahydropyran-6-yl)-1,3:5,7:9,11:13,15:17,19pentakis-O-(1-methylethylidene)eicosane-1,3,5,7,9,11,13,15,17,19decol (12). Lithium metal (80 mg, 11.5 mmol, 112 equiv) was dissolved in 15 mL of ammonia to give a bright blue solution which was cooled to -78 °C. Pentaacetonide nitrile 34 (88.0 mg, 0.103 mmol, 1 equiv) was dissolved in 5 mL of THF and added to the Li/NH3 solution via cannula. After being stirred for 1 h, the reaction was quenched with 740 mg of solid NH4Cl and warmed to room temperature, and the ammonia was allowed to evaporate. The remaining residue was dissolved in 20 mL of H2O and 20 mL of CH2Cl2. The layers were separated, and the aqueous portion was extracted with CH₂Cl₂ (3 × 15 mL). The organic layers were combined and washed with brine, dried with Na2SO4, and concentrated under reduced pressure. Purification by flash chromatography (SiO2, 25% ethyl acetate/hexanes) gave the product (65.5 mg, 81%) as a colorless syrup: $[\alpha]^{27}_D = +20.5^{\circ}$ (c 0.85, CHCl₃); IR (neat) 2989, 2941, 2870, 1379, 1266, 1225, 1199, 1168, 110, 1025, 968, 941, 872, 807 cm⁻¹; ¹H NMR (C₆D₆, 500 MHz) δ 4.29 (m, 1 H), 4.22-4.02 (m, 7 H), 4.02-3.95 (m, 2 H), 3.69-3.63 (m, 2 H), 3.01 (dd, J = 4.2, 7.8 Hz, 1 H), 2.26 (ddd, J = 5.4, 9.2, 13.9)Hz, 1 H), 2.06 (quintet, J = 6.9 Hz, 1 H), 1.84 (m, 1 H), 1.64-1.11 (m, 22 H), 1.603 (s, 3 H), 1.585 (s, 3 H), 1.544 (s, 6 H), 1.487 (s, 3 H), 1.462 (s, 3 H), 1.436 (s, 3 H), 1.417 (s, 3 H), 1.368 (s, 3 H), 1.365 (s, 3 H), 1.02 (d, J = 6.8 Hz, 3 H), 0.99 (d, J = 6.8 Hz, 3 H), 0.93 (m, 1 H), 0.76 (d, J = 6.6 Hz, 3 H); ¹³C NMR (C₆D₆, 75 MHz, DEPT) C 100.3, 98.6, 98.5 (2), 98.3; CH 79.9, 67.6, 66.7, 66.1, 65.6, 65.5, 65.4, 65.0, 64.9, 63.3, 62.5, 31.5, 28.4; CH₂ 60.0, 43.9 (2), 43.1, 43.0, 39.2, 38.7, 38.5, 37.8 (2), 32.2, 28.6, 27.6; CH₃ 30.7 (2), 30.7, 30.4, 25.0, 25.0, 20.7, 20.1, 20.0, 19.9, 19.5, 18.0, 15.8; HRMS (FAB) 767.5298 $(M - CH_3)$. Anal. Calcd for $C_{44}H_{78}O_{11}$: C, 67.49; H, 10.04. Found: C, 67.32; H, 9.89.

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