PALLADIUM MEDIATED ALLYLIC MITSUNOBU DISPLACEMENT: STEREOCONTROLLED SYNTHESIS OF HEPOXILIN A3 AND TRIOXILIN A3 METHYL ESTERS

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Summary: A regio- and stereoselective palladium mediated allylic displacement under Mitsunobu conditions was exploited for the preparation of several hepoxilin A3 and trioxilin A3 stereoisomers.

Hepoxilin A₃ (HxA₃) is generated as a pair of C(8)-diastereomers from 12-hydroperoxyeicosatetraenoic acid (12-HPETE) via an intramolecular rearrangement¹ catalyzed by naturally occurring heme-iron complexes² (eq 1). Enzymatic as well as non-enzymatic hydrolysis³ of the labile *trans*-epoxide gives rise to the corresponding triols, trioxilin A₃ (TxA₃). Metabolites of the hepoxilin/trioxilin pathway have been identified in plants, marine organisms, and several mammalian species⁴ wherein they have been proposed, *inter alia*, to modulate neural signal transduction, stimulate hormone secretion, induced heat shock protein expression, and mobilize intracellular calcium⁵.



The constitution of HxA₃ derived from 12(S)-HPETE was established by Corey and Su⁶, who subsequently clarified the absolute configurations of the C(8)-alcohols⁷. As part of a comprehensive synthetic program to expedite the physiologic evaluation and structural elucidation of novel eicosanoids, we describe herein a stereocontrolled route to HxA₃ and TxA₃ stereoisomers based on a novel palladium mediated allylic displacement⁸ of an acyclic, chiral alcohol⁹ under Mitsunobu conditions.



⁴ DEAD/Ph₃P/PhCO₂H, PdCl₂(CH₃CN)₂, THF, 25 ° C, 30 min.^b 2%HCl/MeOH, 24 ° C, 2h.° TsCl, C₃H₅N/CH₂Cl₂, O ° C, 12h. ^d DDQ, CH ₂Cl₂/H₂O (20:1), O° C, 4h.° NaOMe/MeOH, O ° C, 12h.

Previously, it was noted that Mitsunobu inversion of triol 1, a key intermediate in the synthesis¹⁰ of trioxilin B3, afforded approximately equal amounts of benzoate 2 and a chromatographically separable mixture of transposition products 3a,b (a/b, 4:1). We now report similar treatment of 1 in the presence of freshly prepared PdCl2(CH3CN)2 (0.1 equiv) rapidly resulted in the nearly exclusive production of $3a,b^{11}$ (73%; a/b, 10:1) (Scheme). A mechanistic rationale for this transformation involves palladium interception of the initially formed Mitsunobu oxyphosphonium intermediate¹² to give a π -allyl complex (eq 2). Subsequent *cis*-transfer of coordinated¹³ benzoate to carbon generates the observed major diastercomer 3a. Control experiments using 2 as well as the benzoate of 1 established that neither palladium promoted [3,3]-sigmatropic rearrangement¹⁴ nor allylic displacement at C(8) are significant contributors to product formation under the above reaction conditions.



Access to the hepoxilin series from 3 exploited an unexpectedly specific hydrolysis of only the C(12)benzyl ether. Thus, exposure of 3a to 2% HCl/MeOH under carefully controlled conditions and tosylation of the resultant alcohol led to 4a (TLC: SiO₂, 5% MeOH/CH₂Cl₂, $R_f \sim 0.36$). Cleavage of the remaining benzyl ether using 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) and NaOMe induced ring closure with concomitant benzoate solvolysis gave HxA3 methyl ester $5a^{15}$, $[\alpha]^{24}_{D} = -26^{\circ}$ (c 0.83, acetone). The 8(S)-isomer 5b, $[\alpha]^{24}_{D} = -5.1^{\circ}$ (c 0.41, acetone), was obtained analogously from 3b.

Alternatively, mild acidic hydrolysis of 3a and b as above followed by acetylation (Ac₂O, py, O^oC, 12h) evolved 7a and b, respectively. Sequential DDQ deprotection (77%), tosylation (84%), and NaOMe ring closure/deacetylation (70%) culminated in 8a and b. The enantiomeric pairs 5a/8b and 5b/8a were identical by HPLC and MS with the more polar methyl ester and less polar methyl ester, respectively, of HxA₃ obtained by hematin catalyzed rearrangement of 12-HPETE.



Concurrent DDQ removal of both dimethoxybenzyl ethers from 3a and benzoate solvolysis (NaOMe/MeOH, O°C, 10h) afforded TxA3 methyl ester 6a, $[\alpha]^{24}D = -8.0^{\circ}$ (c 0.65, CCl4). Likewise, 3b furnished 6b, $[\alpha]^{24}D = +2.8^{\circ}$ (c 0.58, CCl4).

Initial studies have demonstrated that other π -allyl palladium complexes can be generated from oxyphosphonium salts. Investigations into the scope and limitations of this methodology are in progress.

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15. Spectral data for 5a: ¹H NMR (C₆D₆, 250 MHz) δ 0.87 (t, J~6.8 Hz, 3H), 1.14-1.40 (m, 6H), 1.55 (dt, J~7, 14Hz, 2H), 1.78-2.35 (complex m, 10H), 2.72 (dt, J~ 2, 5 Hz, 1H), 3.03 (dd, J~ 2, 7.5 Hz, 1H), 3.33 (s, 3H), 3.86-3.98 (m, 1H), 5.13-5.56 (m, 5H), 5.78 (dd, J~ 5, 16 Hz, 1H); mass spec (PICI, CH₄) of 5a TMS ether: m/e (%) 451 (14, M+29), 423 (22, M+1), 407 (100), 333 (84, M-OSiMe₃), 315 (44). 5b: ¹H NMR (C₆D₆, 250 MHz) δ 0.87 (t, J~6.5 Hz, 3H), 1.14-1.40 (m, 6H), 1.55 (dt, J~7, 14 Hz, 2H), 1.82-2.36 (complex m, 10H), 2.72 (dt, J~2, 5Hz, 1H), 3.03 (dd, J~2, 8 Hz, 1H), 3.32 (s, 3H), 3.90 (dd, J~5.5, 11 Hz, 1H), 5.25-5.57 (m, 5H), 5.78 (dd, J~5.4, 15.5 Hz, 1H). 6a: ¹H NMR (C₆D₆, 250 MHz) δ 0.90 (t, J~6.5 Hz, 3H), 1.18-1.42 (m, 6H), 1.60(dt, J~7, 14 Hz, 2H), 1.82-3.95 (m, 1H), 4.15-4.30 (m, 2H), 5.35-5.68 (m, 4H), 5.91 (dd, J~5.5, 16 Hz, 1H), 6.03 (dd, J~5.5, 16 Hz, 1H). MMR (C₆D₆, 250 MHz) δ 0.90 (t, J~6.5 Hz, 3H), 1.18-1.42 (m, 6H), 1.60(dt, J~7, 14 Hz, 2H), 1.92-2.22 (m, 6H), 2.26-2.52 (m, 4H), 3.36 (s, 3H), 3.77-3.88 (m, 1H), 4.15-4.30 (m, 2H), 5.35-5.68 (m, 4H), 5.87 (dd, J~4.8, 15.6 Hz, 1H), 6.02 (dd, J~6.15.6 Hz, 1H).

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