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Synthesis of 7-Methoxyapigeninidin and Its Fungicidal Activity against Gloeocercospora sorghi

Yu Aida^a, Shigeru Tamogami^a, Osamu Kodama^a & Takao Tsukiboshi^{ab}

^a Laboratory of Bio-organic and Pesticide Chemistry, School of Agriculture, Ibaraki University, 3-21-1 Chuo, Ami, Ibaraki 300-03, Japan

^b Plant Pathology Laboratory, National Grassland Research Institute, Nishinasuno, Tochigi 329-27, Japan

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Note

Synthesis of 7-Methoxyapigeninidin and Its Fungicidal Activity against *Gloeocercospora* sorghi

Yu Aida, Shigeru Tamogami, Osamu Kodama,[†] and Takao Tsukiboshi*

Laboratory of Bio-organic and Pesticide Chemistry, School of Agriculture, Ibaraki University, 3–21–1 Chuo, Ami, Ibaraki 300–03, Japan

*Plant Pathology Laboratory, National Grassland Research Institute, Nishinasuno, Tochigi 329–27, Japan Received February 1, 1996

In the structure of sakuranetin, which was isolated as a phytoalexin from the rice plant, the methoxy group at C-7 has been shown to be important for its high activity. Apigeninidin was isolated as a phytoalexin from sorghum, but it had no methoxy group at C-7. We prepared 7-methoxyapigeninidin and compared its fungicidal activity with that of apigeninidin. The 7-methoxyapigeninidin showed higher activity against sorghum fungi than apigeninidin, suggesting that the methoxy group at C-7 was important for the high fungicidal activity.

Key words: phytoalexin; sakuranetin; naringenin; apigeninidin; 7-methoxyapigeninidin

From the rice plant, we have previously reported the isolation of the antifungal flavonoid sakuranetin (1; 5,4-dihydroxy-7methoxyflavanone) as a phytoalexin.¹⁾ Phytoalexins are antimicrobial secondary metabolites which accumulated in plants by microbial invasion. Sakuranetin 1 inhibits spore germination of *Pyricularia oryzae* at approximately 15 ppm.¹⁾ In contrast, naringenin (2; 5,7,4'-trihydroxyflavanone) which is a precursor of sakuranetin 1, shows much less fungicidal activity, scarcely inhibiting spore germination at up to 50 ppm.¹⁾ The only chemicalstructural difference between these two compounds is a hydroxy or methoxy group at the C-7 position. Therefore, we have speculated that methylation of this hydroxy group at C-7 may be important for high fungicidal activity.¹⁾

From the sorghum, apigeninidin (3; 5,7,4'-trihydroxyflavylium chloride) and luteolinidin were isolated as fungitoxic compounds and were considered to be sorghum phytoalexins.²⁾ Apigeninidin 3 and naringenin 2 have three hydroxy groups at the same positions, *i.e.*, the 5, 7, and 4' positions, but no methoxy group at C-7 (Fig. 1). According to our previous speculation, the fungitoxic activity of apigeninidin 3 may be increased by methylation at

C-7. Therefore, we synthesized apigeninidin 3 and 7-methoxy-apigeninidin (4; 5,4'-dihydroxy-7-methoxyflavylium chloride) and compared its fungicidal activity each other.

Apigeninidin 3 was synthesized from naringenin 2 by the method of Sweeny *et al.*³⁾ The 7-methoxyapigeninidin 4 was synthesized from sakuranetin 1 according to the same method (Fig. 2).³⁾ Sakuranetin 1 was prepared from naringenin 2 by treating with ethereal diazomethane under anhydrous conditions in a 75% yield. The analytical data for synthetic sakuranetin 1 showed good accordance with that of natural sakuranetin.¹⁾ After acetylating the hydroxy groups, sakuranetin 1 was converted to 7-methoxyapigeninidin 4 by NaBH₄ reduction and following chloranil dehydrogenation.³⁾ In the ¹H-NMR spectrum of 7-methoxyapigeninidin 4, aromatic protons which are characteristic to



Fig. 1. Structure of the Compounds.



Fig. 2. Synthetic Scheme for 7-Methoxyapigeninidin 4.

[†] To whom correspondence should be addressed.



Fig. 3. Antifungal Activity of 7-Methoxyapigeninidin 4 and Apigeninidin 3

Bars indicate SE. A, against radial growth of Gloeocercospora sorghi; B, against spore germination of Pvricularia orvzae

flavylium compounds and three singlet methyl protons were observed. These aromatic protons showed good accordance with those of apigeninidin 3 reported by Nilsson.⁴⁾ The ultraviolet absorption spectrum of 7-methoxyapigeninidin 4 was dependent on the pH of the solution. The maximum absorption of 7methoxyapigeninidin 4 was observed at 475 nm in a solution at pH 2, and at 550 nm in a solution at pH 10, these values are good accordance with those of apigeninidin 3.5) These characteristic UV absorbance data also validated the structure of synthetic 7methoxyapigeninidin 4

The I₅₀ concentration for inhibiting the radial growth of Gloeocercospora sorghi, by 7-methoxyapigeninidin 4 was 50 ppm, while apigeninidin 3 showed only 25% inhibition at 100 ppm (Fig. 3). Although the radial growth of Pyricularia oryzae was not inhibited by either apigeninidin 3 or 7-methoxyapigeninidin 4 at 100 ppm (data not shown), its spore germination was inhibited by 7-methoxyapigeninidin 4 with an I_{50} concentration of 50 ppm. Apigeninidin 3 was almost inactive against the germination of Pyricularia oryzae, even at a high concentration (Fig. 3). This fairly high fungicidal activity of 7-methoxyapigeninidin 4 against both species is very surprising when compared to that of apigeninidin 3.

These results suggest that the methoxy group at C-7 is very important for high activity in flavylium-type compound as in the case of flavone-type compound (sakuranetin 1 vs. naringenin 2). We have recently succeeded in purifying the enzyme which catalyzes methylation of the hydroxy group at C-7 in naringenin 2 to produce sakuranetin 1 from rice leaves, and discussed the importance of this enzyme to the plant self-defence mechanism.⁶⁾ Although the existence of the enzyme which catalyzes the methylation of apigeninidin 3 has not yet been confirmed nor the presence of 7-methoxyapigeninidin 4 in sorghum, it would be very interesting to investigate if sorghum has the same methylation mechanism as that in the rice plant in future.

Experimental

Fungal inhibition assays. The fungus was cultivated on a potato dextrose agar medium at 25°C for 1 week under dark. Experiments were initiated by placing plugs (1 cm in diameter), taken from the cultures of

Gloeocercospora sorghi, mycelial side up on the medium (8.5 cm diameter), containing appropriate amounts of synthetic apigeninidin 3 and 7-methoxyapigeninidin 4. Synthetic apigeninidin 3 and 7-methoxyapigeninidin 4 were added to the autoclaved medium when its temperature came down to below 60°C. The stability of these synthetic compounds in this medium was confirmed by a TLC analysis with a solvent of ethyl acetate-formic acid-water-HCl (85:6:8:1, v/v/v, upper phase).²⁾ After 1 week, the radial growth of the cultures was measured, and the fungicidal activity was evaluated against an untreated control as a percentage. The spore germination inhibitory activity against Pyricularia oryzae was assayed as previously described.1)

Methylation of naringenin 2. A diazomethane solution (prepared from p-toluenesulfonyl-N-methyl-N-nitrosamide and a 50% KOH solution in ether) was added to 1.0 g of naringenin 2 in 400 ml of dry diethylether. After stirring at room temperature overnight, the solution was concentrated to give a crude product. Silica-gel column chromatography (EtOAc-*n*-hexane = 1:3) gave 790 mg of sakuranetin 1 in 75% yield. ¹Hand ¹³C-NMR, and EI-MS data were in good accord with those of authentic sakuranetin.1)

Acetylation of sakuranetin 1. Five hundred mg of synthetic sakuranetin 1 was dissolved in 1 ml of pyridine and 1 ml of acetic anhydride and stirred at room temperature overnight. Removal of the excess solvents in vacuo gave a crude acylate, which was purified by silica-gel column chromatography (EtOAc-*n*-hexane = 1:3) gave 595 mg of acetylated sakuranetin 5 in a 92% yield. ¹H-NMR δ (500 MHz, CDCl₃): 2.28 (3H, s), 2.36 (3H, s), 2.70 (1H, dd, J = 16.0 Hz and 3.0 Hz), 2.96 (1H, dd, J = 16.0 and 13.0 Hz), 3.78 (3H, s), 5.42 (1H, dd, J = 13.0 and 3.0 Hz), 6.35 (1H, d, J = 2.8 Hz), 6.40 (1H, d, J = 2.8 Hz), 7.15 (2H, br. d, J = 8.8 Hz), 7.45 (2H, br. d, J=8.8 Hz); MS m/z: 370 (M⁺), 328 (100), 285, 167.

Reduction of acetylated sakuranetin 5. To a solution of 300 mg of acetylated sakuranetin 5 in 15 ml of tetrahydrofuran and 15 ml of ethanol, 60 mg of sodium borohydride was added. After stirring for 30 minutes; an additional 60 mg of sodium borohydride was added, and stirring was continued for 2h more at room temperature. The reaction solution was poured into 90 ml of cold 0.5% acetic acid in water and then extracted with chloroform (30 ml, three times). Drying (Na₂SO₄) and evaporating the solvent gave 350 mg of a product. Without purification, this product was treated with pyridine (1 ml) and acetic anhydride (1 ml) in the same manner as that previously described to give 250 mg of compound 6 in an 87% yield after silica-gel column chromatography (EtOAc-n-hexane = 1:3). ¹H-NMR δ (500 MHz, CDCl₃): 1.97 (1H, m), 2.15 (1H, m), 2.28 (6H, s), 2.60 (2H, m), 3.73 (3H, s), 4.98 (1H, br. d, J = 10.5 Hz), 6.28 (1H, d, J=2.5 Hz), 6.38 (1H, d, J=2.5 Hz), 7.10 (2H, d, J=8.2 Hz), 7.42 $(2H, d, J=8.2 \text{ Hz}); \text{ MS } m/z; 356 (M^+, 100), 314, 371, 153.$

Synthesis of 7-methoxyapigeninidin 4. A mixture of the compound 6 (200 mg), 700 mg of chloranil, 24 ml of HOAc, 5 ml of water, and 1.5 ml of 6 N HCl was kept for 1 h with stirring. After cooling, the reaction mixture was diluted with 250 ml of 0.01 N HCl in MeOH and passed through a column of polyclar AT (previously washed 3 times with 3 N HCl). The column was eluted with a second 250 ml of the same solvent. The combined eluates were concentrated in vacuo at 30°C to give 68 mg of 7methoxyapigeninidin 4 as a red solid (40% yield). ¹H-NMR δ (500 MHz, DMSO- d_6): 4.01 (3H, s), 6.82 (1H, br. s), 7.13 (2H, d, J = 10.5 Hz), 7.39 (1H, d, J=2.1 Hz), 8.32 (1H, d, J=10.5 Hz), 8.43 (2H, d, J=10.5 Hz),9.15 (1H, d, J=10.5 Hz). HR-FAB-MS (positive mode): found; 269.0839 $([M-Cl]^+, C_{16}H_{13}O_4)$; calculated, 269.0814.

References

- 1) O. Kodama, J. Miyakawa, T. Akatsuka, and S. Kiyosawa, Phytochemistry, 31, 3807-3809 (1992).
- J. D. Hipskind, R. Hanau, B. Lette, and R. L. Nicholson, Physiol. 2) Molec. Plant Pathol., 36, 381-396 (1990).
- 3) J. G. Sweeny and G. A. Iacobucci, Tetrahedron, 33, 2927-2932 (1977).
- 4) E. Nilsson, Chemica Scripta, 4, 49-55 (1973).
- 5)
- J. M. Baranac and D. S. Amic, J. Agric. Food Chem., 38, 2111-2115 (1990)
- R. Rakwal, M. Hasegawa, and O. Kodama, Biochem. Biophys. Res. 6) Commun., 222, 732-735 (1996).

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