

Synthesis of Indolylalkoxyiminoalkylcarboxylates as Leukotriene Biosynthesis Inhibitors

Teodozyj Kolasa,* Pramila Bhatia, Clint D. W. Brooks, Keren I. Hulkower, Jennifer B. Bouska, Richard R. Harris and Randy L. Bell

Immunoscience Research, D-47K, Abbott Laboratories, 100 Abbott Park, IL 60064-3500, U.S.A.

Abstract—A series of substituted indolylalkoxyiminoalkylcarboxylates were found to be potent leukotriene biosynthesis inhibitors. The structure-activity relationships were investigated. Representative potent inhibitors identified were the quinolyl **3a** (A-86885) and pyridyl **3b** (A-86886) congeners with in vitro IC₅₀s of 21 and 9 nM and in vivo leukotriene inhibition in the rat with oral ED₅₀s of 0.9 and 1.7 mg/kg, respectively. () 1997 Elsevier Science Ltd. All rights reserved.

Introduction

5-Lipoxygenase (5-LO) catalyzes the first two steps of the transformation of arachidonic acid into leukotrienes, known mediators of inflammation.¹ The 5-LO enzyme undergoes activation through an interaction with five-lipoxygenase activating protein (FLAP) at a nuclear membrane site.² Translocation of 5-LO to this activation site and the role of FLAP as an arachidonic acid carrier protein which enhances oxidative catalysis by 5-LO have been described.³ Agents that interfere with the function of FLAP prevent leukotriene biosynthesis in intact cell systems. The reference FLAP



inhibitor, MK-0591 (1) has demonstrated potent inhibition of leukotriene biosynthesis in both preclinical and clinical studies.⁴⁻⁵

In the course of structure–activity evaluations of various substituted indoles, we found that insertion of an oxime function into the alkylcarboxylate substituent of the reference inhibitor provided a congener 2, with improved oral bioavailability.⁶ This paper extends that initial finding by describing the biological properties of an interesting series of inhibitors exemplified by 3, with the regio-isomeric oxime form referred to as the indolylalkoxyiminoalkylcarboxylate derivatives.

Chemistry

The indole intermediate 4 (Scheme 1, see also Experimental for preparation) was the key starting point for the synthesis of several potent leukotriene inhibitors. Alkylation of 4 with the requisite chloromethylheteroaryl group afforded the substituted indole esters 5 that were reduced to provide the corresponding alcohols 6. A Mitsunobu reaction⁷ with N-hydroxyphthalimide followed by hydrazinolysis of the phthaloyl derivative converted the alcohols 6 into the alkoxyamines 7, which were transformed into the indolylalkoxyiminoalkylcarboxylate derivatives 3a-f by reaction with the requisite carbonyl component under standard oxime formation conditions. The compounds were obtained as single isomers, presumably with the E oxime configuration. The absence of isolable amounts of the other isomers (Z) precluded confirmation of the configuration proposed.

The 2-alkyl substituent of the indole was simplified to a methylene unit as in indolylmethoxyiminoalkylcarboxylate analogues (11a, b and 12a, b) that were synthesized from indole 8 (Scheme 2). Treatment with



Scheme 1. Reagents and conditions: (a) chloromethylheterocycle, K₂CO₃, DMF, 60 °C; (b) NaBH₄, CaCl₂, THF, EtOH, 0 °C to rt; (c) PhtNOH, Ph₃P, DIAD, THF; (d) H₂NNH₂, EtOH, reflux; (e) RCOCO₂H, AcOH, THF, H₂O; (f) NaOH, dioxane, MeOH, rt.

AlCl₃ in *t*-butylthiol provided the corresponding hydroxyindole intermediate that was directly alkylated with 2-chloromethylquinoline to provide a mixture of quinolylmethoxyindole 9 in 32% yield and the bisquinoline derivative 10 in 50% yield. These intermediates were separated and further transformed into the desired oxime derivatives 11a, b (isolated as single isomers and the precise stereochemistry was not assigned) and 12a, b according to the previously described methods. The 12a was separated as a single isomer, probably Z, compared to the Z:E (4:1) ratio of less crowded 12b. The preferred configuration as Z for these oximes is probably a result of the 3-thiomethylquinolyl substituent.

A series of analogues was prepared without the 3thioalkyl substituent as shown in Scheme 3. Indolecarboxylate 13 was converted by standard methods into 1-(chlorobenzyl)-5-methoxyindole-2-carboxylic acid methyl ester 14 with 65% overall yield. The 2-quinolylmethoxy substituent was added as before and the resulting intermediate indole ester was saponified and reduced to the corresponding alcohol 15, which was converted to the alkoxyamine by the methods previously described. The oxime derivatives **17a**, **b** were prepared by reaction with the requisite carbonyl derivative under standard conditions. The major isomers were assigned as Z based on NMR studies. The **17a** was isolated from a mixture of 25:1, Z:E isomers and **17b** was separated from a mixture of 49:1, Z:E isomers.

A series of secondary alcohol derivatives **16a–d** was prepared from the primary alcohol **15** by oxidation to the corresponding aldehyde followed by Grignard reaction (Scheme 3). The resulting secondary alcohols **16a–d** were then transformed into the racemic oxime derivatives **17c–g** as described previously. The compounds were isolated as single isomers, probably E (by 300 MHz ¹H NMR).

Results and Discussion

We previously demonstrated that inserting an oxime moiety into the 2-alkylcarboxylate substituent of **1** provided a leukotriene biosynthesis inhibitor **2** with improved oral bioavailability and in vivo pharmacological properties.⁶ This result led us to examine the biological properties of the regioisomeric oxime form



Scheme 2. Reagents and conditions: (a) AlCl₃, *t*-BuSH, CH₂Cl₂, 0 °C, (91%); (b) 2-chloromethylquinoline, K₂CO₃, DMF, rt, 9 (32%) and 10 (50%); (c) NaOH, dioxane, MeOH, rt; (d) ClCO₂Et, Et₃N, THF, -15 °C, then NaBH₄, MeOH, 0 °C; (e) PhtNOH, Ph₃P, DEAD, THF; (f) H₂NNH₂, dioxane, EtOH, reflux; (g) RCOCO₂Et, AcOH, MeOH, dioxane; (h) NaOH, dioxane, MeOH, rt, overall yield (c-h): 11a (21%), 11b (73%), 12a (53%), and 12b (66%).



Scheme 3. Reagents: (a) MeI, NaHCO₃, DMF (69%); (b) *p*-ClBzlCl, NaH, DMF (94%); (c) AlCl₃, *t*-BuSH, CH₂Cl₂ (97%); (d) 2-chloromethylquinoline, K_2CO_3 , DMF (64%); (e) NaOH, dioxane, MeOH (95%); (f) ClCO₂Et, Et₃N, THF, -15 °C, then NaBH₄, MeOH, 0 °C (93%); (g) Ba(MnO₄)₂, toluene, reflux (88%); (h) RMgX, THF, -78 °C; (i) PhtNOH, Ph₃P, DEAD, THF; (j) H₂NNH₂, dioxane, EtOH; (k) CH₃COCO₂CH₃, AcOH, dioxane, methanol; (l) NaOH, dioxane, MeOH. Overall yield from 15: 17a (10%), 17b (4%), 17c (15%), 17d (13%), 17e (35%), 17f (9%), and 17g (11%).

 Table 1. Leukotriene inhibition results

Compound	Human neutrophil ^a IC ₅₀ , nM (95% CL)	In vivo rat anaphylaxis ^b ED ₅₀ , mg/kg (95% CL) or % inh (± SEM)
MK-0591	8 (7-10)	1.6 (0.8-2.5)
A-81834	12 (10-20)	0.24(0.05-0.50)
3a	21 (20-20)	0.9 (0.7–3.7)
3b	9 (10–10)	1.7 (2.1–3.5)
3c	10 (10-20)	$100 (\pm 0.4)$
3d	18 (10-20)	99 (± 0.7)
3e	32 (30-40)	100 (0.0)
3f	8 (10-10)	100 (0.0)
11a	10 (10-20)	65 @ 5 µmol/kg
11b	30 (20-40)	$100 (\pm 1.4)$
12a	70 (60-80)	36 (± 89.8)
12b	110 (90–130)	$0(\pm 32.8)$
17a	17 (12-21)	_
17b	24 (21–27)	_
17c	20 (13-25)	92 (± 9.4)
17d	40 (30-70)	$100 (\pm 0.4)$
17e	100 (80-130)	100 (0.0)
17f	25 (15-35)	_
17g	70 (50–160)	-

^aInhibition of calcium ionophore (A-23187) stimulated LTB₄ formation in human neutrophils, 95% confidence limits shown in parentheses.

^bRat anaphylaxis in the peritoneal cavity, leukotrienes formed measured by enzyme-linked immunoassay, oral dose response determination ED_{50} or percentage leukotriene inhibition at a single dose of 30 µmol/kg with n = 8,95% confidence limits shown in parentheses for oral dose response determinations or SEM for single dose studies. See ref 8 for a detailed description of the biological methods.

represented by the current series of substituted indolylalkoxyiminoalkylcarboxylates. The biological testing included a human neutrophil cellular assay involving calcium ionophore stimulated LTB4 biosynthesis to measure in vitro leukotriene inhibition.⁸ A rat anaphylaxis model involving antigen-antibody-stimulated leukotriene generation in the peritoneal cavity was used to evaluate in vivo leukotriene inhibition after oral administration of test compound.⁸ The biological activity of these analogues in these two assays is shown in Table 1. The indolylalkoxyiminoalkylcarboxylate analogues 3a-f, exhibited comparable in vitro inhibitory activity in the intact human neutrophil assay as 1. These results indicated that several heteroaryl templates including quinolyl, pyridyl, thiazolyl, and benzothiazolyl provided potent leukotriene biosynthesis inhibitors. In the rat anaphylaxis model, 3a was slightly more potent than 1 but less active than the oxime congener 2. Preliminary pharmacokinetic evaluation of 3a in dog, given orally at 7 mg/kg, showed a low plasma concentration $(T_{\text{max}} = 1.5 \text{ h}, C_{\text{max}} = 0.67 \,\mu\text{M})$ and an estimated oral half-life of 3 h. For comparison, 2 given orally at 10 mg/kg provided more than 90% ex vivo inhibition of LTB₄ over an 8-h period. The change in oxime regiochemistry (comparing 2 with 3a) invokes signifcant changes in orientation of the respective heteroatoms which may result in potency differences. However only minor differences between 2 and 3a were found both in vitro (IC₅₀ values of 12 and 21 nM) and in vivo (ED₅₀ values of 0.24 and 0.9 mg/kg). These results would imply that the oxime function in these inhibitors does not have specific strong binding interaction sites with FLAP.

510

Several other structural changes were examined. The nature of the alkyl group in the 2-position of the indole was examined due to the proximity to the newly introduced oxyimino moiety. The methoxyimino analogues 11a and 11b were also found to be potent leukotriene inhibitors. Replacement of the 3-indolyl-St-butyl group with more polar S-2-quinolylmethyl group as in 12a and 12b resulted in a decrease in in vitro inhibitory activity and a loss of in vivo activity. A series of analogues without the 3-indolyl-S-t-butyl substituent was evaluated. Analogues 17a-g had in vitro activity in the 17-100 nM range and for those tested (17c-e) in vivo provided greater than 90% leukotriene inhibition in the rat with a single 30 μ mol/kg oral dose 1 h prior to leukotriene measurement. In conclusion the regioisomeric oxime insertion led to congeners with potent in vitro and in vivo leukotriene inhibitory activity.

Experimental

Melting points were taken on a Thomas-Hoover melting point apparatus and are uncorrected. ¹H NMR spectra were recorded using a Nicolet QE-300 (300 MHz) instrument. Mass spectra were obtained with Hewlett Packard HP5985 spectrometer. Microanalyses were performed by the Robertson Microlit Laboratories, Inc., Madison, NJ, USA. Reagents were obtained from Aldrich and Lancaster chemical companies.

3-[3-t-Butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]-2,2-dimethylpropyloxyiminoacetic acid (3a). To a suspension of 4-methoxyphenylhydrazine:HCl (41 g, 230 mmol) in CH₂Cl₂ (1 L) under nitrogen was added diisopropylamine (79.8 g, 612 mmol), 4-chlorobenzyl chloride (40.25 g, 250 mmol), and tetrabutylammonium bromide (22.8 g, 70 mmol), and the mixture was stirred at rt for 48 h. The reaction mixture was washed with water, brine, dried (MgSO₄) and concentrated in vacuo. The residue was purified by chromatography (silica gel, 2.5%) methanol in CH₂Cl₂) to provide 1-(4-chlorobenzyl)-1-(4-methoxyphenyl)hydrazine (43.5 g, 76%). Mp 55 °C. This hydrazine (38.25 g, 150 mmol) in toluene (250 mL) was treated with acetic acid (175 mL) and ethyl 5-t-butylthio-2,2-dimethyl-4-oxo-pentanoate (38.2 g, 150 mmol) and the resulting mixture was stirred in the dark at rt for four days. Water was added, the organic layer separated and washed with water, brine, dried (MgSO₄), and concentrated in vacuo. The residue was purified by chromatography (silica gel, 4:1, hexane: EtOAc) to provide ethyl 3-[3-t-butylthio-1-(4-chlorobenzyl)-5-methoxyindol-2-yl]-2,2-dimethylpropionate (36.5 g, 50%). To a suspension of AlCl₃ (26.3 g, 50%)198 mmol) in t-butanthiol (60 mL) at 0 °C was added a solution of the ethyl ester (32 g, 66 mmol) in CH_2Cl_2 (90 mL) and the mixture was stirred at 0 °C for 10 min and at rt for 3 h. The mixture was poured on ice and acidified with 10% HCl. The organic layer was washed with water, brine, dried (MgSO₄), and concentrated in vacuo to provide ethyl 3-[3-t-butylthio-1-(4-chlorobenzyl)-5-hydroxyindol-2-yl]-2,2-dimethylpropionate (4) (21.8 g, 70%).

A mixture of chloromethylquinoline hydrochloride $(1.07 \text{ g}, 5 \text{ mmol}), K_2 CO_3 (2.07 \text{ g}, 15 \text{ mmol}) \text{ and } 4$ (2.37 g, 5 mmol) in DMF (30 mL) was refluxed at 60 °C for 3 h and stirred at rt for 15 h. The mixture was poured into water and extracted with ethyl acetate (80 mL), dried (MgSO₄) and concentrated in vacuo to provide ethyl 3-[3-t-butylthio-1-(4-chlorobenzyl)-5-(2quinolylmethoxy)indol-2-yl]-2,2-dimethylpropionate (5) (2.6 g, 84%). To a solution of 5 (2.5 g, 4 mmol) in ethanol (30 mL) and THF (20 mL) was added powdered $CaCl_2$ (0.9 g, 8 mmol) and NaBH₄ (0.6 g, 16 mmol) and the mixture was stirred at 0 °C for 2 h and at rt for 14 h. The mixture was neutralized with 6 N HCl and extracted with ethyl acetate, washed with water, brine, dried (MgSO₄), and concentrated in vacuo to provide 3-[3-t-butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]-2,2-dimethylpropan-1-ol (6) (2.3 g, 99%). To a mixture of the alcohol 6 (2.3 g, 99%)4 mmol), N-hydroxyphthalimide (0.75 g, 4.5 mmol) and triphenylphosphine (1.31 g, 5 mmol) in THF (100 mL) was added dropwise diisopropyl azodicarboxylate (DIAD, 1.01 g, 5 mmol) and the mixture was stirred at rt for 2 h. The mixture was concentrated and purified by chromatography (silica gel, 95:5, CH₂Cl₂:EtOAc) to provide N-phthaloyl-O-[3-[3-t-butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)-indol-2-yl]-2,2-dimethylpropyl]hydroxylamine. This N-phthaloyl derivative and hydrazine hydrate (1 mL) in ethanol was refluxed for 1 h and then concentrated in vacuo. The residue was partitioned between CH₂Cl₂ and 10% aqueous Na_2CO_3 , and the organic layer was washed with water, brine, dried (MgSO₄), and concentrated in vacuo to provide O-[3-[3-t-butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)-indol-2-yl]-2,2-dimethylpropyl]hydroxylamine (7) (1.75 g, 80%). To amine 7 (700 mg, 1.3 mmol) in THF (35 mL) and water (10 mL) was added glyoxylic acid (240 mg, 2.6 mmol) and acetic acid (0.2 mL) and the mixture was stirred at rt for 14 h. The volatile organics were removed in vacuo and the residue was diluted with water to precipate a solid which was collected and purified by chromatography (silica gel, 9:1, CH₂Cl₂:MeOH) to provide **3a** (420 mg, 50%). Mp 152-155 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 0.87 (s, 6 H), 0.97 (s, 9 H), 2.90 (m, 2 H), 3.83 (s, 2 H), 5.39 (s, 2 H), 5.53 (s, 2 H), 6.83 (m, 3 H), 7.13 (d, J = 3 Hz, 1 H), 7.30 (m, 3 H), 7.40 (s, 1 H), 7.65 (m, 2 H), 7.78 (m, 1 H), 7.95 (d, J = 8 Hz, 1 H), 8.05 (d, J = 8 Hz, 1 H), 8.35 (d, J = 8 Hz, 1 H); MS (DCI-NH₃) m/z 644 $(M+H)^+$. Anal. calcd for $C_{36}H_{38}ClN_3O_4S\cdot 2H_2O$: C, 63.52; H, 5.88; N, 6.17. Found: C, 63.04; H, 5.95; N, 5.88.

3-[3-(*t***-Butylthio)-1-(4-chlorobenzyl)-5-(2-pyridylmethoxy)indol-2-yl]-2,2-dimethylpropyloxyiminoacetic acid (3b).** Prepared by the method described for **3a**. ¹H NMR (300 MHz, DMSO-*d*₆) δ 0.91 (s, 6 H), 1.10 (s, 9 H), 2.93 (m, 2 H), 3.85 (s, 2 H), 5.20 (s, 2 H), 5.53 (s, 2 H), 6.83 (d, *J* = 9 Hz, 3 H), 7.09 (d, *J* = 3 Hz, 1 H), 7.32 (m, 4 H), 7.41 (s, 1 H), 7.49 (d, *J* = 8 Hz, 1 H), 7.79 (m, 1 H), 8.56 (m, 1 H); MS (DCI–NH₃) m/z 594 (M+H)⁺. Anal. calcd for C₃₂H₃₆ClN₃O₄S·2H₂O: C, 60.95; H, 6.20; N, 6.66. Found: C, 60.74; H, 5.89; N, 6.54.

3-[3-(*t***-Butylthio)-1-(4-chlorobenzyl)-5-(4-thiazolylmethoxy)indol-2-yl]-2,2-dimethylpropyloxyiminoacetic acid (3c)**. Prepared by the method described for **3a**. Mp 125 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 0.92 (s, 6 H), 1.15 (s, 9 H), 2.95 (m, 2 H), 4.05 (s, 2 H), 5.22 (s, 2 H), 5.53 (s, 2 H), 6.85 (m, 3 H), 7.18 (d, *J*=3 Hz, 1 H), 7.31 (m, 3 H), 7.58 (s, 1 H), 7.73 (s, 1 H), 9.13 (s, 1 H); MS (FAB(-)) *m/z* 598 (M-H)⁺. Anal. calcd for C₃₀H₃₄ClN₃O₄S₂: C, 59.89; H, 5.68; N, 6.99. Found: C, 59.31; H, 5.49; N, 6.81.

3-[3-t-Butylthio-1-(4-chlorobenzyl)-5-(2-benzothiazolylmethoxy)indol-2-yl]-2,2-dimethylpropyloxyiminoacetic acid (3d). Prepared by the method described for **3a**. Mp 154 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 0.92 (s, 6 H), 1.04 (s, 9 H), 2.95 (m, 2 H), 4.03 (s, 2 H), 5.53 (s, 2 H), 5.61 (s, 2 H), 6.85 (m, 3 H), 7.21 (d, *J*=3 Hz, 1 H), 7.31 (m, 3 H), 7.44 (m, 1 H), 7.54 (m, 1 H), 7.57 (s, 1 H), 8.03 (d, *J* = 9 Hz, 1 H), 8.08 (d, *J* = 9 Hz, 1 H); MS (FAB(+)) *m/z* 650 (M+H)⁺. Anal. calcd for C₃₄H₃₆ClN₃O₄S₂: C, 62.79; H, 5.58; N, 6.46. Found: C, 62.70; H, 5.53; N, 6.30.

3-[3-t-Butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]-2,2-dimethylpropyloxyimino-2-propionic acid (3e). A mixture of 7 (700 mg, 1.3 mmol), methyl pyruvate (265 mg, 2.6 mmol), and acetic acid (0.2 mL) in methanol (20 mL), THF (35 mL), and water (10 mL) was stirred at rt for 12 h. The mixture was concentrated in vacuo and the residue was extracted with ethyl acetate, washed with water, dried (MgSO₄), and concentrated in vacuo. The residue was dissolved in methanol (25 mL) and treated with 1 N NaOH (3 mL) at rt for 14 h, concentrated in vacuo and the residue was acidified with 6 N HCl, extracted with ethyl acetate and chromatography purified by (silica gel. 9:1. CH₂Cl₂:MeOH) followed by crystallization from ethyl acetate-hexane to provide 3e (375 mg, 44%). Mp 145-147 °C; ¹H NMR (300 MHz; DMSO- d_6) δ 0.89 (s, 6 H), 0.97 (s, 9 H), 1.85 (s, 3 H), 2.95 (m, 2 H), 3.86 (s, 2 H), 5.40 (s, 2 H), 5.52 (s, 2 H), 6.80 (d, J = 8 Hz, 1 H), 6.86 (d-d, J = 9 Hz and 3 Hz, 1 H), 7.14 (d, J = 3 Hz, 1 H),7.30 (m, 3 H), 7.62 (m, 2 H), 7.78 (m, 1 H), 7.95 (d, J = 8 Hz, 1 H), 8.05 (d, J = 8 Hz, 1 H), 8.35 (d, J = 8 Hz, 1 H); MS (DCI-NH₃) m/z 658 (M+H)⁺. Anal. calcd for C₃₇H₄₀ClN₃O₄S·2H₂O: C, 64.06; H, 6.14; N, 6.06. Found: C, 63.56; H, 5.70; N, 6.01.

3-[3-t-Butylthio-1-(4-chlorobenzyl)-5-(4-thiazolylmethoxy)indol-2-yl]-2,2-dimethylpropyloxyimino-2-propionic acid (3f). Prepared by the method described for **3e**. Mp 110 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 0.95 (s, 6 H), 1.15 (s, 9 H), 1.93 (s, 3 H), 3.0 (m, 2 H), 4.05 (s, 2 H), 5.22 (s, 2 H), 5.53 (s, 2 H), 6.83 (m, 3 H), 7.18 (d, J = 3 Hz, 1 H), 7.30 (m, 3 H), 7.72 (d, J = 1.5 Hz, 1 H), 9.12 (d, J = 1.5 Hz, 1 H); MS (DCI-NH₃) m/z 614 (M+H)⁺. [3-t-Butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]methoxyiminoacetic acid (11a). A mixture of 4-methoxyphenylhydrazine:HCl (5.95 g, 34 mmol) and ethyl 3-(t-butylthio)pyruvate (7.0 g, 34 mmol) in tbutanol (70 mL) was refluxed for 48 h, and then concentrated in vacuo. The residue was partioned between water and ethyl acetate and the organic layer was washed with water, brine, dried (MgSO₄), and concentrated in vacuo. The residue was purified by chromatography (silica gel, 2:1, hexane:EtOAc) to afford ethyl [3-(t-butylthio)-5-methoxyindo-2-yl]carboxylate (3.5 g, 34%). ¹H NMR (300 MHz, CDCl₃) δ 1.32 (s, 9 H), 1.45 (t, J = 7 Hz, 3 H), 3.88 (s, 3 H), 4.44 (q, J = 7 Hz, 2 H), 7.01 (d-d, J = 9 and 3 Hz, 1 H), 7.31(m, 2 H), 9.10 (br s, 1 H); MS (DCI-NH₃) m/z 308 $(M+H)^+$, 325 $(M+NH_4)^+$. To a solution of this ester in DMF (50 mL) at 0 °C was added NaH (60% suspension in mineral oil, 460 mg, 11.5 mmol) and the mixture was stirred at rt for 15 min. Then p-chlorobenzyl chloride (1.93 g, 12 mmol) was added, the mixture was stirred at rt for 14 h, poured into water (200 mL), and extracted with ethyl acetate. The organic extract was washed with water, brine, dried (MgSO₄), and concentrated in vacuo. The residue was purified by chromatography (silica gel, 8:1, hexane:EtOAc) to provide ethyl [3-t-butylthio-1-(4-chlorobenzyl)-5-methoxyindol-2-yl]carboxylate (8) (4.1 g, 84%). ¹H NMR (300 MHz, $CDCl_3$) δ 1.30 (s, 9 H), 1.37 (t, J = 7 Hz, 3 H), 3.88 (s, 3 H), 4.35 (q, J = 7 Hz, 2 H), 5.65 (s, 2 H), 6.95 (m, 3 H), 7.20 (m, 3 H), 7.33 (d, J = 3 Hz, 1 H); MS (DCI-NH₃) m/z 432 (M+H)⁺.

To a suspension of AlCl₃ (4.8 g, 36 mmol) in CH_2Cl_2 (15 mL) at 0 °C was added *t*-butylthiol (12 mL) followed by addition of 8 (3.77 g, 11 mmol) and the mixture was stirred for 45 min. The mixture was poured into ice-cold 1 N HCl (100 mL), extracted with ethyl acetate and concentrated in vacuo to provide crude ethyl [3-t-butylthio-1-(4-chlorobenzyl)-5-hydroxyindol-2-yl]carboxylate (4.2 g). To a solution of this hydroxyindole in DMF (60 mL) was added K₂CO₃ (1.66 g, 12 mmol) and 2-chloromethylquinoline (2.1 g, 12 mmol) and the mixture was stirred at 50 °C for 14 h. The mixture was particulated between water and ethyl acetate, and the organic extract was washed with water, brine, dried (MgSO₄), and concentrated in vacuo. The residue was chromatographed (silica gel, 2:1, hexane:EtOAc) to afford ethyl [3-t-butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl] carboxylate (9) (1.7 g, 32%). ¹H NMR (300 MHz, CDCl₃) δ 1.15 (s, 9 H), 1.35 (t, J = 7 Hz, 3 H), 4.34 (q, J = 7 Hz, 2 H), 5.48 (s, 2 H),5.63 (s, 2 H), 6.95 (d, J = 9 Hz, 2 H), 7.12 (d-d, J = 9and 3 Hz, 1 H), 7.22 (m, 3 H), 7.44 (d, J = 3 Hz, 1 H), 7.56 (t, J = 8 Hz, 1 H), 7.73 (m, 2 H), 7.82 (d, J = 8 Hz, 1 H), 8.11 (m, 1 H), 8.20 (d, J = 8 Hz, 1 H); MS (DCI-NH₃) m/z 559 (M+H)⁺ and ethyl [1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)-3-(2-quinolylmethylthio)indol-2yl]carboxylate 10 (3.0 g, 50%). A mixture of 9 (1.7 g, 3 mmol) and 1 N NaOH (10 mL, 10 mmol) in dioxane (15 mL) and methanol (30 mL) was refluxed for 6 h at 50 °C. The volatile organics were then removed in vacuo and the residue was acidified to pH 3 with

aqueous 10% citric acid. The resulting solid was collected by filtration and dried in vacuo to afford [3t-butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]carboxylic acid (1.15 g, 72%). ¹H NMR (300 MHz, DMSO-*d*₆) δ 0.98 (s, 9 H), 5.43 (s, 2 H), 5.69 (s, 2 H), 7.02 (d, J = 9 Hz, 2 H), 7.11 (d–d, J = 9 and 3 Hz, 1 H), 7.21 (d, J = 3 Hz, 1 H), 7.34 (d, J = 9 Hz, 2 H), 7.50 (d, J = 9 Hz, 1 H), 7.63 (m, 2 H), 7.80 (m, 1 H), 7.96 (m, 1 H), 8.04 (d, J = 8 Hz, 1 H), 8.36 (d, J = 8 Hz, 1 H); MS (DCI-NH₃) m/z 531 (M+H)⁺. To a solution of this acid (360 mg, 0.66 mmol) in THF (20 mL) at -15 °C was added triethylamine (0.1 mL, 0.7 mmol) followed by dropwise addition of ethyl chloroformate (0.07 mL, 0.7 mmol). The mixture was stirred at -15 to -10 °C for 20 min and then NaBH₄ (76 mg, 2 mmol) was added and the mixture was warmed to 0 °C. Methanol (10 mL) was then added very slowly over a 20-min period. The mixture was acidified to pH 4 and extracted with ethyl acetate (70 mL), washed with water, brine, and concentrated in vacuo to provide [3t-butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]methanol (340 mg, 99%). To a mixture of this alcohol, triphenylphosphine (262 mg, 1 mmol), and Nhydroxyphthalimide (108 mg, 0.66 mmol) in THF (25 mL) was added dropwise diethylazodicarboxylate (DEAD, 0.16 mL, 1 mmol). The mixture was left at rt for 15 h and then concentrated in vacuo. The residue was purified by chromatography (silica gel, 3:1, hexane:EtOAc) to afford [3-t-butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]methoxy-Nphthaloylamine (360 mg, 82%). A mixture of the Nphthaloyl intermediate (300 mg, 0.44 mmol) and hydrazine hydrate (0.05 mL, 0.9 mmol) in ethanol (10 mL) and dioxane (5 mL) was refluxed for 30 min and then cooled to rt. Aq 10% Na₂CO₃ was added and the mixture was extracted with ethyl acetate to afford crude [3-t-butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]methoxyamine (230 mg, 98%). ¹H NMR (300 MHz, CDCl₃) δ 1.17 (s, 9 H), 4.97 (s, 2 H), 5.34 (s, 2 H), 5.45 (m, 4 H), 6.92 (d, J = 9 Hz, 2 H), 7.00 (d-d, J = 9 and 3 Hz, 1 H), 7.10 (d, J = 9 Hz, 1 H), 7.24(d, J = 9 Hz, 2 H), 7.40 (d, J = 3 Hz, 1 H), 7.55 (m, 1 H), 7.72 (m, 2 H), 7.81 (m, 1 H), 8.10 (m, 1 H), 8.18 (d, J = 8 Hz, 1 H); MS (DCI-NH₃) m/z 532 (M+H)⁺. To this amine in methanol (25 mL) was added glyoxylic acid (92 mg, 1 mmol) and acetic acid (0.06 mL, 1 mmol) and the mixture was stirred at rt for 14 h, then poured into water and extracted with ethyl acetate. The organic extract was washed with water, brine, dried (MgSO₄), and concentrated in vacuo. The residue was purified by chromatography (silica gel, 4:1, CH₂Cl₂:Et-OH) to afford 11a (100 mg, 37%). Mp 155-157 °C (methylene chloride:ethyl ether); ¹H NMR (300 MHz; $\dot{D}MSO-d_6$) δ 1.00 (s, 9 H), 5.32 (s, 2 H), 5.42 (s, 2 H), 5.5 (s, 2 H), 7.01 (m, 3 H), 7.16 (d, J = 3 Hz, 1 H), 7.19 (s, 1 H), 7.33 (m, 3 H), 7.62 (m, 2 H), 7.79 (m, 1 H), 7.96 (d, J = 8 Hz, 1 H), 8.04 (d, J = 8 Hz, 1 H), 8.35 (d, J = 8 Hz, 1 H); MS (FAB(+)) m/z 588 (M+H)⁺.

[3-t-Butylthio-1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]methoxyimino-2-propionic acid (11b). A mixture of O-(3-(t-butylthio)-1-(4-chlorobenzyl)-5-(2-

quinolylmethoxy)indole-2-methyl)hydroxylamine (266 mg, 0.5 mmol), methyl pyruvate (0.06 mL, 0.6 mmol), and acetic acid (0.04 mL, 0.6 mmol) in dioxane (10 mL), methanol (10 mL), and water (2 mL) was stirred at room temperature for 12 h to provide the desired oxime derivative that was converted by the method described for 3e to provide the desired product 11b (150 mg, 50%). Mp 135–137 °C (decomp); ¹H NMR (300 MHz, DMSO- d_6) δ 1.02 (s, 9 H), 1.53 (s, 3 H), 5.42 (s, 2 H), 5.52 (s, 2 H), 5.57 (s, 2 H), 6.93 (d, J = 9 Hz, 2 H), 7.0 (d-d, J = 9 and 3 Hz, 1 H), 7.20 (d, J = 3 Hz, 1 H), 7.34(m, 3 H), 7.65 (m, 2 H), 7.80 (m, 1 H), 7.96 (d-d, J = 8and 2 Hz, 1 H), 8.05 (d, J = 8 Hz, 1 H), 8.36 (d, J = 8 Hz, 1 H); MS (DCI-NH₃) m/z 602 (M+H)⁺; IR (in CDCl₃) 3440, 1760, 1710, 1620, 1600 cm⁻¹. Anal. calcd for C₃₃H₃₂ClN₃O₄S: C, 65.82; H, 5.36; N, 6.98. Found: C, 65.49; H, 5.30; N, 6.74.

[1-(4-Chlorobenzyl)-5-(2-quinolylmethoxy)-3-(2-quinolylmethylthio)indol-2-yl]methoxyimino-2-propionic acid (12a). Prepared from 10 (2.9 g, 4.5 mmol) according to the methods described for 11b to provide after recrystallization from ethyl acetate:hexane 12a (360 mg, 53% overall). Mp 110–113 °C (dec); ¹H NMR (300 MHz, DMSO-d₆) δ 1.42 (s, 3 H), 4.15 (s, 2 H), 5.16 (s, 2 H), 5.2 (s, 2 H), 5.45 (s, 2 H), 6.8 (d, *J* = 9 Hz, 1 H), 6.93 (m, 1 H), 7.05 (d, *J* = 3 Hz, 1 H), 7.22 (m, 4 H), 7.5 (m, 1 H), 7.63 (m, 3 H), 7.82 (m, 4 H), 8.0 (d, *J* = 8 Hz, 1 H), 8.08 (t, *J* = 8 Hz, 2 H), 8.41 (d, *J* = 8 Hz, 1 H); MS (DCI-NH₃) *m*/*z* 688 (M+H)⁺, 705 (M+NH₄)⁺; IR (KBr pellet): 3440, 1710, 1615, 1600 cm⁻¹. Anal. calcd for C₃₉H₃₁ClN₄O₄S: C, 68.16; H, 4.55; N, 8.15. Found: C, 68.11; H, 4.58; N, 7.79.

[1-(4-Chlorobenzyl)-5-(2-quinolylmethoxy)-3-(2-quinolylmethylthio)indol-2-yl]methoxyimino-4-pentanoic acid (12b). Prepared from 10 according to the methods described for 11b using ethyl levulinate for oxime formation resulting in 12b as a 4:1 mixture of Z and E isomers (120 mg, 66%). ¹H NMR (300 MHz, DMSOd₆) δ 1.36 and 1.70 (two s, 4:1, 3 H), 2.29 (m, 4 H), 4.11 (s, 2 H), 4.92 (m, 2 H), 5.18 and 5.20 (two s, 1:4, 2 H), 5.42 (broad s, 2 H), 6.82 (d, J = 9 Hz, 2 H), 6.90 (m, 1 H), 7.05 and 7.08 (two d, 1:4, J = 3 Hz, 1 H), 7.24 (m, 4 H), 7.50 (m, 1 H), 7.65 (m, 3 H), 7.82 (m, 3 H), 8.00 (d, J = 8 Hz, 1 H), 8.10 (m, 2 H), 8.41 (d, J = 8 Hz, 1 H), 12.09 (br s, 1 H); MS (DCI-NH₃) m/z 715 $(M+H)^+$; IR (in CDCl₃) 3680, 1710, 1620, 1600 cm⁻¹. Anal. calcd for C41H35CIN4O4S: C, 68.85; H, 4.93; N, 7.83. Found: C, 68.11; H, 4.90; N, 7.53.

(Z)-2-{[1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]methoxyimino}propionic acid (17a). A mixture of 5-methoxyindole-2-carboxylic acid (13) (9.55 g, 50 mmol), NaHCO₃ (4.2 g, 50 mmol), and iodomethane (12.5 mL, 200 mmol) in DMF (50 mL) was stirred at rt for 28 h and then poured into ice-water (500 mL). The mixture was extracted with ethyl acetate (200 mL), washed with water, brine, dried (MgSO₄), and concentated in vacuo to provide 5-methoxyindole-2-carboxylic acid methyl ester (7.1 g, 69%). ¹H NMR (300 MHz, DMSO- d_6) δ 3.76 (s, 3 H), 3.86 (s, 3 H), 6.91 (d-d, J = 9 and 3 Hz, 1 H), 7.06 (m, 1 H), 7.11 (d, J = 3 Hz, 1 H), 7.34 (d, J = 9 Hz, 1 H), 11.79 (br s, 1 H); MS (DCI-NH₃) m/z 206 (M+H)⁺. This ester was converted by the methods previously described for **11b** to provide after recrystallization from ethyl acetate:hexane **17a** (80 mg, 10% overall). Mp 158–160 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 1.63 (s, 3 H), 5.36 (s, 4 H), 5.48 (s, 2 H), 6.59 (s, 1 H), 6.94 (m, 3 H), 7.30 (m, 4 H), 7.62 (m, 1 H), 7.70 (d, J = 8 Hz, 1 H), 7.80 (m, 1 H), 8.00 (m, 2 H), 8.40 (d, J = 8 Hz, 1 H), 13.00 (broad s, 1 H); MS (FAB(+)) m/z 514 (M+H)⁺. Anal. calcd for C₂₉H₂₄ClN₃O₄·0.25 H₂O: C, 67.18; H, 4.76; N, 8.10. Found: C, 67.15; H, 4.40; N, 7.95.

(Z)-2-{[1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]methoxyimino}-3-phenylpropionic acid (17b). Prepared by the method described for 17a and oxime formation with phenylpyruvic acid provided after recrystallization from ethyl acetate-hexane 17b (20 mg, 4% overall). Mp 133–136 °C; ¹H NMR $(300 \text{ MHz}, \text{DMSO-}d_6) \delta 3.53 \text{ (s, 2 H)}, 5.35 \text{ (s, 2 H)},$ 5.38 (s, 2 H), 5.40 (s, 2 H), 6.57 (s, 1 H), 6.92 (m, 3 H), 7.02 (m, 2 H), 7.25 (m, 7 H), 7.62 (m, 1 H), 7.70 (d, J = 8 Hz, 1 H), 7.78 (m, 1 H), 8.00 (m, 2 H), 8.40 (d, J = 8 Hz, 1 H); MS (FAB(+)) m/z 590 (M+H)⁺. Anal. calcd for C₃₅H₂₈ClN₃O₄·1.5 H₂O: C, 68.12; H, 5.06; N, 6.81. Found: C, 67.71; H, 4.23; N, 6.53.

{1-[1-(4-Chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]-1-but-3-enyloxyimino}acetic acid (17c). Prepared by the method described for **17e** substituting allylmagnesium chloride for *n*-butylmagnesium chloride. Mp 90–92 °C; ¹H NMR (300 MHz, DMSO-*d*₆) δ 2.80 (m, 2 H), 3.62 (m, 1 H), 5.12 (m, 1 H), 5.45 (m, 5 H), 6.54 (s, 1 H), 6.94 (m, 3 H), 7.18 (d, *J* = 3 Hz, 1 H), 7.30 (m, 4 H), 7.60 (m, 1 H), 7.68 (d, *J* = 9 Hz, 1 H), 7.78 (m, 1 H), 8.0 (m, 2 H), 8.40 (d, *J* = 9 Hz, 1 H); MS (DCI-NH₃) *m/z* 540 (M+H)⁺. Anal. calcd for C₃₁H₂₆ClN₃O₄·H₂O: C, 66.82; H, 5.05; N, 7.53. Found: C, 67.41; H, 5.46; N, 7.23.

{1-[1-(4-Chlorobenzy])-5-(2-quinolylmethoxy)indol-2-yl]-2-phenylethyloxyimino}acetic acid (17d). Prepared according to the method described for 17e substituting benzylmagnesium chloride for *n*-butylmagnesium chloride: mp 94–96 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 1.91 (s, 2 H), 5.36 (m, 4 H), 5.6 (t, J = 7 Hz, 1 H), 6.65 (s, 1 H), 6.85 (m, 3 H), 7.23 (m, 9 H), 7.61 (m, 1 H), 7.68 (d, J = 9 Hz, 1 H), 7.78 (m, 1 H), 8.01 (m, 3 H), 8.40 (d, J = 9 Hz, 1 H); MS (DCI-NH₃) m/z 590 (M+H)⁺. Anal. calcd for C₃₅H₂₈ClN₃O₄·2 H₂O: C, 67.14; H, 5.11; N, 6.70. Found: C, 66.43; H, 5.00; N, 6.37.

{1-[1-(4-Chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]-1-pentyloxyimino}acetic acid (17e). A suspension of [1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]methanol (15) (3.9 g, 9.1 mmol) and barium manganate⁹ (23 g, 90 mmol) in toluene (250 mL) was refluxed at 100 °C for 18 h. The reaction mixture was then cooled to rt and the solids were removed by filtration, washed with ethyl acetate, and THF. The filtrate and washings were combined and concentrated in vacuo to afford [1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]carboxaldehyde (3.4 g, 88%). ¹H NMR (300 MHz, DMSO- d_6) δ 5.40 (s, 2 H), 5.80 (s, 2 H), 7.05 (d, *J* = 9 Hz, 2 H), 7.23 (d–d, *J* = 9 and 3 Hz, 1 H), 7.33 (d, *J* = 9 Hz, 2 H), 7.39 (d, *J* = 3 Hz, 1 H), 7.46 (m, 1 H), 7.60 (m, 2 H), 7.71 (d, *J* = 8 Hz, 1 H), 7.80 (m, 1 H), 8.01 (m, 2 H), 8.41 (d, *J* = 8 Hz, 1 H); MS (DCI-NH₃) *m/z* 427 (M+H)⁺.

To a solution of aldehyde (852 mg, 2 mmol) in THF (50 mL) at -78 °C was added *n*-butylmagnesium chloride (2 M solution in ethyl ether, 1.5 mL, 3 mmol) and the resulting mixture was stirred at rt for 12 h. It was then quenched with aqueous saturated NH₄Cl and extracted with ethyl acetate (100 mL). The organic extract was dried (MgSO₄) and concentrated in vacuo. The residue was chromatographed (silica gel, 2:1, hexane:ethyl acetate) to afford 900 mg (93%) of 1-[1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]-1-pentanol (16c). ¹H NMR (300 MHz, CDCl₃) δ 0.87 (t, J = 7 Hz, 3 H), 1.30 (m, 3 H), 1.61 (m, 1 H), 1.90 (m, 2 H), 4.68 (m, 1 H), 5.44 (m, 4 H), 6.44 (s, 1 H), 6.88 (d, J = 9 Hz, 2 H), 6.95 (d-d, J = 9 and 3 Hz, 1 H), 7.06(d, J = 9 Hz, 1 H), 7.17 (d, J = 3 Hz, 1 H), 7.22 (d, J = 9 Hz, 2 H), 7.55 (m, 1 H), 7.73 (m, 2 H), 7.83 (d, J = 8 Hz, 1 H), 8.10 (d, J = 8 Hz, 1 H), 8.19 (d, J = 8 Hz, 1 H); MS (DCI-NH₃) m/z 485 (M+H)⁺.

To a solution of 16c (650 mg, 1.3 mmol), triphenylphosphine (470 mg, 1.6 mmol) and N-hydroxyphthalimide (260 mg, 1.6 mmol) in THF (45 mL) was added dropwise DIAD (0.35 mL, 1.8 mmol) and the mixture was stirred at rt for 10 h. The THF was removed in vacuo and the residue was chromatographed (silica gel, 3:1, hexane:ethyl acetate) to provide 1-[1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]-1-pentoxyl-Nphthaloylamine (630 mg, 77%). A mixture of the phthaloyl intermediate and hydrazine hydrate (0.2 mL, 3 mmol) in ethanol (20 mL) was refluxed for 30 min and then cooled to rt. Aq 10% Na₂CO₃ was added and the resulting mixture was extracted with ethyl acetate (75 mL). The organic layer was washed with water, brine, dried (MgSO₄), and concentrated in vacuo to provide 450 mg (90%) of 1-[1-(4-chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]1-pentoxyamine. A solution of amine, glyoxylic acid hydrate (165 mg, 1.8 mmol) and acetic acid (0.1 mL) in THF (60 mL), methanol (30 mL) and water (15 mL) was stirred at rt for 12 h. The volatile organics were removed in vacuo and the resulting residue was extracted with ethyl acetate, dried (MgSO₄), concentrated, and chromatographed (silica gel, 19:1, methylene chloride:methanol) followed by crystallization from ethyl acetate:hexane to afford 17e (310 mg, 62%). Mp 125–127 °C. ¹H NMR $(300 \text{ MHz}, \text{DMSO-}d_6) \delta 0.78 \text{ (m, 3 H)}, 1.22 \text{ (m, 4 H)},$ 1.95 (m, 2 H), 5.40 (m, 5 H), 6.54 (s, 1 H), 6.95 (m, 3 H), 7.20 (d, J = 3 Hz, 1 H), 7.30 (m, 4 H), 7.52 (m, 1 H), 7.70 (d, J = 9 Hz, 1 H), 7.80 (m, 1 H), 8.02 (m, 2 H), 8.42 (d, J = 9 Hz, 1 H); MS (DCI-NH₃) m/z 556 $(M+H)^+$. Anal. calcd for $C_{32}H_{30}ClN_3O_4$: C, 69.11; H, 5.43; N, 7.55. Found: C, 69.00; H, 5.50; N, 7.47.

{1-[1-(4-Chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-yl]-1-cyclohexylmethoxyimino}acetic acid (17f). Prepared by the method of 17e using cyclohexylmagnesium chloride. Mp 182–185 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 1.11 (m, 6 H), 1.72 (m, 5 H), 4.98 (d, J = 9 Hz, 1 H), 5.34 (s, 2 H), 5.45 (d, J = 9 Hz, 2 H), 6.36 (s, 1 H), 6.85 (m, 1 H), 7.05 (d, J = 9 Hz, 2 H), 7.15 (m, 2 H), 7.31 (m, 3 H), 7.61 (m, 1 H), 7.69 (d, J = 9 Hz, 1 H), 7.78 (m, 1 H), 8.00 (m, 2 H), 8.40 (d, J = 9 Hz, 1 H); MS (FAB(+)) m/z 582 (M+H)⁺.

{1-[1-(4-Chlorobenzyl)-5-(2-quinolylmethoxy)indol-2-y-I]-1-cyclohexylmethoxyimino}-2-propionic acid (17g). Prepared by the method of 17e with oxime formation as described for 3e. Mp 140 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 1.05 (m, 6 H), 1.55 (m, 3 H), 1.75 (s, 3 H), 1.95 (m, 2 H), 5.1 (d, J = 9 Hz, 1 H), 5.35 (s, 2 H), 5.45 (s, 2 H), 6.38 (s, 1 H), 6.85 (dd, J = 9 and 2 Hz, 1 H), 7.05 (d, J = 9 Hz, 2 H), 7.16 (m, 2 H), 7.30 (m, 2 H), 7.61 (m, 1 H), 7.70 (d, J = 8 Hz, 1 H), 7.78 (m, 1 H), 8.02 (m, 2 H), 8.40 (d, J = 8 Hz, 1 H); MS (DCI-NH₃) m/z 596 (M+H)⁺. Anal. calcd for C₃₅H₃₄CIN₃O₄:H₂O: C, 68.51; H, 5.81; N, 6.85. Found: C, 68.72; H, 5.48; N, 6.75.

References

1. Steinhilber, D. Pharm. Acta Helv. 1994, 69, 3.

2. Woods, J. W.; Evans, J. F.; Ethier, D.; Scott, S.; Vickers,

(Received in U.S.A. 27 August 1996; accepted 15 October 1996)

P. J.; Hearn, L.; Heibein, J. A.; Charleson, S.; Singer, I. I. J. *Exp. Med.* **1993**, *178*, 1935.

3. Abramovitz, M.; Wong, E.; Cox, M. E.; Richardson, C. D.; Li, C.; Vickers, P. J. *Eur. J. Biochem.* **1993**, *215*, 105.

4. (a) Prasit, P.; Belley, M.; Brideau, C.; Chan, C.; Charleson, S.; Evans, J. F.; Fortin, R.; Ford-Hutchinson, A. W.; Gillard, J. W.; Guay, J.; Hutchinson, J. H.; Leger, S.; Riendeau, D.; Young, R. N.; Zamboni, R. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 1395. (b) Brideau, C.; Chan, C.; Denis, D.; Evans, J. F.; Ford-Hutchinson, A. W.; Fortin, R.; Gillard, J. W.; Guay, J.; Hutchinson, J.; Jones, T.; Léger, S.; Mancini, J. A.; McFarlane, C. S.; Pickett, C.; Piechuta, H.; Prasit, P.; Riendcau, D.; Rouzer, C. A.; Tagari, P.; Vickers, P.; Young, R. N. *Can. J. Physiol. Pharmacol.* **1992**, *70*, 799.

5. (a) Diamant, Z.; Timmers, M. C.; Van der Veen, H.; Friedman, B. S.; Smet, M. D.; Depré, M.; Hillard, D.; Bel, E. H.; Sterk, P. J. J. Allergy Clin. Immunol. **1995**, 95, 42. (b) Chapman, K. R.; Freidman, B. S.; Shingo, S.; Heyse, J.; Reiss, T.; Spector, R. Am. J. Respir. Crit. Care Med. **1994**, 149, A215.

6. Woods, K. W.; Brooks, C. D. W.; Maki, R. G.; Rodriguez, K. E.; Bouska, J. B.; Young, P.; Bell, R. L.; Carter, G. W. *Bioorg. Med. Chem. Lett.* (in press).

7. Mitsunobu, O. Synthesis 1981, 1.

8. The biological assay methods are described in detail in: Carter, G. W.; Young, P. R.; Albert, D. H.; Bouska, J.; Dyer, R.; Bell, R. L.; Summers, J. B.; Brooks, D. W. *J. Pharm. Exp. Ther.* **1991**, *256*, 929.

9. Firouzabadi, H.; Ghaderi, E. Tetrahedron Lett. 1978, 839.