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Dedicated to Professor John C. Sheehan on the occasion of his sixty-fifth birthday.

Actinomycin D(C₁) has been synthesized by a route involving the ester formation between two peptide fragments, (2S,3S)-1-(2-nitro-3-benzyloxy-4-methylbenzoyl)-3-methyl-2-aziridinecarbonyl-D-valylproline t-butyl ester and N-benzyloxycarbonylsarcosyl-N-methylvaline, via a ring-opening reaction of aziridine. Cyclization, followed by reduction and oxidation, gave actinomycin $D(C_1)$. The synthetic actinomycin $D(C_1)$ was indistinguishable from natural substance with respect to physical properties and biological activity.

J. Heterocyclic Chem., 17, 1815 (1980). Sir:

A number of cyclic peptide lactone antibiotics such as the actinomycin group, etamaycin, and echinomycin have been reported during the last few years. However, since they possess a cyclic lactone structure and unusual amino acids, the synthetic approaches were limited.

It was found that the reaction of the peptide containing (2S,3S)-3-methyl-2-aziridinecarboxylic acid with N-protected amino acids or dipeptides affords O-acylthreonine peptides as shown in Scheme 1 (1). This paper

$$\begin{array}{c} \text{CH}_3\text{-CH} & \text{CH-CO-Gly-OBz1} \\ & \text{N} \\ & \text{Gly-z} \end{array} \xrightarrow{\text{RCOOH}} \begin{array}{c} \text{O-CO-R} \\ & \text{H-C-CH}_3 \\ \text{z-Gly-NH-CH-CO-Gly-OBz1} \end{array}$$

Scheme 1

reports an application of the ester formation method to the synthesis of the naturally occurring cyclic peptide lactone, actinomycin $D(C_1)$.

Thus far, the general synthetic method for peptide lactones involved initial formation of the ester bond followed by peptide elongation and cyclization by the formation of the amide bond. A new route is designed in which the linear pentapeptide ester (11) is formed directly by the ring-opening reaction of the aziridinecarboxylic acid containing peptide (8) with dipeptide (10) as shown in Scheme 2. The method needs no activating reagent for the preparation of O-acylthreonine peptide, no racemization taking place during the course of direct introduction of the N-protected dipeptide.

(2S,3S)-1-Tri-3-methyl-2-aziridinecarboxylic acid (4) was prepared from threonine by the following route. Tritylthreonine methyl ester (1) was treated with ptoluenesulfonyl chloride in pyridine solution at -10° to give the O-tosyl derivative (2), which was refluxed in THF with triethylamine to form the aziridinecarboxylic acid derivative (3). Saponification of 3 with lithium hydroxide gave the desired amino acid derivative (4).

The aziridine segment 8, (2S,3S)-1-(2-nitro-3-benzyloxy-4-methylbenzoyl)-3-methyl-2-aziridinecarbonyl-Dvalylproline t-butyl ester, was synthesized as follows.

Scheme 2

Table
Physical Properties and Biological Activities of Actinomycin D

	Synthetic	Natural Actinomycin (Lit.)	
Characteristics	Actinomycin D (15)	(a)	(b)
Melting point/(°C)	242-243	241-243	246-247
Optical rotation ($[\alpha]_{\mathbf{D}}^{23}$ (c)	-316	-323 ± 10	-328 ± 10
Uv absorption	24,900 (443)	24,400 (443)	25,000 (443)
$[\epsilon \text{ in methanol } (\lambda, nm)]$	35,000 (240)	34,100 (240)	34,000 (231)
Ir absorption, (cm ⁻¹ potassium bromide)	1745 (lactone C=O)	1745	1760
* '`	1620-1670 (amide)	1620-1670	1620-1670
	1580 (chromophore)	1580	1585
	1195 (lactone COC)	1195	1200 (d)
Antibacterial activity			
(MIC, μ g./ml.)			
B. Subtilis ATCC-6633	0.78	0.78 (e)	
E. Coli NIHJ	100	-	

(a) Reference 3. (b) Reference 5. (c) In methanol at c 0.21. (d) Reference 6. (e) Reference 7.

Coupline of benzyloxycarbonyl-D-valine and proline tbutyl ester using dicyclohexylcarbodiimide (DCC) (2) gave the dipeptide (5). Hydrogenation of 5, followed by coupling with 4 using DCC gave the tripeptide (6) in 91.5% yield. Selective removal of the tirtyl group of 6 with 85% formic acid containing a small amount of methanol gave 7 in 94.6% yield, which was then coupled, in the dark, with 2-nitro-3-benzyloxy-4-methylbenzoic acid N-hydroxysuccinimide ester. Compound 8 was obtained as colorless amorphous powder in 94.7% yield after purification by silica gel column chromatography using benzene-ethyl acetate (1:1 v/v). Benzyloxycarbonylsarcosine was coupled with N-methylvaline t-butyl ester using DCC to give the dipeptide (9). The t-butyl group of 9 was removed by the action of trifluoroacetic acid to give the dipeptide acid (10) as an acid component.

The formation of the ester bond between N-methylvaline and threonine was carried out by heating 8 together with 10 at 110° for 5 hours in the dark. The coupling reaction afforded a mixture of 11 and the hydrolyzed by-product of 8 (threonine derivative), from which 11 was isolated in 45-55% yields by silica gel column chromatography. The t-butyl group of 11 was removed by the action of trifluoroacetic acid, the product 12 being treated with bis(p-nitrophenyl)sulfite in a pyridine solution to give the p-nitrophenyl ester 13. Deprotection of benzyloxycarbonyl group of 13 with hydrogen bromide in dioxane, followed by neutralization, cyclization

(0.98 mmole in 2 l. of pyridine, 60° , 8 hours, and purification on a Sephadex LH-20 column in methanol, gave the cyclic pentapeptide lactone (14) in 21.4% yield. Catalytic hydrogenolysis of 14, followed by oxidation with potassium hexacyanoferrate (III) in the 1:1 mixture of methanol and 0.067M phosphate buffer, pH 7.1 (3), gave actinomycin $D(C_1)$ (15), which was quantitatively crystallized from ethyl acetate-hexane.

The synthetic actinomycin $D(C_1)$ was indistinguishable from the natural substance with respect to physical properties and biological activity against B. Subtilis as shown in the following Table.

REFERENCES AND NOTES

- (1) T. Tanaka, K. Nakajima, T. Maeda, A. Nakamura, N. Hayashi and K. Okawa, Bull. Chem. Soc. Japan, 52, 3579 (1979).
- (2) The abbreviations according to IUPAC-IUB commission, J. Biol. Chem., 247, 977 (1972), are used. "Azyline" is used as the name of an 2-aziridinecarboxylic acid, "Azy" being its abbreviation. 3-MeAzy: (2S,3S)-3-methyl-2-aziridinecarboxylic acid.
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- (7) Natural actinomycin D was purchased from P-L Biochemicals, Inc., Lot No. 610111.