

3,5,7,3',5'-PENTAHYDROXYFLAVAN AND 3 α -METHOXYFRIEDELAN FROM *HUMBOLDTIA LAURIFOLIA*

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Key Word Index—*Humboldtia laurifolia*; Leguminosae; *O*-acetyloleanolic aldehyde; lupeol; 5,7,4'-trihydroxyflavone; (2*R*,3*R*)-3,5,7,3',5'-pentahydroxyflavan; 3 α -methoxyfriedelan.

Abstract—The leaf, bark and timber extractives of *Humboldtia laurifolia* were investigated and the following compounds have been isolated: *O*-acetyloleanolic aldehyde, a sitosterol ester, lupeol, sitosterol, a fatty acid, 5,7,4'-trihydroxyflavone (apigenin), (2*R*,3*R*)-3,5,7,3',5'-pentahydroxyflavan and 3 α -methoxyfriedelan. The latter two compounds are new natural products.

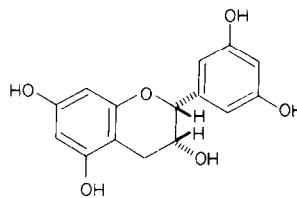
INTRODUCTION

Humboldtia laurifolia belongs to the subfamily Caesalpinoideae of the Leguminosae. The genus *Humboldtia* has not been chemically investigated and we report now the chemical investigation of the leaf, bark and timber of *H. laurifolia*.

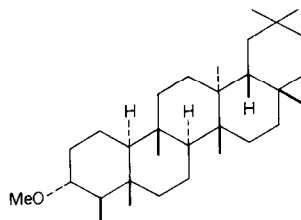
RESULTS AND DISCUSSION

The leaf benzene extract on cooling gave a triterpenoid. The ¹H NMR spectrum showed the triterpenoid to be a methoxy compound (δ 3.33). High resolution mass spectrometry gave a molecular formula of C₃₁H₅₄O (M⁺ at *m/z* 442.4172, requires 442.4174). Demethylation by hydrogen bromide followed by acetylation gave an acetate. Hydrolysis of the acetate yielded a triterpene alcohol, C₃₀H₅₂O (M⁺ at *m/z* 428.4013, requires 428.4017) which was identified as friedelan-3 α -ol. The natural product is thus 3 α -methoxyfriedelan (2). This compound was synthesized from friedelin. Friedelin was reduced with sodium-*n*-propanol to give friedelan-3 α -ol which on treatment with potassium-methyl iodide gave 3 α -methoxyfriedelan (2) which was identical (mmp, IR and co-TLC) with the natural product. This is the first report of the occurrence of 3 α -methoxyfriedelan as a natural product.

The timber extracts on CC over Si gel gave *O*-acetyloleanolic aldehyde and lupeol. The bark extracts on similar separation gave a sitosterol ester, *O*-acetyloleanolic aldehyde, lupeol, sitosterol, a fatty acid, 5,7,4'-trihydroxyflavone (apigenin) and a pentahydroxyflavan, mp 240°, [α]_D = -39.7° (MeOH). The latter compound was tentatively identified to be (2*R*,3*R*)-3,5,7,3',5'-pentahydroxyflavan (1) as follows. It had a molecular formula of C₁₅H₁₄O₆ (M⁺ at *m/z* 290.0787, requires 290.0790). The IR spectrum showed the presence of absorptions at 3545 cm⁻¹ (OH) and 1620 cm⁻¹ (C=C) but had no absorptions for any type of carbonyl group.



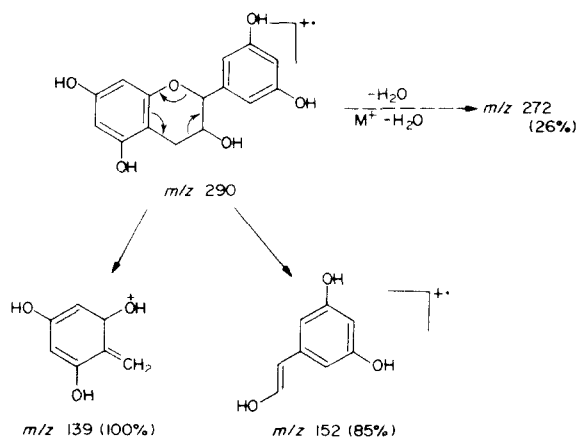
1



2

Methylation with dimethyl sulphate gave a tetramethoxylated compound. The ¹H NMR spectrum of the natural product had a one-proton multiplet at δ 4.21 typical of a -CHOH signal. Two types of benzylic protons appeared at δ 2.78 (2H, *dd*, *J* = 2 and 4 Hz) and at 4.9 (1H, *d*, *J* = 2 Hz). The presence of two types of aromatic system was discerned (a) from the presence of a *meta*-coupled proton system at δ 5.93 (1H, *d*, *J* = 2 Hz) and 6.05 (1H, *d*, *J* = 2 Hz) in one system, and (b) from the two-proton doublet at 6.84 (*J* = 1.2 Hz) and a one-proton doublet at 7.05 (*J* = 1.2 Hz). These protons, which were present in the second aromatic ring, were also *meta*-coupled. These data and the mass spectral fragmentations (Scheme 1) were in keeping with the structure 1 for the pentahydroxyflavan. The configurations at C-2 and C-3 were obtained as follows. The two benzylic protons centred at δ 2.78 in the ¹H NMR spectrum were coupled to each other and to the C-3 proton and appeared as a double doublet (*J* = 4 and 2 Hz) due to their being chemically non-equivalent. This situation is created by the C-3 hydroxy

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Scheme 1. Mass spectral fragmentation of compound 1.

group which takes up an axial conformation. The C-2 and C-3 hydrogen atoms should be in an (a,e) *cis*-orientation or (e,e) *trans*-orientation to give the relatively small coupling effect ($J = 2$ Hz) noticed for the C-2 proton which appeared at δ 4.9 in the ¹H NMR spectrum. The (a,e) *cis*-orientation is more likely since the C-2 aryl ring can then be in an equatorial configuration. An axial aryl arrangement would influence the chemical shift of the C-4 axial proton which was found to have almost the same δ_H value as that of the C-4 equatorial proton. The above information gives the relative stereochemistry at C-2 and C-3 but the absolute configurations at C-2 and C-3 remain to be established and at present they are only tentatively assigned as (2*R*,3*R*).

EXPERIMENTAL

Plant materials were collected from the Kanneliya and Ratnapura forests of Sri Lanka. The leaf, bark and timber of *Humboldtia laurifolia* Vahl. were separately dried and powdered. They were separately extracted with C₆H₆, Me₂CO and finally with MeOH. TLC and prep. TLC were carried out using Si gel. Mp's are uncorr. and were obtained using a Kolfer Hot Stage Reichert model apparatus.

Isolation of 3 α -methoxyfriedelan. The C₆H₆ extract (145 g) of the leaves (2.5 kg) on cooling deposited white crystals of 3 α -methoxyfriedelan (0.52 %); mp 301°, [α]_D -25° (CHCl₃); [M]⁺ at m/z 442.4172, C₃₁H₅₄O requires 442.4174; ¹H NMR (CCl₄): δ 3.33 (3H, s, -OMe) 1.26, 1.16, 2 \times 1.0, 0.93, 0.83, 0.75 (21 H, s, 7 \times Me), 0.86 (3H, d, $J = 7$ Hz); MS m/z (rel. int.): 442(34), 428(43), 410(42), 395(50), 269(49), 257(75), 247(54), 231(79), 205(64), 202(69), 189(81), 177(84), 149(81), 148(86), 125(100).

Demethylation and acetylation of 3 α -methoxyfriedelan. 3 α -Methoxyfriedelan (200 mg) in CHCl₃ (2.5 ml) was refluxed with PhOH (2.0 ml), Ac₂O (2.5 ml) and HBr (47%) (6 ml) for 7 hr. The reaction mixture was evaporated and washed with H₂O to give 180 mg of a crude product. This, on purification by prep. TLC, gave pure 3 α -acetoxyfriedelan (105 mg) mp 302°, lit. [1] 300°, [α]_D -12° (CHCl₃), lit. [1] -12°.

Hydrolysis of 3 α -acetoxyfriedelan. 3 α -Acetoxyfriedelan (55 mg) in MeOH (1.0 ml) was heated with 10% methanolic KOH (0.6 ml) for 6 hr. The hydrolysed product was worked-up in the usual manner to give friedelan-3 α -ol (35 mg) mp 301°, lit. [1] 308°, [α]_D +16° (CHCl₃), lit. [1] +13°; [M]⁺ at m/z 428.4013 (C₃₀H₅₂O requires 428.4017).

Synthesis of 3 α -methoxyfriedelan. Friedelin (250 mg) in *n*-PrOH (2.0 ml) was refluxed with Na (0.5 g) for 1 hr. The solvent was evaporated, the crude product acidified and extracted with CHCl₃. Prep. TLC of the residue yielded pure friedelan-3 α -ol (98 mg) mp 301°, lit. [1] 308°; [identified by comparison with an authentic sample (mmp, co-TLC)].

Friedelan-3 α -ol (300 mg) was dissolved in dry toluene (3.0 ml) and K (1 g) was added to the boiling soln. Reaction was continued for 2 hr and then MeI (5 ml) was added. After another 2 hr the mixture was cooled, excess K was destroyed using MeOH and the solvent evaporated. The residue was partitioned between CHCl₃ and H₂O. The CHCl₃ layer on drying and evaporation gave a residue (170 mg) which was purified by prep. TLC to yield pure 3 α -methoxyfriedelan (80 mg) mp 295°, [α]_D -24° (CHCl₃). It was found to be identical with the natural product isolated from the leaf (mmp, co-TLC).

Isolation of *O*-acetyloleanolic aldehyde. The petrol extract (16 g) of the timber (4.2 kg) was separated by CC over Si gel. Elution with C₆H₆ yielded *O*-acetyloleanolic aldehyde (0.533 g) mp 225°, lit. [2] 225–228°, [α]_D +59° (CHCl₃). In the absence of an authentic sample to compare, the following data were obtained: [M]⁺ at m/z 482.3761 (C₃₂H₅₀O₃ requires 482.3759); ¹H NMR (CCl₄): δ 9.25 (1H, s, CHO), 5.33 (1H, t, $J = 5$ Hz, -CH₂-CH=C), 4.43 (1H, AcO-CH-CH₂-), 1.93 (3H, s, Me-CO-O), 1.06 (3H, s, Me), 0.91 (9H, s, 3Me), 0.83 (6H, s, 2Me), 0.70 (3H, s, Me).

Hydrolysis of *O*-acetyloleanolic aldehyde by alcoholic KOH (10%) gave oleanolic aldehyde mp 171°, lit. [3] 170°, [α]_D +76° (lit. [3] +72°).

Isolation of sitosterol. Continued elution of the column with C₆H₆ gave sitosterol (0.25 g), mp 133–134°, lit. [1] 140°, [α]_D -35°. It was identical with an authentic sample (mmp, co-TLC).

Isolation of lupeol. Further elution with C₆H₆ gave lupeol (380 mg) mp 214°, lit. [4] 212°, [α]_D +33°. It was identical with an authentic sample.

Bark petrol extractives. The dried powdered bark (3.5 kg) gave 36 g of petrol extract. CC over Si gel gave an ester of sitosterol (1.5 g), lupeol (0.75 g), *O*-acetyloleanolic aldehyde (0.75 g), sitosterol (2.09 g) and an aliphatic acid (2.0 g). Known natural products were identified by comparison with authentic samples (mmp, co-TLC).

Bark Me₂CO extract. Defatted bark was extracted with Me₂CO to give 310 g of extract. This extract (70 g) was separated on a Si gel column and elution with 40% EtOAc-petrol gave sitosterol (0.50 g); 5,7,4'-trihydroxyflavone (apigenin) (0.53 g). Apigenin was characterized by its mp 348° (lit. [5] 347°) and by its characteristic ¹H NMR spectrum: δ (DMSO-*d*₆) 12.96 (1H, s, -OH), 7.94 (2H, d, $J = 8$ Hz), 6.94 (2H, d, $J = 8$ Hz), 6.77 (1H, s, =CH), 6.50 (1H, d, $J = 2$ Hz), 6.21 (1H, d, $J = 2$ Hz). Apigenin was acetylated to give a diacetate (mp 185°). The diacetate had [M]⁺ at m/z 354.0739, C₁₉H₁₄O₇ requires 354.0790.

Bark MeOH extract. The bark MeOH extract (10 g) was re-extracted with EtOAc to give 5 g of EtOAc-soluble fraction. Prep. TLC separation of 1 g of this material gave 120 mg of 3,5,7,3',5'-pentahydroxyflavan, mp 240°, [α]_D -39.7° (MeOH); [M]⁺ at m/z 290.0787, C₁₅H₁₄O₈ requires 290.0790; IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3545, 1620, 1510, 1458, 1440, 1250, 1189, 1140, 1090, 1015, 810, 792; ¹H NMR (Me₂CO-*d*₆): δ 7.05 (1H, d, $J = 1.2$ Hz), 6.84 (2H, d, $J = 1.2$ Hz), 6.05 (1H, d, $J = 2$ Hz), 5.93 (1H, d, $J = 2$ Hz), 4.9 (1H, d, $J = 2$ Hz), 4.21 (1H, m), 2.78 (2H, dd, $J = 2$ and 4 Hz); MS m/z (rel. int.): 290(71), 286(45), 272(26), 175(46), 167(46), 163(37), 152(85), 141(54), 140(57), 139(100), 124(68), 123(81), 110(51); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 214(4.47), 225(4.46), 281(3.76), 394(3.16).

Methylation of 3,5,7,3',5'-pentahydroxyflavan. The phenol (0.075 g) in dry Me₂CO (5 ml) was treated with Me₂SO₄ (0.5 ml)

and dry K₂CO₃ (0.20 g). The product, a tetramethyl ether (0.70 g), mp 160° was isolated in the usual manner. It had $[\alpha]_D^{29} -29^\circ$ (CHCl₃); ¹H NMR (CCl₄): δ 6.95 (1H, *d*, *J* = 2 Hz), 6.84(2H, *d*, *J* = 2 Hz), 6.07(1H, *d*, *J* = 2 Hz), 5.95(1H, *d*, *J* = 2 Hz), 4.80(1H, *d*, *J* = 2 Hz), 4.14(1H, *m* -CHOH), 3.85(3H, *s*, OMe), 3.83(3H, *s*, OMe), 3.80(3H, *s*, OMe), 3.74(3H, *s*, OMe), 2.77(2H, *dd*, *J* = 2 and 4 Hz).

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