ANTIMICROBIAL CONSTITUENTS OF THE SPONGE SIPHONODICTYON CORALLIPHAGUM

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Abstract—The burrowing sponge Siphonodictyon coralliphagum contains two phenolic aldehydes, siphonodictyal-A (1) and siphonodictyal-B (2). The structure of siphonodictyal-A (1) was determined by interpretation of spectral data and the synthesis of a derivative. The structure of siphonodictyal-B (2) resulted from interpretation of spectral data and the synthesis of a degradation product. Both compounds exhibit antimicrobial activity.

Siphonodictyon coralliphagum (Rützler)¹ is one of a small group of sponges that burrow into limestone substrata. A study of the burrowing mechanism of Cliona lampa² revealed that 2-3% of the excavated substratum was dissolved while the remaining material was removed as small chips. The chemical mechanism for dissolution of calcium carbonate has not been determined. Unlike most Cliona species which tend to settle and excavate the dead regions of coral, Siphonodictyon coralliphagum is generally found in a living coral head. The major portion of the sponge is inside the coral head with only the oscular chimney protruding. In order to prevent overgrowth by the living coral polyps, the sponge maintains 5-10 mm zone of dead coral polyps around the base of the oscular chimney. Rützler¹ suggested that the dead zone was maintained by the production of mucus that flowed down to the base of the oscular chimney. In this paper, we will report the structure of secondary metabolites from S. coralliphagum that might play a role in either the burrowing process or the inhibition of coral growth.

The crude ethanolic extract of S. coralliphagum was shown to inhibit the growth of Staphylococcus aureus. Solvent partition gave dichloromethane, ethyl acetate, n-butanol and aqueous extracts, all of which showed antimicrobial activity. However, the ethyl acetate extract contained the majority of the material and was highly active. The 'H NMR spectrum of the crude extract suggested the presence of several aldehydes. However, Florisil chromatography of the extract resulted in a very poor recovery of siphonodictyal-A (1) (0.12% dry weight) that showed antimicrobial activity. Much of the material remained firmly adsorbed to the Florisil. Chromatography of the extract of Sephadex LH-20 using methanol as eluant gave siphonodictyal-B (2) (0.9% dry weight) together with smaller quantities of siphonodictyal-A (1) and some products that appear to result from acidcatalyzed rearrangement of siphonodictyal-B (2).

Siphonodictyal-A (1), $[\alpha]_D$ +3.2°, was crystallized from methanol, m.p. 192-3°. High resolution mass measurement gave the molecular formula $C_{22}H_{32}O_4$. The IR (3250, 1645 cm⁻¹) and UV [390 nm (€ 5000), 280 nm (€ [2,000)] spectra were similar to those of 2,5-dihydroxybenzaldehyde. The 'H NMR spectrum contained signals at δ 10.48 (s, 1 H), 7.07 (d, 1 H, J = 8 Hz) and 6.75 (d, 1 H, J = 8 Hz), indicating the presence of a 1,2,3,4 - tetra substituted aromatic aldehyde. Methylation of siphonodictyal - A (1) with dimethyl sulfate and potassium carbonate in dry acetone gave a dimethyl ether 3 [δ 3.83 (s, 3 H) and 3.85 (s, 3 H)] but acetylation with acetic anhydride in pyridine produce a monoacetate 4 [δ 2.36 (s, 3H)] in which a phenolic OH ortho to the aldehyde group had not been acetylated. From five possible 1,2,3,4 - substitution patterns for an alkyl group, a phenol and an o - hydroxyaldehyde, we selected the 2 - alkyl - 3,5 didhydroxybenzaldehyde pattern by comparison of the chemical shifts of the ¹³C NMR signals for the atomatic carbons with calculated values.³

Having proposed the structure of the aromatic portion, we now had to assign a structure to the sesquiterpene portion of siphonodictyal - A (1). The ¹³C NMR spectrum of siphonodictyal - A (1) indicated that the sesquiterpene portion of the molecule must be bicyclic and contain a tertiary alcohol. The ¹H NMR spectrum contained four methyl signals at δ 0.78 (s, 3 H), 0.83 (s, 3 H), 0.98 (s, 3 H) and 1.32 (s, 3 H) and benzylic methylene signals at 2.88 (d, 1 H, J = 16 Hz) and 3.23 (dd, 1 H, J = 16, 8 Hz). These data suggested a drimane ring skeleton and comparison of the ¹³C NMR spectrum with those of model compounds⁵ strongly supported this assignment. Confirmation of this assignment was accomplished using the following reaction sequence.

Dehydration of siphonodictyal-A (1) using *p*-toluenesulfonic acid in refluxing benzene gave a cyclic ether which was methylated using methyl iodide and potassium carbonate in dry acetone to obtain the methyl ether 5. Formylation of chromazonarol methyl ether 6^6 with





dichloromethyl methyl ether and titanium tetrachloride in dry dichloromethane at 0° for 15 min., followed by 1 hr at 20°, gave a mixture of the three possible mono-formyl derivatives, that were separated by LC and differentiated by ¹H NMR data. One of the aldehydes (22%) was identical in all respects to the methyl ether 5 prepared from aldehyde 1. The remaining aldehydes had the formyl group *meta* (36%) or *para* (27%) to the alkyl group. The stereochemistry of the Me group at C-8 in siphonodictyal-A (1) was shown to be axial by comparison of its ¹³C NMR chemical shift (δ 23.9) with the chemical shifts assigned to the C-8 axial (24.9) and equatorial (31.8) Me groups in the alcohols 7 and 8 respectively.⁶



Siphonodictyal-B (2) was obtained as a crystalline solid, m.p. 145-147°. The IR spectrum suggested the presence of phenolic (3100-3500 cm⁻¹) and aldehyde (1685 cm⁻¹) functionalities. The aldehyde gave a green color with methanolic ferric chloride which, together with the UV spectrum [(MeOH) 280 nm (ϵ 6400), 380 nm $(\epsilon 1540);$ (MeOH+OH⁻) 304 nm (ϵ 7300), 403 nm (ϵ 1800)], indicated the presence of a polyhydroxy benzaldehyde as part of the molecule. Since siphonodictyal-B (2) was insoluble in most organic solvents, the remaining spectral data were measured using the trimethoxy derivative 9, obtained from the reaction of the alkehyde 2 with methyl iodide and potassium carbonate in acetone. The trimethoxy derivative 9 had the molecular formula $C_{25}H_{16}O_4$. The 'H NMR spectrum contained three methoxy signals at δ 3.72, 3.85 and 3.91, an aldehvde proton signal at 10.40 (s, 1 H), an aromatic proton signal at 6.91 (s, 1 H), a signal assigned to the C-11 vinyl proton at 6.21 (s, 1 H), an allylic proton signal at 2.65 (m, 1 H) and four Me signals at 1.19 (s, 3 H), 0.91 (s, 3 H), 0.90 (d, 3 H, J = 7 Hz) and 0.89 (s, 3 H). These data could be assigned to a structure having the same carbon skeleton as siphonodictyal-A (1) with a benzylidene olefinic bond as found in spongiaquinone 10.7

Ozonolysis of the trimethyl ether 9 in ethyl acetate at -78° followed by reductive work-up using dimethyl sulfide gave the ketone 11 and the dialdehyde 12. The 'H NMR spectral data for the ketone 11 was identical to that reported for the degradation product of spongiaguinone 10.7 The ¹H NMR spectrum of the dialdehyde 12 contained two aldehyde signals at δ 10.33 (s, 1 H) and 10.25 (s, 1 H), an aromatic proton signal at 7.48 (s, 1 H) and three OMe signals at 4.02 (s, 3 H), 3.97 (s, 3 H), 3.94 (s, 3 H), eliminating symmetrical molecules. Comparison of the chemical shifts of the aromatic proton with calculated values⁸ for the aromatic protons of all possible unsymmetrical formyl - trimethoxy benzaldehydes suggested that the dealdehyde 12 was 3 - formyl - 2,4,5 trimethoxybenzaldehyde. The structure of dialdehyde was confirmed by synthesis. Demethylation of 2,4,5 trimethoxybenzaldehyde (13) with boron trichloride in dichloromethane gave 4.5 - dimethoxysalicaldehyde (14).¹⁰ Treatment of 4.5 - dimethoxysalicaldehyde (14) with dichloromethyl methyl ether and titanium tetra-chloride in dichloromethane¹¹ gave 4,5 - dimethoxy - 3 formylsalicaldehyde (15) as the only dialdehyde together with recovered starting material. Treatment of 4,5 dimethoxy - 3 - formylsalicaldehyde (15) with methyl iodide and potassium carbonate in dry acetone gave 3 formyl - 2,4,5 - trimethoxybenzaldehyde (12), identical in all respects to the product from ozonolysis of the trimethoxy derivative 9.

The ketone 11 and dialdehyde 12 can be formed from either the trimethoxy derivative 9 or the alternative structure 16. In order to differentiate between these alternatives, we calculated⁹ the expected chemical shifts for the aromatic proton in 3 - ethyl - 2,4,5 - trimethoxybenzaldehyde (δ 7.06) and 3 - ethyl - 2,5,6 - trimethoxybenzaldehyde (δ 6.78). Catalytic hydrogenation of the trimethoxy derivative 9 gave the alcohol 17 that was re-oxidized to the aldehyde 18 using manganese dioxide in acetone. The chemical shift of the aromatic proton in aldehyde 18 (δ 7.05) strongly suggested that the trimethoxy derivative had structure 9 rather than the alternative 16.

The stereochemistry at C-8 in these compounds was determined by a ¹H NMR spin decoupling experiment on the trimethoxy derivative 9. Irradiation of the doublet Me signal at δ 0.90 caused the multiplet at 2.65 to collapse to a signal with coupling constants of 6 and 2 Hz, indicating that the allylic proton at C-8 was equatorial. By way of contrast, a similar spin decoupling





experiment on the ketone 11, in which the C-8 Me group had isomerized to the more stable equatorial configuration, gave a C-8 proton signal with coupling constants of 12 and 6 Hz. The geometry about the $\Delta^{9(11)}$ olefinic bond is presumed to be E since models suggest that there is unacceptable steric crowding in the corresponding Z isomer.

Among the minor metabolites of S. coralliphagum was an inseparable 1:1 mixture of two cyclic ethers. The same mixture of cyclic ethers could be generated by acid-catalyzed cyclisation of siphonodictyal-B (2). Methylation of the mixture of cyclic ethers gave two dimethyl ethers that could be separated with difficulty. The ¹H NMR data¹² were compatible with structures of the type 19 but we had insufficient material to complete the structural elucidation.

Both siphonodictyal-A (1) and siphonodictyal-B (2) inhibit the growth of *Staphylococcus aureus* and *Bacillus subtilis* and both have been detected in the mucus from *S. coralliphagum.* However, we have not had the opportunity to determine whether the compounds alone can inhibit the growth of coral polyps. Both compounds, particularly siphonodictyal-B (2) are strongly adsorbed by Florisil and silica gel but we were unable to show that they are involved in calcium carbonate dissolution.

EXPERIMENTAL¹³

Collection and extraction of Siphonodictyon coralliphagum. The protruding portions (oscular chimney) of S. coralliphagum were collected by hand using SCUBA (-12 to -30 m) from coral heads on the leeward side of Lighthouse Reef and Glover Reef, Belize. The sponge, together with as much of the yellow mucus as possible, was frozen and maintained at - 20° until required. The sponge (180 g dry weight) was homogenized in EtOH (11), filtered, steeped in fresh EtOH (11), filtered and the combined EtOH extracts evaporated to obtain an aqueous suspension. The aqueous phase was extracted with CH_2Cl_2 (2×250 ml), EtOAc $(3 \times 250 \text{ ml})$ and n-BuOH (500 ml). The CH₂Cl₂ extract was dried over Na₂SO₄ and evaporated to obtain a yellow oil (1.5 g, 0.8% dry weight). The EtOAc extract was dried over NaSO4 and the solvent evaporated to obtain an orange oil (11.2 g, 6.2% dry weight). Evaporation of the n-BuOH extract gave an orange oil (6 g, 3.3% dry weight).

Isolation of siphonodictyal-A (1). The EtOAc extract (9 g) was preadsorbed onto Florisil (20 g) and applied to a column ($60 \times$ 5 cm diam) of Florisil. Fractions were obtained using eluants of increasing polarity from hexane through ether and EtOAc to 1:1 EtOAc/MeOH. The material eluted with 20% EtOAc in ether was purified on silica gel plates to obtain 1 (175 mg, 0.12% dry weight): m.p. 192-3°; $[\alpha]_D$ + 3.2° (c 0.5, MeOH); UV (MeOH) 390 nm (ϵ 5000), 280 nm (ϵ 12,000); IR (CHCl₃) 3250, 1645, 1580 cm⁻¹; ¹H NMR (CDCl₃) δ 0.78 (s, 3 H), 0.83 (s, 3 H), 0.98 (s, 3 H), 1.32 (s, 3 H), 1.84 (d, 1 H, J = 8 Hz), 2.88 (d, 1 H, J = 16 Hz), 3.23 (dd, 1 H, J = 16.8 Hz), 6.75 (d, 1 H, J = 8 Hz), 7.07 (d, 1 H, J = 8 Hz), 10.48 (s, 1 H); ¹³C NMR (CDCl₃/CD₃OD) δ 197.1 (s), 148.2 (s), 132.6 (s), 126.8 (d), 118.4 (s), 115.9 (d), 74.8 (s), 62.2 (d), 56.6 (d), 44.2 (t), 41.9 (t), 41.3 (t), 40.6 (s), 33.5 (s), 23.9 (t), 20.7 (q), 19.7 (q), 19.3 (q), 17.7 (t), 14.6 (q); high-resolution mass spectrum, obsd 360.233, C₂₂H₃₂O₄ requires: 360.230.

Isolation of siphonodictyal-B (2). The EtOAc extract (3.2 g) was chromatographed on a column (100×5 cm diam) of Sephadex LH-20 using MeOH as eluant. Fractions were screened for antimicrobial activity against *Staphylococcus aureus*. A series of fractions that showed strong anitmicrobial activity were combined and the solvent evaporated to obtain 2 (495 mg, 0.9% dry weight) as yellow crystals from aqueous MeOH: m.p. 145-147°; UV (MeOH) 280 nm (ϵ 6400), 380 (ϵ 1540), (MeOH) 280 nm (ϵ 1800); IR (KBr) 3100-3500, 1685 cm⁻¹; ¹H NMR (CD₃OD) 5 0.82 (d, 1 H, J = 7 Hz), 0.88 (s, 3 H), 0.92 (s, 3 H), 1.21 (s, 3 H), 6.40 (s, 1 H), 6.89 (s, 1 H), 10.27 (s, 1 H).

Methylation of siphonodictyal-A (1). A soln of 1 (7 mg, 0.02 mmol) and Me₂SO₄ (10 μ l) in dry acetone (10 ml) containing anhyd K₂CO₃ (50 mg) was stirred at 80° for 15 hr. The mixture was filtered and the solvent evaporated under vacuum to give an oily residue that was chromatographed on a silica gel plate to obtain 3 (4 mg, 50% theoretical): IR (CHCl₃) 3600, 1685, 1600 cm⁻¹; ¹H NMR (CDCl₃) δ 0.78 (s, 3 H), 0.82 (s, 3 H), 0.98 (s, 3 H), 1.21 (s, 3H), 1.89 (dd, 1 H, J = 8, 4 Hz), 3.11 (dd, 1 H, J = 16, 8 Hz), 3.83 (s, 3H), 3.85 (s. 3 H), 6.77 (d, 1 H, J = 9 Hz), 7.00 (d, 1 H, J = 9 Hz), 10.55 (s, 1 H); mass spectrum, m/z 388, 370, 355, 352.

Acetylation of siphonodictyal-A (1). A soln of 1 (10 mg, 0.03 mmol) in Ac₂O (0.5 ml) and pyride (1 ml) was stirred overnight at 25°. The solvents were evaporated under vacuum and the residue was chromatographed on a silica gel plate to obtain 4 (6 mg, 54% theoretical): IR (CHCl₃) 3450, 1755, 1640 cm⁻¹; UV (MeOH) 260 nm (ϵ 10,000), 345 (ϵ 4600); ¹H NMR (CDCl₃) δ 0.78 (s, 3 H), 0.83 (s, 3 H), 0.98 (s, 3 H), 1.27 (s, 3 H), 2.36 (s, 3 H), 2.88 (dd, 1 H, J = 16, 8 Hz), 3.23 (dd, 1 H, J = 16, 4 Hz), 6.80 (d, 1 H, J = 9 Hz), 10.67 (s, 1 H); mass spectrum, m/z 402, 384, 360, 342.

Dehydration and methylation of siphonodictyal-A (1). A soln of 1 (14 mg, 0.04 mmol) in benzene (10 ml) containing p-toluenesulfonic acid (1 crystal) was boiled under reflux for 2 hr. The cooled soln was washed with sat NaHCO₃ aq (5 ml), then water (5 ml), dried over NaSO₄ and the solvent evaporated to give a cyclic ether. The ether was methylated using the conditions described above to obtain 5 (10 mg, 72% theoretical): IR (CHCl₃) 1690 cm⁻¹; ¹H NMR (CDCl₃) δ 0.84 (s, 3 H), 0.89 (s, 3 H), 0.91 (s, 3 H), 1.14 (s, 3 H), 2.77 (dd, 1 H, J = 18, 12 Hz), 3.08 (dd, 1 H, J = 18, 4 Hz), 3.85 (s, 3 H), 6.79 (d, 1 H, J = 8 Hz), 6.97 (d, 1 H, J = 8 Hz).

Formylation of chromazonarol methyl ether 6. A soln of TiCl₄ (20 mg, 0.1 mmol) in dry CH₂Cl₂ (5 ml) was added to a cooled soln of chromazonarol methyl ether (31 mg, 0.095 mmol) and dichloromethyl methyl ether (20 mg, 0.17 mmol) in dry CH₂Cl₂ (10 mL) and the resulting mixture was stirred at 0° for 15 min then at 22° for 1 hr. The mixture was washed with 3 N HCl (5 ml) and water (5 ml), dried over Na₂SO₄ and the solvent evaporated to obtain a yellow oil. The oil was chromatographed by LC on μ -porasil using 7% ether in hexane as eluant to obtain 5 (7.5 mg, 22% theoretical) together with the alternative mono-formyl derivatives with the formyl group meta (12 mg, 36% theoretical) or para (9 mg, 27% theoretical) to the alkyl group. Compd 5 had identical chromatographic properties and spectral data to the product from 1.

Methylation of siphonodictyal-B (2). A soln of 2 (10 mg, 0.028 mmol) and Me₂SO₄ (0.2 ml) in dry acetone (10 ml) containing anhyd K₂CO₃ (50 mg) was stirred at 80° overnight (> 12 h). The cooled mixture was filtered and the solvent evaporated under vacuum to give an oil that was filtered through a plug of silica gel in ether soln to obtain 9 (10 mg, 90% theoretical): IR (CHCl₃) 1685, 1525 cm⁻¹; ¹H NMR (CDCl₃) δ 0.89 (s, 3 H), 0.90 (d, 3 H, J = 7 Hz), 0.91 (s, 3 H), 1.19 (s, 3 H), 2.65 (m, 1 H), 3.72 (s, 3 H), 3.85 (s, 3 H), 3.91 (s, 3 H), 6.21 (s, 1 H), 6.91 (s, 1 H), 10.40 (s, 1 H); ¹³C NMR (C₂C₂) δ 189.1 (s), 158.3 (s), 152.3 (s), 150.3 (s), 148.9 (s), 130.4 (s), 125.0 (s), 120.5 (d), 114.9 (d), 61.6 (q), 56.1 (q), 50.5 (q), 42.7, 42.4, 41.3, 40.0, 34.1, 33.4, 33.1, 32.7, 22.9, 22.4, 21.9, 20.5, 19.9: high-resolution mass spectrum, obsd 400.2576, C₂₅H₃₆O₄ requires; 400.2613.

Ozonolysis of trimethyl ether 9. A stream of O₃ in O₂ was bubbled into EtOAc at -78° until a blue-colored soln was obtained. The soln was added dropwise to a stirred soln of 9 (10 mg, 0.025 mmol) in EtOAc (10 ml) at -78° until the starting material had completely reacted (as indicated by tlc). Me₂SO₄ (0.5 ml) was added and the mixture was stirred at 25° for 3 hr. The solvent was evaporated under vacuum and the resulting oil was chromatographed on a silica gel plate to obtain 11 (3 mg, 58% theoretical), having identical spectral data to those reported by Kazlauskas et al.,⁷ and 12 (2.5 mg, 45% theoretical): UV (MeOH) 266 nm (ϵ 1920), 233 nm (ϵ 4600); IR (CHCl₃) 1690 cm⁻¹; ¹H NMR (CCl₄) δ 3.94 (s, 3 H), 3.97 (s, 3 H), 4.02 (s, 3 H), 7.48 (s, 1 H), 10.25 (s, 1 H), 10.33 (s, 1 H); mass spectrum, m/z 224, 223.

Synthesis of 3 - formyl - 2,4,5 - trimethoxybenzaldehyde (12). A soln of BCl₃ (~80 mg, excess) in CH₂Cl₂ (4 mL) was added to a cooled soln of 13 (50 mg, 0.26 mmol) in CH₂Cl₂ (10 ml) and the mixture was stirred at -78° for 2½ hr. The mixture was poured into a mixture of ice and 1 N HCl (5 ml) and the organic material was extracted with ether (3 × 20 ml). The combined extracts were evaporated to obtain 14 (40 mg, 86% theoretical) as an oil: IR (CCl₄) 3300, 1665 cm⁻¹; UV (MeOH) 343 nm (ϵ 7800), 289 nm (ϵ 18,500); ¹H NMR (CDCl₃) 3 3.89 (s, 3 H), 3.93 (s, 3 H), 6.46 (s, 1 H), 6.90 (s, 1 H), 9.68 (s, 1 H).

A soln of TiCl₄ (0.4 ml) in dry CH₂Cl₂ (3 ml) was added to a cooled soln of 14 (40 mg) in CH₂Cl₂ (5 ml) and the mixture was stirred at 0°. A soln of dichloromethyl methyl ether (0.2 ml) in dry CH₂Cl₂ (3 ml) was added to the cooled soln over 30 min and the mixture was then stirred at room temp for 2 hr. The mixture was poured into ice-cold 1 N HCl and extracted with ether (3 × 30 ml). The combined ether extracts were washed with water (10 ml), dried over Na₂SO₄ and the solvent evaporated to obtain an oil. The oil was chromatographed on a silica gel plate to obtain 14 (23 mg, 58% recovery) and 15 (3.5 mg, 8% theoretical): IR (CCl₄) 3300, 1670 cm⁻¹; UV (MeOH) 379, 267, 243 nm; ¹H NMR (CDCl₃) δ 3.88 (s, 3 H), 4.15 (s, 3 H), 7.67 (s, 1 H), 10.33 (s, 1 H), 10.39 (s, 1 H).

A soln of 15 (3.5 mg) and MeI (1 ml) in dry acetone (10 ml) containing anhyd K_2CO_3 (100 mg) was stirred at 80° under reflux for 4 hr. The cooled mixture was poured into water (10 ml) and

extracted with EtOAc $(2 \times 20 \text{ mi})$. The EtOAc extract was washed with water, dried over NaSO₄ and evaporated under vacuum to obtain 12 (3.5 mg) identical in all respects to the material obtained from the natural product.

9,11 - Dihydrosiphonodictyal - B trimethyl ether (18). A soln of 9 (10 mg, 0.025 mmol) in ether (5 mi) containing 10% Pd-C catalyst (2 mg) was stirred under an atmosphere of H₂ until a tlc analysis revealed that the starting material had completely reacted. The catalyst was removed by filtration and the solvent evaporated to obtain 17 (10 mg, quantitative): IR (CCl₄) 3400 cm⁻¹; ¹H NMR (CDCl₃) & 0.70 (d, 3 H, J = 7 Hz), 0.84 (s, 3 H), 0.87 (s, 3 H), 0.92 (s, 3 H), 2.34 (dd, 1 H, J = 16, 6 Hz), 2.67 (d, 1 H, J = 16 Hz), 3.73 (s, 3 H) 3.86 (s, 3 H), 3.87 (s, 3 H), 4.75 (bs, 2 H), 6.76 (s, 1 H); mass spectrum, m/z 404 (M⁺).

A soln of 17 (10 mg) in dry acetone (10 ml) containing activated MnO_2^{14} was stirred at 25° for 5 hr. The MnO_2 was removed by filtration and the solvent evaporated to obtain 18 (8 mg, 80% theoretical): IR (CCl₄) 1685 cm⁻¹; UV (MeOH) 334 nm (ϵ 1110), 264 (ϵ 3100); ¹H NMR (CDCl₃) δ 0.71 (d, 3 H, J = 7 Hz), 0.84 (s, 3 H), 0.87 (s, 3 H), 0.92 (s, 3 H), 2.44 (dd, 1 H, J = 16, 6 Hz), 2.67 (dd, 1 H, J = 16, 2 Hz), 3.75 (s, 3 H), 3.89 (s, 3 H), 3.91 (s, 3 H), 7.05 (s, 1 H), 10.41 (s, 1 H); mass spectrum, m/z 402 (M⁺).

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