Efficient Asymmetric Synthesis of Phosphatidyl-D-*myo*-inositol 3,4,5-trisphosphate

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Phosphatidyl-D-myo-inositol 3,4,5-trisphosphate, a candidate of the second messenger in cellular signal transduction, was efficiently synthesized in a homochiral form.

Key words phosphatidyl-D-*myo*-inositol 3,4,5-trisphosphate; phosphatidyl-D-*myo*-inositol 3-kinase; cellular signal transduction; second messenger

Polyphosphoinositides play an extremely important role in cellular signal transduction. Recently, it has been revealed that phosphatidyl-D-myo-inositol 3-kinase (PI 3-kinase) catalyzes the phosphorylation of phosphatidyl-D-myo-inositol 4,5-bisphosphate (PI 4,5-P₂) to phosphatidyl-D-myo-inositol 3,4,5-trisphosphate (PI 3,4,5-P₃: 1). 1) Stephens *et al.* reported that PIP₃ 5-phosphatase dephosphorylates 1 to phosphatidyl-D-myo-inositol 3,4-bisphosphate (PI 3,4-P₂). 2) Interestingly, 1 is resistant to known phospholipase Cs, and no other enzyme that metabolizes 1 has so far been found. Although the role and function of 1 remain unknown, it is a promising candidate for a second messenger in cellular signal transduction.

Many biological studies of PI 3-kinase have been reported, but there are only a few reports on the function of its product 1.3,4) Since the availability of 1 is severely limited, an efficient chemical synthesis is required. Recently, we reported the synthesis of 1-O-alkyl and 1-O-acyl-D-myo-inositol 3,4,5-trisphosphates,5) which might serve as alternatives to 1 in studies of 1 and PI 3-kinase. Here, we present an efficient asymmetric synthesis of 1b⁶⁻⁹⁾ with saturated lipid side chains from the chiral myo-inositol intermediate 3 developed by Prestwich and co-workers. 10)

Results and Discussion

The synthetic scheme of **1b** is depicted in Chart 2. The appropriately protected homochiral key intermediate 3 was synthesized according to Prestwich's method¹⁰⁾ from methyl α-D-glucopyranoside 2 through 9 steps in 26% overall yield. The amidite 7 with two stearoyl groups was obtained from 1,2-stearoyl-sn-glycerol and benzyl N, N, N', N'-tetraisopropylphosphoramidite, which were prepared from (S)-(+)-2,2-dimethyldioxolane-4-methanol and phosphorus trichloride, respectively.^{9,11)} The coupling of the inositol derivative 3 with the amidite 7 followed by m-chloroperbenzoic acids (m-CPBA) oxidation in one pot gave an epimeric mixture of 4.12) Removal of the p-methoxybenzyl (PMB) groups with 2,3-dichloro-5,6dicyano-1,4-benzoquinone (DDQ)¹³⁾ in wet CH₂Cl₂ gave the triol 5. Phosphitylation of the resulting triol with dibenzyl N,N-diethylphosphoramidite, followed by oxidation with m-CPBA in one pot gave fully protected PI 3,4,5-P₃ (6). Deprotection of all benzyl (Bn) and benzyloxymethyl (BOM) groups of 6 was carried out cleanly by hydrogenolysis over Pd black in tert-BuOH in the presence of NaHCO₃. 9) During the hydrogenolysis, partially or fully deprotected sodium salts precipitated. To obtain complete deprotection to **1b**, a small amount of water was added after 2h to dissolve insoluble salts. The overall yield of **1b** from **2** was 21% through 13 steps.

The synthesis described here should provide easy access to a variety of derivatives of **1b**. Clarification of the function of **1** in cellular signal transduction is in progress and the results will be reported elsewhere.

Experimental

 $^{\rm f}$ H- and $^{\rm 31}$ P-NMR spectra were recorded on a JEOL JNM-A500 spectrometer with tetramethylsilane ($^{\rm 1}$ H) and $\rm H_3PO_4$ ($^{\rm 31}$ P) as internal standards, respectively. The chemical shift values are expressed as δ (ppm) values. Multiplicities are indicated by s (singlet), d (doublet), t (triplet), m (multiplet), br (broad). FAB-MS or HR FAB-MS were measured on a JEOL JMS-HX110 spectrometer.

1,2-Di-*O*-stearoyl-sn-glycerol Benzyl *N*,*N*-Diisopropylphosphoramidite (7) According to the procedure described in ref. 9, coupling of 1,2-di-*O*-stearoyl-sn-glycerol (300 mg, 0.48 mmol) with benzyl *N*,*N*,*N'*,*N'*-tetraisopropylphosphoramidite (320 mg, 0.96 mmol) in the presence of 1*H*-tetrazole (68 mg, 0.97 mmol) gave **7** (409 mg, 0.48 mmol, 99%) as a colorless oil. 1 H-NMR (CDCl₃) δ : 0.88 (6H, t, 7.0 Hz), 1.17 (12H, m), 1.24—1.30 (56H, br m), 1.60 (4H, m), 2.28 (2H, t, 6.0 Hz), 2.29 (2H, t, 6.0 Hz), 3.58—3.82 (4H, m), 4.17 (1H, m), 4.34 (1H, m), 4.63—4.76 (2H, m), 5.19 (1H, m), 7.23—7.39 (5H, m).

 ${\bf 2,6\text{-}Di\text{-}}O\text{-}benzyloxymethyl-3,4,5\text{-}tri\text{-}}O\text{-}p\text{-}methoxybenzyl-D\text{-}}myo\text{-}inositol$ 1-O-(1,2-Di-O-stearoyl-sn-glycerol Benzylphosphate) (4) Anhydrous CH₂Cl₂ (10 ml) was added to a mixture of anhydrous 7 (409 mg, 0.48 mmol, dried by azeotropic removal of water with benzene), 2,6di-O-benzyloxymethyl-3,4,5-tri-O-p-methoxybenzyl-D-myo-inositol (3) (185 mg, 0.24 mmol) prepared by Prestwich's procedure 10) and 1H-tetrazole (120 mg, 1.70 mmol). After 30 min, a small amount of water was added. The mixture was further stirred for 30 min, then m-CPBA (330 mg, 1.34 mmol) was added at $-40 \,^{\circ}\text{C}$, and the reaction mixture was allowed to warm to room temperature. It was extracted with AcOEt, and the organic solution was washed with aqueous 10% Na₂SO₃ and aqueous saturated NaHCO3, then dried over anhydrous MgSO4. The solvent was removed in vacuo and the residue was purified by silica gel column chromatography with 5% AcOEt in hexane to give 4. ¹H-NMR (CDCl₃) δ : 0.88 (6H, t, 7 Hz), 1.21—1.31 (56H, br), 1.50—1.59 (4H, m), 2.17—2.25 (4H, m), 3.38—3.43 (2H, m), 3.77 (9H, s), 3.91—4.25 (8H, m), 4.43—5.17 (17H, m), 6.78—6.82 (6H, m), 7.17—7.32 (21H, m).

$$\begin{array}{c} R^2O \longrightarrow OR^1 \\ OH \longrightarrow OH \longrightarrow OH \\ H_2O_3PO \longrightarrow OH \longrightarrow OH \\ OPO_3H_2 \longrightarrow DH \\ \mathbf{1a} \ R^1 = stearoyl; \ R^2 = arachidonoyl \\ \mathbf{1b} \ R^1 = R^2 = stearoyl \end{array}$$

phosphatidylinositol 3,4,5-trisphosphate

Chart 1

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 $\label{eq:Reagents} \textbf{Reagents and conditions:} \ \textbf{a.} \ 1\\ H\text{-tetrazole, CH}_2\text{Cl}_2, \text{r.t., 30 min, then } m\text{-CPBA, -40}^\circ\text{C to r.t.} \\ \textbf{b. DDQ, wet CH}_2\text{Cl}_2, \text{r.t., 1h. } \textbf{c.} \ (\text{BnO})_2\text{PNE}_{12}, 1\\ H\text{-tetrazole, CH}_2\text{Cl}_2, \text{r.t., 30 min; then } m\text{-CPBA, -40}^\circ\text{C to r.t.} \\ \textbf{d. H}_2 \ (50 \text{ psi), Pd black, NaHCO}_3, 85\% \ \text{t-BuOH/H}_2\text{O} \\ \end{cases}$

Chart 2

IR (CHCl₃) cm⁻¹: 2900, 2870, 2830, 1740, 1610, 1590, 1510, 1460, 1360, 1300, 1240—1200, 1170, 1110, 1030, 920. FAB-MS (m-nitrobenzylalcohol (mNBA)) m/z: 1579, 1580 (M+Na). HR FAB-MS m/z: Calcd for $C_{92}H_{133}NaO_{18}P$, 1579.9127, Found 1579.9169.

2,6-Di-O-benzyloxymethyl-D-myo-inositol 1-O-(1,2-Di-O-stearoyl-snglycerol Benzylphosphate) (5) A solution of 4 in wet CH₂Cl₂ 5 ml at room temperature was treated with 95% DDQ (440 mg, 1.84 mmol). After having been stirred for 1 h, the reaction mixture was poured into aqueous saturated NaHCO3 and extracted with AcOEt. The organic layer was washed with brine, dried over anhydrous MgSO4 and concentrated in vacuo. The residue was purified by silica gel column chromatography with 60% AcOEt in hexane to give 5 (254 mg, 0.21 mmol, 90% from 3). ¹H-NMR (CDCl₃) δ : 0.88 (6H, t, 7.0 Hz), 1.23—1.27 (56H, br m), 1.53—1.60 (4H, br), 1.86 (1H, br), 2.22—2.28 (4H, m), 3.01 (1H, br), 3.34—3.44 (2H, m), 3.50 (1H, br), 3.70 (1H, t, 9.0 Hz), 3.78—3.85 (1H, m), 4.04—4.31 (7H, m), 4.53—5.09 (9H, m), 5.15 (1H, m), 7.27—7.35 (15H, m). IR (CHCl₃) cm⁻¹: 2920, 2850, 2400, 1740, 1460, 1380, 1160, 1110, 1020, 920. FAB-MS (mNBA) m/z: 1219 (M+Na). HR FAB-MS m/z: Calcd for $C_{68}H_{109}NaO_{15}P$, 1219.7402, Found 1219.7417.

2,6-Di-O-benzyloxymethyl-3,4,5-tris-O-(dibenzylphosphoryl)-D-myoinositol 1-O-(1,2-Di-O-stearoyl-sn-glycerol Benzyl Phosphate) (6) Dibenzyl N,N-diethylphosphoramidite (0.10 ml, 0.39 mmol) was added to a mixture of **5** (54 mg, 0.042 mmol) and 1*H*-tetrazole (50 mg, 0.71 mmol) in 3 ml of dry CH₂Cl₂ at room temperature. The mixture was stirred for 30 min, then a small amount of water was added. After additional stirring for 10 min, the mixture was treated with m-CPBA (100 mg, 0.41 mmol) at -40 °C, and the whole was allowed to warm to room temperature. It was extracted with AcOEt, washed with aqueous 10% Na₂SO₃, aqueous saturated NaHCO₃ and brine, and dried over anhydrous MgSO₄. The solvent was removed in vacuo and the residue was purified by silica gel column chromatography with 30% AcOEt in CH₂Cl₂ to give 6 (69 mg, 0.035 mmol, 84%). 1 H-NMR δ (CDCl₃): 0.80—0.90 (6H, br), 1.10—1.40 (56H, br), 1.50—1.70 (4H, br), 2.20 (4H, br), 2.90—3.10 (2H, br), 3.90—5.10 (31H, brm), 7.10—7.40 (45H, brm). IR (CHCl₃) cm⁻¹: 2940, 2860, 1740, 1600, 1500, 1460, 1380, 1270, 1080. FAB-MS

(mNBA) m/z: 1999 and 2000 (M+Na), 1977 (M+H). HR FAB-MS m/z: Calcd for $C_{110}H_{148}NaO_{24}P_4$, 1999.9209, Found 1999.9206.

3,4,5-Tris-*O*-phosphoryl-D-*myo*-inositol 1-*O*-(1,2-Di-*O*-stearoyl-sn-glycerol Phosphate) Sodium Salt (1b (Na)) Pd black (28 mg) and NaHCO₃ (6.4 mg, 0.078 mmol) were added to a solution of **6** (22 mg, 0.011 mmol) in 8.3 ml of 85% *tert*-BuOH in water, and the mixture was shaken under H₂ (50 psi) for 3 h. Precipitated insoluble sodium salts were dissolved by the addition of water and the mixture was shaken again under H₂ (50 psi) for a further 3 h. The catalyst was removed by filtration and washed with EtOH and water. The filtrate was concentrated *in vacuo* and lyophilized to give **1b** (Na) as the heptasodium salt. (13.6 mg). ¹H-NMR (D₂O) δ : 0.70 (6H, br), 1.00—1.50 (60H, br), 2.10—2.40 (4H, br), 3.80—4.50 (9H, br m), 5.20 (1H, br). ³¹P-NMR (D₂O) δ : -0.127, 1.984, 3.563, 4.419. Negative FAB-MS m/z: 1105 (M – H), 1127 (M + Na – 2H), 1149 (M + 2Na – 3H), 1171 (M + 3Na – 4H).

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