

Carbohydrate Research 264 (1994) 209-226

CARBOHYDRATE RESEARCH

# Synthesis of neoglycolipids containing a mucin-type core unit

Ryuichi Horie \*, Kouichi Nakano

Biotechnology Research Laboratory, TOSOH Corporation, 2743-1, Hayakawa, Ayase, Kanagawa 252, Japan

Received 30 December 1993; accepted 19 May 1994

#### Abstract

The unnatural glycolipids  $O-\beta$ -D-galactopyranosyl- $(1 \rightarrow 4)$ -O-(2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-(2-acetamido-2-deoxy- $\alpha$ -D-galactopyranosyl)- $(1 \rightarrow 1)$ -ceramide (1), $O-\beta$ -D-galactopyranosyl- $(1 \rightarrow 4)$ - $O-(2-acetamido-2-deoxy-\beta-D-glucopyranosyl)-(1 \rightarrow 6)$ -O-(2-ac-2)etamido-2-deoxy- $\alpha$ -D-galactopyranosyl)-(1 $\rightarrow$ 1)-ceramide (2), and O- $\beta$ -D-galactopyranosyl- $(1 \rightarrow 4)$ -O-(2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-[O- $\beta$ -D-galactopyranosyl- $(1 \rightarrow 4)$ -(2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 6)$ ]-O-(2-acetamido-2-deoxy- $\alpha$ -D-galactopyranosyl)- $(1 \rightarrow 1)$ -ceramide (3), and their  $\beta$ - $(1 \rightarrow 1)$ -linked isomers, were synthesized. The precursor oligosaccharides for 1, 2, and 3 were made by coupling O(2,3,4,6)-tetra-O-acetyl- $\beta$ -Dgalactopyranosyl)- $(1 \rightarrow 4)$ -3,6-di-O-acetyl-2-deoxy-2-phthalimido- $\alpha,\beta$ -D-glucopyranosyl trichloroacetimidate with tert-butyldiphenylsilyl 2-azido-4,6-O-benzylidene-2-deoxy-β-D-galactopyranoside, tert-butyldiphenylsilyl 2-azido-3-O-benzoyl-2-deoxy-B-D-galactopyranoside, and tert-butyl- $O-(2,3,4,6-\text{tetra-}O-\text{acetyl-}\beta-\text{D-galactopyranosyl})-(1 \rightarrow 4)-O-(3,6-\text{di-}O-\text{acetyl-}2-\text{de-})$ diphenylsilyl oxy-2-phthalimido- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-2-azido-2-deoxy- $\beta$ -D-galactopyranoside, respectively. These oligosaccharides were converted into their trichloroacetimidates, which were coupled with 3,2'-di-O-benzoyl ceramide. Deprotection of the coupling products gave the title compounds 1, 2, and 3.

Keywords: Neoglycolipids, synthesis of; Mucin-type core unit in neoglycolipids

## 1. Introduction

The cores of mucin-type carbohydrate chains have unique structures that have not been found in glycolipids. Typical structures such as T [1,2], Tn [3,4], and sialosyl Tn [5] have been chemically identified and considered to be tumor-associated antigens. In contrast

<sup>\*</sup> Corresponding author.



to the situation with the glycolipids, however, it is difficult to determine the distinct structure that is recognized by monoclonal antibodies against mucin-type antigens. To make the characterization of such monoclonal antibodies easier we have established a synthetic route to neoglycolipid [6] antigens such as 1, 2, and 3, which were designed to carry a mucintype core unit on a ceramide foundation [7]. The characteristic feature of a mucin-type core unit is an N-acetyl- $\alpha$ -D-galactosamine residue linked to L-serine or L-threonine. Since the configuration of the functionalized portion of the ceramides is the same as that of Lserine these neoglycolipids are expected to be good analogues of glycoproteins carrying mucin-type carbohydrate chains.

#### 2. Results and discussion

Boron trifluoride etherate-promoted glycosylation [8] of the 2-azido-2-deoxy-D-galactose derivative [9] **4** with lactosamine donor **5** in 1,2-dichloroethane was performed (Scheme 1) in the presence of activated, powdered molecular sieves AW-300 at  $-20^{\circ}$ C to give an 84% yield of **6**. Removal of the benzylidene group [10] of **6** with trifluoroacetic acid in dichloromethane gave an 80% yield of **7**, which was converted into **8** in two steps (*i*, hydrazine hydrate–ethanol [11]; *ii*, Ac<sub>2</sub>O–pyridine, 95% overall). The TBDPS protective group was removed with tetrabutylammonium fluoride in the presence of acetic acid [12] to give an anomeric mixture **9**, which on treatment with 1,8-diazabicyclo[5.4.0] undec-



OR

Bz NHAC AC

H NHAC H

h2

R<sup>3</sup>O

HNAc

12B Bz Na Ac

14

15

R<sup>1</sup> R<sup>2</sup> R<sup>3</sup>

TMSiOTf, molecular sieves CICH<sub>2</sub>CH<sub>2</sub>CI

OAc

HC

TBDPS = tert-butyldiphenylsilyl Phth = phthaloyl TMSiOTf = trimethylsilyl triflate



7-ene and trichloroacetonitrile afforded a trisaccharide imidate **10** (54% overall yield from **8**), as evidenced by <sup>1</sup>H NMR signals at  $\delta$  8.76 (s, 1 H, C=NH) and 6.44 (d, 1 H, J 3.4 Hz, H-1a $\alpha$ ). To serve as a glycosyl acceptor the protected ceramide **11** was prepared in 4 steps from natural ceramide, type IV, (*i*, reduction of the double bond by hydrogenation; *ii*, tritylation; *iii*, benzoylation; and *iv*, removal of the trityl group). The resultant compound was purified by reversed-phase HPLC (Fig. 1), and resolved into 8 peaks. Each peak was analyzed by FD-MS and 500 MHz NMR. The results showed that the material was 3,2'-di-O-benzoylated ceramide in which the  $\alpha$ -hydroxy fatty acid was mainly 24:0, with smaller amounts of 18:0, 22:0, 23:0, 25:0, 26:0, 27:0, and 28:0.

The glycosylation of the protected ceramide 11 with 10 was promoted with trimethylsilyl triflate in 1,2-dichloroethane at room temperature to give a mixture of 12 $\alpha$  and 12 $\beta$ . As judged from the intensities of the NMR signals of H-1a (12 $\alpha$ ,  $\delta_{\rm H}$  4.79, J 3.4 Hz; 12 $\beta$ , 4.02, J 7.8 Hz), the ratio of the  $\alpha$  and  $\beta$  glycosides was 2:3. The anomers were not separated in this stage; instead the azido groups of 12 $\alpha$  and 12 $\beta$  were transformed into acetamido groups in two steps (*i*, NaBH<sub>4</sub>–NiCl<sub>2</sub> in ethanol [13]; *ii*, Ac<sub>2</sub>O–pyridine) to give compounds 13 and 14. These were resolved by PLC on silica gel to give 17% of 13 and 24% of 14, based

C<sub>n</sub>H<sub>2n+1</sub>

C13H27

ŐB<sup>1</sup>

OR1



Fig. 1. Reversed-phase HPLC analysis of 3,2'-di-O-benzoylceramide (11) on TSK ODS-80TM. The solvent was 10:3 THF-H<sub>2</sub>O, pumped at a flow rate of 5 mL/min; temperature 45°C; column, 30 cm  $\times$  21.5 mm i.d.; sample, 21.3 mg.

on 11. Deprotection of both compounds 13 and 14 was carried out by treatment with sodium methoxide to give compound 1 and the  $\beta$  isomer 15. The structures of 1 and 15 were assigned from the reaction sequence, and established by their NMR data (Fig. 2; pyridine- $d_6$ - $D_2O$ , 50°C): 1,  $\delta$  5.36 (d, 1 H, J 3.9 Hz, H-1a); 15,  $\delta$  5.16 (d, 1 H, J 8.3 Hz, H-1a).

For the synthesis of the trisaccharide glycolipid 2 (Scheme 2), the 4,6-unprotected 2-azido-2-deoxygalactose derivative 16, obtained from 4 in two steps (*i*, benzoyl chloride-pyridine; *ii*,  $F_3CCO_2H-CH_2Cl_2$ ), was used as the acceptor. The glycosylation by 5 was promoted with boron trifluoride etherate in the presence of activated, powdered molecular sieves AW-300 in 1,2-dichloroethane at room temperature to give a 29% yield of 17. The 4-*O*-glycosylated regioisomer was not found in this reaction mixture. Conversion of 17 into







Scheme 2.

glycotriosyl donor 20 was carried out (56% overall yield) as described for the preparation of 10 from 7. The coupling of 20 and protected ceramide 11 was promoted with trimethylsilyl triflate in 1,2-dichloroethane at room temperature to give a mixture of 21 $\alpha$  and 21 $\beta$ , which were not separated in this step. As judged from the intensities of the NMR signals of H-1a (21 $\alpha$ ,  $\delta_{\rm H}$  4.93, J 3.9 Hz; 21 $\beta$ , 4.17, J 8.3 Hz), the ratio of the a and  $\beta$  glycosides was 4:3. The azido groups of 21 $\alpha$  and 21 $\beta$  were converted into acetamido groups, and the products were resolved by silica-gel column chromatography to give 11% of 22 and 9% of 23, based on 11. Deprotection of both compounds 22 and 23 was achieved by treatment with sodium methoxide, to give compound 2 and the  $\beta$  isomer 24. The structures of 2 and 24 were supported by their NMR data (Fig. 3; 2:1 CDCl<sub>3</sub>-CD<sub>3</sub>OD): 2,  $\delta$  4.52 (d, 1 H, J 3.4 Hz, H-1a); 24,  $\delta$  4.07 (d, 1 H, J 7.3 Hz, H-1a).

Boron trifluoride etherate-promoted glycosylation of the 2a-azido-2a-deoxy glycotriosyl derivative 7 with lactosamine donor 5 in 1,2-dichloroethane was performed (Scheme 3) in the presence of activated, powdered molecular sieves AW-300 at  $-20^{\circ}$ C to give compound 25. Compound 25 was converted into 26 in two steps(*i*, hydrazine hydrate–ethanol; *ii*, Ac<sub>2</sub>O–pyridine) in 68% overall yield. Assignment of NMR signals to the b and b' rings was confirmed by observation of an NOE between H-1b and H-6a, and comparison of



Scheme 3.

Compound	Chemical shifts ( $\delta$ ) at 400 MHz in CDCl <sub>3</sub>								
	GalNAc			GlcNAc			Gal		
	H-1a	H-2a	H-3a	H-1b	H-2b	H-3b	H-1c	H-2c	H-3c
8	4.21	3.64	3.40	4.55	3.94	5.02	4.54	5.10	4.97
18	4.48	3.79-3.65	4.66	4.27	3.75	4.97	4.45	5.12	4.95
26	4.34	3.62	3.47-3.32	4.54	3.96	5.01	4.53	5.10	4.97
				4.30	3.64	5.04	4.46	5.12	4.95

Table 1 <sup>1</sup>H NMR data

chemical shifts with those of compounds 8 and 18 (Table 1). Compound 26 was converted into the pentasaccharide imidate 28 via 27 (85% overall yield from 26);  $\delta_{\rm H}$  9.25 (s, 0.5 H, C=NH $\alpha$ ), 8.79 (s, 0.5 H, C=NH $\beta$ ), 5.30 (d, 0.5 H, J 3.5 Hz, H-1 $\alpha\alpha$ ), and 5.23 (d, 0.5 H, J 8.2 Hz, H-1 $\alpha\beta$ ). The glycosylation of protected ceramide 11 with 28, as with 10 and 20 was promoted with trimethylsilyl triflate in 1,2-dichloroethane at  $-5^{\circ}$ C to give a mixture of 29 $\alpha$  and 29 $\beta$ . As judged from the intensities of the NMR signals of H-1a (29 $\alpha$ ,  $\delta_{\rm H}$  4.83, J 3.4 Hz; 29 $\beta$ , 4.58, J 7.8 Hz), the ratio of the  $\alpha$  and  $\beta$  glycosides was 2:1. The azido groups of compounds 29 $\alpha$  and 29 $\beta$  were transformed into acetamido groups, and the products were separated by silica-gel column chromatography to give 13% of 30 and 6% of 31, based on 28. Deprotection of both 30 and 31 was accomplished by treatment with sodium methoxide, to give compounds 3 and 32. The structures of 3 and 32 were assigned from the reaction sequence, and established by their NMR data (Fig. 4; Me<sub>2</sub>SO-d<sub>6</sub>-D<sub>2</sub>O): 3,  $\delta_{\rm H}$  4.63 (d, 1 H, J 3.0 Hz, H-1a); 32,  $\delta$  4.27 (d, 1 H, J 8.0 Hz, H-1a).

In conclusion, we have established a facile route to the synthesis of mucin-type neoglycolipids 1, 2, and 3, and now we are in a position to generate specific monoclonal antibodies against mucin-type carbohydrate chains.



Fig. 4. 400 MHz <sup>1</sup>H NMR spectrum of compound 3 in Me<sub>2</sub>SO- $d_6$ - $D_2O$ .

### 3. Experimental

General methods.—<sup>1</sup>H NMR spectra were recorded at 200 MHz with a Varian Gemini 200 and at 400 MHz with a Jeol GX400 instrument for samples dissolved in the solvents noted. Field desorption mass spectra (FD-MS) were recorded with a Hitachi M-80B spectrometer. Column chromatography was carried out on silica gel (Merck Silica 60), with the solvent systems noted. TLC and high performance TLC were performed on Silica Gel 60  $F_{254}$  (Merck). HPLC analysis was performed on columns of TSK gel Silica 60 and ODS-80TM (TOSOH Corporation, Japan).

 $O-(2,3,4,6-tetra-O-acetyl-\beta-D-galactopyranosyl)-(1 \rightarrow 4)-O$ tert-Butyldiphenylsilyl  $(3,6-di-O-acetyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosyl)-(1 \rightarrow 3)-2-azido-4,6-O-ben$ zylidene-2-deoxy-β-D-galactopyranoside (6).-To a stirred mixture of O-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)- $(1 \rightarrow 4)$ -3,6-di-O-acetyl-2-deoxy-2-phthalimido- $\alpha,\beta$ -Dglucopyranosyl trichloroacetimidate [8] (5, 0.869 g, 1.00 mmol), tert-butyldiphenylsilyl 2-azido-4,6-O-benzylidene-2-deoxy-β-D-galactopyranoside [9] (4, 0.426 g, 0.80 mmol), and molecular sieves AW-300 (1.00 g) in 1,2-dichloroethane (11 mL) was added 1.0 M  $BF_3 \cdot E t_2O (0.20 \text{ mL})$  at  $-20^{\circ}C$ . The mixture was stirred for 2 h at  $-20^{\circ}C$ , then filtered, and the solids were washed with CHCl<sub>3</sub>. The filtrate was washed with aq NaHCO<sub>3</sub> and aq NaCl, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The residue was purified by chromatography on silica gel using 3:1 hexane-EtOAc to afford compound 6 (0.827 g, 0.67 mmol, 84%). NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  5.51 (d, 1 H,  $J_{1,2}$  8.3 Hz, H-1b), 5.46 (s, 1 H, PhCH), 4.98 (dd, 1 H, J<sub>3,4</sub> 3.4 Hz, H-3c), 4.76 (dd, 1 H, J<sub>5,6</sub> 2.4 Hz, H-6b), 4.59 (d, 1 H, J<sub>1,2</sub> 7.8 Hz, H-1c), 4.30 (d, 1 H, J<sub>1.2</sub> 7.8 Hz, H-1a), 4.28 (dd, 1 H, J<sub>2.3</sub> 10.3 Hz, H-2b), 4.11 (dd,  $1 \text{ H}, J_{4.5} 0.5 \text{ Hz}, \text{H-4a}, 4.09-4.01 \text{ (m, 2 H, H-6c,6'c)}, 4.02 \text{ (dd, 1 H}, J_{6.6} 12.2 \text{ Hz}, \text{H-6'b)},$ 3.91–3.81 (m, 4 H, H-6a,6'a,4b,5c), 3.71 (m, 1 H, J<sub>5.6</sub>, 3.9 Hz, H-5b), 3.67 (dd, 1 H, J<sub>2.3</sub> 10.7 Hz, H-2a), 3.18 (dd, 1 H, J<sub>34</sub> 3.4 Hz, H-3a), 2.87 (dt, 1 H, H-5a), 2.14, 2.05, 2.04, 2.03, 1.96, 1.92 (6 s, 18 H, Ac), and 1.01 (s, 9 H, t-Bu); FD-MS: m/z 1236.8 (M<sup>+</sup>). Anal. Calcd for C<sub>61</sub>H<sub>68</sub>N<sub>4</sub>O<sub>22</sub>Si: C, 59.21; H, 5.54; N, 4.53. Found: C, 59.3; H, 5.5; N, 4.5.

 $O-(2, 3, 4, 6-tetra-O-acetyl-\beta-D-galactopyranosyl)-(1 \rightarrow 4)-O$ tert-Butyldiphenylsilyl  $(3, 6-di-O-acetyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosyl)-(1 \rightarrow 3)-2-azido-2-deoxy-\beta-$ D-galactopyranoside (7).—To a stirred solution of 6 (0.800 g, 0.65 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (15 mL) was added aq 60% CF<sub>3</sub>COOH (0.5 mL) at 0°C, and the mixture was stirred overnight at room temperature. It was then added to aq NaHCO<sub>3</sub> (50 mL), and extraction was completed after dilution with CHCl<sub>3</sub>. The organic layer was washed with aq NaHCO<sub>3</sub> and aq NaCl, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The residue was purified by chromatography on silica gel using 3:1 hexane-EtOAc to yield compound 7 (0.593 g, 0.52 mmol, 80%). NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{H}$  5.75 (dd, 1 H,  $J_{3,4}$  8.3 Hz, H-3b), 5.49 (d, 1 H,  $J_{1,2}$  8.8 Hz, H-1b), 5.34 (d, 1 H,  $J_{4,5}$  1.0 Hz, H-4c), 5.12 (dd, 1 H,  $J_{2,3}$  10.3 Hz, H-2c), 4.98 (dd, 1 H, J<sub>3,4</sub> 3.4 Hz, H-3c), 4.58 (dd, 1 H, H-6b), 4.56 (d, 1 H, J<sub>1,2</sub> 7.8 Hz, H-1c), 4.32 (d, 1 H,  $J_{1,2}$  7.3 Hz, H-1a), 4.27 (dd, 1 H,  $J_{2,3}$  10.7 Hz, H-2b), 4.12–4.01 (m, 3 H, H-6'b,6c,6'c), 3.89–3.79 (m, 4 H, H-4a,4b,5b,5c), 3.64 (dd, 1 H,  $J_{6,6'}$  11.7 Hz, H-6a), 3.50 (dd, 1 H, J<sub>2.3</sub> 10.3 Hz, H-2a), 3.44 (dd, 1 H, J<sub>5.6</sub>, 3.9 Hz, H-6'a), 3.20 (dd, 1 H, J<sub>3.4</sub> 3.4 Hz, H-3a), 3.04 (dt, 1 H, J<sub>5.6</sub> 7.3 Hz, H-5a), 2.14, 2.09, 2.05, 2.04, 1.96, 1.93 (6 s, 18 H, Ac), and 1.01 (s, 9 H, t-Bu); FD-MS: m/z 1148.8 (M<sup>+</sup>). Anal. Calcd for C<sub>54</sub>H<sub>64</sub>N<sub>4</sub>O<sub>22</sub>Si · H<sub>2</sub>O: C, 55.57; H, 5.70; N, 4.80. Found: C, 55.1; H, 5.3; N, 4.4.

tert-Butyldiphenylsilyl O-(2, 3, 4, 6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)-(1  $\rightarrow$  4)-O-(2acetamido-3, 6-di-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-4, 6-di-O-acetyl-2azido-2-deoxy- $\beta$ -D-galactopyranoside (8).—A solution of 7 (0.254 g, 0.22 mmol) in 50:1 EtOH-H<sub>2</sub>NNH<sub>2</sub>·H<sub>2</sub>O (12.5 mL) was stirred under reflux overnight and then concentrated in vacuo. The residue was dissolved in pyridine (5 mL), and to the solution was added  $Ac_2O(2 \text{ mL})$  and a trace of 4-dimethylaminopyridine. The solution was stirred overnight at room temperature, then diluted with CHCl<sub>3</sub>, successively washed with aq NaHCO<sub>3</sub> and aq NaCl, dried over  $Na_2SO_4$ , and concentrated in vacuo. The resulting residue was purified by chromatography on silica gel using 1:2 hexane-EtOAc to yield compound 8 (0.241 g, 0.21 mmol, 95%). NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{H}$  5.34 (d, 1 H,  $J_{4,5}$  1.0 Hz, H-4c), 5.45 (d, 1 H, J<sub>2,NH</sub> 9.5 Hz, NH-b), 5.20 (dd, 1 H, J<sub>4,5</sub> 0.6 Hz, H-4a), 5.10 (dd, 1 H, J<sub>2,3</sub> 10.5 Hz, H-2c), 5.02 (dd, 1 H,  $J_{3,4}$  8.9 Hz, H-3b), 4.97 (dd, 1 H,  $J_{3,4}$  3.4 Hz, H-3c), 4.55 (d, 1 H, J<sub>1,2</sub> 7.9 Hz, H-1b), 4.54 (d, 1 H, J<sub>1,2</sub> 7.9 Hz, H-1c), 4.45 (dd, 1 H, H-6b), 4.31 (d, 1 H, J<sub>1.2</sub> 7.7 Hz, H-1a), 4.10 (dd, 1 H, H-6c), 4.09 (dd, 1 H, H-6'c), 3.98-3.91 (m, 4 H, H-6a,6'a,2b,6'b), 3.86 (dt, 1 H, H-5c), 3.82 (dd, 1 H, J<sub>4.5</sub> 9.3 Hz, H-4b), 3.64 (dd, 1 H, J<sub>2.3</sub> 10.3 Hz, H-2a), 3.49–3.42 (m, 2 H, H-5a,5b), 3.40 (dd, 1 H, J<sub>3,4</sub> 3.4 Hz, H-3a), 2.15, 2.11, 2.07, 2.06, 2.03, 1.96, 1.94, 1.92, 2.01 (9 s, 27 H, Ac), and 1.01 (s, 9 H, t-Bu); FD-MS: m/z 1167 (M + Na)<sup>+</sup>. Anal. Calcd for C<sub>52</sub>H<sub>68</sub>N<sub>4</sub>O<sub>23</sub>Si: C, 54.54; H, 5.98; N, 4.89. Found: C, 54.6; H, 6.0; N, 5.1.

O-(2, 3, 4, 6-Tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 3)-4, 6-di-O-acetyl-2-azido-2-deoxy-α,β-D-galactopyranose (9).—A solution of compound 8 (0.240 g, 0.21 mmol) and AcOH (14 μL, 0.24 mmol) in dry THF (5 mL) was cooled to  $-10^{\circ}$ C. Tetrabutylammonium fluoride (1 M in THF, 0.55 mL) was added dropwise. The mixture was stirred for 2 h at  $-10^{\circ}$ C, then diluted with H<sub>2</sub>O (50 mL). The mixture was extracted with CHCl<sub>3</sub>, and the organic layer was washed with aq NaCl, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The resulting residue was purified by chromatography on silica gel using 1:2 hexane–EtOAc to yield compound 9 (0.125 g, 0.14 mmol, 66%). NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  5.78 (d, 0.2 H,  $J_{2,\rm NH}$  9.3 Hz, NH-bα), 5.74 (d, 0.8 H,  $J_{2,\rm NH}$  9.3 Hz, NH-bβ), 4.65 (d, 0.8 H,  $J_{1,2}$  6.8 Hz, H-1bβ), 4.54 (d, 1 H,  $J_{1,2}$  8.3 Hz, H-1c), 4.07 (bd, 0.8 H,  $J_{1,2}$  8.6 Hz, H-1aβ), and 4.05 (d, 0.2 H,  $J_{1,2}$  3.5 Hz, H-1aα); FD-MS: m/z 906.9 (M<sup>+</sup>). Anal. Calcd for C<sub>36</sub>H<sub>50</sub>N<sub>4</sub>O<sub>23</sub>·2H<sub>2</sub>O: C, 45.86; H, 5.77; N, 5.94. Found: C, 46.0; H, 5.4; N, 5.6.

3-O-Benzoyl-2-N-[ (2R)-2-benzoyloxyalkanoyl]sphinganine (3, 2'-di-O-benzoylceramide) (11).—A mixture of ceramide (0.248 g, type IV, purchased from Sigma Chemical Co., St. Louis, MO) and 10% Pd–C (0.020 g) in THF (30 mL) was stirred for 15 h at room temperature under H<sub>2</sub>, and filtered through Celite. The filtrate was concentrated in vacuo to give hydrogenated ceramide (0.205 g). The residue and trityl chloride (0.189 mg, 0.678 mmol) were dissolved in pyridine (5 mL) and the mixture was stirred for 6 days, then benzoyl chloride (0.095 mg, 0.678 mmol) was added. This mixture was stirred overnight, and poured into 1 M NaHCO<sub>3</sub>. The product was recovered by extraction with CHCl<sub>3</sub>, and the extract was washed with H<sub>2</sub>O, dried (MgSO<sub>4</sub>), and concentrated in vacuo. Chromatography of the residue over silica gel in 10:1 hexane–EtOAc gave 3,2'-di-O-benzoyl-1-O-trityl ceramide (0.337 mg). This compound was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (10 mL), and added to aq 60% CF<sub>3</sub>COOH (1 mL). The mixture was stirred for 2 h, then poured into 1 M NaHCO<sub>3</sub>. The product was recovered by extraction with CHCl<sub>3</sub>, and the extract was recovered by extraction with CHCl<sub>3</sub>, and the extract was recovered by extraction with CHCl<sub>3</sub>, and the extract was recovered by extraction with CHCl<sub>3</sub>, and the extract was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (10 mL), and added to aq 60% CF<sub>3</sub>COOH (1 mL). The mixture was stirred for 2 h, then poured into 1 washed with H<sub>2</sub>O, dried (MgSO<sub>4</sub>), and concentrated in vacuo. Chromatography of the residue over silica gel in 4:1 hexane–EtOAc gave **11** (0.114 g). NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  7.06 (d, 1 H,  $J_{2,\rm NH}$  8.5 Hz, NH), 5.41 (t, 1 H,  $J_{2',3'\rm A} = J_{2',3'\rm B} = 5.5$  Hz, H-2')<sup>1</sup>, 5.14 (ddd, 1 H,  $J_{2,3}$  7.0,  $J_{3,4\rm A}$  9.5,  $J_{3,4\rm B}$  3.5 Hz, H-3), 4.19 (m, 1 H,  $J_{1,2}$  3.0,  $J_{1,2}$  11.5,  $J_{2,3}$  7.0 Hz, H-2), 3.65 (ddd, 2 H,  $J_{1\rm A,2}$  3.0,  $J_{1\rm B,2}$  11.5,  $J_{1,1}$  25.0 Hz, H-1A,1B), 2.85 (m, 1 H, OH), 2.01 (m, 2 H, H-3'A,3'B), 1.84 (ddd, 1 H,  $J_{3,4\rm A}$  9.5,  $J_{4\rm A,5}$  5.0,  $J_{4\rm A,4\rm B}$  18.5 Hz, H-4A), 1.69 (ddd, 1 H,  $J_{3,4\rm B}$  3.5,  $J_{4\rm B,5}$  8.5,  $J_{4\rm A,4\rm B}$  18.5 Hz, H-4B), 1.25 (m, CH<sub>2</sub>), and 0.88 (t, 6 H, J 6.5 Hz, CH<sub>3</sub>).

Reversed-phase HPLC of the 3,2'-di-O-benzoyl ceramide was performed on TSK gel ODS-80TM (Fig. 1). The mixture was resolved into 8 peaks, which gave the following FD-MS: 1 (18:0,  $C_{50}H_{81}NO_6$ ), m/z 792 (M)<sup>+</sup>; 2 (22:0,  $C_{54}H_{89}NO_6$ ), m/z 848 (M)<sup>+</sup>; 3 (23:0,  $C_{55}H_{91}NO_6$ ), m/z 862 (M)<sup>+</sup>; 4 (24:0,  $C_{56}H_{93}NO_6$ ), m/z 876 (M)<sup>+</sup>; 5 (25:0,  $C_{57}H_{95}NO_6$ ), m/z 890 (M)<sup>+</sup>; 6 (26:0,  $C_{58}H_{97}NO_6$ ), m/z 904 (M)<sup>+</sup>; 7 (27:0,  $C_{59}H_{99}NO_6$ ), m/z 918 (M)<sup>+</sup>; 8 (28:0,  $C_{60}H_{101}NO_6$ ), m/z 932 (M)<sup>+</sup>. All spectra showed a peak at m/z 105 (Bz).

O(2, 3, 4, 6-Tetra-O-acetyl- $\beta$ -D-galactopyranosyl $)-(1 \rightarrow 4)$ -O-(2-acetamido-3, 6-di-Oacetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  3)-O-(4, 6-di-O-acetyl-2-azido-2-deoxy- $\alpha$ -Dgalactopyranosyl)- $(1 \rightarrow 1)$ -3-O-benzoyl-2-N-[(2R)-2-benzoyloxyalkanoyl]sphinganine (12 $\alpha$ ) and O-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 4)-O-(2-acetamido-3,6di-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-(4, 6-di-O-acetyl-2-azido-2-deoxy- $\beta$ -D-galactopyranosyl)- $(1 \rightarrow 1)$ -3-O-benzoyl-2-N- $\{(2R)$ -2-benzoyloxyalkanoyl]-sphinganine  $(12\beta)$ .—To a solution of 9 (0.100 g, 0.11 mmol) in 1,2-dichloroethane (1.0 mL) was added CCl<sub>3</sub>CN (0.11 mL) and 1,8-diazabicyclo[5.4.0] undec-7-ene (0.02 mL, 0.13 mmol). The mixture was stirred for 3 h, and directly subjected to column chromatography on silica gel using 1:2 toluene-EtOAc to afford  $\alpha$ -trichloroacetimidate 10 (0.095 g, 0.090 mmol, 82%). NMR (200 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  8.76 (s, 1 H, C=NH), 6.44 (d, 1 H,  $J_{1,2}$  3.4 Hz, H-1a), 5.53 (dd, 1 H, H-4a), 5.50 (d, 1 H,  $J_{2,NH}$  9.8 Hz, NH-b), 5.35 (dd, 1 H,  $J_{4.5}$  1.0 Hz, H-4c), 5.11 (dd, 1 H, J<sub>2.3</sub> 10.7 Hz, H-2c), 5.05 (dd, 1 H, J<sub>3.4</sub> 8.8 Hz, H-3b), 4.97 (dd, 1 H, J<sub>3,4</sub> 3.4 Hz, H-3c), 4.68 (dd, 1 H, H-6b), 4.67 (d, 1 H, J<sub>1,2</sub> 8.3 Hz, H-1b), 4.52 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 4.30 (dt, 1 H, H-5a), 4.16–3.95 (m, 8 H, H-2a, 3a, 6a, 6'a, 2b, 6'b,6c,6'c), 3.88 (dt, 1 H, H-5c), 3.83 (dd, 1 H, J<sub>4.5</sub> 9.3 Hz, H-4b), 3.59 (m, 1 H, H-5b), 2.15, 2.11, 2.07, 2.06, 2.03, 1.96, 1.94, 1.92 (8 s, 27 H, Ac), and 1.09 (s, 9 H, t-Bu).

To a stirred mixture of 10 (0.091 g, 0.087 mmol), 11 (0.069 g, 0.079 mmol), and powdered molecular sieves AW-300 (0.12 g) in 1,2-dichloroethane (2.0 mL) was added M TMSiOTf in 1,2-dichloroethane (0.20 mL) at  $-20^{\circ}$ C. The mixture was stirred overnight at room temperature, diluted with CHCl<sub>3</sub>, and filtered through Celite. The filtrate was washed with aq NaHCO<sub>3</sub> and aq NaCl, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The resulting residue was purified on silica gel using 1:2 hexane–EtOAc to yield a mixture of anomers 12 $\alpha$  and 12 $\beta$  (0.068 g, 0.39 mmol, 49%). NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  6.95 (d, 0.6 H, Cer NH- $\beta$ ), 6.91 (d, 0.4 H, Cer NH- $\alpha$ ), 4.79 (d, 0.4 H,  $J_{1,2}$  3.4 Hz, H-1a $\alpha$ ), 4.55 (d, 0.6 H,  $J_{1,2}$  7.8 Hz, H-1c $\beta$ ), 4.53 (d, 0.4 H,  $J_{1,2}$  7.8 Hz, H-1c $\alpha$ ), 4.36 (d, 0.6 H,  $J_{1,2}$  8.3 Hz, H-1b $\beta$ ), 4.28 (d, 0.4 H,  $J_{1,2}$  7.8 Hz, H-1b $\alpha$ ), 4.02 (d, 0.6 H,  $J_{1,2}$  7.8 Hz, H-1a $\beta$ ), 1.20 (m, CH<sub>2</sub>), and 0.85 (t, CH<sub>3</sub>).

 $<sup>^{1}</sup>$  The letters A and B are used to designate the two magnetically nonequivalent protons of the pairs attached to C-3 of the ceramide fatty acyl group (i.e., C-3') and to C-1 and C-4 of the sphinganine chain.

O-(2, 3, 4, 6-Tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2-acetamido-3, 6-di-Oacetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  3)-O-(2-acetamido-4, 6-di-O-acetyl-2-deoxy- $\alpha$ -D-galactopyranosyl)- $(1 \rightarrow 1)$ -3-O-benzoyl-2-N-[(2R)-2-benzoyloxyalkanoyl]sphinganine (13) and O-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)-(1  $\rightarrow$  4)-O-(2-acetamido-3,6di-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-(2-acetamido-4,6-di-O-acetyl-2-de $oxy-\beta-D-galactopyranosyl)-(1 \rightarrow 1)-3-O-benzoyl-2-N-f(2R)-2-benzoyloxy-alkanoyl]sphin$ ganine (14).—A mixture of  $12\alpha$  and  $12\beta$  (0.083 g, 0.047 mmol) was dissolved in a solution of NiCl<sub>2</sub> · 6H<sub>2</sub>O (0.183 g, 0.77 mmol) and H<sub>3</sub>BO<sub>3</sub> (0.093 g, 1.50 mmol) in EtOH (4 mL). To this solution was added NaBH<sub>4</sub> (0.050 g, 1.32 mmol) at room temperature, the mixture was stirred for 1.5 h at room temperature, then Ac<sub>2</sub>O was added, and stirring was continued overnight at room temperature. Methanol was then added to the mixture, and it was concentrated in vacuo. The residue was dissolved in CHCl<sub>3</sub>, and the organic layer was washed with aq NaHCO<sub>3</sub> and aq NaCl, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The resulting residue was purified by chromatography on silica gel using 1:2 hexane-EtOAc to afford a mixture of compounds 13 and 14 (0.077 g). This mixture was purified by PLC on silica gel to afford 13 (0.029 g, 0.016 mmol) and 14 (0.040 g, 0.022 mmol) (25% and 48%, respectively, from the mixture  $12\alpha + 12\beta$ ). NMR data for 13 (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$ 7.32 (d, 1 H, Cer NH), 5.35 (dd, 1 H, H-4c), 5.33 (t, 1 H, Cer H-2'), 5.22 (dd, 1 H, Cer H-3), 5.16 (dd, 1 H, H-4a), 5.15 (dd, 1 H, J<sub>3.4</sub> 8.8 Hz, H-3b), 5.12 (dd, 1 H, J<sub>2.3</sub> 10.3 Hz, H-2c), 4.97 (dd, 1 H, J<sub>3,4</sub> 3.4 Hz, H-3b), 4.70 (d, 1 H, J<sub>1,2</sub> 7.8 Hz, H-1b), 4.69 (dd, 1 H, H-6b), 4.63 (d, 1 H, J<sub>1,2</sub> 3.9 Hz, H-1a), 4.54 (d, 1 H, J<sub>1,2</sub> 7.8 Hz, H-1c), 4.44 (m, 1 H, Cer H-2), 4.40 (dd, 1 H,  $J_{2,3}$  10.8 Hz, H-2a), 4.12 (m, 2 H, H-6c,6'c), 4.04 (dd, 1 H, H-6'b), 4.01 (dt, 1 H, H-5a), 3.95 (dd, 1 H, J<sub>3,4</sub> 2.9 Hz, H-3a), 3.92 (m, 2 H, H-6a,6'a), 3.88 (dt, 1 H, H-5c), 3.84–3.80 (m, 2 H, Cer H-1A, H-4b), 3.66 (dd, 1 H, J<sub>2.3</sub> 10.3 Hz, H-2b), 3.58 (m, 1 H, H-5b), 3.26 (dd, 1 H, Cer H-1B), 2.15, 2.14, 2.08, 2.06, 2.05, 2.01, 1.97, 1.95 (8) s, 30 H, Ac), 1.24 (m,  $CH_2$ ), and 0.84 (t,  $CH_3$ ).

NMR data for 14 (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  6.88 (d, 1 H, J 8.1 Hz, Cer NH), 5.35 (dd, 1 H, H-4c), 5.32 (t, 1 H, Cer H-2'), 5.29 (dd, 1 H, H-4a), 5.17 (t, 1 H, Cer H-3), 5.11 (dd, 1 H,  $J_{2,3}$  10.3 Hz, H-2c), 5.01 (dd, 1 H,  $J_{3,4}$  8.8 Hz, H-3b), 4.97 (dd, 1 H,  $J_{3,4}$  3.4 Hz, H-3c), 4.82 (d, 1 H,  $J_{1,2}$  8.3 Hz, H-1a), 4.58 (dd, 1 H, H-6b), 4.51 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1b), 4.51 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 4.51 (dd, 1 H,  $J_{3,4}$  3.4 Hz, H-3a), 4.37 (m, 1 H, Cer H-2), 4.10 (m, 2 H, H-6c,6'c), 4.00 (dd, 1 H, H-6'b), 3.99–3.76 (m, 5 H, H-6a,4b,5c, Cer H-1A,1B), 3.97 (dd, 1 H, H-6'a), 3.91 (dd, 1 H,  $J_{2,3}$  10.3 Hz, H-2b), 3.72 (dt, 1 H, H-5a), 3.52 (m, 1 H, H-5b), 3.09 (dd, 1 H,  $J_{2,3}$  10.7 Hz, H-2a), 2.15, 2.14, 2.06, 2.05, 2.04, 1.99, 1.97, 1.95 (8 s, 30 H, Ac), 1.24 (m, CH<sub>2</sub>), and 0.84 (t, CH<sub>3</sub>).

O-β-D-Galactopyranosyl-(1→4)-O-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1→3)-O-(2-acetamido-2-deoxy-α-D-galactopyranosyl)-(1→1)-2-N-[(2R)-2-hydroxyalkanoyl]sphinganine (1).—A solution of compound 13 (0.016 g, 0.0086 mmol) in 1:1 MeOH-THF (3 mL) containing M NaOMe-MeOH (0.1 mL) was stirred overnight at room temperature, and made neutral with Dowex 50W-X8. The suspension was filtered, and the filtrate was concentrated in vacuo. The resultant residue was purified by PLC on silica gel using 55:40:10 CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O afford to 1 (0.004 g, 0.032 mmol, 37%). NMR (400 MHz, pyridine-d<sub>6</sub>-D<sub>2</sub>O, 50°C):  $\delta_{\rm H}$  5.47 (d, 1 H, J<sub>1,2</sub> 8.3 Hz, H-1b), 5.36 (d, 1 H, J<sub>1,2</sub> 3.9 Hz, H-1a), 5.29 (dd, 1 H, H-2a), 4.97 (d, 1 H, J<sub>1,2</sub> 7.8 Hz, H-1c), 4.74 (m, 1 H, H-4a), 4.73-4.64 (m, 3 H, Cer H-2,3,2'), 4.61 (dd, 1 H, H-3a), 4.55 (dd, 1 H, H-3b), 4.46 (dd, 1 H, H-4c), 4.44–4.05 (m, 14 H, H-5a,6a,6'a,2b,4b,6b,6'b,2c,3c,5c,6c,6'c, Cer H-1A,1B), 3.77 (m, 1 H, H-5b), 2.45, 2.20 (2 s, 6 H, Ac), 1.30 (m,  $CH_2$ ), and 0.88 (t,  $CH_3$ ).

O-β-D-Galactopyranosyl-(1→4)-O-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1→3)-O-(2-acetamido-2-deoxy-β-D-galactopyranosyl)-(1→1)-2-N-[(2R)-2-hydroxyalkanoyl]sphinganine (15).—A solution of compound 14 (0.029 g, 0.016 mmol) in 1:1 MeOH-THF (6 mL) containing 0.1 M NaOMe-MeOH (0.17 mL) was stirred overnight at room temperature, then made neutral with Dowex 50W-X8. The suspension was filtered, and the filtrate was concentrated in vacuo. The resultant residue was purified by PLC on silica gel using 55:40:10 CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O to afford 15 (0.007 g, 0.0057 mmol, 35%). NMR (400 MHz, pyridine-d<sub>6</sub>-D<sub>2</sub>O, 50°C):  $\delta_{\rm H}$  5.52 (d, 1 H, J<sub>1,2</sub> 8.3 Hz, H-1b), 5.16 (d, 1 H, J<sub>1,2</sub> 8.3 Hz, H-1a), 4.98 (d, 1 H, J<sub>1,2</sub> 7.8 Hz, H-1c), 4.75 (dd, 1 H, H-2a), 4.72 (m, 1 H, H-4a), 4.68 (m, 1 H, Cer H-2), 4.67 (dd, 1 H, H-3a), 4.65-4.56 (m, 3 H, H-3b, Cer H-1A,2'), 4.46 (dd, 1 H, H-4c), 4.44 (dd, 1 H, H-2c), 4.41-4.02 (m, 13 H, H-3,5a,6a,6'a,2b,4b,6b,6'b,3c,5c,6c,6'c, Cer H-1B), 3.82 (m, 1 H, H-5b), 2.40, 2.15 (2 s, 6 H, Ac), 1.25 (m, CH<sub>2</sub>), and 0.88 (t, CH<sub>3</sub>).

tert-Butyldiphenylsilyl 2-azido-3-O-benzoyl-2-deoxy- $\beta$ -D-galactopyranoside (16).—To a stirred mixture of compound 4 (0.300 g, 0.56 mmol) and 4-dimethylaminopyridine (12 mg) in CH<sub>2</sub>Cl<sub>2</sub> 20 mL) was added pyridine (0.9 mL) and benzoyl chloride (95 mg, 0.68 mmol). The solution was stirred for 20 h at room temperature, then diluted with CH<sub>2</sub>Cl<sub>2</sub>, successively washed with aq NaHCO<sub>3</sub>, and aq NaCl, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The resulting residue was subjected to column chromatography on silica gel using 4:1 hexane-EtOAc to give a yellow syrup. This residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (15 mL), aq 60% CF<sub>3</sub>COOH (0.5 mL) was added at 0°C, and the mixture was stirred overnight at room temperature. It was then poured into aq NaHCO<sub>3</sub> (50 mL), and extraction was completed after dilution with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed with aq NaHCO<sub>3</sub> and aq NaCl, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The residue was purified by chromatography on silica gel using 3:2 hexane-EtOAc to yield compound 16 (0.168 g, 0.31 mmol, 54% based on 4). NMR (200 MHz,  $CDCl_3$ ):  $\delta_H$  4.80 (dd, 1 H,  $J_{3,4}$  3.0 Hz, H-3), 4.66 (d, l H, J<sub>1,2</sub> 7.8 Hz, H-l), 4.10 (d, l H, H-4), 4.03 (dd, 1 H, J<sub>2,3</sub> 10.8 Hz, H-2), 3.65 (dd, 1 H, J<sub>5,6</sub> 5.8, J<sub>6,6'</sub> 12.0 Hz, H-6), 3.48 (dd, 1 H, J<sub>5,6'</sub> 3.8 Hz, H-6'), 3.21 (m, 1 H, H-5), and 1.11 (s, 9 H, t-Bu). Anal. Calcd for C<sub>29</sub>H<sub>33</sub>N<sub>3</sub>O<sub>6</sub>Si · H<sub>2</sub>O: C, 61.58; H, 6.23; N, 7.43. Found: C, 61.63; H, 6.50; N, 7.22.

tert-Butyldiphenylsilyl O-(2, 3, 4, 6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)-(1  $\rightarrow$  4)-O-(2acetamido-3,6-di-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  6)-3,4-di-O-acetyl-2-azido-2-deoxy- $\beta$ -D-galactopyranoside (18).—To a stirred mixture of 5 (2.26 g, 2.64 mmol), 16 (1.100 g, 2.01 mmol), and 1.00 g of molecular sieves AW-300 in 1,2-dichloroethane was added M BF<sub>3</sub> · Et<sub>2</sub>O (0.65 mL) at  $-20^{\circ}$ C, then the suspension was filtered, and the solids were washed with CHCl<sub>3</sub>. The filtrate was washed with aq NaHCO<sub>3</sub> and aq NaCl, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The residue was purified by chromatography on silica gel using 3:1 hexane–EtOAc to yield compound 17 (0.783 g, 0.59 mmol, 29%). NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  5.67 (dd, 1 H,  $J_{3,4}$  8.3 Hz, H-3b), 5.33 (d, 1 H,  $J_{1,2}$  8.8 Hz, H-1b), 5.31 (d, 1 H, H-4c), 5.11 (dd, 1 H,  $J_{2,3}$  10.7 Hz, H-2c), 4.95 (dd, 1 H,  $J_{3,4}$  3.4 Hz, H-3c), 4.76 (dd, 1 H,  $J_{5,6}$  1.9 Hz, H-6b), 4.70 (dd, 1 H,  $J_{3,4}$  2.9 Hz, H-3a), 4.54 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 4.40 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1a), 4.10–3.96 (m, 5 H, H-2a,4a,2b,6c,6'c), 3.93 (dd, 1 H,  $J_{6,6'}$  12.2 Hz, H-6'b), 3.86 (t, 1 H,  $J_{5,6}$  9.3 Hz, H-6a), 3.83 (dt, 1 H, H-5c), 3.78 (dd, 1 H,  $J_{4,5}$  10.3 Hz, H-4b), 3.70 (m, 1 H, H-5b), 3.45 (dd, 1 H,  $J_{5,6'}$  4.4,  $J_{6,6'}$  9.3 Hz, H-6'a), 3.26 (d, 1 H, J 5.4 Hz, OH-4a), 3.22 (dd, 1 H, H-5a), 2.35, 2.13, 2.03, 1.95, 1.88, 1.86 (6 s, 18 H, Ac), and 1.06 (s, 9 H, t-Bu).

A solution of 17 (0.783 g, 0.59 mmol) in 50:1 EtOH- $H_2NNH_2 \cdot H_2O$  (40 mL) was stirred under reflux overnight and then concentrated in vacuo. The residue was dissolved in pyridine (15 mL), and to the solution Ac<sub>2</sub>O (5 mL) and a trace of 4-dimethylaminopyridine was added. The solution was stirred overnight at room temperature, then diluted with CHCl<sub>3</sub>, successively washed with aq NaHCO<sub>3</sub>, and aq NaCl, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The resulting residue was purified on silica gel using 1:2 hexane-EtOAc to afford compound **18** (0.592 g, 0.52 mmol, 88%). NMR (400 MHz, CDCl<sub>3</sub>): δ<sub>H</sub> 5.35 (dd, 1 H, H-4c), 5.22 (dd, 1 H, J<sub>4.5</sub> 0.5 Hz, H-4a), 5.13 (d, 1 H, J<sub>2.NH</sub> 10.3 Hz, NH-b), 5.12 (dd, 1 H, J<sub>2.3</sub> 10.3 Hz, H-2c), 5.00–4.93 (m, 2 H, H-3b,3c), 4.66 (dd, 1 H, J<sub>3.4</sub> 2.9 Hz, H-3a), 4.48 (d, 1 H, J<sub>1.2</sub> 7.8 Hz, H-1a), 4.45 (d, 1 H, J<sub>1.2</sub> 7.8 Hz, H-1c), 4.40 (dd, 1 H, H-6b), 4.27 (d, 1 H, J<sub>1,2</sub> 7.8 Hz, H-1b), 4.12 (dd, 1 H, H-6c), 4.09 (dd, 1 H, H-6'c), 4.03 (dd, 1 H, H-6'b), 3.86 (dt, 1 H,  $J_{5.6'}$  6.8 Hz, H-5c), 3.79-3.65 (m, 3 H, H-2a,2b,4b), 3.61 (dd, 1 H, H-6a), 3.47 (dt, 1 H, H-5a), 3.44–3.34 (m, 2 H, H-6'a,5b), 2.16, 2.15, 2.08, 2.07, 2.05, 2.03, 2.01, 1.97, 1.73 (9 s, 27 H, Ac), and 1.09 (s, 9 H, t-Bu); FD-MS: m/z 1144.7 (M+1)<sup>+</sup>. Anal. Calcd for C<sub>52</sub>H<sub>68</sub>N<sub>4</sub>O<sub>23</sub>Si: C, 54.54; H, 5.98; N, 4.89. Found: C, 54.3; H, 5.9; N, 4.8.

O-(2, 3, 4, 6-Tetra-O-acetyl-β-D-galactopyranosyl)-(1→4)-O-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1→6)-3, 4-di-O-acetyl-2-azido-2-deoxy-β-D-galactopyranose (19).—Compound 18 (0.592 g, 0.52 mmol) was desilylated as described for synthesis of compound 9, and the product was purified by chromatography on silica gel using 99:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH to yield 19 (0.378 g, 0.40 mmol, 81%). Compound 19 NMR data (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  6.13 (d, 1 H,  $J_{2,\rm NH}$  9.3 Hz, NH-b), 6.10 (bs, 1 H, OH-1a), 4.85 (d, 1 H,  $J_{1,2}$  6.8 Hz, H-1b), 4.72 (bd, 1 H,  $J_{1,2}$  8.6 Hz, H-1a), and 4.48 (d, 1 H,  $J_{1,2}$  8.3 Hz, H-1c); FD-MS: m/z 906.9 (M+1)<sup>+</sup>. Anal. Calcd. for C<sub>36</sub>H<sub>50</sub>N<sub>4</sub>O<sub>23</sub>·CH<sub>3</sub>OH: C, 47.34; H, 5.79; N, 5.97. Found: C, 47.27; H, 5.55; N, 5.73.

O-(2, 3, 4, 6-Tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 6)-O-(3, 4-di-O-acetyl-2-azido-2-deoxy-α-D-galactopyranosyl)-(1 → 1)-3-O-benzoyl-2-N-[(2R)-2-benzoyloxyalkanoyl]sphinganine (21α) and O-(2,3,4,6-tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2-acetamido-3,6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 6)-O-(3,4-di-O-acetyl-2-azido-2-deoxy-β-D-galactopyranosyl)-(1 → 6)-O-(3,4-di-O-acetyl-2-azido-2-deoxy-β-D-galactopyranosyl)-(1 → 1)-3-O-benzoyl-2-N-[(2R)-2-benzoyloxyalkanoyl]-sphinganine (21β).—Compound 17 (0.100 g, 0.11 mmol) was converted into 20 (0.091 g, 0.087 mmol, 79%) as described for synthesis of 10. NMR (200 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  9.30 (s, 0.5 H, C=NH), 8.78 (s, 0.5 H, C=NH), 6.40 (d, 0.5 H,  $J_{1,2}$  3.4 Hz, H-1aα), 5.65 (d, 0.5 H,  $J_{1,2}$  7.8 Hz, H-1aβ).

To a stirred mixture of **20** (0.145 g, 0.22 mmol), **11** (0.100 g, 0.11 mmol), and powdered molecular sieves (0.200 g) in 1,2-dichloroethane (2.5 mL) was added TMSiOTf (0.03 mL, 0.16 mmol) at  $-20^{\circ}$ C. The mixture was stirred overnight at room temperature, diluted with CHCl<sub>3</sub>, and filtered through Celite. The filtrate was washed with aq NaHCO<sub>3</sub> and aq NaCl, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The resulting residue was purified on silica gel using 1:2 hexane–EtOAc to yield a mixture of anomers **21** $\alpha$  and **21** $\beta$  (0.114 g,

0.065 mmol, 57%). A part of mixture was purified with PLC to separate the anomers. NMR data for **21** $\alpha$  (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  6.88 (d, Cer NH), 6.20 (d, 1 H, NH-b), 4.93 (d, 1 H,  $J_{1,2}$  3.9 Hz, H-1a), 4.54 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1b), 4.49 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 1.30 (m, CH<sub>2</sub>), and 0.85 (t, CH<sub>3</sub>). NMR data for **21** $\beta$  (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  7.08 (d, Cer NH), 6.15 (d, 1 H, NH-b), 4.65 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1b), 4.51 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 4.17 (d, 1 H,  $J_{1,2}$  8.3 Hz, H-1a), 1.30 (m, CH<sub>2</sub>), and 0.85 (t, CH<sub>3</sub>).

 $O-(2, 3, 4, 6-Tetra-O-acetyl-\beta-D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-acetamido-3, 6-di-O$  $acetyl-2-deoxy-\beta-D-glucopyranosyl)-(1 \rightarrow 6)-O-(2-acetamido-3, 4-di-O-acetyl-2-deoxy-\alpha-$ D-galactopyranosyl)- $(1 \rightarrow 1)$ -3-O-benzoyl-2-N-f(2R)-2-benzoyloxyalkanoyl]sphinganine (22) and O-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 4)-O-(2-acetamido-3,6di-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 6)$ -2-acetamido-3,4-di-O-acetyl-2-deoxy- $\beta$ -D-galactopyranosyl)- $(1 \rightarrow 1)$ -3-O-benzoyl-2-N- $\int (2R)$ -2-benzoyloxyalkanoyl]-sphinganine (23).—A mixture of compounds  $21\alpha$  and  $21\beta$  (0.026 g, 0.015 mmol) was dissolved in a solution of NiCl<sub>2</sub>·6H<sub>2</sub>O (0.056 g, 0.24 mmol) and H<sub>3</sub>BO<sub>3</sub> (28 mg, 0.45 mmol) in EtOH (1.4 mL). To this solution was added NaBH<sub>4</sub> (0.014 g, 0.37 mmol) at room temperature. The mixture was stirred for 1 h at room temperature, AcOH and EtOH were added, and the solvents were evaporated in vacuo. The residue was dissolved in pyridine and  $Ac_2O_1$ and the mixture was stirred overnight at room temperature, diluted with MeOH, and concentrated in vacuo. The residue was dissolved in CHCl<sub>3</sub>, the organic layer was washed with aq NaHCO3 and aq NaCl, dried over MgSO4, and concentrated in vacuo. The resulting residue was purified on silica gel using 1:2 hexane-EtOAc, and then by HPLC on a TOSOH Silica-60 column (25 cm×4.6 mm) using EtOAc at a flow rate of 1 mL/min to yield compounds 22 (0.005 g, 0.0028 mmol) and 23 (0.004 g, 0.0022 mmol) (19% and 15%, respectively, from the mixture  $21\alpha + 21\beta$ ). NMR data for 22 (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  6.80 (d, 1 H, J 9.8 Hz, Cer NH), 6.06 (d, 1 H, J<sub>2.NH</sub> 9.8 Hz, NH-a), 5.89 (d, 1 H, J<sub>2.NH</sub> 8.8 Hz, NH-b), 5.49 (t, 1 H, Cer H-2'), 5.35–5.27 (m, 2 H, H-4c, Cer H-3), 5.13 (dd, 1 H, H-3b), 5.06 (dd, 1 H, H-2c), 4.93 (dd, 1 H, J<sub>3,4</sub> 3.4 Hz, 1-5-), 4.85 (dd, 1 H, J<sub>3,4</sub> 3.4 Hz, H-3a), 4.76 (dd, 1 H, H-4a), 4.55 (d, 1 H, J<sub>1,2</sub> 3.4 Hz, H-1a), 4.48 (d, 1 H, J<sub>1,2</sub> 6.8 Hz, H-1b), 4.47 (d, 1 H, J<sub>1.2</sub> 7.3 Hz, H-1c), 4.50-4.40 (m, 3 H, H-2a,6b, Cer H-2), 4.16-4.00 (m, 3 H, H-6'b,6c,6'c), 3.84 (dt, 1 H, H-5c), 3.80–3.67 (m, 4 H, H-5a,2b,4b, Cer H-1A), 3.58 (m, 1 H, H-5b), 3.35 (dd, 1 H, Cer H-1B), 3.33 (dd, 1 H, H-6a), 3.26 (dd, 1 H, H-6'a), 2.18, 2.17, 2.10, 2.09, 2.08, 2.03, 1.98, 1.85 (8 s, 30 H, Ac), 1.25 (m, CH<sub>2</sub>), and 0.88 (t,  $CH_3$ ).

NMR data for **23** (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  7.10 (d, 1 H, J 8.1 Hz, Cer NH), 6.11 (d, 1 H,  $J_{2,\rm NH}$  9.3 Hz, NH-b), 5.67 (d, 1 H,  $J_{2,\rm NH}$  8.3 Hz, NH-a), 5.37 (t, 1 H, Cer H-2'), 5.34 (dd, 1 H, H-4c), 5.25 (dd, 1 H, H-4a), 5.22–5.15 (m, 3 H, H-3a,3b, Cer H-3), 5.11 (dd, 1 H,  $J_{2,3}$  10.3 Hz, H-2c), 4.97 (dd, 1 H,  $J_{3,4}$  3.4 Hz, H-3c), 4.71 (d, 1 H,  $J_{1,2}$  8.3 Hz, H-1a), 4.57 (d, 1 H,  $J_{1,2}$  7.3 Hz, H-1b), 4.49 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 4.47 (dd, 1 H, H-6b), 4.41 (m, 1 H, Cer H-2), 4.15–4.03 (m, 3 H, H-6'b,6c,6'c), 3.92–3.80 (m, 3 H, H-2b,5c, Cer H-1A), 3.79–3.50 (m, 7 H, H-2a,5a,6a,6'a,4b,5b, Cer H-1B), 2.18, 2.13, 2.08, 2.06, 2.05, 1.90, 1.88, 1.86 (8 s, 30 H, Ac), 1.25 (m, CH<sub>2</sub>), and 0.88 (t, CH<sub>3</sub>).

O-β-D-Galactopyranosyl- $(1 \rightarrow 4)$ -O-(2-acetamido-2-deoxy-β-D-glucopyranosyl)- $(1 \rightarrow 6)$ -O-(2-acetamido-2-deoxy-α-D-galactopyranosyl)- $(1 \rightarrow 1)$ -2-N-[(2R)-2-hydroxy-alkanoyl]sphinganine (2).—Compound 22 (0.020 g, 11.2 µmol) was converted into 2 (0.010 g, 8.09 µmol, 72%) as described for the synthesis of 1. NMR (400 MHz, 2:1

CDCl<sub>3</sub>–CD<sub>3</sub>OD):  $\delta_{\rm H}$  4.52 (d, 1 H,  $J_{1,2}$  3.4 Hz, H-1a), 4.19 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1b), 4.11 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 3.96 (dd, 1 H, H-2a), 3.79 (dd, 1 H, Cer H-2'), 3.74 (dd, 1 H, H-6c), 3.69 (m, 1 H, Cer H-2), 3.67–3.52 (m, 6 H, H-4a,5a,6b,6'b,5c, Cer H-1A), 3.51–3.37 (m, 5 H, H-3a,6a,2b,3b,6'c), 3.36–3.27 (m, 5 H, H-6'a,4b,2c, Cer H-1B, 3), 3.26 (dd, 1 H, H-3c), 3.17 (m, 1 H, H-5b), 1.79, 1.75 (2 s, 6 H, Ac), 1.08 (m, CH<sub>2</sub>), and 0.63 (t, CH<sub>3</sub>).

O-β-D-Galactopyranosyl-(1→4)-O-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1→6)-O-(2-acetamido-2-deoxy-β-D-galactopyranosyl)-(1→1)-2-N-[ (2R)-2-hydroxyalkanoyl]sphinganine (24).—Compound 23 (0.020 g, 11.2 µmol) was converted into 24 (0.002 g, 1.62 µmol, 14%) as described for synthesis of 15. NMR (400 MHz, 2:1 CDCl<sub>3</sub>– CD<sub>3</sub>OD): δ<sub>H</sub> 4.20 (d, 1 H, J<sub>1,2</sub> 7.3 Hz, H-1b), 4.09 (d, 1 H, J<sub>1,2</sub> 7.3 Hz, H-1c), 4.07 (d, 1 H, J<sub>1,2</sub> 7.3 Hz, H-1a), 1.73, 1.70 (2 s, 6 H, Ac), 0.99 (m, CH<sub>2</sub>), and 0.57 (t, CH<sub>3</sub>).

 $O-(2,3,4,6-tetra-O-acetyl-\beta-D-galactopyranosyl)-(1 \rightarrow 4)-O$ tert-Butyldiphenylsilyl  $(3,6-di-O-acetyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosyl)-(1 \rightarrow 3)-O-[O-(2,3,4,6-te-0)-glucopyranosyl)-(1 \rightarrow 3)-(1 \rightarrow$  $tra-O-acetyl-\beta-D-galactopyranosyl)-(1 \rightarrow 4)-(3, 6-di-O-acetyl-2-deoxy-2-phthalimido-\beta-D-acetyl-2-deoxy-2-dooxy-2-deoxy-2-deoxy-2$ glucopyranosyl)- $(1 \rightarrow 6)$ ]-2-azido-2-deoxy- $\beta$ -D-galactopyranoside (25).—To a stirred mixture of compound 5 (0.272 g, 0.31 mmol), 7 (0.334 g, 0.29 mmol), and molecular sieves AW-300 (0.80 g) in 1,2-dichloroethane (6 mL) was added M BF<sub>3</sub> · Et<sub>2</sub>O (0.06 mL) at  $-20^{\circ}$ C. The mixture was stirred for 2 h at  $-20^{\circ}$ C. The suspension was filtered, and the solids were washed with CHCl<sub>3.</sub> The filtrate was washed with aq NaHCO<sub>3</sub> and aq NaCl, and the organic layer was dried over  $MgSO_4$  and concentrated in vacuo. The residue was purified by chromatography on silica gel using 3:1 toluene-EtOAc to afford compound 25 (0.123 g, 0.066 mmol, 23%) and mixture of compounds 5 and 25 (0.245 g). NMR data for 25 (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  5.58 (dd, 1 H,  $J_{2,3}$  10.8,  $J_{3,4}$  8.3 Hz, H-3b'), 5.46 (dd, 1 H, J<sub>2,3</sub> 11.2, J<sub>3,4</sub> 8.8 Hz, H-3b), 5.35 (dd, 1 H, H-4c), 5.34 (dd, 1 H, H-4c'), 5.28 (d, 1 H,  $J_{1,2}$  8.3 Hz, H-1b'), 5.16 (dd, 1 H,  $J_{2,3}$  10.3 Hz, H-2c), 5.14 (dd, 1 H,  $J_{2,3}$  10.2 Hz, H-2c'), 4.97 (dd, 1 H, J<sub>3,4</sub> 3.9 Hz, H-3c), 4.96 (dd, 1 H, J<sub>3,4</sub> 3.4 Hz, H-3c'), 4.95 (d, 1 H, J<sub>1,2</sub> 8.3 Hz, H-1b), 4.54 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c'), 4.48 (dd, 1 H, H-6b'), 4.45 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 4.22 (dd, 1 H, H-6b), 4.17–3.98 (m, 8 H, H-2b,6'b,2b',6'b',6c,6'c,6c',6'c'), 4.12 (d, 1 H, J<sub>1.2</sub> 7.8 Hz, H-1a), 3.91–3.80 (m, 4 H, H-6a,4b,5c,5c'), 3.83 (dd, 1 H, J<sub>4.5</sub> 8.8 Hz, H-4b'), 3.78 (m, 1 H, H-5b'), 3.54 (dd, 1 H, J<sub>4.5</sub> 0.5 Hz, H-4a), 3.38 (dd, 1 H, J<sub>5,6'</sub> 4.4, J<sub>6,6'</sub> 10.7 Hz, H-6'a), 3.45 (dd, 1 H, J<sub>2,3</sub> 10.3 Hz, H-2a), 3.04 (dd, 1 H, J<sub>3,4</sub> 3.4 Hz, H-3a), 2.98 (dd, 1 H, J<sub>5.6'</sub> 8.8 Hz, H-5a), 2.75 (m, 1 H, H-5b), and 0.95 (s, 9 H, t-Bu). Anal. Calcd for C<sub>86</sub>H<sub>99</sub>N<sub>5</sub>O<sub>39</sub>Si 0.5toluene: C, 56.55; H, 5.46; N, 3.68. Found: C, 56.30; H, 5.90; N, 3.18.

tert-Butyldiphenylsilyl O-(2, 3, 4, 6-tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 3)-O-[O-(2, 3, 4, 6-tetra-Oacetyl-β-D-galactopyranosyl)-(1 → 4)-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 6)]-4-O-acetyl-2-azido-2-deoxy-β-D-galactopyranoside (26).—Compound 25 (0.132 g, 0.070 mmol) was converted into 26 (0.082 g, 0.048 mmol, 68%) as described for the synthesis of 8. Compound 26 had NMR data (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  5.48 (d, 1 H, J 9.8 Hz, NH-b'), 5.34 (m, 2 H, H-4c,4c'), 5.30 (d, 1 H, J 9.3 Hz, NH-b), 5.18 (d, 1 H, J<sub>3,4</sub> 3.4 Hz, H-4a), 5.12 (dd, 1 H, J<sub>2,3</sub> 10.3 Hz, H-2c), 5.10 (dd, 1 H, J<sub>2,3</sub> 10.8 Hz, H-2c'), 5.04 (dd, 1 H, J<sub>3,4</sub> 8.8, J<sub>4,5</sub> 10.2 Hz, H-3b), 5.01 (dd, 1 H, J<sub>2,3</sub> 10.2, J<sub>3,4</sub> 9.3 Hz, H-3b'), 4.98–4.93 (m, 2 H, H-3c,3c'), 4.67 (dd, 1 H, H-6b'), 4.54 (d, 1 H, J<sub>1,2</sub> 8.3 Hz, H- 1b'), 4.53 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c'), 4.46 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 4.41 (dd, 1 H, H-6b), 4.34 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1a), 4.30 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1b), 4.14–4.05 (m, 4 H, H-6c,6'c,6c',6'c'), 4.03 (dd, 1 H, H-6'b), 3.98–3.92 (m, 2 H, H-2b',6'b'), 3.90–3.85 (m, 2 H, H-5c,5c'), 3.82 (dd, 1 H,  $J_{4,5}$  9.3 Hz, H-4b'), 3.70–3.59 (m, 3 H, H-5a,2b,4b), 3.62 (d, 1 H,  $J_{2,3}$  10.2 Hz, H-2a), 3.45–3.33 (m, 5 H, H-3a,6a,6'a,5b,5b'), and 1.10 (s, 9 H, *t*-Bu). Anal. Calcd for C<sub>76</sub>H<sub>101</sub>N<sub>5</sub>O<sub>38</sub>Si: C, 53.05; H, 5.91; N, 4.07. Found: C, 52.83; H, 5.97; N, 4.06.

O-(2, 3, 4, 6-Tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 3)-O-[O-(2, 3, 4, 6-tetra-O-acetyl-β-D-galacto-pyranosyl)-(1 → 4)-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 6)]-4-O-acetyl-2-azido-2-deoxy-α, β-D-galactopyranose (27).—Compound 26 (0.120 g, 0.070 mmol) was desilylated to 27 as described for the conversion of 8 into 9. The yield of 27 was 0.085 g (0.052 mmol, 85% based on 26 consumed). NMR (400 MHz, CDCl<sub>3</sub>): δ<sub>H</sub> 4.85 (d, 0.5 H,  $J_{1,2}$  3.5 Hz, H-1aα), 4.53 (d, 1 H,  $J_{1,2}$  7.9 Hz, H-1b'), 4.53 (d, 1 H,  $J_{1,2}$  8.1 Hz, H-1b), 4.48 (d, 1 H,  $J_{1,2}$  8.1 Hz, H-1c'), 4.44 (d, 1 H,  $J_{1,2}$  8.1 Hz, H-1c), and 4.29 (d, 0.5 H,  $J_{1,2}$  8.2 Hz, H-1aβ). Anal. Calcd for C<sub>60</sub>H<sub>83</sub>N<sub>5</sub>O<sub>38</sub>·3C<sub>2</sub>H<sub>5</sub>OH: C, 48.92; H, 6.28; N, 4.32. Found: C, 49.09; H, 6.68; N, 4.30.

O-(2, 3, 4, 6-Tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 3)-O-[O-(2, 3, 4, 6-tetra-O-acetyl-β-D-galacto-pyranosyl)-(1 → 4)-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 6)]-O-(4-O-acetyl-2-azido-2-deoxy-α-D-galactopyranosyl)-(1 → 1)-3-O-benzoyl-2-N-[(2R)-2-benzoyloxyalkanoyl]sphinganine (**29**α) and O-(2,3,4,6-tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2-acetamido-3,6-di-O-acetyl-2-deoxy-β-D-gluco-pyranosyl)-(1 → 3)-O-[O-(2,3,4,6-tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2-acetamido-3,6-di-O-acetyl-2-deoxy-β-D-gluco-pyranosyl)-(1 → 3)-O-[O-(2,3,4,6-tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-(2-acetamido-3,6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 6)]-O-(4-O-acetyl-2-azido-2-deoxy-β-D-galactopyranosyl)-(1 → 1)-3-O-benzoyl-2-N-[(2R)-2-benzoyloxyalkanoyl]-sphinganine (**29**β).—The above preparation of **27** was converted into the trichloroacetimidate **28** as described for the transformation of **9** into **10**. NMR data for **28** (200 MHz, CDCl<sub>3</sub>): δ<sub>H</sub> 9.25 (s, 0.5 H, C=NHβ), 8.79 (s, 0.5 H, C=NHα), 5.30 (d, 0.5 H, J<sub>1,2</sub> 3.5 Hz, H-1aα), 5.23 (d, 0.5 H, J<sub>1,2</sub> 8.2 Hz, H-1aβ), 4.55 (d, 1 H, J<sub>1,2</sub> 7.9 Hz, H-1b'), 4.49 (d, 1 H, J<sub>1,2</sub> 8.1 Hz, H-1b), 4.63 (d, 1 H, J<sub>1,2</sub> 8.1 Hz, H-1c'), and 4.60 (d, 1 H, J<sub>1,2</sub> 8.6 Hz, H-1c).

To a stirred mixture of **28** (0.082 g, 0.050 mmol), **11** (0.050 g, 0.057 mmol), and powdered molecular sieves (0.100 g) in 1,2-dichloroethane (2.0 mL) was added 0.1 M TMSiOTf in 1,2-dichloroethane (0.5 mL, 0.05 mmol) at  $-5^{\circ}$ C. The mixture was stirred for 1.5 h, diluted with CHCl<sub>3</sub>, and filtered through Celite. The filtrate was washed with aq NaHCO<sub>3</sub> and aq NaCl, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The resulting residue was purified on silica gel using 4:1 CH<sub>2</sub>Cl<sub>2</sub>-acetone to afford a mixture of compounds **29** $\alpha$ and **29** $\beta$  (0.029 g, 0.012 mmol, 25%). NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  4.83 (d, 0.67 H,  $J_{1,2}$ 3.4 Hz, H-1a $\alpha$ ), 4.63 (d, 1 H,  $J_{1,2}$  7.3 Hz, H-1b), 4.58 (d, 0.33 H,  $J_{1,2}$  7.8 Hz, H-1a $\beta$ ), 4.52 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c'), 4.49 (d, 1 H,  $J_{1,2}$  7.8 Hz, H-1c), 4.23 (d, 1 H,  $J_{1,2}$  8.3 Hz, H-1b'), 1.22 (m, CH<sub>2</sub>), and 0.86 (t, CH<sub>3</sub>).

O-(2, 3, 4, 6-Tetra-O-acetyl-β-D-galactopyranosyl)-(1 → 4)-O-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 3)-O-[O-(2, 3, 4, 6-tetra-O-acetyl-β-D-galacto-pyranosyl)-(1 → 4)-(2-acetamido-3, 6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 6)]-O-(2-acetamido-4-O-acetyl-2-deoxy-α-D-galactopyranosyl)-(1 → 1)-3-O-benzoyl-2-N-

 $[(2\mathbf{R})-2-benzoyloxyalkanoyl]$ sphinganine (30) and O- $(2,3,4,6-tetra-O-acetyl-\beta-D-ga$ lactopyranosyl)- $(1 \rightarrow 4)$ -O-(2-acetamido-3,6-di-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-[O-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)-(1  $\rightarrow$  4)-(2-acetamido-3,6di-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 6)$ ]-O-(2-acetamido-4-O-acetyl-2-deoxy- $\beta$ -D-galactopyranosyl)- $(1 \rightarrow 1)$ -3-O-benzoyl-2-N-f(2R)-2-benzoyloxy-alkanoyl]sphinganine (31).—A mixture of compounds  $29\alpha$  and  $29\beta$  (38 mg, 0.016 mmol) was converted into **30** (19 mg, 0.008 mmol) and **31** (9 mg, 0.004 mmol) (50% and 24%, respectively, from the mixed precursor) as described for the synthesis of 13 and 14. Compound 30 had NMR data (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H}$  5.39–5.31 (m, 3 H, H-4c,4c', Cer H-2'), 5.31–5.24 (m, 1 H, Cer H-3), 5.24–5.03 (m, 5 H, H-4a,3b,3b',2c,2c'), 5.03–4.90 (m, 2 H, H-3c,3c'), 4.66 (d, 1 H, J<sub>1,2</sub> 3.0 Hz, H-1a), 4.65–4.42 (m, 3 H, H-6b,6b', Cer H-2), 4.56 (d, 1 H, J<sub>1,2</sub> 8.0 Hz, H-1b'), 4.52 (d, 1 H, J<sub>1,2</sub> 8.0 Hz, H-1b), 4.49 (d, 1 H, J<sub>1,2</sub> 8.0 Hz, H-1c), 4.49 (d, 1 H, J<sub>1.2</sub> 8.0 Hz, H-1c'), 4.35 (dd, 1 H, H-2a), 4.19–3.96 (m, 6 H, H-6'b,6'b',6c,6'c,6c',6'c'), 3.94-3.60 (m, 10 H, H-3a,6a,6'a,2b,4b,2b',4b',5c,5c', Cer H-1A), 3.59-3.46 (m, 2 H, H-5b, 5b'), 3.36-3.26 (m, 2 H, H-5a, Cer H-1B), 1.30 (m, CH<sub>2</sub>), and 0.90 (t,  $CH_3$ ).

NMR data for **31**:  $\delta_{\rm H}$  6.84 (d, 1 H, J 8.0 Hz, Cer NH), 6.36 (d, 1 H, NH-b'), 6.34 (d, 1 H, J 8.4 Hz, NH-a), 5.79 (d, 1 H, NH-b), 5.28 (dd, 2 H,  $J_{4,5}$  0.4 Hz, H-4c,4c'), 5.26–5.16 (m, 2 H, H-3b', Cer H-2'), 5.22 (dd, H-4a), 5.14 (m, 1 H, Cer H-3), 5.06 (dd, 1 H,  $J_{2,3}$  10.4 Hz, H-2c'), 5.04 (dd, 1 H,  $J_{2,3}$  10.4 Hz, H-2c), 4.91 (m, 1 H, H-3b), 4.90 (dd, 1 H,  $J_{3,4}$  3.4 Hz, H-3c), 4.89 (dd, 1 H,  $J_{3,4}$  3.4 Hz, H-3c'), 4.73 (d, 1 H,  $J_{1,2}$  8.0 Hz, H-1a), 4.55 (d, 1 H,  $J_{1,2}$  7.6 Hz, H-1b'), 4.53 (dd, 1 H, H-6b), 4.45 (d, 1 H,  $J_{1,2}$  8.0 Hz, H-1b'), 4.43 (d, 1 H,  $J_{1,2}$  8.0 Hz, H-1c'), 4.43 (d, 1 H,  $J_{1,2}$  8.0 Hz, H-35 (m, 2 H, H-3a,6b'), 4.28 (m, 1 H, Cer H-2), 4.10–3.98 (m, 5 H, H-6'b',6c,6'c,6c',6'c'), 3.95 (dd, 1 H, H-6'b), 3.91–3.50 (m, 11 H, H-6a,6'a,2b,4b,2b',4b',5b',5c,5c', Cer H-1A,1B), 3.48–3.34 (m, 2 H, H-5a,5b), 3.05 (m, 1 H, H-2a), 1.19 (m, CH<sub>2</sub>), and 0.80 (t, CH<sub>3</sub>).

O-β-D-Galactopyranosyl-(1→4)-O-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1→3)-O-[O-β-D-galactopyranosyl-(1→4)-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1→6)]-O-(2-acetamido-2-deoxy-α-D-galactopyranosyl)-(1→1)-2-N-[(2R)-2-hydroxyalkanoyl]sphinganine (3).—Compound 30 (0.008 g, 3.5 µmol) was converted into 3 (0.004 g, 2.7 µmol, 75%) as described for synthesis of 1. Compound 3 had NMR (400 MHz, Me<sub>2</sub>SO-d<sub>6</sub>): δ<sub>H</sub> 4.63 (d, 1 H, J<sub>1,2</sub> 3.0 Hz, H-1a), 4.57 (d, 1 H, J<sub>1,2</sub> 8.5 Hz, H-1b'), 4.31 (d, 1 H, J<sub>1,2</sub> 7.0 Hz, H-1b), 4.20 (d, 2 H, J<sub>1,2</sub> 7.5 Hz, H-1c,1c'), 4.08 (dd, 1 H, J<sub>2,3</sub> 11.0 Hz, H-2a), 1.85, 1.81, 1.78 (3 s, 9 H, Ac), 1.22 (m, CH<sub>2</sub>), and 0.85 (t, CH<sub>3</sub>).

O-β-D-Galactopyranosyl-(1→4)-O-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1→3)-O-[O-β-D-galactopyranosyl-(1→4)-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1→6)]-O-(2-acetamido-2-deoxy-β-D-galactopyranosyl)-(1→1)-2-N-[(2R)-2-hydroxyalkanoyl]sphinganine (**32**).—Compound **31** (0.008 g, 3.5 µmol) was converted into **32** (0.004 g, 2.7 µmol, 75%) as described for synthesis of **15**. Compound **32** had NMR (400 MHz, Me<sub>2</sub>SO-d<sub>6</sub>):  $\delta_{\rm H}$  4.57 (d, 1 H,  $J_{1,2}$  8.3 Hz, H-1b'), 4.40 (d, 1 H,  $J_{1,2}$  8.0 Hz, H-1b), 4.27 (d, 1 H,  $J_{1,2}$  8.0 Hz, H-1a), 4.23 (d, 2 H,  $J_{1,2}$  7.0 Hz, H-1c,1c'), 1.83, 1.81, 1.80 (3 s, 9 H, Ac), 1.23 (m, CH<sub>2</sub>), and 0.84 (t, CH<sub>3</sub>).

#### References

- [1] G.F. Springer, P.R. Desai, and I. Banatwala, Naturwissenschaften, 61 (1974) 457-458.
- [2] A.F.R. Rahman and B.M. Longenecker, J. Immunol., 129 (1982) 2021–2024; B.M. Longenecker, A.F.R. Rahman, J.B. Leigh, R.A. Purser, A.H. Greenberg, D.J. Willans, O. Keller, P.K. Petrik, T.Y. Thay, M.R. Suresh, and A.A. Noujaim, Int. J. Cancer, 33 (1984) 123–129.
- [3] W. Dahr, G. Uhlenbruck, and G.W.G. Bird, Vox Sang., 27 (1974) 29-42.
- [4] S. Hirohashi, H. Clausen, T. Yamada, Y. Shimosato, and S. Hakomori, Proc. Natl. Acad. Sci. U.S.A., 82 (1985) 7039–7043; D.J. Roxby, A.A. Morley, and M. Burpee, Br. J. Haematol., 67 (1987) 153–156.
- [5] M. Nuti, Y.A. Teramoto, R. Mariani-Costantini, P.H. Hand, D. Colcher, and J. Schlom, Int. J. Cancer, 29 (1982) 539–545; A. Kurosaka, S. Fukui, H. Kitagawa, H. Nakada, Y. Numata, I. Funakoshi, T. Kawasaki, and I. Yamashina, FEBS Lett., 215 (1987) 137–139; T. Kjeldsen, H. Clausen, S. Hirohashi, T. Ogawa, H. Iijima, and S. Hakomori, Cancer Res., 48 (1988) 2214–2220.
- [6] P.W. Tang, H.C. Gool, M. Hardy, Y.C. Lee, and T. Feizi, Biochem. Biophys. Res. Comm., 132 (1985) 474– 480.
- [7] R. Horie, K. Hara, and K. Nakano, Carbohydr. Res., 230 (1992) C11-C15.
- [8] G. Grundler and R.R. Schmidt, Carbohydr. Res., 135 (1985) 203-218.
- [9] Y. Nakahara, H. Iijima, S. Sibayama, and T. Ogawa, Tetrahedron Lett., 31 (1990) 6897-6900.
- [10] R.M. Ratcliffe, D.A. Baker, and R.U. Lemieux, Carbohydr. Res., 93 (1981) 35-41.
- [11] R.U. Lemieux, T. Takeda, and B.Y. Chung, ACS Symp. Ser., 39 (1976) 90-115.
- [12] W. Kinzy and R.R. Schmidt, Liebigs Ann. Chem., (1985) 1537-1545.
- [13] H. Paulsen, and J.-P. Hölck, Carbohydr. Res., 109 (1982) 89–107; H. Paulsen and V. Sinnwell, Chem. Ber., 111 (1978) 879–889.