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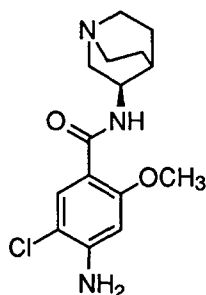
**(R)-3-(6-CHLORO-1-ISOPROPYLBENZIMIDAZOLE-4-CARBOXAMIDO)QUINUCLIDINE: A HIGH AFFINITY LIGAND FOR THE (R)-ZACOPRIDE BINDING SITE**

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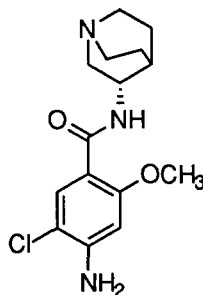
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**Abstract:** The (R)-3-amido quinuclidine **6** (RS-16566) was found to be a high affinity ligand for the (R)-zacopride binding site.

Zacopride is a well-known racemic serotonergic agent that exhibits properties of both 5HT<sub>3</sub> antagonism<sup>3</sup> and 5HT<sub>4</sub> agonism.<sup>4</sup> The order of binding affinity of the zacopride antipodes is (S)-zacopride > (R)-zacopride at both of these serotonin receptor subtypes; however, recent reports have described a non-serotonergic binding site in rat cerebral cortex and NG 108-15 clonal cells that exhibits (R)-zacopride > (S)-zacopride selectivity.<sup>5,6</sup> The extant pharmacology and distribution studies of the so-called (R)-zacopride binding site suggest a novel, high affinity locus with a wide distribution in central and peripheral tissues.<sup>5b</sup> This binding site has so far been primarily characterized by radioligand binding studies with [<sup>3</sup>H]-(R)-zacopride; however, emerging in vivo data suggest that it may exhibit a functional role in the behavioral actions of (R)-zacopride that is not simply accounted for by serotonergic properties.<sup>7</sup>

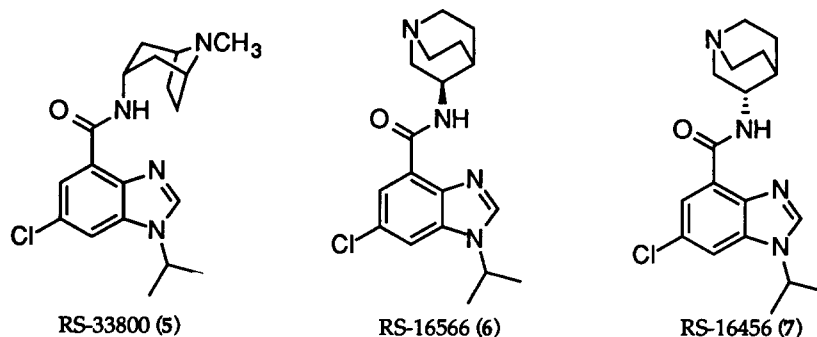


(R)-zacopride



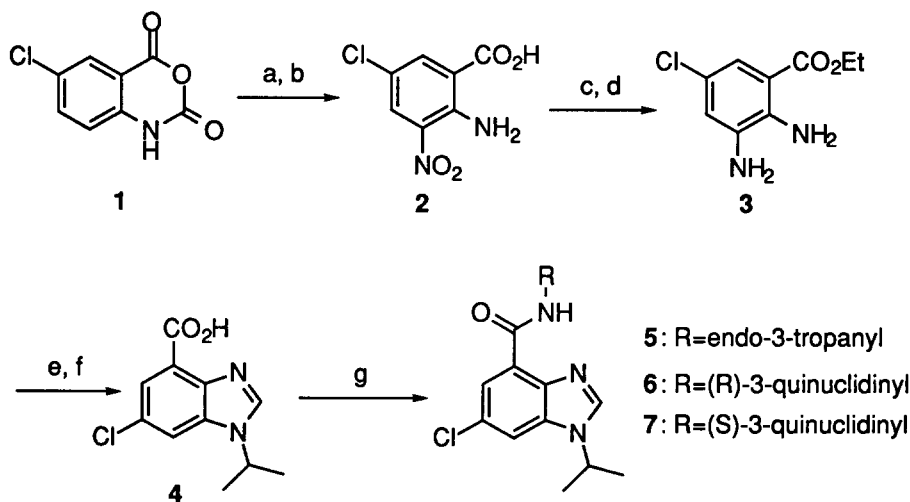
(S)-zacopride

During the course of a program aimed at discovery of 5HT<sub>3</sub> and 5HT<sub>4</sub> receptor ligands we noted that the achiral lead RS-33800, endo-3-(6-chloro-1-isopropylbenzimidazole-4-carboxamido)tropane (**5**), displayed a relatively high affinity (16 nM) for the (R)-zacopride binding site ((R)-ZBS) in NG 108-15 clonal cells. The binding affinity of **5** is roughly comparable to that of (R)-zacopride itself using the NG 108-15 cell line; therefore, we briefly explored the effect of substituting (R)-3-quinuclidinyl and (S)-3-quinuclidinyl moieties for the tropane ring system to give the enantiomeric amides RS-16566 (**6**) and RS-16456 (**7**), respectively.



The 6-chloro-1-isopropylbenzimidazole-4-carboxamides **5-7** were prepared from commercially available 5-chloroisatoic anhydride (**1**) as shown in Scheme I. Nitration of **1**, followed by hydrolysis of the anhydride group, gave 5-chloro-3-nitroanthranilic acid **2** in 80 % yield. Esterification of compound **2** with satd. ethanolic HCl followed by reduction of the nitro group with  $H_2$ -10 % Pd/C afforded ethyl 5-chloro-3-aminoanthranilate **3** in quantitative yield. The 3-amino group was selectively monoalkylated by warming a solution of **3** in 1:1 2-iodopropane-DMF to 50 °C for 6 h; chromatographic purification of the N-monoalkylated product, followed by treatment with formic acid in hot aq. HCl, gave 6-chloro-1-isopropylbenzimidazole-4-carboxylic acid (**4**) in 35 % yield. Carboxylic acid **4** was coupled with endo-3-aminotropane, (R)-3-aminoquinuclidine, or (S)-3-aminoquinuclidine using carbonyl diimidazole in DMF solution to afford amides **5-7** respectively. Compounds **5-7** were converted to their hydrochloride salt forms for all subsequent uses.

Scheme I



**a:**  $NaNO_3/H_2SO_4$  **b:**  $H_2O$ , reflux **c:** EtOH-HCl **d:**  $H_2$ -10 % Pd/C **e:** 1:1 DMF-2-iodopropane (50 °C, 6 h) **f:**  $HCO_2H$ -aq. HCl, reflux **g:** CDI-DMF; then  $RNH_2$

Selected receptor binding affinities for compounds 5-7 and the zacopride enantiomers are given in the Table.

<b>TABLE. Receptor Binding Affinities of 5-7 and (R)- and (S)-zacopride</b>			
Compound	Binding $pK_i^a$		
	5HT <sub>3</sub> <sup>b</sup>	5HT <sub>4</sub> <sup>c</sup>	(R)-ZBS <sup>d</sup>
<b>5</b>	9.0±0.1	6.80±0.07	7.70±0.10
<b>6</b>	9.27±0.03	7.62±0.01	9.84±0.42
<b>7</b>	9.86±0.08	6.8 <sup>e</sup>	6.22±0.30
(S)-zacopride	9.74±0.03	6.36±0.12	5.3 <sup>f</sup>
(R)-zacopride	8.43 <sup>g</sup>	5.55±0.13	8.30±0.22

<sup>a</sup>Except as noted, radioligand binding assays were carried out as described in reference 8. Values are mean  $pK_i \pm \text{SEM}$  ( $n \geq 3$ ).  
<sup>b</sup>Displacement of [<sup>3</sup>H]-BRL 43694 from NG-108-15 cell membranes.  
<sup>c</sup>Displacement of [<sup>3</sup>H]-GR 113808 from Guinea pig brain striatum.  
<sup>d</sup>Displacement of [<sup>3</sup>H]-(R)-zacopride from ondansetron-treated NG-108-15 cell membranes. <sup>e</sup> $pK_b$  estimate from relaxation of carbachol-contracted rat esophageal muscularis mucosae. <sup>f</sup>Reference 5.  
<sup>g</sup>Displacement of racemic [<sup>3</sup>H]-zacopride from rat cortex (reference 3b).

All of the compounds in the Table exhibit high affinity for the 5HT<sub>3</sub> receptor and low-to-moderate affinity for the 5HT<sub>4</sub> receptor; however, the range of affinities of these compounds at the (R)-zacopride binding site ((R)-ZBS) is remarkable. Thus, the (R)-antipode of zacopride shows a 1000-fold higher (R)-ZBS affinity than does (S)-zacopride.<sup>5</sup> (R)-Amido quinuclidine 6, which shows 4000-fold higher affinity at the (R)-zacopride binding site than the corresponding (S)-antipode 7, also exhibits a 35-fold improvement in (R)-ZBS binding affinity over (R)-zacopride itself. Unfortunately, while both (S)-zacopride and compound 7 show excellent 5HT<sub>3</sub>/(R)-ZBS selectivity, no ligands are known to date which exhibit useful (R)-ZBS/5HT<sub>3</sub> selectivity. The achievement of *ca.* 100-fold (R)-ZBS/5HT<sub>3</sub> selectivity in a high affinity ligand would clearly provide a powerful pharmacological tool for the further assessment of physiological functions of the (R)-zacopride binding site; thus, additional structure-activity studies are needed toward this end.

## References and Notes

\*This paper is dedicated to Professor Clayton H. Heathcock on the occasion of his 60th birthday.

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2. Division of Neurobiology
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8. **5HT<sub>2</sub> receptor binding assay:** NG-108 cells were grown in Dulbecco's modified Eagle's medium supplemented with 10 % bovine calf serum and 1 X hypoxanthine-aminopterin-thymidine. Confluent cells were harvested from 250 mL flasks using a 1 min exposure to trypsin. The cell suspension was centrifuged (200 x G; 5 min), resuspended in a Tris (50 mM) EDTA (5 mM) buffer (pH 7.4 @ 4 °C), homogenized using a Polytron P10 tissue disrupter (setting 5 for 10 sec), and recentrifuged for 15 min. For competition binding assays the NG-108 cell membranes were incubated with 1.0 nM [<sup>3</sup>H]-BRL 43694 and competing ligands in 0.25 mL of Tris-Krebs buffer (154 mM NaCl, 5.4 mM KCl, 1.2 mM K<sub>2</sub>PO<sub>4</sub>, 2.5 mM CaCl<sub>2</sub>, 1 mM MgCl<sub>2</sub>, 11 mM D-glucose and 10 mM Tris; pH 7.4 @ 25 °C) for 45 min. Reactions were terminated by vacuum filtration through filters pretreated with 0.3 % PEI. Nonspecific binding was determined with 1 μM BRL 43694. **(R)-zacopride binding site assay:** NG-108 cell membranes, prepared as above, were incubated with 2 nM [<sup>3</sup>H]-(R)-zacopride and competing ligands in 0.5 mL of Tris buffer (25 mM Tris-HCl; pH 7.4 @ 30 °C). 1 μM Ondansetron was included in the incubation to prevent [<sup>3</sup>H]-(R)-zacopride binding to 5HT<sub>2</sub> receptors. Reactions were terminated after 30 min by vacuum filtration. Nonspecific binding was defined with 100 μM mianserin. **5HT<sub>4</sub> receptor binding assay:** Guinea pig brain striata were dissected from Guinea pig brains (Rockland Inc., Gilbertsville, PA). Membranes were prepared by homogenization in a Tris buffer (10 mM Tris, 250 mM sucrose, 5 mM EDTA; pH 7.4 @ 4 °C), filtration through nylon mesh, and centrifugation of the filtrate (1000 x G for 15 min). Membranes were incubated with 0.1 nM [<sup>3</sup>H]-GR 113808 and competing ligands in 0.5 mL of a HEPES buffer (50 mM HEPES, 130 mM choline chloride, 5 mM D-glucose, 5.4 mM KCl, 0.5 mM EDTA; pH 7.4 @ 25 °C). Reactions were terminated after 1 h by vacuum filtration. Nonspecific binding was defined with 1 μM GR 113808.

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