



Pergamon

SCIENCE @ DIRECT®

Bioorganic & Medicinal Chemistry Letters 13 (2003) 3083–3086

BIOORGANIC &
MEDICINAL
CHEMISTRY
LETTERS

2,6-Disubstituted Pyran-4-one and Thiopyran-4-one Inhibitors of DNA-Dependent Protein Kinase (DNA-PK)

Jonathan J. Hollick,^a Bernard T. Golding,^a Ian R. Hardcastle,^a Niall Martin,^b
Caroline Richardson,^b Laurent J. M. Rigoreau,^{a,b} Graeme C. M. Smith^b
and Roger J. Griffin^{a,*}

^aNorthern Institute for Cancer Research, School of Natural Sciences-Chemistry, Bedson Building, University of Newcastle, Newcastle Upon Tyne NE1 7RU, UK

^bKuDOS Pharmaceuticals Limited, 327 Cambridge Science Park, Milton Road, Cambridge CB4 0WG, UK

Received 9 April 2003; accepted 16 May 2003

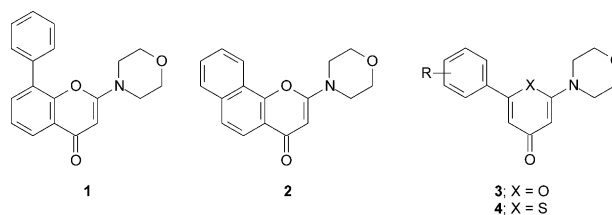
Abstract—6-Aryl-2-morpholin-4-yl-4*H*-pyran-4-ones and 6-aryl-2-morpholin-4-yl-4*H*-thiopyran-4-ones were synthesised and evaluated as potential inhibitors of the DNA repair enzyme DNA-dependent protein kinase (DNA-PK). Several compounds in each series exhibited superior activity to the chromenone LY294002, and were of comparable potency to the benzochromenone NU7026 (IC₅₀ = 0.23 μM). Importantly, members of both structural classes were found to be selective inhibitors of DNA-PK over related phosphatidylinositol 3-kinase-related kinase (PIKK) family members. A multiple-parallel synthesis approach, employing Suzuki cross-coupling methodology, was utilised to prepare libraries of thiopyran-4-ones with a range of aromatic groups at the 3'- and 4'-positions on the thiopyran-4-one 6-aryl ring. Screening of the libraries resulted in the identification of 6-aryl-2-morpholin-4-yl-4*H*-thiopyran-4-ones bearing naphthyl or benzo[*b*]thienyl substituents at the 4'-position, as potent DNA-PK inhibitors with IC₅₀ values in the 0.2–0.4 μM range.

© 2003 Elsevier Ltd. All rights reserved.

Introduction

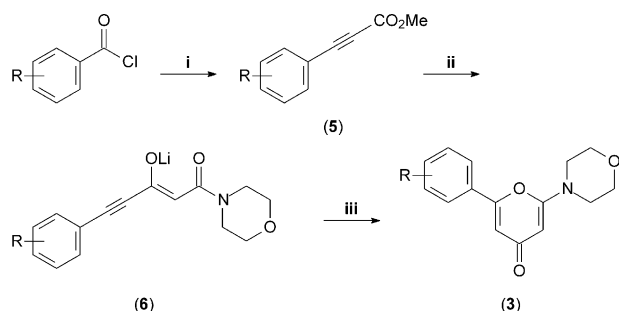
The nuclear serine/threonine protein kinase DNA-dependent protein kinase (DNA-PK), a member of the phosphatidylinositol 3-kinase-related kinase (PIKK) family, comprises a large catalytic subunit (DNA-PKcs), and a heterodimeric subunit (Ku).^{1–3} DNA-PK is activated and assembles at the site of DNA double strand breaks (DSBs), and is a crucial component of the cellular DNA DSB repair machinery.⁴ Phosphorylation of a number of target proteins has been observed in vitro, including those involved in the cell cycle and apoptosis.⁵ DNA-PK plays an essential role in the physiological process of V(D)J recombination,² and is also involved in the repair of DNA DSBs induced by ionising radiation and certain cancer chemotherapeutic agents. Importantly, cells without competent DNA-PK are hypersensitive to ionising radiation and radio-mimetic agents, and inhibition of DNA-PK activity has

been shown to potentiate the in vitro cytotoxicity of ionising radiation^{5,6} and a number of anticancer drugs.^{7–9} DNA-PK inhibitors may thus have clinical utility as radio- and chemo-potentiators in the treatment of cancer.



Although reportedly a selective ATP-competitive PI 3-kinase (PI 3-K) inhibitor (IC₅₀ = 1.5–2.0 μM),^{10,11} the chromenone LY294002 (**1**) has also been shown to exhibit comparable inhibitory activity against DNA-PK (IC₅₀ = 1.4 μM, K_i = 6.0 μM).¹² As part of a research programme to develop potent and selective DNA-PK inhibitors suitable for eventual clinical evaluation, we have utilised **1** as a structural lead. In the absence of suitable crystal structure information for DNA-PK, a

*Corresponding author. Tel.: +44-191-222-8591; fax: +44-191-222-8591; e-mail: r.j.griffin@ncl.ac.uk



Scheme 1. General synthesis of pyran-4-ones. Reagents and conditions: (i) (a) $\text{Ph}_3\text{P}=\text{CHCO}_2\text{Me}$, PhMe , reflux; (b) 250°C ; (ii) $n\text{-BuLi}$, $(i\text{-Pr})_2\text{NH}$, N -acetylmorpholine, $-78^\circ\text{C}\rightarrow 0^\circ\text{C}$; (iii) MeSO_3H , 25°C .

pharmacophore mapping approach has been exploited for the delineation of structure–activity relationships (SARs) around the core structure of **1**. These studies have resulted in the identification of a number of structurally diverse inhibitors, which are more potent and selective than **1**, as exemplified by NU7026 (**2**) ($\text{IC}_{50}=0.23\ \mu\text{M}$).¹³ Further refinement of the pharmacophore model indicated that, whereas the 2-morpholin-4-yl substituent appears to be a prerequisite for inhibitory activity, ‘minimalisation’ of the chromenone core structure might be tolerated. In this paper we describe the synthesis and preliminary biological evaluation of DNA-PK inhibitors derived from the monocyclic pyran-4-one (**3**) and thiopyran-4-one (**4**) templates.

Chemical Synthesis

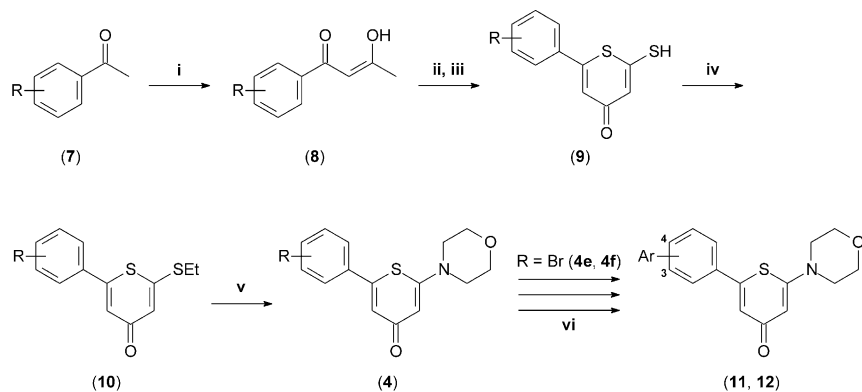
The required 2,6-disubstituted pyran-4-ones (**3**) were prepared by a literature procedure as summarised in Scheme 1.¹⁴ Briefly, treatment of the appropriate benzoyl chloride with methyl triphenylphosphoranylidene, gave the phenylpropiolate ester (**5**), which furnished the phenylacetylenic β -ketoamide (**6**) on reaction with the *O*-lithio derivative of acetylmorpholine. Ring closure of **6** to the required pyran-4-one (**3**) was achieved with methanesulfonic acid. To the best of our knowledge, the corresponding 6-aryl-2-morpholin-4-ylthiopyran-4-ones (**4**) have not been reported previously, and the synthesis of this heterocycle is summarised in Scheme 2. The

substituted aroylacetone (**8**) was readily prepared by acylation of the appropriate acetophenone (**7**) with ethyl acetate–sodium ethoxide. Treatment of the dianion of **8** with carbon disulfide afforded, upon carefully controlled workup, the 2-mercapto-6-arylthiopyran-4-one (**9**), and alkylation of the thiol group of **9** with iodoethane gave **10** quantitatively. Final displacement of the 2-thioethyl substituent was effected by heating **10** with morpholine in ethane-1,2-diol to yield the target thiopyran-4-ones **4a–f**.¹⁵

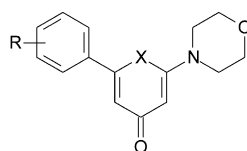
Although unambiguous SARs are not apparent from the initial compounds synthesised, the nature and position of the 6-aryl substituent clearly influences DNA-PK inhibitory activity in both series (Table 1). In addition, preliminary in vitro studies with the pyran-4-ones (**3**) and thiopyran-4-ones (**4**) suggest that the latter series might have superior cellular activity, despite the slightly higher potency of the pyran-4-ones against DNA-PK. In order to investigate this further, a multiple-parallel synthesis (MPS) approach was utilised to generate two libraries with a range of aromatic groups at the 3'- and 4'-positions on the thiopyran-4-one 6-aryl ring. The required libraries were prepared by a Suzuki cross-coupling reaction between the 3'- and 4'-bromophenyl substituted thiopyran-4-ones (**4e** and **4f**), and a range of commercially available boronic acids and esters (Scheme 2).¹⁶ A total of 97 thiopyran-4-one derivatives (42 from **4e** and 55 from **4f**) were deemed of suitable purity ($\geq 85\%$) for evaluation in a pre-screen DNA-PK assay, from which six compounds emerged as sufficiently potent for IC_{50} values to be determined (Table 2).

Results and Discussion

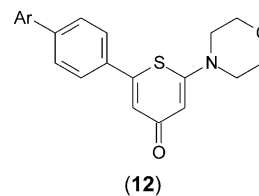
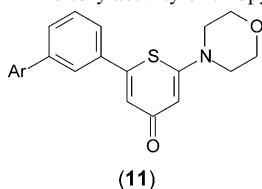
Our previous studies have demonstrated that selective inhibition of DNA-PK, over other PIKK family members, is achievable through modification of the chromenone pharmacophore of **1**. For example, NU7026 (**2**) is approximately 6-fold more potent than **1** as a DNA-PK inhibitor, and at least 70-fold more selective for DNA-PK over PI 3-K ($\text{pI}10\alpha$), whereas **1** is essentially equipotent against both kinases.^{13,17} However, the benzo[*h*]chromenone template of **2** is not readily amenable



Scheme 2. General synthesis of thiopyranones. Reagents and conditions: (i) NaOEt , EtOAc , THF , 25°C ; (ii) LDA , THF , CS_2 , $-78\rightarrow 25^\circ\text{C}$; (iii) H_2O , $0^\circ\text{C}\rightarrow 25^\circ\text{C}$, HCl (aq); (iv) K_2CO_3 , EtI , $(\text{Me})_2\text{CO}$, reflux; (v) morpholine, $(\text{CH}_2\text{OH})_2$, $120\rightarrow 150^\circ\text{C}$; (vi) $\text{Pd}(\text{PPh}_3)_4$, K_2CO_3 , $\text{ArB}(\text{OH})_2$, dioxane, 90°C .

Table 1. Inhibition of DNA-PK by selected pyranone-4-ones and thiopyran-4-ones^a

Compd	X	R	IC ₅₀ (μM) ^b	Compd	X	R	IC ₅₀ (μM) ^b
1	—	—	1.4	3g	O	4-MeO	0.22
2	—	—	0.23	3h	O	4- ^t Bu	0.48
3a	O	H	1.10	4a	S	H	0.72
3b	O	3-F	0.53	4b	S	4-Cl	0.53
3c	O	4-F	0.35	4c	S	4-MeO	0.28
3d	O	4-Cl	0.18	4d	S	4- ^t Bu	0.92
3e	O	2-MeO	0.38	4e	S	3-Br	0.80
3f	O	3-MeO	0.54	4f	S	4-Br	0.68

^aRef 17.^bValues are the means of at least three separate determinations.**Table 2.** DNA-PK-inhibitory activity of thiopyran-4-ones identified from library screens^a

Compd	Ar	IC ₅₀ (μM) ^a	Compd	Ar	IC ₅₀ (μM) ^a
11a		0.92	12a		0.33
11b		0.35	12b		0.19
11c		0.84	12c		0.26

^aLibrary compounds were assayed for % inhibition at 1.0, 0.5 and 0.1 μM. IC₅₀ values were determined for compounds exhibiting > 50% inhibition at 1.0 μM, according to ref 17.

to extensive chemical modification, and in particular the introduction of substituents onto the fused aromatic ring system. Accordingly, it was envisaged that a 6-arylpyran-4-one or 6-arylthiopyran-4-one template, bearing the essential 2-morpholin-4-yl group, would offer more opportunities for introducing structural diversity in the aromatic region, while retaining the core pharmacophore elements common to **1** and **2**.

All of the pyran-4-ones (**3a–3h**) and thiopyran-4-ones (**4a–4f**) synthesised initially are more potent than LY294002 (**1**), with the pyran-4-ones generally tending to be at least equipotent (compare **3g** with **4c**) or slightly more potent (compare **3d** with **4b**, and **3h** with **4d**) than their thiopyran-4-one counterparts. An exception arises with the parent 6-phenyl derivatives, where the thiopyran-4-one (**4a**) is marginally more active than the corresponding pyran-4-one (**3a**). Although definitive SAR correlations are clearly not possible, it is evident

that a lipophilic substituent at the 4'-position of the 6-aryl ring is favoured for both series, with two pyran-4-ones (**3d** and **3g**) and one thiopyran-4-one (**4c**) exhibiting activity comparable with NU7026 (**2**). Perhaps more importantly, the pyran-4-one/thiopyran-4-one template was found to retain the selectivity for DNA-PK observed for the benzo[*h*]chromenone scaffold. For example, **3g** was found to be at least 200-fold more potent an inhibitor of DNA-PK (IC₅₀ = 0.22 μM) compared with the other PIKK family members PI 3-K (p110α), ATM and ATR (IC₅₀ > 50 μM). A similar degree of selectivity has also been observed in the thiopyran-4-one series.

Initial evaluation of the two thiopyran-4-one compound libraries (**11** and **12**) for DNA-PK inhibitory activity revealed a wide range of activities, with library members bearing an aryl substituent at the 4'-position (**12**) generally proving more potent than those with a 3'-aryl

group (**11**). Three compounds from each library (**11a–11c**) and (**12a–12c**) were identified from this pre-screen as being sufficiently potent to warrant IC₅₀ determinations (Table 2). Although there are no large differences in activity within this compound set, those thiopyran-4-ones with a 4'-aryl substituent (**12a–12c**) tend to be the more potent, consistent with the overall trend observed for the initial two libraries. Perhaps more interesting are the clear structural similarities between the 4'-aryl substituents on **12a–12c**, which comprise 1-naphthyl and 1-(4-methyl)naphthyl (**12b** and **12c**), and the isosteric 3-thianaphthene (3-benzo[*b*]thienyl) group (**12a**). These three thiopyran-4-ones exhibit DNA-PK inhibitory activity comparable with that of NU7026 (**2**), and are among the most potent DNA-PK inhibitors reported to date. In summary, we have established that pyran-4-ones and thiopyran-4-ones are versatile platforms for the development of potent and selective DNA-PK inhibitors, and have utilised a simple library approach for the identification of a potentially interesting class of inhibitors derived from the thiopyran-4-one pharmacophore. Further studies are underway to optimise the biological and pharmaceutical properties of this new series of DNA-PK inhibitors.

Acknowledgements

The authors thank Cancer Research UK and the BBSRC (Studentship to J.J.H.) for financial support.

References and Notes

1. Doherty, A. J.; Jackson, S. R. *Curr. Biol.* **2001**, *11*, R920.
2. Smith, G. C. M.; Jackson, S. P. *Genes Dev.* **1999**, *13*, 916.
3. Shiloh, Y. *Nat. Rev. Cancer* **2003**, *3*, 155.
4. Jackson, S. P. *Carcinogenesis* **2002**, *23*, 687.
5. Jackson, S. P. *Int. J. Biochem. Cell Biol.* **1997**, *29*, 935.
6. Rosenzweig, K. E.; Youmell, M. B.; Palayoor, S. T.; Price, B. D. *Clin. Cancer Res.* **1997**, *3*, 1149.
7. Boulton, S.; Kyle, S.; Durkacz, B. W. *Eur. J. Cancer* **2000**, *36*, 535.
8. Kim, C. H.; Park, S. J.; Lee, S. H. *J. Pharmacol. Exp. Ther.* **2002**, *303*, 753.
9. Oliveira, N. G.; Castro, M.; Rodrigues, A. S.; Gil, O. M.; Toscano-Rico, J. M.; Rueff, J. *Teratog. Carcinog. Mutag.* **2002**, *22*, 343.
10. Vlahos, C. J.; Matter, W. F.; Hui, K. Y.; Brown, R. F. *J. Biol. Chem.* **1994**, *269*, 5241.
11. Ward, S.; Sotsios, Y.; Dowden, J.; Bruce, I.; Finan, P. *Chem. Biol.* **2003**, *10*, 207, and references therein.
12. Izzard, R. A.; Jackson, S. P.; Smith, G. C. M. *Cancer Res.* **1999**, *59*, 2581.
13. Griffin, R. J.; Calvert, A. H.; Curtin, N. J.; Durkacz, B. W.; Golding, B. T.; Hardcastle, I. R.; Leahy, J. J. J.; Martin, N.; Newell, D. R.; Rigoreau, L.; Smith, G. C. M.; Stockley, M.; Veuger, S.; Hickson, I. *Proc. Am. Assoc. Cancer Res.* **2002**, *43*, 4210.
14. Morris, J.; Wishka, D. G. *Synthesis-Stuttgart* **1994**, 43.
15. All new compounds exhibited spectral (¹H NMR, IR, UV) and analytical (elemental analysis and/or LC-MS) data fully consistent with the assigned structures.
16. A general procedure for the MPS of thiopyran-4-ones is as follows: A solution of **4e** or **4f** (20 mg, 0.057 mmol) in dioxane (1 mL) was sonicated under N₂ for 5 min, before addition to a reaction tube containing the appropriate arylboronic acid (0.0625 mmol) and K₂CO₃ (19 mg, 0.14 mmol). A solution of tetrakis(triphenylphosphine)palladium(0) (3 mg) in dioxane (0.3 mL) was added, and the reaction mixture was stirred under N₂ at 90 °C for 18 h. After cooling, the product was isolated by chromatography on silica (isolute Si 500 mg cartridge), eluting with CH₂Cl₂:MeOH (3:1, 3 mL). Further purification by preparative HPLC furnished the required thiopyranones in 5–60% yield.
17. The DNA-PK used for in vitro assays was purified from HeLa cell nuclear extract. The known ability of DNA-PK to phosphorylate the serine-15 residue of a p53 peptide in vitro was exploited in a classic ELISA style assay using an antibody that only recognises the p53 serine-15 site when phosphorylated (Cell Signalling Technology). The primary antibody to p53 phosphoserine-15 was detected using an HRP conjugated goat anti-rabbit antibody (Pierce) with ECL reagent (NEN) being used for the readout. The ability of compounds to inhibit this phosphorylation event was monitored over a concentration range with IC₅₀ values generated from these results. Full details of the assay will be published elsewhere.