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Neighboring group participation Part 15. Stereoselective synthesis of some steroidal tetrahydrooxazin-2-ones, as novel presumed inhibitors of human 5α-reductase^[†]

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Abstract

During the alkaline methanolysis of 3β -acetoxy-21-chloromethyl-pregn-5-ene- 20β -*N*-phenylurethane, and its *p*-substituted phenyl derivatives, cyclization occurs, in the course of which 17β -[3-(*N*-phenyl)tetrahydrooxazin-2-on-6-yl]androst-5-en-3 β -ol and its *p*-substituted phenyl derivatives are formed. The cyclization takes place with (N⁻-6) neighboring group participation. Oppenauer oxidation of the 3β -hydroxy-*exo*-heterocyclic steroids yielded the corresponding Δ^4 -3-ketosteroids. The structures of the new compounds were proved by IR, ¹H and ¹³C NMR spectroscopy, using up-to-date measuring techniques such as 2D-COSY, HMQC, and HMBC. The inhibitory effects (CI₅₀) of the Δ^4 -3-ketosteroids on 5α -reductase were studied.

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1. Introduction

Prostatic cancer is the second leading cause of cancerrelated mortality worldwide. Considerable attention has been devoted to the development of drugs that are active against prostate cancer by intervening in the processes of tumor growth, and especially in the steroid metabolism pathways of testosterone production [2]. Androgen deprivation by the inhibition of human cytochrome P450_{17α} (17α-hydroxylase/C_{17,20}-lyase), the enzyme responsible for the conversion of C₂₁ steroids to C₁₉ androgens, has been proposed as a potential approach for the therapeutic treatment of this disease, which is androgen-dependent in the majority of cases [3]. The syntheses of abiraterone [17-(3-pyridyl)androsta-5,16-dien-3β-ol] and its 4-en-3-one analog were reported recently; these display a high inhibitory activity against P450_{17 α}. It has been suggested that this activity is related to the presence of the heterocyclic moiety in ring D [4]. Guarna et al. have reported on the synthesis of (+)-17-(3-pyridyl)-(5 β)-10-azaestra-1,16-dien-3-one, which may be a fundamental 5 α -reductase inhibitor [5]. Since 5 α -reductase converts testosterone to the more potent androgen dihydrotestosterone in prostatic tissues, the dual inhibition of 5 α -reductase and P450_{17 α} might be particularly beneficial in the treatment of prostate cancer [6]. Therefore, it was hypothesized that anchoring a 3'-pyridyl moiety on the D ring of 19-nor-10-azasteroid would produce novel steroid derivatives, as presumed inhibitors of both 5 α -reductase and P450_{17 α} [7].

We set out to synthesize a novel series of steroidal tetrahydrooxazin-2-ones containing heterocycles involving O and N heteroatoms at position 17 β of androst-5-en-3 β -ol and position 17 β of androst-4-en-3-one, respectively. The present work was stimulated by the importance

 $[\]stackrel{\scriptscriptstyle{\rm th}}{\to}$ For a preceding paper in this series, see reference [1].

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of pentacyclic steroids offering novel binding sites and binding distances to the receptors by introducing heteroatoms and various functionalities into the *exo*-heterocycles.

The discovery of two different genes encoding for two 5α -reductase isoenzymes, types 1 and 2 [8], with different biochemical characteristics and tissue distributions [9], added a further impulse to this research. In human, the type 1 isoenzyme is present mainly in the nongenital skin and in the liver whereas the type 2 is active in the prostate, genital skin, epididymis, seminal vesicle, and also in the liver. Cancerous and hyperplastic prostate are abundant in the type 2 isoenzyme, therefore tissue specimens of these tumors are suitable enzyme sources for the in vitro study of the 5α -reductase type 2 inhibitors. In vitro experiments on the newly synthesized steroid compounds **8a**–g were carried out with homogenates of human prostates.

2. Experimental

2.1. General

Melting points (mp) were determined on a Kofler block and are uncorrected. Specific rotations were measured in CHCl₃ (c 1) at 20 °C with a POLAMAT-A (Zeiss-Jena) polarimeter and are given in units of 10^{-1} degree cm² g⁻¹. Elemental analysis was carried out in the analytical laboratory of the University of Szeged with a Perkin-Elmer model 2400 CHN analyzer. The reactions were monitored by TLC on Kieselgel-G (Merck Si 254 F) layers (0.25 mm thick); solvent systems (ss) (A): acetone/benzene/light petroleum (30:35:35 v/v), (B): ethyl acetate/chloroform (10:90 v/v), (C): ethyl acetate/chloroform (5:95 v/v), (D): ethyl acetate/chloroform (1:99), and (E): dichloromethane. The spots were detected by spraying with 5% phosphomolybdic acid in 50% aqueous phosphoric acid. The $R_{\rm f}$ values were determined for the spots observed by illumination at 254 and 365 nm. Flash chromatography: silica gel 60, 40–63 μ m. All solvents were distilled prior to use. The IR spectra were run in KBr disks on a Bruker IFS-113v vacuum optic FT-spectrometer with an Aspect 2000 computer (3d and 6-8a, b, d, f, h) or on a Bruker IFS-55 FT-spectrometer controlled by Opus 3.0 software (4d, 5a, b and 6-8c, e, g). The ¹H and ¹³C NMR spectra were recorded in CDCl₃ or in (CD₃)₂SO solution in 5 or 10 mm tubes at room temperature, on a Bruker WM-250 and/or WP-80-SY FT-spectrometer controlled by an Aspect 2000 computer at 250.13 MHz (¹H) and 62.89 or 20.14 MHz (¹³C), and on a Bruker DRX 500 spectrometer at 500.13 (¹H) and 125.76 (^{13}C) MHz, with the deuterium signal of the solvent as the lock and TMS as internal standard. DEPT spectra were run in a standard manner, using only the $\Theta = 135^{\circ}$ pulse to separate CH/CH₃ and CH₂ lines phased "up" and "down," respectively. The HMQC and HMBC spectra were obtained by using the standard Bruker pulse programs.

The characteristic IR band frequencies and the ¹H and ¹³C NMR data on the compounds investigated (**3d**, **4d**, **5a**, **b**, **6a–h**, **7a–h**, and **8a–h**) are listed in Tables 1–3.

2.2. 21-Hydroxymethylpregn-5-ene-3β,20-diol (3a)

21-Hydroxymethylenepregn-5-en-3 β -ol-20-one (**2**) (3.44 g, 10 mmol), prepared by the procedure of Hirsch and Fujimoto [10] (mp 209–211 °C), was suspended in ethanol (50 ml) and cooled in an ice-bath, and KBH₄ (1.62 g, 0.03 mol) was added in small portions. The mixture was allowed to stand for 6 h, and was then neutralized with dilute HCl. The resulting solution was poured onto ice (300 g), and the precipitate that separated out was filtered off, dried over P₂O₅, and crystallized from ace-tone/water to give **3a** (3.05 g, 87%), mp 184–186 °C, $R_{\rm f} = 0.30$ (ss A); [α]_D – 26 (*c* 1 in methanol) ([10] mp 183–185 °C).

2.3. 3β -Acetoxy-21-hydroxymethylpregn-5-en-20 β -ol (**3d**) and 3β -acetoxy-21-hydroxy methylpregn-5-en-20 α -ol (**4d**)

3β-Acetoxypregn-5-ene-21-hydroxymethyl-20-hydroxy cyclic acetonide (**3b**) (4.30 g, 10 mmol), prepared by the procedure of Hirsch and Fujimoto [10] (mp 154–166 °C), was dissolved in methanol (60 ml) and *p*-toluenesulfonic acid (0.5 g) in water (3 ml) was added to the solution. The reaction mixture was allowed to stand for 6 h and was then poured onto water (500 ml). The precipitate that separated out was filtered off, dissolved in chloroform, and subjected to chromatographic separation on silica gel in ethyl acetate/chloroform (5:95), yielding pure **3d** (3.1 g, 79%), mp 165–168 °C, *R*_f = 0.35 (ss C); $[\alpha]_D^{20} - 53$ (*c* 1 in chloroform) (found: C, 73.76; H, 9.95. C₂₄H₃₈O₄ requires C, 73.81; H, 9.81%); continued elution resulted in a mixture of **3d** and **4d** (500 mg, 13%).

Continued elution furnished **4d** (170 mg, 4%), mp 183–186 °C, $R_{\rm f} = 0.25$ (ss B); $[\alpha]_{\rm D}^{20} - 45$ (*c* 1 in chloroform) (found: C, 73.58; H, 9.65. C₂₄H₃₈O₄ requires C, 73.81; H, 9.81%).

2.4. 3β -Acetoxy-21-p-toluenesulfonyloxymethylpregn-5ene-20 β -ol (5a)

Compound **3d** (3.9 g, 10 mmol) was dissolved in pyridine (30 ml), and a solution of *p*-toluenesulfonyl chloride (2.1 g, 11 mmol) dissolved in pyridine (10 ml) was added during cooling in ice. The reaction mixture was allowed to stand at room temperature for 20 h, and was then saturated with water. The precipitate was filtered off, dissolved in chloroform and chromatographed on silica gel with a mixture of ethyl acetate/chloroform (2.5:75.5) to obtain **5a** (4.8 g, 88%), mp 176–178 °C, $R_f = 0.45$ (ss C); $[\alpha]_D^{20} - 20$ (*c* 1 in chloroform) (found C, 68.42; H, 8.02. C₃₁H₄₄O₆S requires C, 68.35; H, 8.14%).

Table 1 ¹H NMR data^a and characteristic IR frequencies^b of compounds 3d, 4d, 5a, b, 6a-h, 7a-h, and 8a-h

Compound	CH ₃ ^c (OAc)	CH3 ^d (Ar)	CH ₃ ^c Pos. 18	CH ₃ ^c Pos. 19	H-3 ^e <i>m</i> (1H)	$\frac{\text{H-6/4}^{\text{f}}}{\text{(1H)}} d$	OCH ^g m (1H)	XCH ₂ ^{g,h} <i>m</i> (2H)	OH/NH br (1H)	ArH-2',6' $\sim d (2H)^{i}$	ArH-3',5' $\sim d (2H)^{i}$	νC=O band ^j	vNCOO band	$\gamma C_A r H$ band
3d	2.03	_	0.79	1.03	4.62	5.40	~3.85	~3.85	$\sim 2.65^{h}$	_	_	1733	_	_
4d	2.03	_	0.70	1.02	4.60	5.38	3.85	3.85	~ 2.65	_	_	1732	_	_
5a	2.02	2.44	0.73	1.01	4.59	5.36	3.63	4.14		7.78	7.34	1713	_	903
5b	2.02	_	0.78	1.02	4.59	5.36	3.75	3.70		_	_	1717	_	-
6a	2.03	2.36	0.70	0.98	4.62	5.36	4.95	4.12	6.69	7.74	$7.05 - 7.4^{k}$	1731	1731	813 ¹
6b	2.03	2.30	0.74	0.99	4.60	5.3	5.00	3.56	6.73	7.10	7.30	1740	1723	817
6c	2.04	1.22	0.75	1.00	4.61	5.37	5.02	3.57	6.73	7.33	~7.14	1721	1721	832
6d	2.03	3.79	0.74	0.99	4.62	5.38	5.00	3.57	6.62	~ 7.35	6.85	1740	1722	829
6e	2.03	1.39	0.74	1.02	4.60	5.37	5.10	3.56	~ 6.6	~7.3	6.83	1731	1731	830
6f	2.04		0.74	1.00	4.60	5.36	5.00	3.56	6.95	7.00	\sim 7.4	1735	1716	833
6g	2.04		0.74	0.99	4.61	5.36	5.02	3.56	6.91	7.26	\sim 7.38	1734	1714	828
6h	2.04		0.74	0.99	4.65	5.40	5.05	3.56	~ 6.8	7.2–7.5	7.2–7.5	1740	1724	822
7a	-	-	0.78	0.97	$\sim 3.25^{\rm m}$	5.27	4.38	3.55	4.56	7.22 ^k	$\sim 7.35^k$	_	1667	759 ¹
7b	-	2.29	0.78	0.97	$\sim 3.25^{\rm m}$	5.27	4.35	3.55	4.50	\sim 7.20	\sim 7.20		1676	823
7c	_	1.24	0.85	1.03	3.52	5.35	4.36	3.62		\sim 7.22	~ 7.22		1672	840
7d	_	3.80	0.84	1.03	$\sim 3.6^{n}$	5.35	4.35	$\sim 3.6^{n}$		7.25	6.90	_	1680	832
7e	_	1.40	0.76	1.01	3.51	5.35	4.32	3.57		7.19	6.87	_	1686	830
7f	-	-	0.84	1.03	$\sim 3.65^{n}$	5.40	~ 4.4	$\sim 3.65^{n}$		7.21	7.49	_	1692	827
7g	-	-	0.84	1.03	3.53	5.36	4.36	3.62	~ 1.6	7.26	7.35	-	1702	826
7h	_	_	0.84	1.03	~ 3.55	5.35	4.36	~ 3.75		~ 7.05	~7.35	_	1673	837
8a	-	-	0.88	1.19	_	5.72	4.36	\sim 3.7	_	\sim 7.3	~7.3	1675	1695	754 ¹
8b	_	2.33	0.87	1.19	_	5.72	4.35	~ 3.65	_	~ 7.18	~ 7.18	1680	1680	816
8c	_	1.22	0.86	1.19	_	5.71	4.34	3.62	_	\sim 7.20	\sim 7.20	1670	1686	840
8d	_	3.80	0.87	1.20	_	5.72	4.35	~ 3.72	_	7.21	6.90	1683	1683	850
8e	-	1.40	0.80	1.20	_	5.74	4.34	3.58	_	7.19	6.88	1686	1686	840
8f	-	_	0.87	1.20	_	5.73	$\sim \!\! 4.4$	~ 3.7	_	7.20	7.49	1670	1700	824
8g	-	_	0.86	1.19	_	5.72	4.36	3.62	_	7.25	7.33	1682	1682	845
8h	-	-	0.87	1.20	_	5.72	4.38	~ 3.65	_	\sim 7.1	~7.3	1670	1692	850

^a Measuring frequency: 500 (4d, 5a, b, and 6–8c, e, g) or 250 MHz (3d and 6–8a, b, d, f, h). Solvent: CDCl₃ or DMSO-d₆ (for 7a, b). Chemical shifts in ppm ($\delta_{TMS} = 0$ ppm), coupling constants in Hz. Assignments were supported by 2D-HMQC (for 4d, 5a, 6c, 7g, and 8c) and for 4d also by 2D-COSY measurements.

^b In KBr discs (cm⁻¹). Further bands, vOH (doubled for 7e, h): 3380–3500 (3d, 4d, 5a, b, and 7a–h), vNH: 3315–3430 (6a–h, doubled for 6d, e), vSO₂: 1358, 1190, 1178 (5a and 6a). ^c s (3H).

^d s (3H), OMe group (6–8d), (Ar)Et group (6–8c, e): t (J: 7.0), CH₂ (ArEt): 2.62 (6–8c), OCH₂ (ArEt): 4.02 (7e and 8e).

^e For **5a**, **b** and **6a-h** tt with coalesced lines.

^f Pos. 6 (3–7), Pos. 4 (8),~d, J: 4.5 \pm 0.6 (5, 6), 5.1 \pm 0.1 (7), ~1 (8).

^g In the side-chain or oxazinone ring, X: O (3d, 4d, and 5a, 6a), Cl (5b and 6b-h), or N (7 and 8).

^h Two *m*'s (2 × 1H) with the second signal at ~2.95 (2H, 3d), 4.25 (5a), 3.72 ± 0.03 (7a, b, c, g, and 8c, g), 3.64 ± 0.02 (7e and 8e).

ⁱ Rudimentaly AA'BB'-type spectrum: J: 8.3 (**5a** and **6a–c**), 8.8 (**5d**, e, g, **7d–g**, and **8d–g**), $\sim s$, broad signals with coalesced fine structure (due to F,H-couplings for **6–8h**): $2 \times m$ ($2 \times 2H$, **6–8h**), m (4H) for **6h**, **7b**, c, and **8b**, c, m (5H) for **8a**.

^j Ester (3d, 4d, 5a, b, and 6a-h) or ketone group (8a-h).

^k Intensity: 7/1/4H.

¹*p*-Disubstituted benzene ring, 748 (phenyl), γC_{Ar}C_{Ar} (phenyl): 698 (**6a**, **7a**, and **8a**).

^m Hidden by the water signal of the solvent.

ⁿ Overlapping signals.

Table 2		
¹³ C NMR chemical shifts ^a	of compounds 3d, 4d, 5a,	b, 6a-h, 7a-h, and 8a-h ^{b,c}

Compound	C-1	C-2	C-3	C-4	C-5	C-6	C-7	C-8	C-9	C-10	C-11	C-12	C-13	C-14	C-15	C-16	C-17	C-18	C-19
3d	38.0 ^d	37.1	74.0 ^d	39.8	139.8	122.4	31.9	31.8	50.1	36.6	21.0	38.1 ^d	42.4	56.2 ^d	25.4	27.8	56.5 ^d	12.3	19.3
4d	38.4 ^d	37.4	74.3	39.3	140.1	122.9	32.2	31.9	50.4	37.0	21.1	38.5 ^d	42.1	56.9 ^d	25.4	28.2	57.1 ^d	13.1	19.7
5a	37.4	36.3	74.3	38.5	140.1	122.8	32.3	32.1	50.4	37.0	21.3	40.3	42.8	56.9	25.6	28.1	56.4	12.8	19.7
5b	38.5	37.4	74.3	40.0 ^d	140.1	122.8	32.3	32.1	50.4	37.0	21.3	40.3 ^d	42.9	57.0	25.7	28.1	56.5	12.8	19.7
6a	38.2	37.1	74.0	39.1	140.0	122.3	31.9 ^e	31.9 ^e	50.3	36.7	21.0	33.6	42.4	56.1	25.4	27.9	53.4	12.3	19.2
6b	38.1 ^d	37.0	74.0	39.1 ^d	139.9	122.3	31.9	31.8	50.2	36.6	21.0	38.2 ^d	42.4	56.1	25.5	27.8	53.6	12.4	19.3
6c	38.4 ^d	37.4	74.4	38.5 ^d	139.9	122.8	32.3	32.1	50.5	37.0	21.4	39.5	42.8	56.4	25.9	28.2	54.0	12.8	19.7
6d	38.1 ^d	37.1	74.0 ^d	39.1	139.9	122.3	31.8 ^e	31.8 ^e	50.2	36.6	21.0	38.2 ^d	42.5	56.1	25.5	27.8	53.7	12.3	19.3
6e	38.5 ^d	37.4	74.3 ^e	39.1	140.1	120.8	32.2	31.9	50.4	37.0	21.1	38.7 ^d	42.2	56.8	25.1	28.1	54.6	12.9	19.7
6f	38.0 ^d	37.1	74.0 ^d	39.2	139.9	122.3	31.9	31.8	50.2	37.0	21.1	38.7 ^d	42.2	56.8	25.1	28.1	54.6	12.9	19.7
6g	38.5 ^d	37.4	74.4	39.5 ^d	140.2	120.3	32.2	32.1	50.5	37.0	21.4	38.3 ^d	42.8	56.4	25.8	28.2	53.9	12.8	19.7
6h	38.2 ^d	37.0	74.0	39.1 ^d	139.9	122.3	31.83	31.75	50.2	36.7	210	37.9 ^d	42.4	56.1	25.5	27.8	53.5	12.4	19.3
7a	38.7	33.1 ^e	71.7	43.9	143.2	121.8	33.1 ^e	33.1 ^e	51.6	37.8	22.0	40.4	43.7	57.4	25.7 ^d	28.1	54.9	13.7	20.8
7b	38.6	33.0 ^e	71.7	43.8	143.1	121.6	33.0 ^e	33.0 ^e	51.5	37.8	21.9	40.3	43.6	57.3	25.5 ^d	28.0	54.9	13.6	20.6
7c	37.6	32.1 ^d	72.2	42.7		121.7	32.4 ^d	32.2	50.6	37.0	21.2	39.4	43.0	56.7	25.1 ^d	27.3	54.0	12.7	19.8
7d	37.4	32.1 ^d	71.9	42.4	143.3	121.3	31.9 ^d	32.0	50.5	36.7	20.9	39.2	42.7	56.5	24.8 ^d	27.1	53.9	12.3	19.5
7e	37.7	32.2 ^d	72.0	42.6	141.2	121.8	32.0 ^d	31.9	50.5	36.9	21.2	39.3	42.2	56.8	26.0 ^d	28.1	54.9	13.0	19.2
7f	38.5	33.0 ^e	71.6	43.8		121.5	33.0 ^e	33.0 ^e	51.5	37.7	21.9	40.2	43.5	57.2	25.5 ^d	27.9	54.8	13.5	20.5
7g 7h	37.6 37.5	32.1 32.2 ^d	72.2	42.7	141.5		32.4 32.0 ^d	32.2	50.6	37.0	21.2	39.4	43.0	56.7	24.9 ^d 24.9 ^d	27.2 27.1	54.1 54.0	12.7	19.8
7h 8-	37.5 35.6 ^d	32.2 ^d 33.9 ^d	71.8 199.1	42.6 123.8	141.4 170.9	121.3 32.8 ^d	32.0 ^d	32.1 ^d 35.7 ^d	50.6 53.7	36.8	21.0 20.8	39.3	42.8	56.6	24.9 ^d 24.6 ^d	27.1 26.9 ^d	54.0 54.0	12.3 12.3	19.5 17.4
8a 8b	35.7 ^d	34.0 ^d	199.1 199.5	123.8	170.9	32.8 ^d	32.2 ^d	35.6 ^d	53.7 53.7	38.7 38.7	20.8	39.0 38.9	42.7 42.7	55.5 55.5	24.0 24.7 ^d	26.9 ^d	54.0 53.9	12.3	17.4
80 80	36.1	34.0 34.4	200.0	125.8	171.5	33.3	32.6	35.9	55.7 54.3	38.7 39.1	20.8	39.3	42.7	55.5 55.8	24.7 24.8 ^d	20.9 ^d	55.9 54.0	12.5	17.4
ac 8d	35.7 ^d	33.9	200.0 199.4	124.2	171.7	32.8	32.0	35.5 ^d	53.6 ^d	38.6	20.8	38.9	43.0	55.8 ^e	24.6 ^d	27.2 ^d 26.9 ^d	53.9 ^d	12.7	17.8
ou 8e	36.1	33.9 34.4	199.4	123.8	171.3	33.2	32.1	35.5 35.6	55.0 54.1 ^d	39.0	20.8	38.9 39.1	42.0	55.4 55.9	24.0 24.5 ^d	20.9 28.2 ^d	54.9 ^d	12.5	17.4
8f	35.6	33.8	199.8	124.3	171.4	32.7 ^d	32.3 32.0 ^d	35.0	53.6 ^d	39.0	20.7	38.8	42.5	55.3	24.3 24.4 ^d	26.2 26.7 ^d	53.8 ^d	12.2	17.8
8g	36.1	34.4	200.0	123.7	171.0	33.3 ^d	32.0 ^d	35.9	54.0 ^d	39.1	20.7	39.3	43.0	55.8	24.4 24.8 ^d	20.7 27.2 ^d	54.3 ^d	12.2	17.8
og 8h	35.8	34.4 34.0	200.0 199.4	124.2 123.9	171.1	33.3 32.9 ^d	32.3 ^d	35.6	53.7 ^d	39.1	20.8	39.3 39.0	43.0	55.8 55.5	24.8 24.4 ^d	27.2 26.9 ^d	54.0 ^d	12.8	17.8

^a In ppm ($\delta_{TMS} = 0$ ppm) at 126.7, 62.9 (3d, 6b, f, h, 7b, d, f, h, and 8b, d, f, h) or 20.1 MHz (6a, d, 7a, and 8a). Solvent: CDCl₃ or DMSO-d₆ (for 7a, b), respectively.

^b Assignments were supported by DEPT (except for 3d, 6a, 7b, f, h, and 8a, b, d), and for 4d, 5a, 6c, 7g, and 8c also by HMQC and HMBC measurements.

^c Further lines: see Table 3 and the footnote to it.

^d Interchangeable assignments.

e Overlapping lines.

2.5. 3β -Acetoxy-21-chloromethylpregn-5-ene-20 β -ol (5b)

Compound **3d** (3.9 g, 10 mmol) was suspended in CCl₄ (40 ml) and Ph₃P (2.88 g, 3.1 mmol) was added. After an 6-h reflux, the solvent was evaporated off. The resulting crude product was chromatographed on silica gel with a mixture of ethyl acetate/chloroform (2.5:97.5) to obtain **5b** (2.9 g, 74%), mp 183–184 °C, $R_f = 0.50$ (ss C); $[\alpha]_D^{20} - 31$ (*c* 1 in chloroform) (found C, 70.56; H, 8.98. C₂₄H₃₇O₃Cl requires C, 70.48; H, 9.12%).

2.6. 3β-Acetoxy-21-p-toluenesulfonyloxymethylpregn-5ene-20β-N-phenylurethane (**6a**) and 3β-acetoxy-21chloromethylpregn-5-ene-20β-N-phenylurethane (**6b**)

2.6.1. General procedure

Compound **5a** (1.09 g, 2 mmol) or **5b** (818 mg; 2 mmol) was dissolved in dichloromethane (25 ml) and phenyl isocyanate (960 mg, 8 mmol) and triethylamine (0.1 ml) were added. The reaction mixture was refluxed for 6 h, and was then poured into 10% NaHCO₃ solution (50 ml). The

dichloromethane fraction was washed thoroughly with water, dried and evaporated to dryness. The residue was subjected to chromatographic separation with dichloromethane.

Compound **5b** (818 mg; 2 mmol) was dissolved in benzene (25 ml) and triethylamine (0.3 ml) and a solution of phosgene in benzene (5 ml; 10%) were added. The reaction mixture was allowed to stand at room temperature for 24 h. Diethyl ether (30 ml) was then added, and the triethylamine HCl that separated out was filtered off and the filtrate was evaporated in vacuo. The residue was dissolved in benzene (30 ml), and triethylamine (0.3 ml) and the corresponding substituted aniline (2.2 mmol) were added. The mixture was allowed to stand for 6 h, and was then diluted with water. The precipitate that separated out was filtered off and dried over P₂O₅. The resulting crude product was chromatographed on silica gel with dichloromethane.

2.6.2. 3β -Acetoxy-21-p-toluenesulfonyloxymethylpregn-5-ene-20 β -N-phenylurethane (**6a**)

6a (1.18 g; 85%), mp 166–167 °C, $R_{\rm f} = 0.6$ (ss D); $[\alpha]_{\rm D}^{20} - 51$ (*c* 1 in chloroform) (found C, 68.82; H, 7.55. C₃₈H₄₉O₇SN requires C, 68.75; H, 7.44%).

Table 3	
¹³ C NMR chemical shifts ^a of compounds 3d, 4d, 5a, b, 6a-h, 7a-h, and 8a-h	b,c

Compound	CH ₃ (OAc)	CH ₃ (Ar)	Side chain or oxazinone ring			Aryl gro	oup	NCOO	C=O (OAc)		
			CH(OH)	CCH ₂ C	XCH ₂	C-1′	C-2′,6′	C-3′,5′	C-4′		
3d	21.3	_	74.3 ^d	24.6	61.4	_	_	_	_	_	170.6
4d	21.8	_	74.6	24.5	62.3	_	_	_	_	_	171.0
5a	21.8	22.0	70.8	24.9	68.3	133.6	128.3	130.3	145.2	_	170.9
5b	21.8	_	71.9	24.9	42.3	_	_	_	_	_	170.6
6a	21.3 ^d	21.5 ^d	73.6	24.3	67.1	133.5 ^e	129.0 ^e	129.8 ^e	144.6 ^e	152.9	170.4
6b	21.4	20.7	74.5	24.4	40.7	133.0	119.0	129.5	135.5	153.2	170.5
6c	21.8	16.2	74.8	24.7	41.1	136.0	119.2	128.8	140.2	153.5	170.6
6d	21.3	55.0	74.5	24.3	40.6	131.3	121.0	114.4	156.2	153.5	170.4
6e	21.8	15.3	74.3 ^d	24.4	41.3	131.4	122.8	115.3	155.7	154.1	170.6
6f	21.4	_	74.8	24.4	0.6	134.1	$\sim \! 120.7$	115.6 ^f	159.1 ^f	153.3	170.6
6g	21.9	_	75.2	24.4	41.0	137.1	122.7	129.4	128.8	153.4	171.2
6h	21.4	_	74.8 ^d	24.3	40.6	137.4	120.4	131.9	115.9	153.0	170.7
7a	_	_	80.4	25.6 ^d	48.7	145.2	127.4	130.3	127.5	153.2	_
7b	_	22.0	80.2	25.6 ^d	48.7	142.6	127.1	130.6	136.7	153.6	_
7c	_	15.8	79.6	24.9 ^d	48.2	141.1 ^d	126.1	129.0	141.6 ^d	153.1	_
7d	_	55.6	79.3	24.8 ^d	48.2	136.4	127.2	114.6	158.3	152.8	_
7e	_	15.2	80.7	24.6 ^d	48.8	136.2	127.6	115.4	157.9	153.8	_
7f	_	_	80.4	25.4 ^d	48.3	144.3	129.2	132.9	119.7	152.9	_
7g	_	_	79.9	25.1 ^d	48.0	142.0	127.4	129.6	132.4	152.8	_
7h	_	_	79.5	24.6 ^d	48.0	139.2	127.6 ^f	116.0 ^f	161.0 ^f	152.6	_
8a	_	_	79.1	26.9	47.6	143.3	125.7	129.0	126.4	152.3	_
8b	_	21.0	79.1	26.9	47.8	140.6	125.7	129.8	136.4	152.6	_
8c	_	15.8	79.5	27.2	48.2	141.1	126.1	129.0	143.1	153.0	_
8d	_	55.4 ^g	79.1	26.9	48.1	136.1	127.1	114.4	158.1	152.8	_
8e	_	15.2	80.2	28.2	48.8	136.1	127.6	115.5	158.0	153.7	_
8f	_	_	79.2	26.7	47.3	142.1	127.1	131.9	119.5	152.0	_
8g	_	_	79.8	27.2	48.0	142.0	127.4	129.6	132.5	152.7	_
8h	_	_	79.3	26.9	48.0	139.1 ^f	127.6 ^f	116.0 ^f	161.0 ^f	152.5	_

^a In ppm ($\delta_{TMS} = 0$ ppm) at 125.7 and 62.9 (3d, 6b, f, h, 7b, d, f, h, and 8b, d, f, h) or 20.1 MHz (6a, d, 7a, and 8a). Solvent: CDCl₃ or DMSO-d₆ (for 7a, b), respectively.

^b Assignments were supported by DEPT (except for **3d**, **6a**, **7b**, **f**, **h**, and **8a**, **b**, **d**), for **4d**, **5a**, **6c**, **7g**, and **8c** also by HMQC and HMBC measurements. ^c Further lines (see also Table 2): phenyl (**6a**), C-1: 138.3, C-2,6: 118.9, C-3,5: 127.9, C-4: 123.4, ArCH₂: 28.7 (**6–8c**), OCH₂: 64.1 (**5–7e**).

^d Interchangeable assignments.

^e Tosvl group.

^f Doublet due to F,C-couplings, ¹*J*: 242.5 (6f) and 246.2 (7h and 8h), ²*J*: 22.4 (6f) and 22.7 (7h and 9h), ³*J*: 8.4 (7h and 8h), ⁴*J*: 3.2 (8h). ^g Two overlapping lines.

2.6.3. 3β -Acetoxy-21-chloromethylpregn-5-ene-20 β -N-4-tolylurethane (**6b**)

6b (960 mg; 88%), mp 162–164 °C, $R_{\rm f} = 0.6$ (ss E); $[\alpha]_{\rm D}^{20} + 8$ (*c* 1 in chloroform) (found C, 70.95; H, 8.05. C₃₂H₄₄O₄NCl requires C, 70.89; H, 8.18%);

2.6.4. 3β -Acetoxy-21-chloromethylpregn-5-ene-20 β -N-4-ethylphenylurethane (**6c**)

6c (910 mg; 81%) mp 129–131 °C, $R_f = 0.55$ (ss E); $[\alpha]_D^{20} + 4$ (*c* 1 in chloroform) (found C, 71.35; H, 8.22. C₃₃H₄₆O₄NCl requires C, 71.26; H, 8.34%).

2.6.5. *3β*-Acetoxy-21-chloromethylpregn-5-ene-20β-N-4-methoxyphenylurethane (**6***d*)

6d (895 mg; 80%), mp 105–107 °C, $R_{\rm f} = 0.40$ (ss E); $[\alpha]_{\rm D}^{20} + 4$ (*c* 1 in chloroform) (found C, 68.75; H, 8.02. C₃₂H₄₄O₅NCl requires C, 68.86; H, 7.95%).

2.6.6. 3β -Acetoxy-21-chloromethylpregn-5-ene-20 β -N-4-ethoxyphenylurethane (**6**e)

6e (980 mg; 85%), mp 157–159 °C, $R_{\rm f} = 0.50$ (ss E); $[\alpha]_{\rm D}^{20} - 50$ (*c* 1 in chloroform) (found C, 69.36; H, 8.28. C₃₃H₄₆O₅NCl requires C, 69.27; H, 8.10%).

2.6.7. 3β -Acetoxy-21-chloromethylpregn-5-ene-20 β -N-4-chlorophenylurethane (**6f**)

6f (900 mg; 80%), mp 116–118 °C, $R_f = 0.70$ (ss E); $[\alpha]_D^{20} + 11$ (*c* 1 in chloroform) (found C, 66.27; H, 7.35. C₃₃H₄₁O₄NCl₂ requires C, 66.18; H, 7.35%).

2.6.8. 3β -Acetoxy-21-chloromethylpregn-5-ene-20 β -N-4-bromophenylurethane (**6g**)

6g (950 mg; 78%), mp 140–142 °C, $R_f = 0.60$ (ss E); $[\alpha]_D^{20} + 13$ (*c* 1 in chloroform) (found C, 61.52; H, 6.72. C₃₁H₄₁O₄BrClN requires C, 61.34; H, 6.81%).

2.6.9. 3β -Acetoxy-21-chloromethylpregn-5-ene-20 β -N-4-fluorophenylurethane (**6h**)

6h (920 mg; 84%), mp 107–110 °C, $R_{\rm f} = 0.65$ (ss E); $[\alpha]_{\rm D}^{20} - 5$ (*c* 1 in chloroform) (found C, 68.23; H, 7.67. C₃₁H₄₁O₄NFCl requires C, 68.18; H, 7.57%).

2.7. 17β -[3-(N-Aryl)-tetrahydrooxazin-2-on-6-yl]androst-5-en-3 β -ol (**7a**-**h**)

2.7.1. General procedure

The individual compounds **6a**–**h** (2 mmol) were dissolved in methanol (30 ml), NaOCH₃ (432 mg, 8 mmol) was added and the solution was kept at the boiling point. The progress of the reaction was monitored by TLC. Refluxing was continued until the starting material had been fully consumed (about 120 min). The reaction mixture was then neutralized with dilute HCl, and diluted with water. The resulting precipitate was filtered off, dissolved in dichloromethane and chromatographed on silica gel with a mixture of ethyl acetate/chloroform (5:95).

2.7.2. 17β -[3-(N-Phenyl-tetrahydrooxazin-2-on-6-yl)] and rost-5-en-3 β -ol (**7***a*)

7a (786 mg; 86%), mp 212–214 °C, $R_{\rm f} = 0.45$ (ss B); $[\alpha]_{\rm D}^{20} - 79$ (*c* 1 in chloroform) (found C, 77.58; H, 8.57. C₂₉H₃₉O₃N requires C, 77.47; H, 8.74%).

2.7.3. 17β -[3-(N-4-Tolyl)tetrahydrooxazin-2-on-6-yl] androst-5-en-3 β -ol (**7b**)

7b (810 mg; 87%), mp 240–243 °C, $R_{\rm f} = 0.45$ (ss B); $[\alpha]_{\rm D}^{20} - 82$ (*c* 1 in chloroform) (found C, 77.93; H, 9.05. C₃₀H₄₁O₃N requires C, 77.71; H, 8.91%).

2.7.4. 17β-[3-(N-4-Ethylphenyl)tetrahydrooxazin-2-on-6-yl]androst-5-en-3β-ol (**7c**)

7c (805 mg; 84%), mp 190–192 °C, $R_{\rm f} = 0.40$ (ss B); $[\alpha]_{\rm D}^{20} - 73$ (*c* 1 in chloroform) (found C, 77.82; H, 8.98. C₃₁H₄₃O₃N requires C, 77.95; H, 9.07%).

2.7.5. 17β -[3-(N-4-Methoxyphenyl)tetrahydrooxazin-2on-6-yl]androst-5-en-3 β -ol (**7d**)

7d (790 mg; 82%), mp 232–234 °C, $R_{\rm f} = 0.35$ (ss B); $[\alpha]_{\rm D}^{20} - 81$ (*c* 1 in chloroform) (found C, 75.25; H, 8.55. C₃₀H₄₁O₄N requires C, 75.12; H, 8.62%).

2.7.6. 17β -[3-(N-4-Ethoxyphenyl)tetrahydrooxazin-2-on-6-yl]androst-5-en-3 β -ol (**7**e)

7e (832 mg; 84%), mp 207–210 °C, $R_{\rm f} = 0.45$ (ss B); $[\alpha]_{\rm D}^{20} + 2$ (*c* 1 in chloroform) (found C, 75.55; H, 8.87. C₃₁H₄₃O₄N requires C, 75.42; H, 8.78%).

2.7.7. 17β -[3-(N-4-Bromophenyl)tetrahydrooxazin-2-on-6-yl]androst-5-en-3 β -ol (**7f**)

7f (910 mg; 86%), mp 213–214 °C, $R_f = 0.45$ (ss B); $[\alpha]_D^{20} - 72$ (*c* 1 in chloroform) (found C, 66.08; H, 7.32. C₂₉H₃₈O₃NBr requires C, 65.90; H, 7.25%).

2.7.8. 17β-[3-(N-4-Chlorophenyl)tetrahydrooxazin-2on-6-yl]androst-5-en-3β-ol (**7g**)

7g (915 mg; 94%), mp 179–183 °C, $R_{\rm f} = 0.30$ (ss B); $[\alpha]_{\rm D}^{20} - 70$ (*c* 1 in chloroform) (found C, 72.06; H, 7.83. C₂₉H₃₈O₃NCl requires C, 71.96; H, 7.91%).

2.7.9. 17β-[3-(N-4-Fluorophenyl)tetrahydrooxazin-2on-6-yl]androst-5-en-3β-ol (**7h**)

7h (876 mg; 93%), mp 235–237 °C, $R_{\rm f} = 0.35$ (ss B); $[\alpha]_{\rm D}^{20} - 69$ (*c* 1 in chloroform) (found C, 74.63; H, 8.02. C₂₉H₃₈O₃NF requires C, 74.49; H, 8.19%).

2.8. 17β-[3-(N-Aryl)tetrahydrooxazin-2-on-6-yl]androst-4-en-3-one (**8a-h**)

2.8.1. General procedure

The individual compounds 7a-h (2 mmol) were dissolved in toluene (60 ml), aluminium isopropoxide (2.04 g, 10 mmol) and cyclohexanone (20 ml) were added, and the mixture was stirred at the boiling point. The disappearance of the starting material was followed by TLC. When the reaction was complete, dilute H₂SO₄ (1%, 200 ml) was added to the reaction mixture. Most of the organic solvent was removed in vacuo and the residual emulsion was extracted with chloroform. The organic phase was evaporated down and the resulting crude product was chromatographed on silica gel with ethyl acetate/chloroform (5:95).

2.8.2. 17β -[3-(N-Phenyl)tetrahydrooxazin-2-on-6-yl] androst-4-en-3-one (**8a**)

8a (720 mg; 80%), mp 234–235 °C, $R_f = 0.55$ (ss B); $[\alpha]_D^{20} + 46$ (*c* 1 in chloroform) (found C, 77.64; H, 8.45. C₂₉H₃₇O₃N requires C, 77.82; H, 8.33%).

2.8.3. 17β -[3-(N-4-tolyl)tetrahydrooxazin-2-on-6-yl] androst-4-en-3-one (**8b**)

8b (685 mg; 74%), mp 208–209 °C, $R_{\rm f} = 0.45$ (ss B); $[\alpha]_{\rm D}^{20} + 45$ (*c* 1 in chloroform) (found C, 77.94; H, 8.67. C₃₀H₃₉O₃N requires C, 78.05; H, 8.52%).

2.8.4. 17β-[3-(N-4-Ethylphenyl)tetrahydrooxazin-2on-6-yl]androst-4-en-3-one (8c)

8c (692 mg; 75%), mp 183–186 °C, $R_{\rm f} = 0.50$ (ss B); $[\alpha]_{\rm D}^{20} + 52$ (*c* 1 in chloroform) (found C, 78.37; H, 8.74. C₃₁H₄₁O₃N requires C, 78.28; H, 8.69%);

2.8.5. 17β-[3-(N-4-Methoxyphenyl)tetrahydrooxazin-2on-6-yl]androst-4-en-3-one (**8d**)

8d (756 mg; 79%), mp 228–231 °C, $R_{\rm f} = 0.50$ (ss B); $[\alpha]_{\rm D}^{20} + 29$ (*c* 1 in chloroform) (found C, 75.62; H, 8.25. C₃₀H₃₉O₄N requires C, 75.44; H, 8.23%).

2.8.6. 17β-[3-(N-4-Ethoxyphenyl)tetrahydrooxazin-2-on-6-yl]androst-4-en-3-one (**8**e)

8e (872 mg; 88%), mp 198–201 °C, $R_f = 0.45$ (ss B); $[\alpha]_D^{20} + 126$ (*c* 1 in chloroform) (found C, 75.61; H, 8.65. C₃₁H₄₁O₄N requires C, 75.73; H, 8.41%).

2.8.7. 17β-[3-(N-4-Bromophenyl)tetrahydrooxazin-2on-6-yl]androst-4-en-3-one (8f)

8f (874 mg; 83%), mp 222–225 °C, $R_{\rm f} = 0.60$ (ss B); $[\alpha]_{\rm D}^{20} + 47$ (*c* 1 in chloroform) (found C, 65.98; H, 6.67. C₂₉H₃₆O₃NBr requires C, 66.16; H, 6.89%).

2.8.8. 17β -[3-(N-4-Chlorophenyl)tetrahydrooxazin-2on-6-yl]androst-4-en-3-one (**8g**)

8g (790 mg; 81%), mp 210–213 °C, $R_{\rm f} = 0.40$ (ss B); $[\alpha]_{\rm D}^{20} + 46$ (*c* 1 in chloroform) (found C, 72.43; H, 7.41. C₂₉H₃₆O₃NCl requires C, 72.26; H, 7.53%).

2.8.9. 17β -[3-(N-4-Fluorophenyl)tetrahydrooxazin-2on-6-yl]androst-4-en-3-one (**8h**)

8h (780 mg; 83%), mp 237–239 °C, $R_{\rm f} = 0.55$ (ss B); $[\alpha]_{\rm D}^{20} + 50$ (*c* 1 in chloroform) (found C, 74.65; H, 7.91. C₂₉H₃₆O₃NF requires C, 74.81; H, 7.79%).

2.9. Determination of 5α -reductase activity and its inhibition in the human prostate

Human prostates were obtained surgically from patients with benign prostatic hyperplasia and homogenized [11]. The protein content was determined according to Lowry et al. [12]. The homogenate was incubated with 200 nM [4-¹⁴C]testosterone and the test compound (in four different concentrations) in the presence of 1 mM NADPH for 20 min at 37 °C. The control experiments were performed without the test substances in every series. The enzymatic reaction was stopped by the addition of ethyl acetate and freezing. After extraction the radioactive testosterone and dihydrotestosterone were separated by TLC and a Packard Radiochromatogram Scanner was used to trace the separated steroids. Following measurement of the radioactivity of the dihydrotestosterone formed and the substrate testosterone, the 5 α -reductase activity was calculated and expressed in terms of pmol dihydrotestosterone per mg of protein for a 20-min incubation. The inhibitory effects of the compounds investigated are given in terms of IC₅₀ values, i.e. the concentration of inhibitor at which the 5α -reductase activity was decreased to 50%.

3. Results and discussion

3.1. Synthetic studies

For the synthesis of tetrahydrooxazinone-2 ring at position 17 β of the sterane ring, we chose the basic cyclization of the α , γ -haloalkyl-*N*-arylurethanes in alkaline media. Scott et al. examined the ring-closure reactions of α , β - and α , γ -haloalkyl *N*-arylurethanes in alkaline media [13]. They found that the *N*-arylurethane group and its substituted derivatives reacted at different rates, depending on the inductive and conjugative effects of the substituents. The cyclization typically occurred with neighboring group participation, to yield *N*-phenyloxazolidone-2 or *N*-phenyltetrahydrooxazinone-2 derivatives, depending on the starting compounds [14]. In the solvolysis of vicinal halourethanes, oxazolidinone derivatives condensed to the estrane skeleton are formed, which are likewise potential pharmacons [15]. The cyclization of the four possible isomeric 16-*p*-toluene-sulfonyloxymethyl-17-unsubstituted phenylurethanes on the androstane and estrane skeletons has already been investigated [16,17].

To prepare the α , γ -diol system on the side chain of the sterane skeleton, we chose 3β -acetoxypregn-5-en-20-one (1) as starting compound. Elongation of the side-chain with a hydroxymethyl group on C-21 in 1 was accomplished by condensation with ethyl formate in the presence of NaOMe, followed by reduction with KBH₄ to the triols [10].

The simultaneous development of the new chiral centers at C-20 permits the formation of 21-hydroxymethylpregn-5-ene-3 β ,20 β -diol (3a) and its 20 α epimer (4a) in a ratio of 95:5. The separation of the two isomeric compounds in this form was not possible. The cyclic acetonides of the mixture of 21-hydroxymethyl-20-hydroxy compounds were acetylated in the 3β position, and the subsequent acidic hydrolysis of the cyclic ketal produced a chromatographically separable mixture of 3\beta-acetoxy-21-hydroxymethylpregn-5-en-20-ol isomers (3d and 4d). The separation of the two isomeric compounds in this form was possible. All ¹³C NMR chemical shifts of the epimeric pair 3d and 4d are the same within 0.5 ppm. Only the shifts of C-14, C-17, and C-18, carbons near to the chiral center (in position C-20) with different configurations, differ by 0.7, 0.6, and 0.8 ppm, respectively. Similarly, the ¹H NMR chemical shifts of the diastereomers are practically identical ($\Delta \delta \leq 0.02 \text{ ppm}$). The only exception is the 18-CH₃ group, which is similar close to C-20, the shifts of which differ by 0.09 ppm for the isomers.

For the further reactions, we used only the pure 20β isomer (**3d**). In the earlier literature it was supposed that the reduction of the C-20 ketone is stereoselective and that C-20 has the *R* configuration [10].

The selective *p*-tosyl ester formation from **3d** resulted in 5a, which, in a subsequent reaction with phenyl isocyanate, was converted to 3\beta-acetoxy-21-p-toluenesulfonyloxymethylpregn-5-ene-20\beta-phenylurethane (**6a**). For the preparation of substituted phenylurethane derivatives **6b-h**, the starting compound was 3B-acetoxy-21chloromethylpregn-5-en-20\beta-ol (5b), which was prepared by the Appel reaction of 3d [18]. The OH \rightarrow Cl change in 5b as compared with 3d is confirmed by the very large upfield shift (by 19.1 ppm) of the ¹³C NMR line of the neighboring carbon. 5b Reacted with phosgene in the presence of triethylamine to furnish the desired 3B-acetoxy-21-chloromethylpregn-5-ene-20\beta-chlorocarbonic acid ester. Without isolation, this compound was reacted with the appropriately substituted anilines to yield 6b-h. The presence of the O-tosyl group in 5a and 6a is demonstrated by the characteristic spectral data on this substituent (e.g.

the ν SO₂ IR bands, the methyl singlet with 3H intensity and the AA'BB' multiplet of the *p*-disubstituted aromatic ring in the ¹H NMR spectra and the ¹³C NMR lines of the methyl and aromatic carbons in the ¹³C NMR spectra) and also by the downfield shifts (by 0.29 and 0.4 ppm) of the 21-methylene signal in the ¹H NMR spectra. The urethane carbonyl line appears in the expected interval [19], between 152.3 and 154.1 ppm, for all compounds in the series **6–8**. The urethane NH in series **5** gives a broad signal of 1H intensity in all (**a–h**) cases (6.6–7.0 ppm).

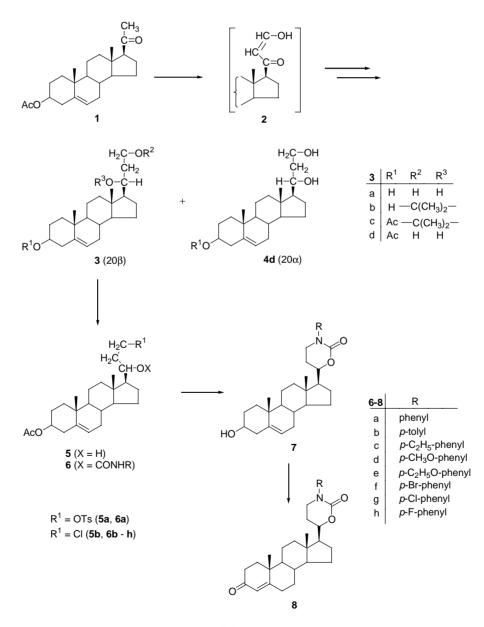
Compounds **6a–h** were subjected to methanolysis in the presence of four equivalents of NaOCH₃. Under these experimental conditions, *N*-phenyl-tetrahydrooxazinone-2 rings (**7a–h**) were formed in rapid reactions. The cyclization can be explained by the nucleophilic attack of the nitrogen atom

of the deprotonated acid amide. This cyclization process is typical neighboring group participation. In the notation proposed by Winstein and Boschan [14], the process can be characterized by the symbol $(N^- - 6)$.

The 3-acetoxy group in **3d**, **4d**, **5a**, **b**, and **6a–h** gives a high ν C=O frequency (1713–1740 cm⁻¹) in the IR spectrum, expected for saturated esters [20], a ¹H NMR methyl signal (2.03 ± 0.01 ppm) and a ¹³C NMR carbonyl line (170.8 ± 0.4 ppm).

Oppenauer oxidation of **7a**-**h** resulted in good yields of the 17β -[3-(*N*-aryl)tetrahydrooxazin-2-on-6-yl]androst-4-en-3-ones **8a**-**h**.

The change OAc \rightarrow OH causes a moderate upfield shift of the C-3 line, which appears at 74.2 \pm 0.2 ppm (series **6** and **3d**, **4d**, and **5**, **a**, **b**) or at 71.9 \pm 0.3 ppm (series **7**). This



Scheme 1.

Table 4 In vitro inhibition of 5α -reductase by steroidal tetrahydrooxazin-2-ones **(8a-g)**

Compounds	IC ₅₀ value (nM)	Relative inhibition IC ₅₀ finasteride IC ₅₀ compound
Finasteride	55	1
8a	270	0.20
8b	>1000	< 0.055
8c	260	0.21
8d	600	0.09
8e	245	0.22
8f	500	0.11
8g	420	0.13

line is dramatically shifted downfield, of course, for series **8**, where it appears at 199.5 \pm 0.5 ppm, values characteristic of ketones [21a]. The H-3 signal is also shifted downfield (due to the $-I_S$ effect of the *O*-acetyl group) by ca. 1.2 ppm, as expected [21b] for series **5** as compared with series **7** and **8**. The polarization of the enone moiety leads [21c] to a very great difference (>47 ppm) between the C-4 and C-5 shifts (δ C-4: 124.0 \pm 0.3 and δ C-5: 171.3 \pm 0.4) in series **8**.

The ¹H and ¹³C NMR signals of the 19-CH₃ group are sensitive to the change of the Δ^5 -double bond (series **6** and **7**) to the Δ^4 -en-3-one moiety in series **8**: δ H(19-CH₃) is 1.00 ± 0.03 for series **6** and **7**, but 1.19-1.20 for series **8**, while δ C(19-CH₃) is 20.0 ± 0.8 for series **6** and **7**, but 17.6 ± 0.2 for series **8**. The freely rotating aryl-urethane group in series **6** can lead to a shielding effect of ca. 0.1 ppm on the 18-CH₃ hydrogen relative to series **7** and **8** (where the ring structure forces the aryl group far from 18-CH₃) due to the anisotropic neighboring effect of the benzene ring [21d]. The spectral data characteristic of the substituents (Me, Et, OMe, OEt, halogens) on the aryl group are observed in series **6–8** (Tables 1–3; Scheme 1).

It is interesting to note that in the more rigid ring B of the steroidal skeleton in series 6 and 7 (caused by the Δ^5 -double bond) the C-9 line is shifted upfield by ca. 3.5 ppm as compared with series 8, probably because there is no steric hindrance in the latter series (field effect) [21e,22].

3.2. Effects of steroidal tetrahydrooxazin-2-ones on 5α -reductase activity

The inhibitory effects of compounds **8a–g** on human prostatic 5α -reductase were investigated with an in vitro incubation technique. Finasteride (17 β -(*N-tert*-butylcarba-moyl)-4-aza- 5α -androst-1-en-3-one), a potent 5α -reductase inhibitor already applied in medical practice, was used as a reference compound.

IC₅₀ values and relative inhibitory efficiencies as compared with the IC₅₀ of finasteride are presented in the Table 4. The investigated steroids (**8a–g**) exhibit some 5α reductase inhibitory effect. The IC₅₀ values of **8a–g** vary between 270 and 600 nM, with the exception of **8b**, which is a very weak inhibitor. The relative inhibitory effect of the unsubsituted *N*-phenyl compound **8a** is 0.20. As concerns the effects of substituents at position 4 of the phenyl ring in **8a**, the introduction of an ethyl (**8c**) or ethoxy (**8e**) group resulted in a weak enhancement of 5α -reductase inhibition. Substitution with halogens (**8f** and **8g**), methyl (**8b**) or methoxy (**8d**) caused a drop-off in inhibition ability.

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