excellent yield.¹⁹ Deprotection of DMPM groups with DDQ in the presence of MPM protections usually proceeded with excellent selectivity,^{1,7,10} but unfortunately 13 gave only unsatisfactory results $(3.0-4.3:1 \text{ selectivity}).^{20}$ The C-3 hydroxy compound 14, $[\alpha]_D^{13.5}$ -2.6° , was readily converted to the C-3 keto compound 18 by Swern oxidation. The final conversion of 18 into 1 proceeded efficiently without any detectable formation of kromycin; namely, when 18 was retreated with a large excess of DDQ at room temperature, rapid deprotection of the MPM group occurred within 5 min and then the Bn group was gradually removed to give pikronolide $(1)^{21}$ in high yield²² (Scheme II).

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Supplementary Material Available: $[\alpha]_D$, ¹H NMR, mass, and IR data for 1, 3, 5-9, 12-14, 18 (6 pages). Ordering information is given on any current masthead page.

(19) The ester 15, synthesized similarly via 5, was also subjected to the macrocyclization.¹⁸ The reaction required a rather long time (20 h) and the

14-membered ring enone (16) was isolated in moderate yield (66%). (20) When the O-acetate of 7 was treated with DDQ (1.2 equiv) in tolu-ene-H₂O (20:1) at -10 to -5 °C for 5.5 h, deprotection of the DMPM group

(21) Mp 140–141.5 °C (*n*-hexane–EtOAc), $[\alpha]_{D}^{18.5}$ +66° (*c* 0.187, MeOH) [lit.^{3b} mp 139 °C, $[\alpha]_{D}$ +70°(MeOH)].

(22) So far, attempts to obtain 1 by oxidation of 17 derived from 16 have been unsuccessful; i.e., Swern oxidation gave only the C-5 keto compound, which was also obtained very slowly by RuCl₂(PPh₃)₃ oxidation.²³ (23) Tomita, H.; Takai, K.; Oshima, K.; Nozaki, H. Tetrahedron Lett.

1981, 22, 1605.

Enantioselective Total Synthesis of (+)-Negamycin and (-)-Epinegamycin by an Asymmetric 1,3-Dipolar Cycloaddition

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Negamycin (1),¹ a structurally unique peptide-like natural product which exhibits striking activity against Gram-negative bacteria, including Pseudomonas aerginosa,² has attracted considerable synthetic interest^{3,4} since its structure elucidation in 1971.⁵ Herein we report an efficient chiral entry into (+)-negamycin in natural form (1) and the unnatural isomer (-)-3-epinegamycin (2). Our strategy for the synthesis of (+)-1 is outlined retrosynthetically in Scheme I. The key step envisioned would involve a highly enantioselective 1,3-dipolar cycloaddition of an appropriate chiral nitrone⁶ (4) to the allylamine. This cyclo-

(3) For syntheses of racemic negamycin, see: (a) Streicher, W.; Reinshagen, H.; Turnowsky, F. J. Antibiol. 1978, 31, 725. (b) Pierdet, A.; Nédélec, L.; Delaroff, V.; Allais, A. Tetrahedron 1980, 36, 1763. (c) Pasquet, G.; Boucherot, D.; Pilgrim, W. R. Tetrahedron Lett. 1980, 21, 931.
(4) For synthesis of (+)-negamycin in natural form, see: (a) Shibahara, S.; Kondo, S.; Maeda, K.; Umezawa, H.; Ohno, M. J. Am. Chem. Soc. 1972, 94, 4353. (b) Wang, Y.-F.; Izawa, T.; Kobayashi, S.; Ohno, M. Ibid. 1982, 104, 6465.

104, 6465.

(5) Kondo, S.; Shibahara, S.; Takahashi, S.; Maeda, K.; Umezawa, H.; Ohno, M. J. Am. Chem. Soc. 1971, 93, 6305.

(6) For cycloaddition with chiral nitrones, see: (a) Vasella, A. Helv. Chim. (6) For cycloaddition with chiral nitrones, see: (a) Vasella, A. Helv. Chim. Acta 1977, 60, 426, 1273. (b) Oppolzer, W.; Petrzilka, M. Ibid. 1978, 61, 2755. (c) Belzecki, C.; Panfil, I. J. Org. Chem. 1979, 44, 1212. (d) Workulich, P. M.; Uskoković, M. R. J. Am. Chem. Soc. 1981, 103, 3956. (e) Vasella, A.; Voeffray, R. J. Chem. Soc., Chem. Commun. 1981, 97. (f) Vasella, A.; Voeffray, R. Helv. Chim. Acta 1982, 65, 1134. (g) Vasella, A.; Voeffray, R. Ibid. 1983, 66, 1241. (h) Tice, C. M.; Ganem, B. J. Org. Chem. 1983, 48, 5048. (i) Kametani, T.; Nagahara, T.; Honda, T. Ibid. 1985, 50, 2327 2327.

Scheme I



^{*a*}(a) 1,1-Dimethoxycyclohexane, TsOH, benzene, reflux, 10 h; (b) DIBAL, toluene/THF (1:1), -78 °C, 1 h; (c) NH₂OH·HCl, py, room temperature, 2 h; (d) $(7 \rightarrow 10)$ methyl glyoxylate, 9, toluene, reflux, 14 h; (e) 10% HCl/MeOH (3:8), 90 °C, 4 h; (f) PhCH₂Br, K_2CO_3 , DMF, 50 °C, 1 h; (g) LiAlH₄, Et₂O, room temperature, 30 min.

addition would simultaneously create two new asymmetric centers adaptable to the 3R, 5R stereochemistry of (+)-1. Our first objective was to develop a suitable, chiral nitrone and to demonstrate acceptable diastereoselection during the cycloaddition. Toward this end, we chose the carbohydrate as the chiral template (Schemes II).

D-Gulono- γ -lactone (5) was first converted to 2,3:5,6-di-Ocyclohexylidene-D-gulo-furanose (6), $[\alpha]^{20}_D - 12.3^\circ$ (CHCl₃), by treatment with 1,1-dimethoxycyclohexane (benzene, TsOH) followed by DIBAL reduction in 88% yield from 5. Compound 6 was converted quantitatively to the oxime 7, $[\alpha]^{20}_{D} + 45.5^{\circ}$ $(CHCl_3)$. The nitrone 8, generated in situ by the reaction of 7 with methyl glyoxylate probably as a mixture of E and Z isomers, was allowed to react with the allylamine derivative 9 (toluene, reflux, 14 h) to produce an inseparable mixture of the 3R,5R-trans (10a) and 3S,5R-cis (10b) adducts in 84% yield. After removal of the D-gulosyl auxiliary group by acid hydrolysis, the product was subjected to N-benzylation (PhCH₂Br, K₂CO₃, DMF) followed by $LiAlH_4$ reduction to provide the chromatographically separable (silica gel, 50:1 CHCl₃/MeOH) trans alcohol 11a, mp 99–100 °C, $[\alpha]^{25}_{D}$ –16.7° (CHCl₃), and cis alcohol **11b**, $[\alpha]^{24}$ -29.0° (CHCl₃), in a ratio of 2:3 (55% overall yield from 10a +10b). Thus utilization of the D-gulosyl chiral template in this process resulted in a highly stereobiased synthesis of 11a and 11b

⁽¹⁾ Hamada, M.; Takeuchi, T.; Kondo, S.; Ikeda, Y.; Naganawa, H.;

Maeda, K.; Umezawa, H. J. Antibiot. 1970, 23, 170.
 (2) Korzybski, T.; Kowszyk-Gindifer, Z.; Kurytowicz, W. Antibiotics;
 American Society of Microbiology: Washington, DC, 1978; Vol. 1, pp 343-346.

⁽⁷⁾ Ness, R. K.; Diehl, H. W.; Flecher, Jr., H. G. J. Am. Chem. Soc. 1954, 76, 763.

Scheme III"



^a(a) TsCl, (i-Pr)₂NEt, Et₃N/CH₂Cl₂ (1:1), 0 °C \rightarrow room temperature, 10 h; (b) NaCN, (Me)₂SO, room temperature (2 h) \rightarrow 50 °C (10 h); (c) HCl/EtOH, 0 °C \rightarrow room temperature, 12 h; (d) 4% aqueous NaOH/MeOH (1:2), room temperature, 3 h; (e) ClCO₂ Et, Et₃N, toluene, 0 °C, 25 min, then benzyl (1-methylhydrazino)acetate, 0 °C (2 h) \rightarrow room temperature (10 h); (f) H₂, Pd/C, 10% aqueous AcOH/MeOH (1:2), 3 atm, 12 h.

in 93.7% ee and 94.2% ee (determined as the (+)-MTPA esters⁸), respectively.

Both trans (10a) and cis (10b) products obtained in this cycloaddition using the nonconjugated olefin⁹ as the dipolarophile must arise (applying Diels-Alder terminology) from the exo transition state;¹⁰ the E isomer of the nitrone 8 yields the trans adduct 10a, while the Z isomer yields the cis adduct 10b. The facial selectivity observed in this cycloaddition with the E and Z nitrones may be interpreted in terms of "O-endo" transition-state model^{6a,11} as shown in A, wherein, by analogy to recent reports,¹²⁻¹⁴



the electron-donating group (secondary alkyl) rather than the polar group (alkoxy) is perpendicular to the plane of the nitrogen-carbon double bond to permit the maximum orbital overlap of the participating centers, leading to the favored re face approach at the prochiral olefin. A similar approach to a prochiral diene has been observed in pericyclic cyclocondensation reactions of chiral sugars.15

(8) Dale, J. A.; Dull, D. L.; Mosher, H. S. J. Org. Chem. 1969, 34, 2543. (9) The nitrone cycloadditions with monosubstituted electron-rich dipolarophiles, incapable of secondary orbital interactions, proceed through exo transition states and are described as dipole LUMO controlled, resulting in 5-substituted isoxazolidines (cf.: Tufariello, J. J. In 1,3-Dipolar Cycloaddition Chemistry; Padwa, A., Ed.; Wiley-Interscience: New York, 1984; Vol. 2,

bonded interactions between the furan ring oxygen atom and the CHCO₂Me group

(12) DeShong, P.; Leginus, J. M. J. Am. Chem. Soc. 1983, 105, 1686.

(12) Deshong, F., Leginds, J. M. J. Am. Chem. Soc. 1983, 105, 1650.
 (13) Jäger, V.; Schohe, R.; Paulus, E. F. Tetrahedron Lett. 1983, 24, 5501.
 (14) Houk, K. N.; Susan, R. M.; Wu, Y.-D.; Rondan, N. G.; Jäger, V.;
 Schohe, R.; Fronczek, F. R. J. Am. Chem. Soc. 1984, 106, 3880.
 (15) Danishefsky, S. J.; Marring, C. J.; Barbachy, M. R.; Segmuller, B.

E. J. Org. Chem. 1984, 49, 4565.

Compound 11a was converted to (+)-negamycin in six steps (Scheme III). Tosylation of 11a followed by substitution (NaCN, Me₂SO) gave the nitrile **12a**, $[\alpha]^{17}_{D}$ +31.4° (CHCl₃), in 72% overall yield. Compound 12a was converted to the carboxylic acid 13a, $[\alpha]^{14}_{D}$ +31.7° (CHCl₃), in 79% yield via ethanolysis and subsequent saponification. Condensation of 13a with benzyl (1-methylhydrazino)acetate was carried out using the mixed carboxylic acid anhydride method (ClCO₂Et, Et₃N)¹⁶ affording the hydrazide 14a, $[\alpha]^{16}_{D}$ +20.4° (CHCl₃), in 67% yield. Hydrogenolysis resulted in combined debenzylation and N-O bond cleavage; purification of the crude product by silica gel chromatography¹⁷ gave (+)-negamycin (1), mp 108-115 °C dec (lit.⁵ mp 110–120 °C dec), $[\alpha]^{20}_{D}$ +2.3° (c 4.07, H₂O) (lit.⁵ $[\alpha]^{29}_{D}$ +2.5° (c 2, H_2O), in 75% yield. This material was found to be identical with natural negamycin (TLC, ¹H NMR, and antibacterial activity18).

We then completed the synthesis of optically active 3-epinegamycin (2) by transformation of the 3S, 5R-cis isomer 11b (Scheme III). Compound 11b was converted in four steps to the carboxylic acid 13b, $[\alpha]^{20}_{D}$ +26.0° (CHCl₃), which was then worked up in a manner similar to that described for 13a, giving rise to the hydrazide 14b, $[\alpha]^{20}_{D}$ +17.4° (CHCl₃), in 34.4% overall yield from 11b. Hydrogenolysis of 14b followed by silica gel chromatography¹⁷ afforded (-)-3-epinegamycin (2) in 65% yield, $[\alpha]^{20}_{D}$ -3.2° (c 4.42, H₂O), mp 165-195 °C dec (for (±)-2 lit.^{3c} mp 150-180 °C dec), which had an identical ¹H NMR spectrum in a D_2O solution with that of (\pm) -2. Antibacterial activity for synthetic (-)-2 is under investigation.

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(18) Kono, M.; O'hara, K.; Ohmiya, K.; Iida, H.; Kibayashi, C.; Kasahara, K. Jpn. J. Antibiot. 1986, 39, 247.

On the Characterization of Intermediates in the Mitomycin Activation Cascade: A Practical Synthesis of an Aziridinomitosene

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Mitomycin C (mutamycin) is already a significant resource in cancer chemotherapy.^{1,2} Potential second generation mitomycins are in various stages of preclinical development. It has long been recognized that mitomycins (1) are not per se biologically potent but require reductive priming.³ One mode of action of suitably primed mitomycins involves the alkylation and cross-linking of DNA.⁴ Furthermore, the reductive process seems to generate

Chapter 9). (10) Endo transition states would greately be restricted by suffering from unfavorble steric interactions between the CH₂NHCbz group in the incoming dipolarophile 9 and the furan ring oxygen atom of the nitrone 8. (11) "O-Exo" transition states should be disfavored due to serious non-

⁽¹⁶⁾ Vaughan, J. R., Jr.; Osato, R. L. J. Am. Chem. Soc. 1952, 74, 676. (17) Elution was initiated with CHCl₃/MeOH (4:1), containing 0.5% of concentrated NH4OH, continued with gradient solvent system and finally conducted with MeOH including 1% concentrated NH4OH.

⁽¹⁾ Carter, S. K.; Crooke, S. T. Mitomycin C: Current Status and New Developments; Academic Press: New York, 1979.

^{(2) (}a) Cassady, J. M.; Duorose, J. D. Anticancer Agents Based On Natural Product Models; Academic Press: New York, 1980. (b) Remers,

<sup>Natural Product Models; Academic Press: New York, 1980. (b) Remers,
W. A. Antineoplastic Agents; Wiley: New York, 1980. (c) (3) (a) Iyer, V. N.; Szybalski, W. Proc. Natl. Acad. Sci. U.S.A. 1963, 50, 355. (b) Iyer, V. N.; Szybalski, W. Science (Washington, D.C.) 1964, 145, 55. (c) Nagata, C.; Matsuyama, A. Prog. Antimicrob. Anticancer Chemother. Proc. Int. Congr. Chemother., 6th, 1969 1970, 2, 423.</sup>