

Bioorganic & Medicinal Chemistry Letters 9 (1999) 1599-1600

BIOORGANIC & MEDICINAL CHEMISTRY LETTERS

## SYNTHESIS OF 25-AMINOSTEROLS, NEW ANTIFUNGAL AGENTS

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Received 15 March 1999; accepted 28 April 1999

Abstract : 25-aminolanostenol 1 and 25-aminocholesterol 2 were hemisynthesized from natural sterols and tested *in vitro* against *Candida albicans*. The biological activity of compound 1 was rather weak, whereas 2 exhibited *in vitro* antifungal activity with MIC value of 4  $\mu$ M. © 1999 Elsevier Science Ltd. All rights reserved.

Introduction : Since a few decades, there has been an increasing demand for new fungicidal agents, due especially to resistances to current azole antifungals, enhanced by immunodeficiencies, metabolic derangements, or suppression of competitor organisms.<sup>1</sup> Antifungal chemotherapy mainly implies systemic treatment with inhibitors of ergosterol, the dominant sterol in yeasts and fungi.<sup>2</sup> Azole derivatives inhibit the P450-dependent lanosterol 14 $\alpha$ -demethylase and cause accumulation of 14-methylated sterols; as a result, the lack of ergosterol modifies the membrane's fluidity, creates vesicles, deformation of buds, and abnormal thickening which leads to destruction by the phagocytes<sup>3</sup>. The  $\Delta^7$ -5-desaturase is another target of inhibition, widely studied<sup>4</sup> because of its specificity in the ergosterol biosynthesis pathway. We have already reported<sup>5</sup> potent activities of hemisynthesized aminosterol derivatives as potential transition state's mimics of desaturation.

In ergosterol biosynthesis, another specific step different from mammalian cholesterol synthesis, is the side chain C24-methylation by the sterol methyl transferase (24-SMT).<sup>2</sup> Thus, 24,25-epiminolanostenol was reported to inhibit the growth of *Gibberella fujikuroi*,<sup>6</sup> whereas azasterols (with nitrogen at C23, C24 or C25)<sup>7</sup> inhibited *Saccharomyces cerevisiae*. Few others lanosterol or cholesterol derivatives with nitrogen functionalities at C24 displayed fungistatic properties.<sup>8</sup> On the other hand, cytoxicity was noticed for 24,25-iminocholesterol.<sup>9</sup> In the scope of our studies on primary amine sterol derivatives, we performed the hemisynthesis of 25-aminolanostenol 1 and 25-aminocholesterol **2** (figure 1) as new potential 24-SMT inhibitors.

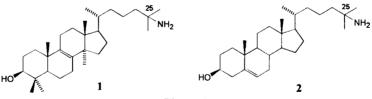
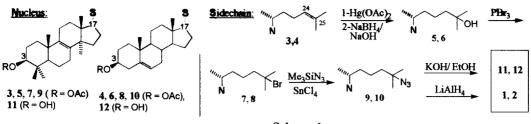


Figure 1

**Chemistry**: The synthetic route to 1 and 2 is outlined in scheme 1. Lanosterol and desmesterol of natural origin were acetylated at C3 by the usual method followed by acetoxymercuration/demercuration<sup>10</sup> to give respectively 25-hydroxylanostenyl acetate 5 and 25-hydroxycholesteryl acetate 6 in 90% yields. The tertiary alcohols were then treated with phosphorus tribromide in chloroform and led to the corresponding bromosteryl acetate derivatives (7,8) in 89-90% yield. Treatment with trimethylsilyl azide in excess and a catalytic amount of SnCl<sub>4</sub> in toluene<sup>11</sup> gave in 4 days 69 to 77 % of 25-azidosteryl acetate 9 and 10. Saponification of 9 and 10 finally produced the 25-azidosterols 11 and 12 in quantity, or by reduction with LiAlH<sub>4</sub> in dry diethylether and subsequent acetate cleavage, afforded the title compounds<sup>12</sup> 1 and 2 in good yields. All compounds were fully described by IR, <sup>1</sup>H (400 MHz), <sup>13</sup>C (100 MHz) NMR, or MS spectroscopy experiments.

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## Scheme 1

**Biological results and conclusion :** Results of the antifungal *in vitro* screening<sup>13</sup> are summarized in table 1. No appreciable activity was found for 11 and 12; conversely, 1 and 2 inhibited *Candida albicans*. Surprisingly, 25-aminocholesterol (MIC value = 4  $\mu$ M) was found to be 15 times more potent than 25-aminolanostenol (MIC value = 60  $\mu$ M). In addition, 2 at 4  $\mu$ M was still inhibitory on *C. albicans* after 48h incubation. Bioassays on three bacterial strains (*E. hirae*, *S. aureus* and *E. coli*) displayed no activity on these microorganisms. These results are in perfect concordance with the reported fungicidal specific activity of 24-aminocholesterol and 24-aminolanostenol on *Candida sp.*<sup>8</sup> Antifungal activity is clearly in relation with the primary amine function and with the sterol tetracyclic structure. We postulate that the aminosterol 2 give rise to an ammonium form species that mimics the transition state (positive charge) of the methylation. In conclusion, we consider that *in vivo* bioevaluation should be performed in order to validate further development of 2.

Table 1 : Growth inhibition\* of C. albicans induced by 25-nitrogen substituted sterols

Fungus strain :	1	2	11	12
Candida albicans (IP 1180.79)	60	4	> 250	> 250
			. 14	1. 11. 14

\* MIC (inhibitory concentration in  $\mu$ M). C. albicans was cultured in liquid Sabouraud medium at 30°C containing 2% v/v of an appropriate solvent. Innoculum size : 2% v/v.

Acknowledgments : We acknowledge the Region Poitou Charentes Council for financial support.

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- 12. **25-Aminocholest-5-en-3**β-ol (2): M.p=177 °C. IR (v, cm<sup>-1</sup>): 3500-3300 (OH); 3200-3250 (NH<sub>2</sub>). <sup>1</sup>H NMR (δ, ppm): 0.68 (s, 3H, Me-18); 0.93 (d, 3H, Me-21, J=6.4 Hz); 1.01 (s, 3H, Me-19); 1.09 (s, 6H, Me-26 and Me-27); 3.48-3.56 (m, 1H, H-3α); 5.35 (d, 1H, H-6, J=6 Hz). <sup>13</sup>C NMR (δ, ppm): 11.8 (C-18); 18.7 (C-21); 19.4 (C-19); 20.9 and 21.1 (C-23 and C-11); 49.5 (C-25); 71.7 (C-3); 121.7 (C-6); 140.7 (C-5). MS, m/z (%, disconnection): 401[M<sup>-1</sup>]; 384 (54; M-NH<sub>3</sub>); 369 (27; M-NH<sub>3</sub>-CH<sub>3</sub>); 366 (13; M-NH<sub>3</sub>-H<sub>2</sub>O); 351 (23; M-NH<sub>3</sub>-CH<sub>3</sub>-H<sub>2</sub>O); 299 (35; M-NH<sub>3</sub>-C<sub>6</sub>H<sub>11</sub>-2H); 271 (100; M-NH<sub>3</sub>-sidechain); 253 (27; 271- H<sub>2</sub>O); 213 (38; M-NH<sub>3</sub>-H<sub>2</sub>O-D cycle). **25-Aminolanost-8-en-3**β-ol (1): IR (v, cm<sup>-1</sup>): 3500-3100 (OH); 3200-3250 (NH<sub>2</sub>). <sup>1</sup>H NMR (δ, ppm): 0.66 (s, 3H, Me-18); 0.71 and 0.81 (2s, 6H, Me-4β et Me-14α); 0.89-1.00 (m, 9H, Me-4α, Me-21, Me-19); 1.16 (s, 6H Me-26 and Me-27); 3.18-3.39 (m, 1H, H-3α).
- 13. The fungal growth was measured in vitro using a liquid-phase turbimetric system (Bioscreen®, Labsystem) and automatically evaluated every 30 minutes for 16 hours using various concentrations of drugs. Dei-Cas, E.; Dujardin, L.; Ribeiro Pinto, M.E.; Fruit, J.; Poulain, D.; Camus, D.; Vernes, A. Mycoses, 1991, 34, 167.