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Carbonucleotoids and Carbopeptoids: New Carbohydrate Oligomers

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Abstract: Carbonucleotoids (phosphate-bond linked carbohydrates) and carbopeptoids (peptidebond linked carbohydrates) are proposed as new oligosaccharide ligands. The ready access to these analogues is demonstrated by the facile synthesis of the tetrameric carbonucleotoid **1**.

Oligonucleotides, peptides and oligosaccharides are three classes of oligomers whose chemical synthesis has been extensively studied.^{1,2,3} Such molecules can now be routinely synthesized both in solution and in the solid phase, manually or on automated systems with the exception of oligosaccharides whose construction is considerably less efficient. This state of affairs reflects the availability of techniques for the construction of the phosphate and peptide bonds in nearly quantitative yields and the absence of stereocenters associated with these linkages. In contrast, the relatively lower yields and the complication of two isomeric glycosidation products in glycoside bond forming reactions renders the synthesis of conventional oligosaccharides rather problematic.

The rapidly growing importance of carbohydrates in biology⁴ and the emergence of chemical libraries⁵ as potential tools for ligand lead and drug discovery prompted us to design two groups of carbohydrate-based oligomers I and II (Figure 1), using the nucleotide and peptide bond, respectively, to link component units.



Several advantages were envisioned for these molecules termed "carbonucleotoids" (I) and

"carbopeptoids" (II), respectively, including: (a) ease of formation; (b) possibility of library generation; (c) choice of solution or solid phase synthesis; and (d) polyfunctionality. In this communication, we demonstrate the validity of these concepts by the synthesis of the tertrameric carbonucleotoid 1 from readily available starting materials.

Scheme 1 summarizes the synthesis of the tetrameric compound 1 starting with glucose pentaacetate (2). Thus, manipulation of 2 by a known sequence afforded methylester 3 in large quantities.⁶ The latter compound was then selectively silylated at the primary position, benzylated at the three secondary positions,⁷ and reduced under standard conditions to afford key intermediate 4 in 51% overall yield.⁸ Compound 4 served as a common intermediate to prepare the two requisite carbohydrate units 6 and 7.⁹ The naphthoyl group on 7 was chosen to serve as a convenient UV-detectable marker for chromatographic analysis of subsequent intermediates and to mimic potential solid phase chemistry. The conversion of 4 to the targeted *N*,*N*-diisopropyl-aminophosphoramidite (6) was accomplished using 2-cyanoethyl-*N*,*N*-diisopropylchlorophosphoramidite 5¹⁰ in the presence of diisopropylethylamine (DIPEA) in methylene chloride under strictly anhydrous conditions. Purification of the crude product, formed in a few minutes in essentially quantitative yield, could be carried out by quick filtration through a short silica gel column using petroleum ether: ethylacetate: triethylamine (75: 20: 5) as eluent.

The hydroxy component **7** was prepared in 90% overall yield from **4** by esterification with 2naphthoyl chloride in the presence of dimethylaminopyridine (DMAP), followed by acid-induced desilylation. Coupling of the two intermediates **6** and **7** proceeded smoothly under anhydrous conditions in methylene chloride in the presence of 1*H*-tetrazole. Upon completion of the reaction as indicated by TLC, addition of *m*-CPBA to the reaction mixture resulted in oxidation of phosphorus as judged by a lower Rf value. Standard work-up with aqueous NaHCO₃ solution, followed by flash chromatography afforded pure dimer **8** in 97% yield.⁹ Removal of the silyl group from **8** under acid conditions gave the hydroxy compound **9** in 95% yield which was now ready for further extension via coupling with another molecule of **6**.⁹

Repetition of the sequence allowed the preparation of the trimeric compounds 10 and 11 and the tetrameric compounds 12 and 13 in similarly high yields (Scheme 1). Removal of the cyanoethylene group under mildly basic conditions (NH₄OH), followed by hydrogenolysis (Pd/C-H₂) of the benzyl ethers led to the naphthoate ester 1 in 80% overall yield.

It is envisioned that the described chemistry could be used to produce higher homologs of a variety of carbonucleotoid structures (I) similar to 1, and that extensions in the corresponding carbopeptoid area would generate structures of type II. Mixing different component units in these reactions may, of course, result in a variety of chemical libraries of predesigned molecular structures. The latter are expected to find applications in ligand-receptor binding studies and biological screening assays.

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^aReagents and conditions: (a)-(d) ref. 6, 41% overall; (e) 1.2 equiv of ¹BuMe₂SiCl, 2.5 equiv of imid., DMF, 2 h, 0 °C, 93%; (f) 9 equiv of BnBr, 6 equiv of Ag₂O, DMF, 20 h, 25 °C, 83%; (g) 3 equiv of DIBALH, THF, 1 h, 0 °C, 66%; (h) i. 3 equiv of naphthoyl chloride, 10 equiv of pyr., 45 min., 25 °C; ii. AcOH/THF/H₂O (3:1:1) 15 h, 25 °C, 90% overall; (i) 1.5 equiv of 5, 4 equiv of ¹Pr₂EtN, CH₂Cl₂, 5 min, 25 °C, 95%; (j) 3-5 equiv of 6 (0.1M solution in CH₂Cl₂), 10 equiv of 1 *H*-tetrazole, CH₂Cl₂, 20 min., 25 °C, then 4.5-7.5 equiv of *m*-CPBA, 5 min, 97%; (k) AcOH/THF/H₂O (3:1:1), 18 h, 25 °C, 95%; (l) 1:1 conc. aq. NH₄OH / CH₃CN, 2 h, 50 °C, 88%; (m) H₂, Pd/C, EtOH/THF/AcOH (2:1:1),25 °C, 72 h, 78%.

References and Notes

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- 8. All compounds exhibited satisfactory analytical and spectroscopic data. Yields refer to homogeneous materials.
- Phosphoramidate 6 (2 diastereomers): IR, (neat) cm⁻¹: 3089, 2964, 2927, 2856, 2253, 1497, 1455, 1396, 1363, 1253, 1184, 1156, 1094, 1028, 978, 876, 836, 779, 735; ¹H-NMR (400 MHz, C₆D₆): δ 7.34 (m, 5 H, Ph), 7.14 (m, 10 H, Ph), 4.97 (m, 4 H, CH₂Ph), 4.78 (m, 2 H, CH₂Ph), 4.07-3.24 (m, 13 H, OCH, OCH₂, CH₂CN), 1.81 (m, 2 H, CH(CH₃)₂), 1.16 (m, 12 H, CH₃CH), 1.03, 1.02 (2 s, 9 H, ^tBuSi), 0.20, 0.18, 0.16, 0.15 (4 s, 6 H, Me₂Si); HRMS: C₄₃H₆₃O₇N₂PSi, Calc. (M+Cs⁺): 911.3197; found: 911.3185.

Naphthoylester 7: IR, (neat) cm⁻¹: 3494, 3062, 2919, 1716, 1630, 1600, 1454, 1355, 1284, 1228, 1197, 1091,779, 736; ¹H-NMR (250 MHz, CDCl₃): δ 8.58 (s, 1 H, Ar), 8.00 (m, 2 H, Ar), 7.89 (m, 2 H, Ar), 7.59 (m, 2 H, Ar), 7.32 (m, 15 H, Ph), 4.95 (m, 3 H), 4.90 (d, *J*=4.5 Hz, 1 H), 4.69 (m, 3 H), 4.52 (dd, *J* = 3.9, 12.0 Hz, 1 H), 3.91 (dd, *J* = 2.6, 12.0, 1 H), 3.83 (d, *J*=8.3, 1 H), 3.70 (m, 4 H), 3.96 (m, 1 H), 2.25 (s, 1 H, OH). HRMS: C₃₉H₃₈O₇ Calc. (M+Cs⁺): 751.1672; found: 751.1668.

Dimer 9: IR, (neat) cm⁻¹: 3397, 3030, 2923, 2254, 1718, 1653, 1629, 1497, 1453, 1355, 1284, 1227, 1197, 1094, 1029, 780.¹H-NMR (400 MHz, C_6D_6): δ 8.82 (s, 1 H, Ar), 8.26 (d, 1 H, Ar), 7.72 (m, 1 H, Ar), 7.61 (m, 1 H, Ar), 7.48 (m, 1 H, Ar), 7.37-6.95 (m, 32 H, Ar, Ph), 4.89-4.18 (m, 21 H, CH₂Ph, CH₂-Ar, -CH₂CH₂CN, CHCH₂-Ar and CH₂OH), 3.95-3.45 (m, 13 H, CH- and CH₂-sugar), 1.71 (s, 1 H, OH); HRMS: C₁₇₀H₇₂O₅NP calc. (M+H⁺): 1198.4718; found: 1198.4715.

Tetramer 13: IR, (neat) cm⁻¹: 3420, 3064, 2924, 2255, 1721, 1497, 1455, 1357, 1278, 1028, 737. ¹H-NMR (400 MHz, CDCl₃): δ 8.41 (s, 1 H, Ar), 8.00 (m, 2 H, Ar), 7.91 (m, 2 H, Ar), 7.55 (m, 2 H, Ar), 7.30 (m, 60 H, Ph), 4.93-4.05 (m, 39 H, CH₂Ph, CH₂-Ar, CH₂CH₂CN and CH₂OH), 3.88-3.27 (m, 23 H, CH- and CH₂-sugar), 2.58 (s, 1 H, OH). HRMS: C₁₃₂H₁₄₀-O₃₁N₃P₃ Calc. (M+Cs⁺): 2488.7738; found: 2488.7758.

Tetramer 1: IR, (neat) cm⁻¹: 3376, 2934, 1450, 1244, 1110, 1088. ¹H-NMR (400 MHz, D₂O): δ 8.41 (s, 1 H, Ar), 8.00 (m, 2 H, Ar), 7.91 (m, 2 H, Ar), 7.55 (m, 2 H, Ar), 4.93-4.05 (m, 4 H, CH₂-Ar and CH₂OH), 3.88-3.27 (m, 32 H, CH- and CH₂-sugar); HRMS: C₃₉H₅₉O₃₁P₃ Calc. (M+H⁺):1117.2331; found: 1117.2350.

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