## CHEMOENZYMATIC SYNTHESIS OF (-)-CARBOCYCLIC 7-DEAZAOXETANOCIN G

Xing Chen, Suhaib M. Siddiqi and Stewart W. Schneller\* Department of Chemistry, University of South Florida Tampa, Florida 33620-5250

**Summary:** The enantioselective synthesis of (-)-(1R,2S,3S)-2-amino-7-[2,3-bis(hydroxymethyl)-cyclobuty]]pyrrolo[2,3-d]pyrimidin-4(3H)-one (5) as the 7-deaza analogue of carbocyclic oxetanocin G is described in 13 steps from*trans*-3,3-diethoxy-1,2-bis(hydroxymethyl)cyclobutane (8) in an overall yield of 6%. Included in this route is the use of*Pseudomonas cepacia*lipase for enzymatic resolution.

The naturally occurring oxetanocin A  $(1)^1$  and synthetic oxetanocin G  $(2)^2$  represent a novel class of nucleosides possessing antiviral properties.<sup>3</sup> Modification of the unique oxetanosyl-N-glycoside structural feature of 1 and 2 into the carbocyclic nucleoside framework<sup>4</sup> led to the synthesis of 3 and 4 as racemates<sup>5</sup> and individual enantiomers,<sup>6</sup> which have shown activity against herpesviruses and HIV.<sup>5b,c,6,7</sup> One particular interest to this laboratory is the development of 7-deazapurine carbocyclic nucleosides that can be effective against human cytomegalovirus (HCMV). In view of the potent and selective anti-HCMV properties of the C-1' *R*-stereoisomer of 4,<sup>6b</sup> we desired a synthesis of the corresponding *R*-stereoisomer 5.



Related 7-deazapurine carbocyclic nucleoside syntheses in our laboratory indicated that the most efficacious route to 5 would be via reaction of the protected chiral amine 6 with the dimethylacetal of 2-(2-amino-4,6-dichloropyrimidin-5-yl)acetaldehyde  $7^8$  followed by ring closure, hydrolysis and deprotection. A review of the literature revealed two enantioselective routes to cyclobutyl derivatives used in the synthesis of chiral carbocyclic oxeanocins<sup>6</sup> that seemed suitable for preparing a precursor of 6. In one case, <sup>6a</sup> however, the initial step involved a [2+2]-cycloaddition reaction of not easily obtainable reagents in the presence of a chiral titanium compound as catalyst. In the other case, <sup>6b</sup> resolution of a chiral mixture of cycloadducts was done through diastereomeric amides that added steps to the synthesis. In view of the ease with which enzymes<sup>9</sup> can produce re-

solution of a racemic mixture, we chose to use *Pseudomonas cepacia* lipase<sup>10</sup> on the  $(\pm)$ -acetate 13 as a more efficient means to 6.

Scheme



*Reaction conditions: a, (i)* NaH in DMF; (ii) BnBr; *b,* 0.5% H<sub>2</sub>SO<sub>4</sub> in MeCN; *c,* LS-Selectride in THF, -78 °C; *d,* Ac<sub>2</sub>O/ pyridine; *e, Pseudomonas cepacia* lipase in phosphate buffer; *f,* KOH in MeOH; *g,* TsCl/pyrldine; *h,* NaN<sub>3</sub> in DMF, 100 °C; *i,* BH<sub>3</sub>•THF in THF; *j,* 7 and Et<sub>3</sub>N in 1-BuOH, 100 °C; *k,* 2 N HCl; *l,* 5% aq. NaOH in MeOH; *m,* EtSH and BF<sub>3</sub>•Et<sub>2</sub>O

Thus, benzylation of *trans*-3,3-diethoxy-1,2-bis(hydroxymethyl)cyclobutane (8)<sup>5a,b</sup> to *trans*-3,3diethoxy-1,2-bis[(benzyloxy)methyl]cyclobutane (9,<sup>11</sup> 100%, oil) (Scheme) was followed by hydrolysis to *trans*-2,3-bis[(benzyloxy)methyl]cyclobutanone (10,<sup>11,12</sup> 100%, oil). Reduction of 10 with LS-Selectride<sup>13</sup> to ( $\pm$ )-(1 $\alpha$ ,2 $\alpha$ ,3 $\beta$ )-2,3-bis[(benzyloxy)methyl]cyclobutanol (11,<sup>14</sup> 81%, oil) occurred with no indication (by <sup>1</sup>H and <sup>13</sup>C nmr) of the diastereomeric alcohol 12 having been formed. Acetylation of 11 gave ( $\pm$ )-(1 $\alpha$ ,2 $\alpha$ ,3 $\beta$ )-2,3bis[(benzyloxy)methyl]-1-cyclobutyl acetate (13,<sup>14</sup> 92%, oil), which, when subjected to treatment with *Pseudomonas cepacia* lipase yielded (+)-(1*S*,2*S*,3*S*)-bis[(benzyloxy)methyl]-1-cyclobutyl acetate ((+)-13,<sup>11</sup> oil, 48%, >99% ee,<sup>15</sup>, {[ $\alpha$ ]<sub>D</sub><sup>25</sup> +14.57° (*c* 0.508, CH<sub>2</sub>Cl<sub>2</sub>)} and (-)-(1*R*,2*R*,3*R*)-2,3-bis[(benzyloxy)methyl]-1-cyclobutanol ((-)-11,<sup>11</sup> oil, 50%, >99% ee,<sup>15</sup> {[ $\alpha$ ]<sub>D</sub><sup>25</sup> -27.06° (*c* 0.436, CHCl<sub>3</sub>)}). The structural assignments for (+)-13 and (-)11 were accomplished by conversion into the enantiomeric alcohols 14 and 15 and comparing the optical rotation data for each product with the reported<sup>16</sup> values for these latter compounds.



Saponification of (+)-13 to (+)-(15,25,35)-2,3-bis[(benzyloxy)methyl]cyclobutanol ((+)-11,<sup>11</sup> 100%, oil, {[ $\alpha$ ]p<sup>25</sup> +27.14° (*c* 0.7, CHCl<sub>3</sub>)}) was followed by tosylation to (+)-(15,25,35)-2,3-bis[(benzyloxy)methyl]-1-cyclobutyl tosylate (16,<sup>14</sup> 90%, oil). Treatment of 16 with sodium azide (to 17) followed by reduction provided (+)-(1*R*,25,35)-2,3-bis[(benzyloxy)methyl]cyclobutylamine (6,<sup>11</sup> 54% from 16, oil), which, in turn, upon reaction with 7 gave the desired intermediate 18. Ring closure of 18 with dilute acid produced (+)-(1*R*,25,35)-2-amino-7-{2,3-bis[(benzyloxy)methyl]cyclobutyl}-4-chloropyrrolo[2,3-*d*]-pyrimidine (19,<sup>14</sup> 63% from 6, oil) that underwent subsequent hydrolysis to the protected 7-deazaguanine derivative (+)-(1*R*,25,35)-2-amino-7-{2,3-bis[(benzyloxy)methyl]cyclobutyl}pyrrolo[2,3-*d*]pyrimidin-4(3*H*)-one (20,<sup>14</sup> 78%, oil). Attempted hydrogenolytic debenzylation of 20 produced the 5,6-dihydro derivative of 5. On the other hand, use of boron trifluoride/ethylthiol on 20 led to the target compound (-)-(1*R*,25,35)-2-amino-7-[2,3-bis(hydroxymethyl)cyclobutyl]pyrrolo[2,3-*d*]pyrimidin-4(3*H*)-one (5,<sup>14</sup> 64%, mp 250-252 °C from MeOH, {[ $\alpha$ ]<sub>D</sub><sup>25</sup> -12.29° (*c* 0.358, DMSO)}). It should be noted that attempts to react 16 with 2-amino-4-chloropyrrolo[2,3-*d*]pyrimidine<sup>8</sup> resulted in recovery of starting material.

The biological properties of 5 and other uses of (+)-13 for preparing various carbocyclic oxetanocin derivatives will be reported in the future.

Acknowledgments. This research was supported by funds from the Department of Health and Human Services (NO1-AI-72645) and this is appreciated. We are grateful to Dr. K. Tanaka of the Institute for Chemical Research, Kyoto University, Kyoto, Japan for his assistance in determining the enantiomeric excess of (-)-11 and (+)-13.

## **References and Notes**

- (a) Shimada, N.; Hasegawa, S.; Harada, T.; Tomisawa, T.; Fujii, A.; Takita, T. J. Antibiot. 1986, 39, 1623-1625.
  (b) Nakamura, H.; Hasegawa, S.; Shimada, N.; Fujii, A.; Takita, T.; Iitaka, Y. J. Antibiot. 1986, 39, 1626-1629.
- 2. Shimada, N.; Hasegawa, S.; Saito, S.; Nishikiori, T.; Fujii, A.; Takita, T. J. Antibiot. 1987, 40, 1788-1790.
- (a) Hoshino, H.; Shimizu, N.; Shimada, N.; Takita, T.; Takeuchi, T. J. Antibiot. 1987, 40, 1077-1078.
  (b) Nishiyama, Y.; Yamamoto, N.; Takahashi, K.; Shimada, N. Antimicrob. Agents Chemother. 1988.

32, 1053-1056. (c) Nishiyama, Y.; Yamamoto, N.; Yamada, Y.; Fujioka, H.; Shimada, N.; Takahashi, K. J. Antibiot. 1989, 42, 1308-1311.

- 4. Marquez, V.E.; Lim, M.-I. Med. Res. Rev. 1986, 6, 1-40.
- (a) Honjo, M.; Maruyama, T.; Sato, Y.; Horii, T. Chem. Pharm. Bull. 1989, 37, 1413-1415.
  (b) Slusarchyk, W.A.; Young, M.G.; Bisacchi, G.S.; Hockstein, D.R.; Zahler, R. Tetrahedron Lett. 1989, 30, 6453-6456.
  (c) Norbeck, D.W.; Kern. E.; Hayashi, S.; Rosenbrook, W.; Sham, H.; Herrin, T.; Plattner, J.J.; Erickson, J.; Clement, J.; Swanson, R.; Shipkowitz, N.; Hardy, D.; Marsh, K.; Arnett, G.; Shannon, W.; Broder, S.; Mitsuya, H. J. Med. Chem. 1990, 33, 1281-1285.
- (a) Ichikawa, Y.-i.; Narita, A.; Shiozawa, A.; Hayashi, Y.; Narasaka, K. J. Chem. Soc., Chem. Commun. 1989, 1919-1921. (b) Bisacchi, G.S.; Braitman, A.; Cianci, C.W.; Clark, J.M.; Field, A.K.; Hagen, M.E.; Hockstein, D.R.; Malley, M.F.; Mitt, T.; Slusarchyk, W.A.; Sundeen, J.E.; Terry, B.J.; Tuomari, A.V.; Weaver, E.R.; Young, M.G.; Zahler, R. J. Med. Chem. 1991, 34, 1415-1421.
- (a) Hayashi, S.; Norbeck, D.W.; Rosenbrook, W.; Fine, R.L.; Matsukura, M.; Plattner, J.J.; Broder, S.; Mitsuya, H. Antimicrob. Agents Chemother. 1990, 34, 287-294. (b) Field, A.K.; Tuomari, A.V.; McGeever-Rubin, B.; Terry, B.J.; Mazina, K.E.; Haffey, M.L.; Hagen, M.E.; Clark, J.M.; Braitman, A.; Slusarchyk, W.A.; Young, M.G.; Zahler, R. Antiviral Res. 1990, 13, 41-52.
- 8. Siddiqi, S.M.; Schneller, S.W. unpublished results.
- 9. Davies, H.G.; Green, R.H.; Kelly, D.R.; Roberts, S.M. Biotransformations in Preparative Organic Chemistry. The Use of Isolated Enzymes and Whole Cell Systems in Synthesis; Academic Press: San Diego, CA, 1989.
- (a) Hughes, D.L.; Bergan, J.J.; Amato, J.S.; Bhupathy, M.; Leazer, J.L.; McNamara, J.M.; Sidler, D.R.; Reider, P.J. Grabowski, E.J.J. J. Org. Chem. 1990, 55, 6252-6259. (b) A recent review on *Pseudomonas fluorescens* lipase (Xie, Z.-F. Tetrahedron: Asymmetry 1991, 2, 733-750) may (see footnote 7 of reference 10a) be a review on *Pseudomonas cepacia* lipase.
- 11. Satisfactory <sup>1</sup>H and <sup>13</sup>C nmr data was obtained for this compound.
- 12. Hsiao, C.-N.; Hannick, S.M. Tetrahedron Lett. 1990, 31, 6609-6612 reports the synthesis of the (+)enantiomer of 10 in a rather lengthy process.
- 13. Reference 5b reports LS-Selectride provided the greatest configurational selectivity at the hydroxyl bearing carbon when reducing *trans*-2,3-bis[(benzoyloxy)methyl]cyclobutanone.
- 14. Satisfactory microanalytical and nmr (<sup>1</sup>H and <sup>13</sup>C) data were obtained for this compound.
- 15. Enantiomeric purity was determined by HPLC using a Chiracell OJ column (0.46 cm x 25 cm) and eluting with 15% 2-PrOH-hexane.
- 16. Compound 14:  $[\alpha]_D^{25}$  -12.7° (c 1.21, CHCl<sub>3</sub>); lit.<sup>6b</sup>  $[\alpha]_D^{25}$  -13.9° (c 1.18, CHCl<sub>3</sub>). Compound 15:  $[\alpha]_D^{25}$  +12.3° (c 1.03, CHCl<sub>3</sub>); lit.<sup>6b</sup>  $[\alpha]_D^{25}$  +12.6° (c 1.00, CHCl<sub>3</sub>).

(Received in USA 21 November 1991; accepted 3 February 1992)