α-HYDROXYDIHYDROCHALCONES AND RELATED 1,3-DIARYLPROPAN-2-ONES FROM XANTHOCERCIS ZAMBESIACA

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Abstract—In addition to some previously described 7,8-dioxy-isoflavonoids and 2-benzylbenzo[b]furan-3 (2H)-ones the heartwood of Xanthocercis zambesiaca has been shown to contain 7,8,3'-trihydroxy-4'-methoxyisoflavone. These are accompanied by the novel α ,3,4,4'-tetrahydroxy-2'-methoxydihydrochalcone and its 3-deoxy analogue, the latter of which presumably served as biogenetic precursor for the unique 1-hydroxy-3(4-hydroxyphenyl)-1-(4-hydroxy-2methoxyphenyl) propan-2-one and its 1-deoxy analogue. Such possible biosynthetic relationship was demonstrated by the facile *in vitro* transformations of the appropriate α -hydroxydihydrochalcone into the different 1,3diarylpropan-2-ones.

INTRODUCTION

Previous investigations [1, 2] of the heartwood extractives of Xanthocercis zambesiaca (Baker) Dumaz-le-Grand [3], the protected Nyala tree, revealed the presence of a variety of tri-, tetra-, and penta-oxygenated isoflavonoids. The 7,8-dioxy analogues amongst these may serve as precursors for xanthocercin A and B, the first members of the novel class of isoflavono-lignoids [2]. These compounds contain the 2-aryl-3-hydroxymethyl-1,4-benzodioxane moiety, a common feature of a variety of natural neolignans [4] and presumably originate by oxidative phenol coupling of substituted catechols with p-hydroxystyrenes. The potential implied by the versatile 7,8-dihydroxy functionality of the A-ring of the isoflavones thus prompted more detailed examination of heartwood metabolites related to the above classes of natural products.

RESULTS AND DISCUSSION

The methanol extract of the heartwood of Xanthocercis zambesiaca afforded xanthocercin A (1) and B (2) [2], the known 7-hydroxy-8,4'-dimethoxyisoflavone (8-Omethylretusin) [1, 5], 7-hydroxy-8,3',4'-trimethoxyisoflavone [1], 7,3'-dihydroxy 8,4'-dimethoxyisoflavone [6], 7,3',4'-trihydroxyflavonol (fisetin) [7], 2-benzyl-2,3',4',6tetrahydroxybenzo[b]furan-3(2H)-one [8] and its 4methoxy-3'-deoxy analogue [9, 10]—the latter three compounds being obtained from this natural source for the first time. These metabolites are accompanied by a novel derivative (3) of the 7,8,3',4'-tetrahydroxyisoflavone, two novel α -hydroxydihydrochalcones (5) and (7), and two unique 1,3-diarylpropan-2-ones (9) and (11).

The ⁱH NMR spectrum (300 MHz, $CDCl_3$) of the tri-O-acetyl derivative (4) of the 7,8,3'-trihydroxy-4'- methoxyisoflavone (3) exhibited the diagnostic H-2 vinylic singlet in the low-field aromatic region (δ 8.35). The aromatic region of the spectrum additionally displayed an AB-system (δ 8.12, 7.40, both d, J = 9.0 Hz), characteristic of the A-ring of 7,8-dioxygenated isoflavones [1] and an ABX-pattern (δ 7.50, dd, J = 2.0, 8.5 Hz; 7.42, d, J= 2.0 Hz; 7.16, d, J = 8.5 Hz) consistent with a 3,4- or 2,4disubstituted B-ring. Differentiation between these possibilities was effected by application of nuclear Overhauser effect (NOE) difference spectroscopy which indicated the selective association (12.3%) of methoxy protons with the o-doublet at δ 7.16 only.

Comparison of ¹H NMR data of the O-acetyl derivatives 6 and 8 of racemic α , 3, 4, 4'-tetrahydroxy-2'methoxydihydrochalcone 5 and its 3-deoxy analogue 7 revealed their close structural resemblance. Both displayed AMX-systems [δ 2.91, dd, J = 9.4, 14.5 Hz; 3.25, dd, J = 3.0, 14.5 Hz; 6.10, dd, J = 3.0, 9.4 Hz for 8: δ 2.91, dd, J = 9.7, 14 Hz; 3.26, dd, J = 3.0, 14.6 Hz; 6.08, dd, J= 3.0, 9.7 Hz for 6] in the heterocyclic regions, as well as aromatic ABC-patterns for their respective A-rings. Differences between the spectra include replacement of the aromatic ABC-system (δ 7.23, dd, J = 2.0, 8.0 Hz; 7.19, d, J = 2.0 Hz; 7.17, d, J = 8.0 Hz) for the B-ring of 6 with an AA'BB'-pattern (δ 7.33, 7.06, both d, J = 8.8 Hz) in the case of the trioxy derivative (8). The heterocyclic AMXsystems for both derivatives may, however, be indicative of the presence of a β -acetoxydihydrochalcone moiety for both these analogues. Detection of mutual benzylic coupling between H-2/6 (B-ring) and the methylene protons clearly defined these compounds 6 and 8 as α hydroxydihydrochalcones (cf. refs [11 and 12]). Finally the unambiguous placement of the methoxy function at C-2' in both 6 and 8 followed from the observed NOE association of these protons with H-3' (15.2 and 22.9% resp.), H-a (1.9 and 2.1% resp.), H-6 (1.2 and 1.3*% resp.), and with H-2 (1.2 and 1.9*% resp.).

The α -hydroxydihydrochalcones (5) and (7) appropriately 'protected' at their 2'-oxygen functions and hence incapable of cyclization, should presumably be suscep-

^{*}Approximate values due to signal overlap.











tible to preferential modification of the aliphatic α -ketol moiety. Identification of the isomerized α -hydroxydihydrochalcone, 1-hydroxy-3-(4-hydroxyphenyl)-1-(4hydroxy-2-methoxyphenyl)-propan-2-one (9) and its 1deoxy analogue (11), the first members of the novel class of naturally occurring 1,3-diaryl-propan-2-ones could, therefore, be anticipated. These metabolites were characterized by the physical data of the phenols (9 and 11) and of those of their O-acetyl derivatives (10 and 12). ¹H NMR spectra revealed marked differences between the two α -ketols (9 and 7) in the heterocyclic regions. Whereas the spectrum of the α -hydroxydihydrochalcone (7) clearly exhibits an AMX-system (see above), the product of acyloin rearrangement (9) shows two doublets (δ 3.60 and 3.51, J = 15.4 Hz) for the diastereotopic methylene hydrogens, as well as vicinal coupling (J = 5.2 Hz)between H-1 (δ 5.36) and the C-1 hydroxyl proton (δ 4.36), the latter signal disappearing on D₂O exchange and acetylation. In the peracetate (10) H-1 is deshielded to δ 6.47 with the chemical shift of the acetoxy resonance (δ 2.13) indicative of its benzylic nature. The 1,3diarylpropan-2-one arrangement for 9 was confirmed by appropriate spin-decoupling experiments which demonstrated association of the isolated proton at C-1 with the ABC-system of the A-ring and of the methylene protons with the AA'BB'-system of ring B. Location of the methyl group at the C-2" oxygen in 10 was again verified by an NOE experiment which indicated strong association (15.1%) of the methyl protons and H-3".

The ¹H NMR spectrum of 3-(4-hydroxyphenyl)-1-(4-

hydroxy-2-methoxyphenyl)propan-2-one (11) exhibited aromatic ABC- and AA'BB' spin systems, two isolated methylene singlets (δ 3.58, 3-CH₂; δ 3.54, 1-CH₂) and a single aromatic O-methyl resonance (δ 3.69), shown by its NOE association (17.5%) with H-3" to be located at the C-2" oxygen function. Spin decoupling of the methylene singlets clearly showed their adjacency to the respective aromatic rings and thus their benzylic nature (cf. refs [11, 12]). This, when taken in conjunction with the presence of a ¹³C carbonyl resonance at δ 206.2 and fragments (13) and (14) in the mass spectrum of diacetate (12) defined the constitution of the novel 1,3-diarylpropan-2-one (11).

[scheme 1 for (9) and 2 for (11)]. Thus, reduction of the α ,4,4'-tri-O-acetyl-2'-O-methyldihydrochalcone (8) by means of catalytic hydrogenation afforded the benyzyl alcohol (15) which was not isolated but treated with ptoluenesulphonic acid in anhydrous benzene to give the propan-2-one (12) presumably via intermdiacy of the enol-acetate (16) and enol (17). The 1,3-diaryl-1hydroxypropan-2-one was synthesized by utilization of the same protocol as was desbribed for base-catalysed α ketol rearrangements of 3,3-diaryl-2-hydroxypropiophenones into isomeric 1-hydroxypropan-2-ones [13]. Thus, treatment of α -hydroxy-2',4,4'-trimethoxydihydrochalcone (18) with mild aqueous base under nitrogen afforded 1-hydroxy-1-(2,4-dimethoxyphenyl)-3-

The proposed structures (9) and (11) for the 2-propanones were unequivocally confirmed by synthesis



Scheme 2.

(4-methoxyphenyl)propan-2-one (20) via enolate (19). Acetylation of the α -ketol (20) afforded the monoacetate (21) with physical data identical to those of the corresponding derivative of the natural product.

In view of their identical phenolic substitution patterns the tri-oxygenated α -hydroxydihydrochalcone (7) presumably served as biogenetic precursor to both the 1,3diarylpropan-2-ones (9) and (11). This contrasts with the suggested biosynthetic formation of reduced forms like (22-25) from flavan-3-ols and flavans [14-16]. The above facile *in vitro* transformations of α -hydroxydihydrochalcones (8) and (18) into the respective derivatives (12) and (20) of the natural products and the proven ease of conversion of α -hydroxydihydrochalcones to α -hydroxychalcones *via* the 1-hydroxy-1,3-diarylpropan-2-one of type (20) [17] may be taken as additional evidence for the presumed central role of the α -hydroxychalcone- α -hydroxydihydrochalcone pair in flavanoid biosynthesis.

EXPERIMENTAL

Mps: uncorr. ¹H NMR spectra were recorded at 300 MHz in CDCl₃ or Me₂CO- d_6 (19°) with the solvent signal (δ 7.24 or 2.04) as reference, unless otherwise stated, and mass spectral data on a Varian CH-5 instrument. Prep. TLC was carried out using 20 × 20 cm plates with 1.0 mm layers of silica gel PF₂₅₄. Zones were detected by UV and eluted with Me₂CO. CC was done with Kieselgel 60 (230–400 mesh) as stationary phase. Prep. PC comprised of 46 × 57 cm Whatman No. 3 sheets (100 mg/sheet), which were developed in 2% aq. HOAc. Bands were detected by UV and ammoniacal AgNO₃ solution and eluted with 80% EtOH. Acetylations were performed in Ac₂O-pyridine and methylations with MeI in Me₂CO/K₂CO₃.

Extraction and isolation of compounds. Drillings (2.1 kg) of the dried heartwood of Xanthocercis zambesiaca were extracted with MeOH (2 × 15 l) at room temp. (72 hr each). The combined extracts were evapd to 5 l, defatted with hexane (5 × 1 l) and evapd to give a dark-brown powder (135 g). A portion (2 × 40 g) of this material was subjected to counter-current distribution in a Quickfit Model 20 machine (25 ml lower phase; 103 transfers) in H₂O-sec-BuOH-hexane (5:4:1). The contents of tubes 66–103 (30 g) were resolved by means of paper chromatography to afford seven fractions. Fraction 1 (R_f 0.76, 2.0 g) and 6 (R_f 0.15, 0.4 g) were further fractionated by prep. TLC [C₆H₆-Me₂CO-MeOH (16:3:1) × 2 and C₆H₆-MeOH (9:1)] to yield subfractions 1.1–1.4 and 6.1–6.4 respectively.

3-(4-Hydroxyphenyl)-1-(4-hydroxy-2-methoxyphenyl) propan-2-one (11). TLC purification $[C_6H_6$ -EtOAc-MeOH (7:2:1)] of subfraction 1.2 (R_f 0.37, 164 mg) afforded propan-2-one (11) (R_f 0.58) as a pale yellow amorphous solid (36 mg); ⁻¹H NMR (Me₂CO- d_6): δ 8.30 and 8.23 (each 1H, each s, 2 × OH), 6.97 (2H, d, J = 8.5 Hz, H-2', 6'), 6.88 (1H, d, $J_{5^{\prime\prime},6^{\prime\prime}}$ = 8.1 Hz, H-6''), 6.75 (2H, d, J = 8.5 Hz, H-3',5'), 6.44 (1H, d, $J_{3^{\prime\prime},5^{\prime\prime}}$ = 2.5 Hz, H-3''), 6.36 (1H, dd, $J_{3^{\prime\prime},5^{\prime\prime}}$ = 2.5 Hz, $J_{5^{\prime\prime},6^{\prime\prime}}$ = 8.1 Hz, H-5''), 3.71 (3H, s,

> HO R^{1} OH R^{2} OH R^{2} 22 $R^{1} = OH, R^{2} = OMe$ 23 $R^{1} = R^{2} = H$ 24 $R^{1} = OH, R^{2} = H$

2"-OMe), 3.58 (2H, s, 3-CH₂), 3.54 (2H, s, 1-CH₂); MS m/z (rel. int.): 272 [M]⁺ (18), 137 (100), 107 (43).

3-(4-Acetoxyphenyl)-1-(4-acetoxy-2-methoxyphenyl)-propan-2one (12). Acetylation of ketone (11) (9 mg) yielded the diacetate (12) as an amorphous solid (8 mg). (Found: M⁺, 356.1246. $C_{22}H_{20}O_6$ requires: M, 356.1260); ¹H NMR (CDCl₃): δ 7.13 (2H, d, J = 8.6 Hz, H-2',6'), 7.06 (1H, d, $J_{5'',6''} = 8.0$ Hz, H-6''), 7.00 (2H, d, J = 8.6 Hz, H-3',5'), 6.64 (1H, dd, $J_{3'',5''} = 2.5$ Hz, $J_{5'',6''} = 8.0$ Hz, H-5''), 6.57 (1H, d, $J_{3'',5''} = 2.5$ Hz, H-3''), 3.69 (3H, s, 2''-OMe), 3.68 and 3.66 (each 2H, each s, $2 \times CH_2$), 2.28 and 2.27 (each 3H, each s, $2 \times OAc$); MS m/z (rel. int.): 356 [M]⁺ (8.6), 314 [M-CH₂CO]⁺ (5.4), 179 (19), 137 (100), 107 (24).

 (\pm) -1-Hydroxy-3-(4-hydroxyphenyl)-1-(4-hydroxy-2-methoxyphenyl)propan-2-one (9). The 1-hydroxy-2-propanone (9) (R_f 0.36) was obtained by TLC purification [CHCl₃-MeOH (47:3) × 2] of subfraction 1.3 (R_f 0.28, 156 mg) as a pale yellow amorphous solid (22 mg); ¹H NMR (Me₂CO-d₆): δ 8.65 and 8.30 (each 1H, each br s, 2 × OH), 7.05 (1H, d, $J_{5^{(*)},6^{(*)}}$ = 8.2 Hz, H-6"), 6.88 (2H, d, J = 8.5 Hz, H-2',6'), 6.70 (2H, d, J = 8.5 Hz, H-3',5'), 6.51 (1H, d, $J_{3^{(*)},5^{(*)}}$ = 2.1 Hz, H-3"), 6.43 (1H, dd, $J_{3^{(*)},5^{(*)}}$ = 2.1 Hz, $J_{5^{(*)},6^{(*)}}$ = 8.2 Hz, H-5"), 5.36 (1H, d, J = 5.2 Hz, H-1), 4.36 (1H, d, J = 5.2 Hz, 1-OH), 3. 79 (3H, s, 2"-OMe), 3.60 and 3.51 (each 1H, each d, J = 15.4 Hz) (3-CH₃).

 (\pm) -1-*Acetoxy*-3-(4-*acetoxyphenyl*)-1-(4-*acetoxy*-2-*methoxy-phenyl*)*propan*-2-*one* (10). Acetylation of compound (9) (9 mg) yielded the triacetate (10) as an amorphous solid (7 mg).(Found: M^+ , 414.1337. $C_{22}H_{22}O_8$ requires: M, 414.1315); 1H NMR (CDCl₃): δ 7.27 (1H, *d*, $J_{5'',6''}$ =8.2 Hz, H-6''), 7.02 (2H, *d*, *J* = 9.0 Hz, H-2',6'), 6.94 (2H, *d*, *J* = 9.0 Hz, H-3',5'), 6.70 (1H, *dd*, $J_{3'',5''}$ =2.1 Hz, $J_{5'',6''}$ =8.2 Hz, H-5''), 6.66 (1H, *d*, $J_{3'',5''}$ =2.1 Hz, H-3''), 6.47 (1H, *s*, H-1), 3.79 (3H, *s*, 2''-OMe), 3.72 and 3.63 (each 1H, each *d*, *J* = 16.0 Hz) (3-CH₂), 2.30 and 2.26 (each 3H, each *s*, 2 × OAc), 2.13 (3H, *s*, 1-OAc); MS *m/z* (rel. int.): 414[M]⁺ (3.0), 237 (29), 195 (89), 153 (100), 135 (3.0), 107 (21).

 (\pm) -1-Acetoxy-1-(2,4-dimethoxyphenyl)-3-(4-methoxyphenyl) propan-2-one (21). Methylation followed by acetylation and TLC separation [hexane-CHCl₃-Me₂CO (7:2:1)] of propan-2one (9) (9 mg) gave derivative (21) (R_f 0.28) as a solid (5 mg); ³H NMR (CDCl₃): δ 7.16 (1H, d, $J_{5'',6''} =$ 7.0 Hz, H-6''), 6.94 (2H, d, J = 8.8 Hz, H-2'.6'), 6.77 (2H, d, J = 8.8 Hz, H-3'.5'), 6.48 (1H, dd, $J_{3'',5''} =$ 2.0 Hz, $J_{5'',6''} =$ 7.0 Hz, H-5''), 6.47 (1H, d, $J_{3'',5''} =$ 2.0 Hz, H-3''), 6.43 (1H, s, H-1), 3.81, 3.80 and 3.75 (each 3H, each s, 3 × OMe), 3.63 and 3.56 (each 1H, each d, J = 16.0 Hz, 3-CH₂), 2.11 (3H, s, 1-OAc).

(±)-2-Benzyl-2,3'.4',6-tetra-acetoxybenzo[b]firan-3(2H)-one [8]. Acetylation and prep. TLC separation [hexane-C₆H₆-1,2dichloroethane-MeOH (9:6:4:1) × 2] of a portion (100 mg) of subfraction 1.4 (R_f 0.19, 172 mg) afforded two amorphous solids R_f 0.63 (12 mg) and 0.57 (16 mg) respectively. The former (R_f 0.63) was identified as 2-benzyl-2,3',4'.6-tetra-acetoxybenzo[b]furan-3(2H)-one; ¹H NMR (CDCl₃): δ 7.59 (1H. d, $J_{4,5}$ = 8.4 Hz, H-4), 7.13 (1H, dd, $J_{2',6'}$ = 2.0 Hz, $J_{5',6'}$ = 8.1 Hz, H-6'), 7.09 (1H, d, $J_{2',6'}$ = 2.0 Hz, H-2'), 7.04 (1H, d, $J_{5',6'}$ = 8.1 Hz, H-5'). 6.81 (1H, d, $J_{5,7}$ = 2.0 Hz, H-7), 6.78 (1H, dd, $J_{5,7}$ = 2.0 Hz, $J_{4,5}$ = 8.4 Hz, H-5), 3.24 and 3.02 (each 1H, each d, J = 14.1 Hz)



(CH₂), 2.29, 2.25 and 2.24 (each 3H, each s, $3 \times OAc$), 2.07 (3H, s, 2-OAc); MS m/z (rel. int.): 456 [M]⁺ (2.9), 414 [M $-CH_2CO$]⁺ (7.9), 397 [M-OAc]⁺ (1.9), 270 [M $-HOAc-3 \times OAc$]⁺ (23), 206 (30), 179 (46), 165 (33), 151 (7.4), 137 (100), 123 (83).

(±)-2-Benzyl-2,4',6-triacetoxy-4-methoxybenzo [b] furan-3(2H)-one (2,4',6-tri-O-acetylcarpusin [9, 10]. The second compound (R_f 0.57; vide supra) was identified as 2,4',6-tri-Oacetylcarpusin; ¹H NMR (CDCl₃): δ 7.25 (2H, d, J=8.8 Hz, H-2',6'), 6.95 (2H, d, J=8.8 Hz, H-3',5'), 6.37 (1H, d, J_{5,7} = 1.8 Hz, H-7), 6.21 (1H, d, J_{5,7} = 1.8 Hz, H-5), 3.87 (3H, s, 4-OMe), 3.25 and 3.04 (each 1H, each d, J = 14.0 Hz) (CH₂), 2.28 and 2.25 (each 3H, each s, 2 × OAc), 2.05 (3H, s, 2-OAc); MS m/z (rel. int.): 428[M]⁺ (8.8), 386 [M-CH₂CO]⁺ (2.6), 368 [M-HOAc]⁺ (12), 237 (15), 209 (43), 195 (60), 167 (100), 107 (60).

(±)-α,4,4'-Trihydroxy-2'-methoxydihydrochalcone (7). Prep. TLC separation [CHCl₃-Me₂CO-MeOH (17:2:1) of fraction 3 from paper chromatography (R_f 0.57, 1.5 g) gave two compounds R_f 0.43 (172 mg) and 0.24 (150 mg) respectively. The first compound (R_f 0.43) comprised of the α-hydroxydihydrochalcone (7) as a yellow solid; ¹H NMR (Me₂CO-d₆): δ 7.74 (1H, d, $J_{5',6'}$ = 8.9 Hz, H-6'), 7.01 (2H, d, J = 8.7 Hz, H-2,6), 6.70 (2H, d, J = 8.7 Hz, H-3,5), 6.60 (1H, d, $J_{3',5'}$ = 2.0 Hz, H-3'), 6.56 (1H, dd, $J_{3',5'}$ = 2.0 Hz, $J_{5',6'}$ = 8.9 Hz, H-5'), 5.14 (1H, dd, J = 3.2, 8.0 Hz, H-α), 3.91 (3H, s, 2'-OMe), 2.97 (1H, dd, $J_{\alpha,\beta}$ = 3.2 Hz, $J_{\beta,\beta}$ = 13.9 Hz), 2.51 (1H, dd, $J_{\alpha,\beta}$ = 8.0 Hz, $J_{\beta,\beta}$ = 13.9 Hz) (β-CH₂).

 (\pm) -α,4,4'-Triacetoxy-2'-methoxydihydrodrochalcone (8). Acetylation of α-hydroxydihydrochalcone (7) (72 mg) yielded the triacetate (8) as a solid (70 mg). (Found: C, 63.7; H, 5.4. $C_{22}H_{22}O_8$ requires: C, 63.8; H, 5.4%), ¹H NMR (Me₂CO-d₆, TMS as internal standard): δ 7.81 (1H, d, $J_{5',6'}$ =8.6 Hz, H-6'), 7.33 (2H, d, J=8.8 Hz, H-2,6), 7.06 (2H, d, J=8.8 Hz, H-3,5), 7.02 (1H, d, $J_{3',5'}$ =1.8 Hz, H-3'), 6.88 (1H, dd, $J_{3',5'}$ =1.8 Hz, $J_{5',6'}$ =8.6 Hz, H-5'), 6.10 (1H, dd, J=3.0, 9.4 Hz, H-α), 4.03 (3H, s, 2'-OMe), 3.25 (1H, dd, $J_{\alpha,\beta}$ =3.0 Hz, $J_{\beta,\beta}$ =14.5 Hz), 2.91 (1H, dd, $J_{\alpha,\beta}$ =9.4 Hz, $J_{\beta,\beta}$ =14.5 Hz) (β-CH₂), 2.29 and 2.24 (each 3H, each s, 2 × OAc), 2.02 (3H, s, α-OAc); MS m/z (rel. int.): 354 [M -HOAc]⁺ (13), 312 [M-HOAc-CH₂CO]⁺ (10), 270 [M -HOAc-2 × CH₂CO]⁺ (3.8), 151 (100), 119 (1).

(±)-α-Hydroxy-2',4,4'-trimethoxydihydrochalcone (18). Methylation of α-hydroxydihydrochalcone (7) (100 mg) followed by TLC purification [CHCl₃-Me₂CO (19:1)] gave the methyl ether (18) (R_f 0.64) as a yellow solid (90 mg). (Found: C, 68.5; H, 6.3. C₁₈H₂₀O₅ requires: C, 68.3; H, 6.4%); ¹H NMR (CDCl₃): δ 7.81 (1H, d, $J_{5',6'}$ = 8.8 Hz, H-6'), 7.03 (2H, d, J = 8.8 Hz, H-2,6), 6.72 (2H, d, J = 8.8 Hz, H-3,5), 6.56 (1H, dd, $J_{3',5'}$ = 2.2 Hz, $J_{5',6'}$ = 8.8 Hz, H-5'), 6.44 (1H, d, $J_{3',5'}$ = 2.2 Hz, H-3'), 5.25 (1H, m, Hα), 3.91 (1H, s, α-OH), 3.88, 3.84 and 3.72 (each 3H, each s, 3 × OMe), 3.06 (1H, dd, $J_{\alpha,\beta}$ = 3.8 Hz, $J_{\beta,\beta}$ = 13.8 Hz), 2.63 (1H, dd, $J_{\alpha,\beta}$ = 7.5 Hz, $J_{\beta,\beta}$ = 13.8 Hz) (β -CH₂); MS m/z (rel. int.): 298 [M - H₂O]⁺ (60), 195 (69), 167 (90), 165 (100), 151 (31), 139 (40), 137 (49), 121 (79), 107 (22).

(±)-α-3,4,4'-*Tetra-acetoxy*-2'-methoxydihydrochalcone (6). Acetylation and TLC purification [hexane-Me₂CO-EtOAc (11:6:3)] of the second compound [R_f 0.24; cf. (7)] gave the title compound (6) (R_f 0.42) as a solid (64 mg). (Found: C, 60.8; H, 5.0. C₂₄H₂₄O₁₀ requires: C, 61.0; H, 5.1%); ¹H NMR (Me₂CO-d₆, TMS as int. standard): δ 7.82 (1H, d, $J_{5',6'}$ = 8.5 Hz, H-6'), 7.23 (1H, dd, $J_{2,6}$ = 2.0 Hz, $J_{2,6}$ = 2.0 Hz, $J_{5,6}$ = 8.0 Hz, H-6), 7.19 (1H, d, $J_{2,6}$ = 2.0 Hz, H-2), 7.17 (1H, dd, $J_{3',5'}$ = 2.1 Hz, H-3'), 6.87 (1H, dd, $J_{3',5'}$ = 2.1 Hz, $J_{5',6'}$ = 8.5 Hz, H-5'), 6.08 (1H, dd, $J_{=3,0}$ = 7.42, $J_{5,\beta}$ = 14.6), 2.91 (1H, dd, $J_{a,\beta}$ = 9.7 Hz, $J_{\beta,\beta}$ = 14.6 Hz) (β -CH₂), 2.28, 2.26 and 2.25 (each 3H, each s, 3 × OAc), 2.02 (2H, s, α-OAc); MS m/z (rel. int.): 412 [M -HOAc]⁺ (7.0), 370 [M-HOÅc-CH₂CO]⁺ (6.6), 328 [M $-HOAc-2 \times CH_2CO]^+$ (12), 286 [M-HOAc-3 $\times CH_2CO]^+$ (4.3), 151 (100), 136 (2.0).

7-Acetoxy-8,4'-dimethoxyisoflavone (7-O-acetyl-8-O-methylretusin [1, 5]. Acetylation and subsequent TLC purification [hexane-Me₂CO-EtOAc (13:4:3)] of subfraction 6.1 (R_f 0.31, 19 mg) yielded 7-acetoxy-8,4'-dimethoxyisoflavone (R_r 0.41) as a solid (15 mg); ¹H NMR (CDCl₃, TMS as internal standard: $\delta 8.03$ (1H, d, $J_{5,6} = 8.5$ Hz, H-5) 8.02 (1H, s, H-2), 7.49 (2H, d, J = 9.0 Hz, H-2',6'), 7.12 (1H, d, $J_{5,6} = 8.5$ Hz, H-6), 6.97 (2H, d, J = 9.0 Hz, H-3',5'), 4.01 (3H, s, 8-OMe), 3.83 (3H, s, 4'-OMe), 2.39 (3H, s, 7-OAc); MS m/z (rel. int.): 340 [M]⁺ (49), 298 [M - CH₂CO]⁺ (100), 166 (12), 132 (26).

7-Acetoxy-8,3',4'-trimethoxyisoflavone [1]. Acetylation and TLC purification [C_6H_6 -MeOH (92:8)] of subfraction 6.2 (R_f 0.26, 22 mg) afforded the 7-acetoxy-8,3',4'-trimethoxyisoflavone (R_f 0.27) as a solid (16 mg); ¹H NMR (CDCl₃, TMS as int. standard: δ 8.07 (1H, s, H-2), 8.04 (1H, d, $J_{5,6}$ = 9.0 Hz, H-5), 7.20 (1H, d, $J_{2',6'}$ = 2.1 Hz, H-2'), 7.14 (1H, d, $J_{5,6}$ = 9.0 Hz, H-6), 7.06 (1H, dd, $J_{2',6'}$ = 2.1 Hz, $J_{5',6'}$ = 8.0 Hz, H-6'), 6.94 (1H, d, $J_{5',6'}$ = 8.0 Hz, H-5'), 4.03 (3H, s, 8-OMe), 3.94 and 3. 92 (each 3H, each s, 3' and 4'-OMe), 2.40 (3H, s, 7-OAc); MS m/z (rel. int.): 370 [M]⁺ (91), 328 [M-CH₂CO]⁺ (100), 167 (3.0), 166 (1.3), 162 (6.4).

7,3'-Diacetoxy-8,4'-dimethoxyisoflavone [6]. Acetylation follwed by TLC purification [hexane–Me₂CO–EtOAc (11:6:3)] of subfraction 6.3 (R_f 0.20, 41 mg) gave 7,3'-diacetoxy-8,4'-dimethoxyisoflavone (R_f 0.48) as a solid (23 mg); ¹H NMR (Me₂CO- d_6 , TMS as int. standard): δ 8.43 (1H, s, H-2), 7.93 (1H, d, $J_{5',6'} = 9.0$ Hz, H-5), 7.52 (1H, dd, $J_{2',6'} = 2.1$ Hz, H-2'), 7.25 (1H, d, $J_{5,6} = 8.7$ Hz, H-6'), 7.43 (1H, d, $J_{2',6'} = 8.7$ Hz, H-5'), 4.02 (3H, s, 8-OMe), 3.87 (3H, s, 4'-OMe), 2.38 and 2.27 (each 3H, each s, 2 × OAc); MS m/z (rel. int.): 398 [M]⁺ (30), 356 [M-CH₂CO]⁺ (64), 314 [M-2×CH₂CO]⁺ (100), 167 (2.9), 148 (2.3).

7,8,3'-Triacetoxy-4'-methoxyisoflavone (4). The title compound (4) (R_f 0.42) was obtained from subfraction 6.4 (R_f 0.10, 32 mg), after acetylation and TLC purification [hexane-Me₂CO-EtOAc (11:6:3)], as an amorphous solid (29 mg). (Found: M⁺, 426.0975. C₂₂H₁₈O₉ requires: M, 426.0951); ¹H NMR (Me₂CO-d₆, TMS as int. standard): δ 8.35 (1H, s, H-2), 8.12 (1H, d, J_{5.6} = 9.0 Hz, H-5), 7.50 (1H, dd, J_{2'.6'} = 2.0 Hz, J_{5'.6'} = 8.5 Hz, H-6'), 7.42 (1H, d, J_{2'.6'} = 2.0 Hz, H-2'), 7.40 (1H, d, J_{5.6} = 9.0 Hz, H-6), 7.16 (1H, d, J_{5'.6'} = 8.5 Hz, H-5'), 3.86 (3H, s, 4'-OMe), 2.41, 2.36 and 2.26 (each 3H, each s, 3 × OAc); MS m/z (rel. int.): 426 [M]⁺ (46), 384 [M-CH₂CO]⁺ (92), 342 [M-2 × CH₂CO]⁺ (68), 300 [M-3 × CH₂CO]⁺ (100), 152 (2.9), 148 (2.6).

3,7,3',4'-Tetra-O-acetylfisetin. CC [CHCl₃-MeOH (19:1), RR_t 153 hr] of a portion (2 g) of fraction 7 (R_f 0.02, 9.5 g) followed by acetylation and TLC purification [hexane-Me₂CO-EtOAc (11:6:3)] yielded tetra-O-acetylfisetin (R_f 0:39) as a solid (9 mg); mp 194–196° (lit. [7] mp 196–198°).

Synthesis of (\pm) -3-(4-acetoxyphenyl)-1-(4-acetoxy-2-methoxyphenyl)propan-2-one (12). Catalytic hydrogenation [EtOH (20 ml), 10% Pd/C (13 mg), 6 hr] of α ,4,4'-triacetoxy-2'methoxydihydrochalcone (8) (50 mg) followed by TLC purification [C₆H₆-MeOH (19:1)] yielded an intermediate of R_f 0.31, which was refluxed (22 hr) in C₆H₆ (20 ml) containing ptoluenesulphonic acid (5 mg). Evapn of the solvent, followed by TLC separation [CHCl₃-EtOAc (9:1)] afforded the title propan-2-one (12) (R_f 0.65) as an amorphous solid (12 mg) identical to that obtained from the natural source.

Synthesis of (\pm) -1-hydroxy-1-(2,4-dimethoxyphenyl)-3-(4methoxyphenyl)propan-2-one (20). 1 N aq. NaOH (5 ml) was added under N₂ to a solution of α -hydroxy-2',4,4'-trimethoxydihydrochalcone (18) in THF (80 mg in 5 ml) and the mixture stirred at room temperature for 22 hr. After stirring at 35° for another 3 hr, ice was added, the reaction mixture acidified to pH 5 (1 N HCl) and extracted with EtOAc (3 × 20 ml). The combined extracts were washed with water (3 × 60 ml), until acid-free, dried (Na₂SO₄) and purified by TLC [hexane-CHCl₃-Me₂CO (7:2:1) × 2] to give the 1-hydroxy-2-propanone (**20**) (R_f 0.43) as a pale yellow solid (11 mg). (Found: M⁺, 316.1372. C₁₈H₂₀O₅ requires: M, 316.1341); ⁻¹H NMR (Me₂CO-d₆): δ 7.17 (1H, d, $J_{5'',6''}$ = 8.2 Hz, H-6''), 6.97 (2H, d, J = 8.6 Hz, H-2',6'), 6.80 (2H, d, J = 8.6 Hz, H-3',5'), 6.59 (1H, d, $J_{3'',5''}$ = 2.6 Hz, H-3''), 5.39 (1H, dd, $J_{3'',5''}$ = 2.6 Hz, $J_{5'',6''}$ = 8.2 Hz, H-5''), 5.39 (1H, d, J = 5.5 Hz, H-1), 4.43 (1H, d, J = 5.5 Hz, 1-OH), 3.83, 3.81 and 3.74 (each 3H, each s, 3 × OMe), 3.66 and 3.56 (each 1H, each d, J = 15.8 Hz, 3-CH₂); MS m/z (rel. int.): 316 [M]⁺ (1.1), 195 (2.6), 167 (100), 149 (1.6), 121 (20).

 (\pm) -1-Acetoxy-1-(2,4-dimethoxyphenyl)-3-(4-methoxyphenyl)propan-2-one (21). Acetylation of 1-hydroxy-2-propanone (20) (8 mg) gave the monoacetate (21) (R_f 0.28, 5 mg) identical to that obtained from the natural product.

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