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Synthesis of Hexahydrocyclopentimidazol-2-(1*H*)-one Derivatives Displaying Selective DP-Receptor Agonist Properties

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Abstract—The rationale for investigating conformationally restricted analogues of BW245C as DP-receptor ligands and the syntheses of three such racemic bicyclic imidazolidinone analogues are described. Compounds 7 (BW587C), 8 (BW480C85), and 9 (BW572C85) were found to be potent inhibitors of human platelet aggregation and selective DP-receptor agonists in washed platelet and jugular vein isolated tissue assays.

Introduction

Blood platelets play a key role in the pathophysiology of arterial thrombosis. Both prostaglandin (PG) I₂ and PGD₂ are potent inhibitors of human platelet aggregation although they act via different receptors^{1,2} (IP and DP respectively). Indeed, PGI₂ (Flolan, epoprostenol sodium) has been marketed for use as an antithrombotic agent.³ The DP-receptor agonist, BW245C (1), has also been studied⁴ in human volunteers. The major limitation to the clinical use of these agents as antithrombotics has been their short duration of action and their vasodepressor side effects. In view of their favourable antithrombotic profile, the identification of platelet-selective agonists, devoid of effects on vascular receptors is therefore an attractive goal.

DP-selective ligands are required to determine whether DP-receptors on human platelets and vascular smooth muscle are of the same type. DP-receptor properties are known to vary between species^{5,6} and animal models are therefore of limited value for assessing vasodilatation side effects which manifest as facial flushing and headache. The hypothesis that selective DP-agonists will display potent platelet effects with minimal vascular actions can therefore probably only be tested in man.

As a first step towards testing this hypothesis we obviously required a selective DP-receptor agonist. The DP-agonist 1^7 was taken as the lead compound for optimisation. Studies of the structure-activity relationships (SARs) of BW245C have shown⁸ that when the 9'-carbonyl group of 1 is replaced by CH₂ (to give 2) the inhibitory effects on human platelet aggregation are retained. Similarly, replacement of the 5'-6' CH_2CH_2 moiety of 1 by CH_2S (to give 3) leads to only a small variation in potency with respect to inhibition of platelet aggregation.⁹ The 13'-aza analogues 4 and 6 are more potent inhibitors of platelet aggregation than 1 and their effects are of longer duration in vivo.¹⁰ Compounds 1–4 and 6 are non-selective DP-receptor agonists, but which display additional agonist effects at other prostanoid (possibly EP₂) vascular receptors. Nevertheless, replacement of the N-10 H of 4 by a benzyl group, to give 5 (BWA868C), leads to a highly selective DP-receptor in addition to a complete loss of affinity at the other vascular receptor.

As a result of our SAR and modelling studies⁹ we speculated that the DP-receptor-bound conformations of PGD₂ and BW245C are ones in which C-6 (and C-6', PG numbering) is folded back toward the main ring. We therefore designed the heterobicyclic analogues of BW245C, **7-9**, in which C-6' is tethered by a stable methylene linked to the ring at C-9'. Bicycles **7-9** cannot adopt the fully extended α - ω -parallel conformation ('Hairpin')¹¹ of PGF_{2a}, but are capable of adopting conformations similar to the C-6 folded crystal conformation¹² of PGE₂ reasonably well. It seemed possible, therefore, that these conformationally restricted analogues might display selective DP-receptor agonism especially if extended conformations^{13,14} are a prerequisite for agonism at other PG receptors.

Although analogues 7-9 are more rigid than 1 they are more flexible than PGI_2 . Nonetheless, our main



concern with these conformational restraints was that the broad structural similarity between these analogues and PGI_2 might lead to effects at IP-receptors.

Synthesis

The eleven step synthesis of racemic 7, from the unstable bromide 14^{15} and nitrile $12^{16,17}$ is shown in Scheme 1. The key step is the in situ generation of enone 17 and subsequent conversion into bicycles 18 and 19 via an intramolecular Michael type addition. Diastereomers 18 and 19 are readily separated by flash chromatography.¹⁸ Their relative stereochemistries were determined by converting each diastereomer into the corresponding prostanoid. Since 7 displayed far more potent (>100×) PG mimetic activity than its 15'-epimer 23, 7 and 18 (1 (\mathbb{R}^*) 3aa, 6aa) were assigned the same configurations as PGD₂. The reduction of the O-acetate of 18 with NaBH₄ gave a single alcohol epimer which was assigned as 20 (i.e., the 5- α configuration) on the basis of NOE experiments.¹⁹ Reaction of mesylate 21 with NaS(CH₂)₃CO₂Me²⁰ was also stereospecific and gave, as expected, diester 22 with the inverted 5- β configuration.²¹

The 13'-aza analogues (\pm) -8 and 9 were obtained from bromide 14¹⁵ and benzaldehyde hydrazone 24²² by the route shown in Scheme 2. Stereochemical assignments²³ were analogous to those of the 13'-carbo series. Thus, hydrazone 27 was converted into the bicyclic N-amino imidazolidinone 29 and then to alcohol 30. Since this alcohol could not be stored satisfactorily for more than a few days separation of the diastereomers was carried out²⁴ on the mesylate intermediates 31 and 32. Conversion of (\pm) -14 into (\pm) -8 was achieved in 2.4 overall yield,²⁵ compared with 0.6% of (\pm) -7 from (\pm) -14.²⁶

Pharmacological Results and Discussion

The PG mimetic activities of analogues 1-9 were determined by previously described methods^{5.27} and are shown in Table 1. Bicyclic sulphides 7-9 are more potent inhibitors of platelet aggregation than sulphide 3 but only the 13'-aza analogue 9 was found to be much more potent than BW245C. Thus, structural changes (conformational restriction, loss of carbonyl, and replacement of 13'-CH₂ by NH) in going from $3 \rightarrow 7 \rightarrow 9$ lead to improved potency of platelet effects. Analogue 7 displays much weaker vasodilatory actions than 3, 8, or 9. Interestingly, the duration of the hypotensive effects of 8 are greater than those of 9 or 7, whereas 4 is longer acting than 6. Further work will be required to determine if their metabolic deactivation occurs by different pathways.



Scheme 1. Reagents: (i) NBS, AIBN, CCl₄ [ref. 15]; (ii) *n*-BuLi, CH₃CN, -78 °C (52%); (iii) LiAlH₄, Et₂O, 25 °C (58%); (iv) Toluene, 25 °C for 24 h; 40 °C for 3 h; NEt₃ (32%); (v) KCNO, 1 N HCl, EtOH-H₂O (85%); (vi) 2 N HCl, THF, 25 °C, 2 h (45% of 18; 40% of 19); (vii) Ac₂O, C₃H₃N, rt, 18 h (84%); (viii) NaBH₄, EtOH (54%); (ix) MeSO₂Cl, C₃H₃N, 0-5 °C, 24 h (50%); (x) Thiobutyrolactone, NaOMe, MeOH; DMSO (63%); and (xi) 2 N NaOH, MeOH (88%).



Scheme 2. Reagents: (i) Toluene -40 °C→0 °C; NEt₃, reflux 2 h (55%); (ii) COCl₂, C₅H₅N, toluene 0 °C; (iii) aqueous NH₃ (d 0.88) (90% overall); (iv) Camphor sulphonic acid, Me₂CO (55%); (v) NaBH₄, EtOH (85%); (vi) H₂, 10% Pd-C, EtOH, AcOH, 70 °C, 90 atmos (90%); (vii) C₆H₁₁CH(OAc)CHO [ref. 30], NaOAc, MeOH, 50 °C, 3 h (75%); (viii) MeSO₂Cl, C₅H₅N, CH₂Cl₂ (35% of **31**, 36% of **32**); (ix) Thiobutyrolactone, NaOMe, MeOH; DMSO (50%); (x) LiOH, H₂O-MeOH (86%); (xi) Na(CN)BH₃, AcOH, MeOH (90%).

Table 1. Pharma	acological	activities	of BW245C	and	analogues
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The sulphide 7 proved to be a potent and selective DP-receptor agonist (Table 2). In the washed human platelet assay⁷ it was found to be a full agonist at the DP-receptor. It behaves as a partial agonist at the DP-receptor in the rabbit jugular vein assay⁷ and shows greater selectivity than compound 1 since the effects of 7 could be completely antagonised by BWA868C, which is selective for DP-receptors, with no evidence of actions at other PG receptors. The 13'-aza analogues BW480C85 (8) and BW572C85 (9) also proved to be selective DP-receptor agonists and behaved as full agonists in the platelet assay. In the jugular vein assay both 8 and 9 behaved as partial agonists, with increased potencies relative to those of 7. As with 7 the compounds did not activate a second vascular receptor mediating relaxation. EP₂- and IP-receptor mediated relaxations and TP-receptor mediated constrictions in jugular vein were unaffected by these compounds.²⁸ Also they did not effect IP- or TP-receptor mediated actions on platelet aggregation or smooth muscle contraction at FP- or EP₁-receptors.²⁸ The DPreceptor selectivity of these bicycles is probably a reflection of their restricted conformations. A better understanding of any unique features of DP-receptor bound ligands would be facilitated by detailed studies of structure-activity relationships,²⁹ crystallographic investigations, and development of other selective IPand EP₂-receptor ligands. Unfortunately, selective antagonists for IP- and EP-receptors are not available at present so the sub-type of PG-receptor mediating the DP-antagonist insensitive effects of BW245C cannot be assigned.

In summary bicycles **7–9** should find utility in PGreceptor classification studies²⁸ as selective DP-agonists. Investigations of the dose–response relationships for inhibition of platelet aggregation and vasodilatation in

Compound	Inhibition of ADP-in aggre	duced human platelet gation	Blood pressure lowering activity in rat		
	$\frac{\text{Relative}}{\text{PGD}_2 = 1}$	potencies* (PGI ₂ =1)	Relative potency ^b (PGI ₂ =1)	t _{1/2} (min) ^c	
1 BW245C	8	(0.20)	0.06	4	
2	6	(0.14)	0.01	3	
3	4.4	(0.1)	0.15	3	
4 BW361C	24	(0.6)	0.02	28	
5 BWA868C	no agonism		d		
6 BW68C	12	(0.3)	0.04	17	
7 BW587C	10	(0.26)	< 0.001	1	
8 BW480C85	9.6	(0.24)	0.01	25	
9 BW572C85	29	(0.72)	0.07	7	

^aPotencies relative to PGD₂ are approximate. BW245C is approximately 8 and 0.2 times as potent as PGD₂ and PGI₂, respectively. The relative potencies were calculated from the IC₅₀ values of the compounds. IC₅₀ is the concentration required to reduce the aggregation to 50% of its control amplitude. Compounds 1, 4, 6, 7, 8 and 9 have IC₅₀ values (ng mL⁻¹) of 1.5 ± 0.1 (10), 0.50 ± 0.06 (8), 1.1 ± 0.2 (6), 1.2 ± 0.26 (5), 1.25 ± 0.2 (6), and 0.41 ± 0.04 (8) respectively for (n) experiments. Inactive compounds have IC₅₀'s > 200 ng mL⁻¹. All compounds were incubated with human platelet rich plasma (PRP) for 1 min. PGI₂ was used as a standard for each batch of PRP. Relative potencies were confirmed by comparing the effects of groups of analogues (up to 5 per experiment) with BW245C and PGI₂ on the same batch of PRP.

^b Values are relative to prostacyclin in the same anaesthetized animal, $n \ge 3$.

^cHalf-life (standard error mean \pm 20%) of hypotensive effects at a dose that causes a fall in systemic arterial blood pressure of 20 mmHg. ^dTachycardia and some blood pressure lowering is observed in conscious rats.

Table 2.	DP-receptor	effects	of	BW245C	and	analogues
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Compound	pA_{50}^{a}		pK_A^d		Agonist/Antagonist ^e		DP-selectivity ^f
	Platelet ^b	Jugular ^c	Platelet ^b	Jugular ^c	Platelet	Jugular	
1 BW245C	8.24	7.09	n.d. ^g	n.d. ^g	F	F	No
5 BWA868C	_	_	9.25 ^h	8.7	Ant.	Ant.	Yes
7 BW587C	6.53	5.29	n.d. ^g	n.d. ^g	F	P (0.5)	Yes
8 BW480C	8.00	7.37	6.69	6.75	F	P (0.6)	Yes
9 BW572C	8.43	7.34	7.21	6.63	F	P (0.7)	Yes

^aParameter describing potency of agonist response. $pA_{50} = -\log_{10}A_{50}$ where A_{50} is the concentration of compound that produces 50% of the maximum agonist response. Typical error: **9**, 8.43 ± 0.15 (n = 4).

^bAgonists inhibit the aggregation of human washed platelets induced by 50 μM ADP.

^cAgonists produce relaxation of tone in rabbit jugular vein ring segments after prior contraction with 1 µM histamine.

⁴Parameter describing affinity of compound for DP-receptor. $pK_A = -\log_{10}K_A$ is the dissociation constant for the agonist at the DP-receptor. Typical errors: 9, 7.21 ± 0.29; 6.63 ± 0.13: 8, 6.69 ± 0.40, 6.75 ± 0.15. Values are the average of 3–6 estimates.

*Classification of compound as agonist or antagonist. F = Full agonist; P = partial agonist; Ant. = antagonist. In the platelet assay all agonists gave a maximal response (100% inhibition of aggregation). BW245C was the only agonist to give a maximal response (100% relaxation) in the rabbit jugular vein. For partial agonists the approximate mean efficacy is given in brackets. Thus compound 8 ($\alpha = 0.6$) gives $61 \pm 25\%$ (n=3) of the response observed for BW245C.

Inferred from experiments with the selective DP-antagonist, **5**. If the effects of the agonist were totally blocked in the presence of 10 μ M **5** the agonist was assumed to be DP-selective. Non-selective agonists show a response which is partly resistant to BWA868C antagonism. The DP-receptor selectivity of compounds **7–9** was confirmed by their absence of effects at other prostanoid receptors. All experiments ($n \ge 2$) were performed in the presence of 10 μ m BWA858C to prevent binding to the DP-receptor. None of the compounds, at concentrations up to 10 μ m, exerted any effect on baseline tone in the rabbit jugular vein preparation nor on histamine (1 μ m)-induced tone in this assay. It has previously been shown⁷ that this jugular vein preparation contains TP-receptors which mediate vasoconstriction, IP-receptors² and EP₂-receptors³¹ mediating smooth muscle contraction, compounds **7–9** were without effect at concentrations up to 10 μ M. In the platelet assay neither IP-receptor-mediated inhibition of aggregation (by PGI₂, PGE₁ and carbacyclin) nor TP-receptor-mediated aggregation (by U46619) was affected by concentrations up to 10 μ m.

⁸n.d. = not determined. PGD₂ and BW245C elicit non-DP-receptor effects (e.g., vasorelaxation through presumed EP₂-receptors). Displacements of agonist-concentration effect curves may also be confounded by slowing of agonist responses in the presence of BWA868C.

^hp K_B value of the antagonist. p K_B is the dissociation constant of the antagonist.

man will allow a test of the hypothesis that selective inhibition of platelet aggregation by DP-receptor ligands is possible.

Experimental

Mps were taken in open capillary tubes and are uncorrected. ¹H NMR spectra were obtained on Bruker HFX90, AM-200, or WM-360 spectrometers. Ei mass spectra were obtained on an A.E.I. MS 902 spectrometer, interfaced to a VG MULTISPEC data system at 70 eV. Fast atom bombardment mass spectrometry (FABMS) were obtained from a Kratos MS50 mass spectrometer equipped with an RF magnet. The instrument was calibrated to 2,000 amu with caesium iodide. The samples were admixed with thioglycerol on the probe tip, introduced via the vacuum lock to the source on the mass spectrometer, and bombarded with fast Xe atoms. Thin-layer chromatograms (TLC) were performed on Merck silica gel 60 F_{254} and developed with iodine or phosphomolybdic acid, gravity column chromatography with Merck silica gel (60–120 mesh), and flash chromatography with Merck silica gel (230-400 mesh).

Several of the intermediates prepared which are described as oils, syrups or waxes may be crystalline solids when analytically pure. Crystallisation of many pure compounds was often very slow and inhibited by small amounts of occluded solvents. Melting points of the bicycles were often of poor diagnostic value since they appeared to vary with heating rate and trace impurities. Compounds in this series are probably capable of existing in several interconvertible crystalline forms. **Hazard**: Bioactive compounds should be handled with care since absorption on skin may cause facial flushing.

 (\pm) -3-Cyclohexyl-3-hydroxypropylamine (13). A solution of 3-cyclohexyl-3-hydroxyproprionitrile $(12)^{16,17}$ (40.8 g, 0.27 mol) in dry ether (360 mL) was added dropwise, over 45 min, to a stirred solution of lithium aluminium hydride (10.4 g, 0.27 mol) in dry ether (450 mL) at 0 °C under N₂. The mixture was stirred at rt for 2 h, a saturated aqueous solution of Na₂SO₄ added slowly followed by CHCl₃ (1500 mL). Inorganic material was removed by filtration and washed with more CHCl₃. Evaporation of the filtrate gave a pale yellow oil. Purification by distillation yielded 24.6 g (58%) of 13, bp 82–92 °C/0.3 mmHg, as a colourless syrup which solidified to a wax on cooling. 'H NMR (200 MHz, CDCl₃) δ: 0.80-2.08 (13H, complex m, aliphatic H), 2.40 (2H, brs, NH₂, exchang.) 2.64-3.20 (2H, m, NCH₂), 3.30-3.75 (1H, m, OCH). FABMS m/z 158 (M^+ + H). Anal. calcd for C₉H₁₉NO requires C, 68.7; H, 12.2; N, 8.9. Found C, 68.5; H, 12.4; N, 8.7%.

[1(*R*,*S*)-N(*R**,*S**)]-N-(3-Cyclohexyl-3-hydroxypropyl)-4,4-ethylenedioxycyclopent-2-enamine (15). 5-Bromo3-ethylenedioxycyclopent-2-ene (14) was prepared from the ketal 10 (15.0 g, 0.12 mol) by the method of DePuy¹⁵ and dissolved in dry toluene (150 mL). The solution was added dropwise with stirring to the amine 13 (15.0 g, 0.096 mol) in dry toluene (240 mL) at -25 °C. The brown solution was stirred for 26 h at rt then at 40 °C for 3.5 h then triethylamine (18.0 g) was added. The resulting cloudy solution was washed with three portions of water, then dried (MgSO₄) and the solvent evaporated in vacuo giving a brown oil. Purification by column chromatography (silica, chloroform: methanol, 100:7) gave the mixture of diastereomers of 15 as a pale red oil. Yield 8.58 g (32%) ¹H NMR (200 MHz, $CDCl_3$) δ : 0.80–2.02 (14H, complex m, aliphatics + ring C<u>H</u>), 2.14–2.56 (1H, m, ring CH<u>H</u>), 2.60–3.58 (5H, complex m, NHCH₂+CHOH, 2H exchang.), 3.64-3.90 (1H, m, CHN), 3.92 (4H, s, OCH₂CH₂O), 5.56–5.80 (1H, m, olefinic H), 5.86–6.10 (1H, m, olefinic H). FABMS m/z 282 (M⁺ + H).

 $[1(R^*),3a\alpha,6a\alpha] - (+) - 1 - (3 - Cyclohexyl-3 - hydropropyl)$ tetrahydro-2, 5(1H, 3H)-cyclopentimidazoledione (18) $[1(S^*), 3a\alpha, 6a\alpha] \cdot (\pm) \cdot 1 \cdot (3 \cdot Cyclohexyl-3 \cdot hydroxy$ and propyl)tetrahydro-2,5(1H,3H)-cyclopentimidazoledione (19). The amine 15 (6.60 g, 23.4 mmol) in ethanol (140 mL) was treated with potassium cyanate (2.86 g, 35.4 mmol) in water (15 mL) at 0-5 °C and then 1 N aqueous hydrochloric acid (20 mL, 20 mmol). The mixture was stirred at 0-5 °C for 2 h and then at rt. After 18 h, the solvent was evaporated, the residue diluted with water and extracted with chloroform. The extract was dried then decolourised and the solvent was removed in vacuo to give crude N-(3-cyclohexyl-3-hydroxypropyl)-N-(4.4-ethylenedioxycyclopent-2-envl)-urea (16) as a tan glass (6.45 g ca. 85%). This material was dissolved in tetrahydrofuran (90 mL) and 2.0 N aqueous hydrochloric acid (75 mL) and the solution stirred at rt for 2 h. The solvent was removed in vacuo, the residue diluted with water and the oily mixture extracted with $CHCl_3$ (2×). The combined extracts were dried (MgSO₄) and evaporated to give a tan oil. This material was purified and separated into its diastereomers by column chromatography on silica. Elution with MeOH: EtOAc (1:9) gave 1.26 g (45%) of the less polar diastereomer 18 as a pale yellow glass. ¹H NMR (360 MHz, CDCl₃) δ: 0.90–1.48 (7H, complex m, aliphatic H), 1.58-1.92 (6 H, complex m, aliphatic H), 2.32-2.70 (4H, complex m, CH₂ C(O)CH₂), 2.85-2.95 (1H, m NCHH), 3.17-3.27 (1H, m, OCH), 3.56 (1H, d, J=6Hz, OH, exchang.), 3.67-3.80 (1H, m, NCHH), 4.28-4.42 (2H, m, NCHCHN), 5.85 (1H, s, NH, exchang.). FABMS m/z 281 (M⁺+H) R_f 0.3 (silica, EtOAc: MeOH, 9:1). 1.11 g (40%) of the more polar diastereomer 19 was also obtained as a pale yellow glass 'H NMR (360 MHz, CDCl₃) δ: 0.90-1.48 (7H, complex m, aliphatic H), 1.58-1.92 (6H, complex aliphatic H), 2.34-2.70 (4H, complex m, m. CH₂C(O)CH₂), 3.18-3.28 (1H, m, NCHH), 3.40-3.62 (2H, m, NCH<u>H</u>+OCH), 3.80 (1H, brs, OH, exchang.), 4.32-4.45 (2H, m, NCHCHN), 6.16 (1H, s, NH, exchang.). FABMS m/z 281 (M⁺+H), R_f 0.2 (silica, EtOAc: MeOH, 9:1).

 $[1(R^*), 3a\alpha, 5\alpha, 6a\alpha] \cdot (\pm) \cdot 1 \cdot [3 \cdot Acetyloxy) \cdot 3 \cdot cyclohexyl$ propyl]hexahydro-5-hydroxy-2(1H)-cyclopentimidazolone (20). The keto-alcohol 18 (less polar isomer) (1.18 g) was dissolved in pyridine (6.0 mL) and acetic anhydride (6.0 mL) and stirred at rt for 18 h. The solution was evaporated, the residue diluted with chloroform and washed with water, N hydrochloric acid, aqueous sodium bicarbonate solution, then finally water. After drying $(MgSO_4)$ the organic layer the solvent was removed in vacuo to give 1.14 g (84%) of $[1(R^*), 3a\alpha, 6a\alpha] - (\pm) - 1 - [3 - acetoxy - 3 - cyclohexylpropy]]$ tetrahydro-2,5(1H,3H)-cyclopentimidazoledione as a yellow glass. This keto-ester (1.14 g, 3.5 mmol) in ethanol (50 mL) was treated at +5 °C with sodium borohydride (250 mg, 6.6 mmol) with stirring. After 4 h at rt the ethanol was removed in vacuo, the residue diluted with water, and the product extracted into ether. The extract was dried $(MgSO_4)$, the solvent removed in vacuo and the residue chromatographed on silica. Elution with EtOAc: MeOH (19:1) gave 619 mg (54%) of 20 as a colourless oil. ¹H NMR (360MHz, CDCl₃) δ : 0.90–2.04 (17 H, complex m, aliphatic H), 2.05 (3H, s, CH₃), 2.48 (1H, brd, J=6 Hz, OH, exchang.), 2.88-3.68 (2H, m, NCH₂), 4.05-4.30 (2H, m, NCHCHN), 4.32–4.48 (1H, m, CHOH), 4.60 (1H, brs, NH, exchang.), 4.70-5.08 (1H, m, CHOAc). FABMS m/z 325 (M⁺ + H).

The relative stereochemistry of alcohol **20** was established by a full NMR analysis of the *O*-Tosyl derivative **35** which was prepared by an analogous series of reactions. Details of the preparation and assignments are given below.

 $[1(R^*), 3a\alpha, 5\alpha, 6a\alpha] \cdot (\pm \cdot 1 \cdot [3 \cdot (Acetyloxy octyl) hexa \cdot$ hydro-5-(4-methyl phenyl sulphonyloxy)-2(1H)-cyclopentimidazolone (35). $[1(R^*), 3a\alpha, 5\alpha, 6a\alpha] - (\pm) - 1 - [3-$ (Acetyloxy octyl) hexahydro-5-hydroxy-2(1H) cyclopentimidazolone (36), (312 mg) in dry pyridine (2.4 mL) was treated at rt with p-toluenesulphonyl chloride (240 mg). The orange solution was set aside at rt for 24 h then diluted with chloroform and washed with excess 2 N hydrochloric acid. The organic extract was dried (MgSO₄) and the solvent removed in vacuo and the residual gum purified by column chromatography (silica, elution with ether: methanol, 23:2) giving 0.28 g (60%) of the O-Tosyl derivative 35 as colourless crystals, mp 51-52 °C (from ether: hexane). ¹H NMR (360 MHz, CDCl₃) δ : 0.88 (3H, t, J=6.8 Hz, aliphatic CH_3), 1.20–1.36 (6H, m, 3× CH_2), 1.50–1.61 (2H, m, CH₂), 1.70-1.80 (2H, m, N-C-CH₂), 1.85-1.97 (1H, m, $J_{6\alpha,6\beta} = 15.0, J_{6\alpha,6\beta} = 6.6, J_{5\beta,6\beta} = 5.3$ Hz, H-6 β), 1.97 (111, 111, $J_{6\alpha,6\beta} = 15.0, J_{6\alpha,6\beta} = 6.6, J_{5\beta,6\beta} = 5.3$ Hz, H-6 β), 1.98–2.10 (2H, m, H_{4 α}, H_{4 β}), 2.22–2.30 (1H, m, $J_{6\alpha,6\beta} = 15.0, J_{5\beta,6\alpha} = 2.6, J_{6\alpha,6\alpha} = J_{4\alpha,6\alpha} = 2$ Hz, H6 α), 2.44 (3H, s, Arcut) = 2.72 (4H = 2.52) (2.44) (3H, s, Arcut) = 2.72 (2.52) (2.52 ArCH₃), 2.77-2.86 (1H, m, NCHH), 3.42-3.53 (1H, m, NCH<u>H</u>), 4.09–4.17 (1H, m, $J_{3a,6a} = 8.8$, $J_{3a,4\beta} = 6.6$ Hz, $J_{3a,4\alpha} = 2$ Hz, H-3a), 4.22-4.30 (2H, m, $J_{3a,6\alpha} = 8.8$, $J_{6a,6\beta} = 6.6, J_{6a,6\alpha} = 2$ Hz, H-6a + NH, exchang.), 4.84 (1H, quintet, J=6.3 Hz, CHOAc), 5.13-5.19 (1H, m, $J_{4\beta,5\beta} = J_{5\beta,6\beta} = 5.3$ Hz, $J_{4\alpha,5\beta} = J_{5\beta,6\alpha} = 2.6$ Hz, H-5 β), 7.34 (2H, d, J = 8.2 Hz, aromatics), 7.70 (2H, d, J = 8.2 Hz, aromatics).

NOE experiments: irradiation of the δ 4.13 signal (H-3a) gave enhancements of the δ 4.27 (H-6a) and δ 1.98–2.10 (H_{42,4β}) signals; irradiation of the δ 4.27 signal (H-6a) gave enhancements of the δ 4.13 (H-3a), 2.26 (H-6\alpha), 1.90 (H-6\beta), 1.74 (N-C-CH₂), 3.48 (NCHH and 4.84 (CHOAc) signals; irradiation of the δ 5.15 (H-5β) signal gave enhancements of the δ 1.98–2.10 (H₄₂₂H_{4β}), 2.26 (H-6α), 1.90 (H-6β), and 7.70 (aromatic) signals; irradiation of the δ 1.90 (H-6β) signal gave enhancements of the δ 1.98–2.10 (H-4) signals gave enhancements of the δ 1.98–2.10 (H-4) signals gave enhancements of the δ 1.98–2.10 (H-64) signal gave enhancements of the δ 1.98–2.10 (H-4) signals gave enhancements of the δ 1.98–2.10 (H-64) signals gave enhancements of the δ 1.90 (H-66), 2.82 (NCHH) and 5.15 (H-5β signals).

Spin decoupling expts: irradiation of the δ 3.48 (NC<u>H</u>H) signal simplified the multiplets at δ 2.82 (NC<u>H</u><u>H</u>) and 1.74 (N-C-CH₂); irradiation of the δ 4.13 (H-3a) signal simplified the multiplets at δ 1.98–2.10 (H-4) and 1.90 (H-6 β); irradiation of the δ 4.84 (CHOAc) signal simplified the δ 1.74 (N-C-CH₂) signal; irradiation of the δ 5.15 (H-5 β) signal simplified the δ 2.26 (H-6 α , dt, J=15, 2, 2Hz) and δ 1.90 (H-6 β , dd, J=15, 6.6 Hz) signals; irradiation of the δ 2.26 (H-6 α , dt, J=15.0, 2, 2 Hz) and 4.13 (H-3a, dt, J=8.8, 2, 2 Hz); irradiation of the δ 2.26 (H-6 α) signal simplified the δ 1.90 (H-6 β) and 4.27 (H-6 α) signals. FABMS m/z 467 (M⁺ + H). Anal. cald for C₂₃H₃₄N₂O₆S requires C, 59.2; H, 7.34; N, 6.02. Found C, 59.0; H, 7.52; N, 5.81%.

 $[1(R^*), 3a\alpha, 5\alpha, 6a\alpha] \cdot (\pm) \cdot 1 \cdot (3 \cdot Acetyloxy \cdot 3 \cdot cyclohexyl$ propyl) hexahydro - 5 - methanesulphonyloxy - 2(1H) cyclopentimidazolone (21). Methanesulphonyl chloride (0.70 mL, 9.0 mmol, redistilled) was added dropwise to a stirred solution of acetoxy-alcohol (20) (1.10 g, 3.4 mmol) in dry dichloromethane (20 mL) and dry pyridine (0.70 mL) at 0 °C. The reaction mixture was stirred at rt for 2 h, set aside at +5 °C for 16 h, and then ice-water (50 mL) added. The mixture was extracted with CH_2Cl_2 (3 ×), the extracts washed with brine, 5% hydrochloric acid, 5% NaHCO₃ solution, and finally brine. After drying (MgSO₄) the solvent was removed in vacuo and the residual oil chromatographed on silica. Elution with EtOAc: MeOH (9:1) and crystallisation from EtOAc-Et₂O yielded 0.68 g (50%) of **21**, mp 125 °C 'H NMR (200 MHz, CDCl₃) δ: 0.85-2.68 (17H, complex m, aliphatic H), 2.06 (3H, s, C-CH₃), 2.72-3.14 (1H, m, NCHH), 2.97 (3H, s, SCH₃), 3.24-3.52 (1H, m, NCHH), 4.03-4.36 (2H, m, NCHCHN), 4.50-4.84 (1H, m, CHOAc), 5.01-5.36 (2H, m, CHOMes+NH, 1H exchang.). FABMS m/z403 (M⁺ + H). Anal. cald for $C_{18}H_{30}N_2O_6S$ requires C, 53.7; H, 7.51; N, 6.96. Found C, 53.5; H, 7.68; N, 6.76%.

 $[1(R^*), 3a\alpha, 5\beta, 6a\alpha] - (\pm)$ -Methyl-4-[[1-(3-Acetyloxy 3-cyclohexylpropyl)octahydro-2-oxo-5-cyclopentimidazolyl]thio]butanoate (22). Thiobutyrolactone (326 mg, 3.2 mmol) was dissolved in methanolic sodium methoxide [from sodium (75 mg, 3.3 g.a.) and dry methanol (7.0 mL)] and set aside at rt for 2 h. The solvent was removed in vacuo and the residue dissolved in dry dimethylsulphoxide (7.0 mL). The mesylate 21 (0.60 g, 1.5 mmol) in dimethylsulphoxide (3.0 mL) was added in one portion to the thiolate solution prepared²⁰ above and the mixture set aside at rt for 18 h. The solution was then diluted with 0.5% aqueous sodium dihydrogen phosphate solution (45 mL) and the product extracted into ethyl acetate. The extract was washed with water, dried (MgSO₄), the solvent removed in vacuo and the product purified by column chromatography (silica, elution with methanol:ether, 3:100) giving 0.41 g (63%) of 22 as a colourless syrup. ¹H NMR [200 MHz, CDCl₃] δ: 0.80-2.20 (19 H, complex m, aliphatic H), 2.03 (3H, s, C-CH₃), 2.28-2.80 (4H, complex m, CH₂C(O), CH₂S), 2.85-3.87 (3H, m, NCH₂+CHS), 3.66 (3H, s, OCH₃), 4.08–4.35 (2H, m, NCHCHN), 4.63–5.04 (2H, m, CHOAc+NH, 1H exchang.). FABMS m/z 441 (M⁺ + H).

 $[1(R^*), 3a\alpha, 5\beta, 6a\alpha] - (+) - 4 - [[1 - (3 - Cyclohexy] - 3$ hydroxypropyl)octahydro-2-oxo-5-cyclopentimidazolyl]thio]butanoic acid (7). The ester 22 (0.40 g, 0.90 mmol) in methanol (8.5 mL) was treated with 2 N aqueous sodium hydroxide (5.2 mL, 10.4 mmol) and set aside at rt for 2 h. The solution was diluted with water, washed with ether, and the aqueous phase acidified to pH 3 with dilute hydrochloric acid. The product was extracted into chloroform, the extract dried (MgSO₄), the solvent removed in vacuo and the residue recrystallized from chloroform:ether giving 0.30 g (88%) of acid 7 as colourless prisms, mp 148-149.5 °C. 'Η NMR (360 MHz, CDCl₃) δ: 0.90-1.98 (17H, complex m, aliphatics), 2.12-2.19 (1H, m, H-4 α), 2.21–2.28 (1H, m, H-6 α), 2.42 (2H, t, J = 7.0Hz, CH₂CO), 2.60 (2H, t, J=7.0 Hz, CH₂S), 2.90-3.13 (NCHH), 3.19–3.32 (2H, m, CHS+OCH), 3.60–3.78 (1H, m, NCHH), 4.07–4.28 (2H, m, H-3a, H-6a), 4.3–5.2 (2H, rise in baseline, $2 \times OH$, exchang.), 5.80 (1H, brs, NH, exchang).

NOE experiments: irradiation of the δ 2.90–3.13 (NC<u>H</u>H) signal gave an enhancement of the δ 3.60–3.78 (NCH<u>H</u>), and δ 2.21–2.28 (H-6 α) signals and vice-versa; irradiation of the δ 5.80 (NH) signal gave an enhancement of the δ 4.24 (H-3a) and δ 2.12–2.19 (H-4 α) signals.

COSY2D and spin decoupling experiments allowed the following assignments: δ : 1.55 (H-6 β), 1.68 (H-4 β), 2.16 (H-4 α), 2.24 (H-6 α), 3.27 (H-5 α), 4.17 (H-6 α), 4.24 (H-3 α); *J* values 3a-N1=1, 4 α -6 α =1.8, 4 α -4 β =13.5, 6 α -6 β =13.7, 4 α -5 α =6.0, 4 β -5 α =6.0, 4 β -5 α =11.3, 5 α -6 α =6.0, 5 α -6 β =11.5, 3a-4 α =1, 3a-4 β =6.2, 6a-6 α =1, 6a-6 β =6.2, 3a-6a=8.7 Hz. Anal. cald for C₁₉H₃₂N₂O₄S requires C, 59.4; H, 8.39; N, 7.29. Found C, 59.2; H, 8.53; N, 7.09%

 $[1 (S^*), 3a\alpha, 5\beta, 6a\alpha] - (\pm) - 4 - [1 - (3 - Cyclohexyl - 3 - hydroxypropyl)octahydro-2-oxo-5-cyclopentimidazolyl-thio]butanoic acid (23). It was found that the sequence of reactions described above for the synthesis$

of acid 7 could be carried out on the mixture of diastereomeric ketones 18 and 19 utilising only a purification of the mesylate intermediates (21 and epimer) and final acid products. This gave a 36% overall yield mixture of 7 and 23, which were separated by column chromatography (silica 100 × wt. of mixture, CHCl₃:MeOH:HOAc, 93:3.5:3.5). The seperation of these acid diastereomers proved more difficult than that for ketone intermediates 18 and 19. Nevertheless this procedure gave a 6% overall yield of 23 as a colourless glass ¹H NMR (360 MHz, CDCl₃) δ: 0.90-1.98 (17H, complex m, aliphatics), 2.06-2.15 (1H, m, H-4 α), 2.16–2.22 (1H, m, H-6 α), 2.40 (2H, t, J=7.0 Hz, CH₂CO), 2.60 (2H, t, J = 7.0 Hz, CH₂S), 3.13–3.29 $(2H, m, NCHH + H-5\alpha), 3.30-3.42$ (1H, m, NCHH), 3.50-3.62 (1H, m, CHOH), 4.13-4.24 (2H, m, H-3a, H-6a), 6.10 (1H, brs, NH, exchang), 6.5-7.5 (2H, vbr peak, $2 \times OH$, exchang.).

NOE, COSY and spin decoupling experiments permitted the following assignments: δ 1.58 (H-6 β), 1.66 (H-4 β), 2.11 (H-4 α), 2.18 (H-6 α), 3.24 (H-5 α); $J 4\alpha - 6\alpha = 1.8, 4\alpha - 4\beta = 6\alpha - 6\beta = 13.5, 4\alpha - 5\alpha = 6.0, 4\beta - 5\alpha = 6.0$ 11.1, $5\alpha - 6\alpha = 6.0$, $5\alpha - 6\beta = 11.1$, $3a - 4\alpha = 1$, $3a - 4\beta = 6.0$, $6a-6\beta = 6.0.$ R_f 0.30 (Silica, CHCl₃: $6a-6\alpha = 1$, MeOH:HOAc, 93:3.5:3.5). Epimer 7 could more readily be obtained from the mixture of 7 and 23 by seeding it with pure 7 in ether and recrystallisation yield 12% of 7, mp 147–149 °C, R_f 0.32 (silica, CHCl₃: MeOH: HOAc), 93:3.5:3.5). The relative stereochemistry of 7 was confirmed by analogy with that of acid 37, obtained by a similar sequence of reactions.²⁶ This compound was more amenable to a full NMR analysis.

[1(*R**),3aα,5β,6aα]-(±)-4-[1-(3-Hydroxyoctyl)octahydro-2-oxo-5-cyclopentimidazolyl] thio] butanoic acid (37). Mp 79–80 °C. ¹H NMR (360 MHz, CDCl₃) δ: 0.89 (3H, t, *J*=6.8 Hz, CH₃), 1.22–1.38 (8H, m, $4 \times$ CH₂), 1.40–1.60 (2H, N-C-CHH, H-6_β), 1.62–1.74 (2H, m, N-C-CHH, H-4_β), 1.83–2.02 (2H, m, S-C-CH₂-C-CO₂), 2.17 (1H, br dd, *J*_{4x,4β}=13.5, *J*_{4x,5x}=5.8, *J*_{3a,4x} ca. 1, *J*_{4x,6x} ca. 2 Hz, H-4α), 2.25 (1H, br dd, *J*_{6x,6β}=13.5, *J*_{5,6x}5.8, *J*_{6a,6x} ca. 1, *J*_{4x,6x} ca. 2 Hz, H-6α), 2.38–2.54 (2H, m, CH₂CO₂), 2.63 (2H, t, *J*=7.0 Hz, SCH₂), 2.94–3.03 (1H, m, NCHH), 3.23–3.34 (1H, 7 line m, *J*_{4β,5x}=*J*_{5x,6β}=11.6, *J*_{4x,5x}=*J*_{5x,6x}=5.8 Hz, H-5α), 3.48–3.56 (1H, m, OCH), 3.62–3.74 (1H, m, NCHH), 4.19 (1H, br dd, *J*_{3a,6a}=8.8, *J*_{6a,6β}=6.2, *J*_{6a,6x} ca. 1Hz, H-6a), 4.25 (1H, br dd, *J*_{3a,6a}=8.8 *J*_{3a,4β}=6.2, *J*_{3a,4x} ca. 1Hz), 5.5–6.0 (2H, vbr peak, NH, OH, exchang.).

NOE Experiments: irradiation of the δ 2.17 (H-4 α) signal gave an enhancement of the δ 1.69 (H-4 $_{\beta}$), δ 3.28 (H-5 α) signals and δ 4.25 (H-3 α) signals; irradiation of the δ 2.25 (H-6 α) signal gave enhancements of the δ 1.55 (H-6 $_{\beta}$), 3.28 (H-5 α), 4.19 (H-6 α) and 2.98 (NCHH) signals; irradiation of the δ 3.28 (H-5 α) signal gave enhancements of the δ 2.17 (H-4 α), 2.25 (H-6 α) and 2.63 (SCH₂) signals; irradiation of the δ 1.55 (H-6 $_{\beta}$), 1.69 (H-4 $_{\beta}$), 2.98 (NCHH), and 3.68 (NCHH) signals;

irradiation of the δ 4.25 (H-3a) signal gave an enhancement of the δ 1.69 (H-4_{β}) signal; irradiation of the δ 5.5–6.0 (NH, OH) signal gave enhancements of the δ 4.25 (H-3a), 3.50 (OCH), 3.68 (NCH<u>H</u>) and δ 2.17 (H-4 α) signals.

Spin decoupling experiments: irradiation of the δ 2.25 (H-6 α) signal simplifies the multiplets at δ 1.55 (H-6_{β}), 4.19 (H-6a) and 3.28 (td, $J_{5\alpha,6\beta} = J_{4\beta,5\alpha} = 11.6$, $J_{4\alpha,5\alpha} =$ 5.8Hz, H-5 α); irradiation of the δ 3.28 (H-5 α) signal simplifies the multiplets at δ 1.55 (H-6_{β}), 1.69 (H-4_{β}), 2.17 (br d, $J_{4_{\alpha},4\beta}$ = 13.5 Hz, H-4 α), δ 2.25 (br d, $J_{6\alpha,6\beta} = 13.5$ Hz, H-6 α); irradiation of the δ 1.69 (H-4_{β}) signal simplifies the multiplets at δ 2.17 (H-4 α), 2.98 (NCHH), 3.28 (H-5 α), 3.68 (NCHH) and 4.25 (br d, $J_{3a,6a} = 8.8$ Hz, H-3a); irradiation of the δ 1.55 signal (H-6_{β}) simplifies the multiplets at δ 2.25 (br d, $J_{5\alpha,6\alpha} = 5.8$ Hz, H-6 α), 3.28 (H-5 α), 3.50 (OCH) and 4.19 (br d, $J_{3a,6a} = 8.8$ Hz, H-6a); irradiation of the δ 2.17 (H-4 α) signal simplifies the multiplets at δ 1.69 (H-4 $_{\beta}$), 3.28 (td, $J_{5x,6\alpha} = J_{4\alpha,5\alpha} = 5.8$; $J_{5x,6\beta} = 11.6$ Hz, H-5 α) and 4.25 (H-3a). Anal. cald for $C_{18}H_{32}N_2O_4S$ requires C, 58.0; H, 8.66; N, 7.52. Found C, 57.8; H, 8.72; N, 7.36%.

Benzaldehyde 1,4-dioxaspiro [4.4] non-6-en-8-ylhydrazone (25). The allylic bromide 14^{15} prepared from ketal 10 (22.5 g, 0.178 mol), was dissolved in toluene (50 mL) and the solution cooled to -40 °C with stirring. Benzaldehyde hydrazone 24²² (17.0 g, 0.141 mol) in toluene (50 mL) was added rapidly over 5 min, followed by triethylamine (25 g). The solution was warmed slowly to 15 °C and stirred at this temperature for 16 h. The mixture was then heated to a gentle reflux for a further 5 h. After cooling, the triethylamine hydrobromide formed was filtered off under reduced pressure and the resulting filtrate concentrated in vacuo. The residual dark oil was chromatographed on silica (450 g). Elution with ether-hexane (1:1) and then ether gave 18.9 g (55%) of 25 as a yellow oil. ¹H NMR (90 MHz, CDCl₃) δ 1.88 (1H, dd, J=14, 5 Hz, C-C<u>H</u>H-C), 2.46 (1H, dd, J = 14, 7 Hz, C-CH<u>H</u>-C), 3.84 (4H, s, OCH₂CH₂O), 4.36-4.84 (1H, m, CHN), 5.28 (1H, brs, NH), 5.68-5.92 (1H, m, olefinic H), 5.93-6.20 (1H, m, olefinic H), 7.04-7.82 (5H, m, aromatics), 7.48 (1H, s, CHAr). FABMS m/z 245 (M⁺+H). This compound decomposes on standing at rt for several days, but could be stored satisfactorily at -20 °C for 3 days. It is important to purify the crude product as soon as possible.

(\pm)-1-Benzylideneamino-tetrahydrocyclopentimidazol-2,5(1*H*,3*H*)-dione (27). The hydrazone 25 (7.42 g, 30.4 mmol) and pyridine (3.0 mL, 37.1 mmol) were added to toluene (175 mL) and the resulting solution cooled to 0 °C. To this stirred mixture, a solution of phosgene in toluene (12%, 40 mL, 44.2 mmol) was rapidly added over 10 min, after which the solution was warmed to 15 °C and stirred for a further 20 min. Aqueous ammonia solution (190 mL, d=0.88) was added to the solution at 0 °C over 10 min and after stirring for a further 30 min, water (200 mL) was added. The organic layer was separated, dried (sodium evaporated in vacuo sulphate) and to give Benzaldehyde 2-(1,4-dioxaspiro[4.4]non-6-en-8-yl)semicarbazone (26) as a brown oil (7.85 g, 90%). An acetone solution (150 mL) of semicarbazone 26 (7.85 g) was stirred at 15 °C while camphorsulphonic acid (0.75 g) was added portionwise over 10 min. The mixture was heated at reflux for 5 h, cooled and concentrated in vacuo. Water (150 mL) and chloroform (100 mL) were added and the organic phase separated. The latter was washed with dilute aqueous NaHCO₃ solution (100 mL), dried (MgSO₄) and concentrated in vacuo to give a brown oil, (21.0 g). This material was chromatographed on silica (260 g). Elution with CHCl₃: MeOH (19:1) gave 3.66 g (55%) of 27, mp 49–52 °C ¹H NMR (200 MHz, (CD₃)₂SO) δ: 2.12-2.35 (2H, m, CH₂C(O)), 2.65-3.04 (2H, m, CH₂C(O)), 4.35-4.49 (1H, m, CHN), 4.73-4.90 (1H, m, CHN), 7.29-7.52 (3H, m, aromatics), 7.62-7.70 (2H, m, aromatics), 7.74 (1H, s, CHPh). FABMS m/z244 (M⁺+H). Anal. cald for $C_{13}H_{13}N_3O_2$ requires C, 64.2; H, 5.39; N, 17.3. Found C, 64.4; H, 5.50; N, 17.1%.

 $[3a\alpha, 5\alpha, 6a\alpha] \cdot (\pm) \cdot 1 \cdot Benzylideneamino \cdot 5 \cdot hydroxyhexa$ hydrocyclopentimidazol-2(1H)-one (28). The ketone 27 (3.60 g, 14.8 mmol) was dissolved in ethanol (60 mL) and the solution cooled to 0 °C while sodium borohydride (0.80 g, 21.1 mmol) was added portionwise over 15 min. When addition was complete the mixture was stirred at rt for 1 h. The solvent was removed in vacuo, water (100 mL) and chloroform (100 mL) added and the organic phase separated. After drying (Na₂SO₄) concentration in vacuo gave the crude product as a brown syrup. This was purified by column chromatography (silica, CHCl₃:MeOH, 9:1) to give 3.09 g (85%) of 28 as a yellow solid, mp 206-207 °C. ¹H NMR (200 MHz, (CD₃)₂SO) δ: 1.62–1.92 (2H, m, C-CH₂-C), 1.90-2.22 (2H, m, C-CH₂-C), 4.02-4.15 (1H, m, CHN), 4.16-4.27 (1H, m, OCH), 4.40-4.52 (1H, m, CHN), 5.60 (1H, d, J=5 Hz, OH, exchang.), 7.26-7.48 (3H, m, aromatics), 7.62-7.73 (3H, m, aromatics + CHPh). FABMS m/z 246 (M⁺ + H). Anal. cald for C₁₃H₁₅N₃O₂ requires C, 63.7; H, 6.16; N, 17.1. Found C, 63.5; H, 6.37; N, 16.8%.

[$3a\alpha, 5\alpha, 6a\alpha$]-(\pm)-1-Amino-5-hydroxyhexahydrocyclopentimidazole-2(1*H*)-one (29). The hydrazone 28 (3.00 g, 12.2 mmol) was added to a solution of ethanol (70 mL) and acetic acid (0.7 mL) containing 10% Pd on carbon (1.3 g) and the resulting mixture stirred at 70 °C under 90 atmospheres of hydrogen for 3 days. The catalyst was removed by filtration through Celite and concentration of the filtrate in vacuo gave 1.73 g (90%) of 29 as an unstable yellow oil FABMS *m*/*z* 158 (M⁺ + H). This material was utilised for the following reaction as soon as possible—it appears unstable to air and decomposes on standing.

 $[1(R^*,S^*),3a\alpha,5\alpha,6a\alpha] \cdot (\pm) \cdot 5$ -Hydroxy-1-(2-cyclohexyl-2-acetoxyethylideneamino)-hexahydrocyclopentimidazole-2(1H)-one (30). A solution of the hydrazine **29** (1.73 g, 11.0 mmol) and 2-acetoxy-2-cyclohexylacetaldehyde³⁰ (2.06 g, 11.2 mmol)) in methanol (175 mL) containing sodium acetate (1.03 g, 12.6 mmol) was stirred at 25 °C under a nitrogen atmosphere for 16 h. The alcohol was removed in vacuo and water (50 mL) and chloroform (100 mL) added. The organic phase was separated and washed with brine (50 mL), then dried (Na₂SO₄). Concentration of the filtrate in vacuo gave a yellow oil. Column chromatography of the crude product (silica, CHCl₃:MeOH (12:1) gave 2.71 g (75%) of **30** (mixture of diastereomers) as a colourless gum, FABMS m/z 324 (M⁺ + H). This compound decomposed on standing at rt for a few days.

 $[1(R^*), 3a\alpha, 5\alpha, 6a\alpha] \cdot (\pm) \cdot 5$ -Methanesulphonyloxy-1-(2cyclohexyl - 2 - acetyloxyethylidene - amino) hexahydro cyclopentimidazole - 2 (1H) - one (31) and $[1(S^*), 3a\alpha,$ $5\alpha, 6a\alpha$] - (\pm) - 5 - Methanesulphonyloxy - 1 - (2 - cyclo hexyl - 2 - acetyloxyethylidene - amino) hexahydrocyclo pentimidazole-2(1H)-one (32). To a solution of the acetoxy-alcohol 30 (2.60 g, 8.05 mmol) and pyridine (3.6 g) in dichloromethane (50 mL) at 0 °C under an atmosphere of nitrogen was added methanesulphonyl chloride (1.25 mL, 16.2 mmol). After stirring for 3 days at 25 °C, water (50 mL) was added and the organic layer separated. The latter was washed with 1 N HCl (50 mL) and again separated and dried (MgSO₄). Evaporation in vacuo gave 3.16 g of crude product containing both diastereomers as a tan oil. The individual diastereomers, were separated by flash chromatography (silica, EtOAc: MeOH, 9:1) to give 0.56 g (35%) of **31** as white crystals, mp 147-148 °C. NMR (360 MHz, $CDCl_3$ δ: ^{1}H 0.94 -1.38 (5 H, complex m, aliphatics), 1.56-1.90 (6 H, complex m, aliphatics), 1.93-2.18 (2H, m, H-4β, H-6β), 2.08 (3H, s, C-CH₃), 2.41 (1H, dd, $J_{4\alpha,4\beta} = 15$, $J_{4\alpha,5\beta} = 2.5$ Hz, H-4 α), 2.53 (1H, dd, $J_{6\alpha,6\beta} = 15$, $J_{5\beta,6\alpha} = 2.5$ Hz, H-6 α), 2.97 (3H, s, S-CH₃), 4.36 (1H, dd, $J_{3\alpha,6\alpha} = 8.8$, $J_{3a,3b} = 6.6$ Hz, H-3a), 4.54 (1H, dd, $J_{3a,6a} = 8.8$, $J_{6a,6b} = 6.6$ Hz, H-6a), 5.17 (1H, brt, J=5 Hz, CHOAc), 5.26–5.35 (1H, m, $J_{4\beta,5\beta} = J_{5\beta,6\beta} = 5$, $J_{4\alpha,5\beta} = J_{5\beta,6\alpha} = 2.5$ Hz, H-5 β), 5.92 (1H, s, NH, exchang.), 7.22 (1H, d, J=6 Hz, CH = N).

NOE experiments: irradiation of the δ 2.41 (H-4 α) and δ 2.53 (H-6 α) signals gave an enhancement of the δ 5.30 (H-5 β) signal and vice-versa. FABMS *m/z* 402 (M⁺ + H). *R_f* 0.3 (silica, EtOAc:MeOH, 9:1). Anal. cald for C₁₇H₂₇N₃O₆S requires C, 50.9; H, 6.78; N, 10.5. Found C, 50.6; H, 6.89; N, 10.3%.

Also obtained from this experiment were 0.58 g (36%) of **32**, mp 78–82 °C. ¹H NMR (360 MHz, CDCl₃) δ : 0.94–1.38 (5H, complex m, aliphatics), 1.56–1.90 (6 H, complex m, aliphatics), 1.95–2.20 (2H, m, H-4 β , H-6 β), 2.08 (3H, s, C-CH₃), 2.40 (1H, dd, $J_{4x,4\beta}=15$, $J_{4x,5\beta}=2.5$ Hz, H-4 α), 2.50 (1H, dd, $J_{6\alpha,6\beta}=15$, $J_{5\beta,6\alpha}=2.5$ Hz, H-6 α), 2.97 (3H, s, S-CH₃), 4.36 (1H, dd, $J_{3a,6a}=8.8$, $J_{3a,3\beta}=6.6$ Hz, H-3a), 4.54 (1H, dd, $J_{3a,6a}=8.8$, $J_{6a,6\beta}=6.6$ Hz, H-6a), 5.20 (1H, brt, J=6Hz, CHOAc), 5.25–5.36 (1H, m, H-5 β), 5.93 (1H, s, NH, exchang.) 7.18 (1H, d, J=6 Hz, CH=N). FABMS m/z 402 (M⁺ + H). R_f 0.2

(silica, EtOAc: MeOH, 9:1). Anal. cald for $C_{17}H_{27}N_3O_6S$ requires C, 50.9; H, 6.78; N, 10.5. Found C, 50.6; H, 6.99; N, 10.2%.

 $[1(R^*), 3a\alpha, 5\beta, 6a\alpha] \cdot (\pm)$ -Methyl-4-1-(2-acetyloxy-2cyclohexylethylideneamino) octahydro-2-oxo-5-cyclopentimidazolyl]thio]butanoate (33). Under an atmosphere of nitrogen, thiobutyrolactone (0.26 g, 2.55 mmol) was added to methanolic sodium methoxide (from sodium, 60 mg, 2.61 g.a) in dry methanol (6 mL) at 25 °C and the resulting solution²⁰ stirred for 2 h. The solvent was removed in vacuo and the residue dissolved in dry dimethylsulphoxide (5 mL) under nitrogen. The mesylate **31** (0.50 g, 1.25 mmol) in dimethylsulphoxide (5 mL) was added in one portion to the thiolate solution and the resulting mixture stirred at 25 °C for 16 h. The solution was diluted with water (100 mL) and the crude product extracted into ether. After drying (Na_2SO_4) , the organic extract was concentrated in vacuo to give an oil. Column chromatography (silica, EtOAc: MeOH, 95:5) gave 274 mg (50%) of 33 as a colourless oil ¹H NMR (360 MHz, CDCl₃ δ: 0.93-1.37 5H, complex m, aliphatics), 1.53-1.99 (10 H, complex m, aliphatics), 2.08 (3H, s, C-CH₃), 2.18 (1H, dd, $J_{4\alpha,4\beta} = 13$, $J_{4\alpha,5\alpha} = 6$ Hz, H-4 α), 2.26 (1H, dd, $J_{6\alpha,6\beta} = 13$, $J_{5\alpha,6\alpha} = 6$ Hz, H-6 α), 2.41 (2H, t, J = 7 Hz, CH₂CO), 2.60 $(2H, t, J=7 Hz, SCH_2), 3.15-3.37 (1H, m, H-5\alpha), 3.67$ $(3H, s, OCH_3), 4.25-4.37 (1H, dd, J_{3a,6a}=9, J_{3a,4\beta}=6 Hz,$ H-3a), 4.37–4.51 (1H, dd, $J_{3a,6a}=9$, $J_{6a,6\beta}=6$ Hz, H-6a), 5.21 (1H, t, J=6 Hz, CHOAc), 5.52 (1H, brs, NH, exchang.), 7.09 (1H, d, J=6 Hz, CH = N). FABMS m/z440 $(M^+ + H)$.

 $[1(R^*), 3a\alpha, 5\beta, 6a\alpha] - (+) - 4 - [[1 - (2 - Cyclohexyl - 2 - hy$ droxyethylideneamino)octahydro-2-oxo-5-cyclopentimidazolyl]thio]butanoic acid (8). The ester 33 (0.26 g, 0.60 mmol) was dissolved in methanol:water (2:1, 8 mL) and to the stirred solution was added lithium hydroxide (0.10 g, 2.4 mmol). The resulting mixture was stirred at rt for 3 h. The pH was adjusted to 5.0 with 2 N HCl, water (20 mL) was added and the product was extracted into chloroform (50 mL). After drying (Na₂SO₄) the solvent was removed in vacuo to give 0.19 g (86%) of **8**, mp 176–178 °C (dec) (CHCl₃–Et₂O) (1.15 g). 'H NMR (360 MHz, $(CD_3)_2SO)$ δ : 0.88–1.87 (15H, complex m, aliphatics), 1.96-2.04 (1H, m, H-4 α), 2.08-2.17 (1H, m, H-6 α), 2.31 (2H, t, J = 7.0 Hz, CH_2CO_2), 2.56 (2H, t, J = 7.0Hz, CH₂S), 3.02-3.14 (1H, m, H-5α), 3.70-3.78 (1H, m, CHOH), 4.12-4.19 (1H, m, H-3a), 4.40-4.45 (1H, m, H-6a), 4.91 (1H, d, J = 4.5 Hz, OH, exchang.), 6.83(1H, d, J=6.6 Hz, CH=N), 7.01 (1H, brs, NH,exchang.), 11.8 (1H, brs, CO₂H, exchang.).

Spin decoupling experiments: on irradiation of the δ 7.01 (NH) signal the δ 4.17 (H-3a) signal changed to ddd ($J_{3a,6a} = 8.2$, $J_{3a,4\beta} = 6$, $J_{3a,4\alpha} = 1$ Hz); resolution enhancement of the NH signal (δ 7.01) showed it as a d ($J_{3a,NH} = 1.5$ Hz): on irradiation of the δ 4.17 (H-3a) signal the δ 3.08 (H-5 α) signal changed to dt ($J_{5\alpha,6\beta} = J_{4\beta,5\alpha} = 11.3$, $J_{5\alpha,6\alpha} = J_{4\alpha,5\alpha} = 5.8$ Hz), the δ 2.01 (H-4 α) signal changed to dddd ($J_{4\alpha,4\beta} = 13.3$, $J_{4\alpha,5\alpha} = 5.8$,

 $J_{4\alpha,6\alpha} = 1.8$ Hz; $J_{3a,4\alpha} = 1$ Hz) and the δ 1.622 (H-4 β) signal changed to dd ($J_{4\alpha,4\beta} = 13.3$, $J_{4\beta,5\alpha} = 11.3$ Hz; $J_{3a,4\beta} = 6$ Hz); on irradiation of the δ 4.44 (H-6a) signal the δ 2.13 (H-6 α) signal changed to dddd ($J_{6\alpha,6\beta} = 13.3$, $J_{5\alpha,6\alpha} = 5.8$, $J_{4\alpha,6\alpha} = 1.8$ Hz; $J_{6a,6\alpha} = 1$ Hz) and the δ 1.632 (H-6 β) signal changed to dd ($J_{6\alpha,6\beta} = 13.3$, $J_{5\alpha,6\beta} = 11.3$ Hz; $J_{6a,6\beta} = 6.6$ Hz). R_f 0.28 (Silica, CHCl₃: MeOH:HOAc, 90:5:5). Anal. cald for C₁₈H₂₉N₃O₄S requires C, 56.4; H, 7.62; N, 11.0. Found C, 56.2; H, 7.86; N, 10.7%.

 $[1(S^*), 3a\alpha, 5\beta, 6a\alpha] \cdot (\pm) - 4 \cdot [[1 - (2 - Cyclohexyl - 2 - hy$ droxyethylideneamino)octahydro-2-oxo-5-cyclopentimidazolyl]thio]butanoic acid (34). This acid was obtained from mesylate 32 by an identical series of reactions to that used for the preparation of 8. Compound 34 was obtained in 16% overall yield as a white wax. ¹H NMR (360 MHz, CDCl₃) δ : 0.97–2.10 (15H, complex m, aliphatics), 2.20-2.40 (2H, m, H-6a, H-4 α), 2.45 (2H, t, J = 7 Hz, CH₂CO), 2.60 (2H, t, J = 7Hz, CH_2S), 3.20–3.35 (1H, m, H-5 α), 4.08–4.18 (1H, m, OCH), 4.30-4.40 (1H, m, H-3a), 4.42-4.52 (1H, m, H-6a), 7.06 (1H, d, J=6 Hz, CH=N). The ¹H NMR spectrum of 8 in CDCl₃ was identical to the above except for the CH==N (δ 7.12) and OCH signal (δ 4.18–4.27). FABMS m/z 384 (M⁺ + H) R 0.28 (Silica, CHCl₃: MeOH: HOAc, 90:5:5).

 $[1(R^*), 3a\alpha, 5\beta, 6a\alpha] \cdot (\pm) \cdot 4[[-1 \cdot (2 \cdot cyclohexyl - 2 \cdot hy$ droxyethylamino)-octahydro-2-oxo-5-cyclopentimidazolyl]thiobutanoic acid (9). The hydrazone 8 (50 mg; 0.13 mmol) was dissolved in a mixture of methanol (1 mL) and acetic acid (1 mL) at rt and to the stirred solution was added sodium cyanoborohydride (12 mg, 0.19 mmol). After 1 h, the solution was concentrated in vacuo, water and chloroform were added and the organic phase separated. After drying (Na_2SO_4) the solvent was removed in vacuo to give 45 mg (90%) of 9 as a white wax ¹H NMR (200 MHz, $CDCl_3$) δ : 0.91-2.02 (15H, complex m, aliphatics), 2.05-2.22 (2H, m, H-4 α , H-6 α), 2.46 (2H, t, J=7 Hz, CH₂CO), 2.52-2.76 (3H, m, CH₂S+NC<u>H</u>H), 2.95 (1H, dd, J = 14, 2 Hz, NCH<u>H</u>), 3.11–3.42 (2H, m, CHS+OCH), 4.11-4.30 (2H, m, NCHCHN), 5.87 (1H, brs, NH, exchang.), 5.0-6.5 (2H, vbr baseline rise, $2 \times OH$, exchang.). FABMS m/z 386 (M⁺ + H). R_f 0.30 (silica, CHCl₃: MeOH: HOAc, 90:5:5).

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18. The less polar isomer is 18; the $R_{/S}$ of 18 and 19 are 0.3 and 0.2 (silica, ethyl acetate:methanol, 9:1) respectively.

19. NOE and spin decoupling experiments helped to provide a detailed analysis of the 360 MHz ¹H NMR spectrum (CDCl₃) of the 5 α -O-tosylate **35**. This analogue was prepared by tosylation of **36**, which was obtained by an identical series of reactions to that used for synthesizing **20**. Particularly diagnostic was the enhancement between H-3a and H-4_{β}, H-4_{β} and H-5_{β}, H-5_{β} and H-6_{β}, H-6_{β} and H-6a, and between H-6a and H-3a; δ ppm H-3a (4.13), H-4_{β} (2.01), H-5_{β} (5.15), H-6_{β} (1.90), H-6a (4.27); $J_{3a,4\beta}=J_{6a,6\beta}=6.6$, $J_{4\beta,5\beta}=J_{5\beta,6\beta}=5.3$, $J_{4\pi,5\beta}=J_{5\beta,6\gamma}=2.6$, $J_{4\pi,4\beta}=J_{6\pi,6\beta}=15.0$, J_{3a} , $J_{6a}=8.8$ Hz.



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21. Based on the nOe, spin decoupling experiments, and analysis of the 'H NMR spectrum of **37**, prepared as for 7. Enhancements observed between H-3a and H-4_{β}, H-6a and H-6_{β}, H-5_{χ} and H-6_{χ}, H-5_{χ} and H-4_{χ}. δ (CDCl₃) ppm H-3a (4.25), H-4_{χ}(2.17), H-4_{β} (1.69), H-5_{χ} (3.28), H-6a (4.19), H-6_{χ} (2.25), H-6_{β} (1.55); $J_{5\chi,6\beta}=J_{5\chi,4\beta}=11.6$, $J_{5\chi,4\chi}=J_{5\chi,6\chi}=5.8$, $J_{3\alpha,4\beta}=6.2$, $J_{4\chi,4\beta}=J_{6\chi,6\beta}=13.5$, $J_{3\alpha,6\alpha}=8.8$ Hz.

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23. The ¹H NMR spectra of **31** and **33** were analysed in detail as before and the chemical shifts and coupling constants of the cyclopentane ring protons were similar to those of **35** and 7, respectively.

24. Flash chromatography was employed; the R_{i} s of **31** and **32** are 0.3 and 0.2 (silica, ethyl acetate:methanol, 19:1).

25. Some preliminary experimental details for Scheme 2 are given in Eur. Pat. Appl. 458 642 (*Chem. Abstr.* 1992, *116*; 106293b).

26. Some preliminary experimental details for Scheme 1 are given in Eur. Pat. Appl. 46 597 (*Chem. Abstr.* 1982, 97; 144568w).

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29. Interestingly N-alkyl derivatives of 4, 8 and 9 display lower efficacy agonism than the parent compound and in the latter two series DP-receptor selectivity is retained. In addition sulphide 37 (mp 79–80 °C) is a much less potent PG mimetic (0.1×7 , platelets) than 7. SAR Studies would be facilitated by the synthesis and pharmacological evaluation of olefins 38 and 39. To date olefin 38 has only been obtained as a mixture of *E*- and *Z*-isomers (by reaction of 18 with Ph₃P(CH₂)₄ CO₂H Br (5 equiv), KOCMe₃ (10 equiv), dry benzene, N₂, reflux and purification by the methyl ester). Attempts to obtain 39 by a similar route have so far proved unsuccessful. Preliminary evaluation of 38 indicate it is a weak non-selective DP-agonist.



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