ORIGINAL RESEARCH

Synthesis and antiproliferative properties of novel naringenin derivatives

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Abstract In order to improve the antiproliferative activity of naringenin, a naturally occurred flavonoid in citrus fruits, a series of naringenin derivatives with a tertiary amino side chain were prepared. The antiproliferative activities of these naringenin derivatives were evaluated on four human cancer cell lines, namely, MCF-7, HCT116, Hela, and A549. Compounds **4a**, **9a**, and **10a** exhibited remarkably enhanced growth inhibition activity. Based on the observed results, the structure–activity relationship of these derivatives was discussed.

Keywords Naringenin · Antiproliferative activity · Synthesis · Derivatives

Introduction

Naringenin (NG) is one of the most abundant flavonoids in citrus fruits (Kawaii et al. 1999). It has been shown to exert a variety of biological effects, such as antioxidant, antiinflammatory, cholesterol-lowing, antitumor, etc (Lee et al. 1999; Chen et al. 2002; Fuhr et al. 1993; Benavente-Garcia

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et al. 1997; Heo et al. 2004). Owing to its low-toxicity, (Moon et al. 2006) the anticancer activity of NG is especially attractive. Both in vitro and in vivo studies had revealed that NG can effectively inhibit the growth of several types of human tumor cell lines (Kanno et al. 2005; Arul and Subramanian 2013; Sabarinathan et al. 2010). Mechanistic studies indicated that the proapoptotic activity of NG on cancer cell lines may involve the inhibition of PI3K/Akt signaling pathways and caspase-3 activation (Sabarinathan and Vanisree 2013; Bak et al. 2011; Park et al. 2008). However, the antineoplastic activity of NG is not strong enough for clinical application. Furthermore, the clinical potential of NG is seriously restricted by its low oral bioavailability (Semalty et al. 2010; Felgines et al. 2000). Several NG derivatives had been designed in order to improve the anticancer potential (Yoon et al. 2013; Lee et al. 2007; Liu et al. 2013; Kim et al. 2007). For instance, NG derivatives modified at position 7 with bulky substituents exhibited enhanced inhibitory effects on HCT116 human colon cancer cells and Human Lung epithelial carcinoma A549 cells (Yoon et al. 2013; Lee et al. 2007).

In this study, eleven NG derivatives with an aliphatic tertiary amino side chain were synthesized. Their antiproliferative effects were evaluated on four human cancer cell lines, namely MCF-7, HCT116, Hela, and A549. Based on the observed results, the structure–activity relationship of these derivatives was discussed.

Material and methods

Chemistry

All chemicals were of analytical grade and used as received unless otherwise stated. ¹H and ¹³C NMR spectra were

recorded on a Bruker DRX 400 NMR instrument. MS were recorded on a Qstar mass spectrometer.

Synthesis of 7-(2-Bromoethoxy)-5-hydroxy-2-(4hydroxyphenyl)chroman-4-one (1)

A mixture of NG (2.72 g, 10 mmol), potassium carbonate (0.69 g, 5 mmol) and 1,2-dibromoethane (21.6 mL, 250 mmol) in 100 mL DMF was stirred at 50 °C for 3 days. The resultant mixture was cooled to room temperature and filtrated. The filtrate was mixed with 1 L H₂O, and then extracted with ethyl acetate (150 mL \times 3). The organic layer was dried over Na₂SO₄ and concentrated. The residue was purified with a silica gel column and was eluted with Petroleum ether/EtOAc (2:1) to afford 1 (0.95 g, 25%): mp 180.0–181.0 °C. ¹H nuclear magnetic resonance (NMR) (400 MHz, dimethylsulfoxide (DMSO)-d₆): δ 2.73 (dd, J =2.4, 17.2 Hz, 1H, H-3cis), 3.33 (m, 1H, H-3trans), 3.78 (t, J = 5.2 Hz, 2H, H-2"), 4.38 (t, J = 5.2 Hz, 2H, H-1"), 5.49 (dd, J = 2.4, 14.4 Hz, 1H, H-2), 6.10 (d, J = 1.6 Hz, 1H, H-8), 6.12 (d, J = 1.6 Hz, 1H, H-6), 6.79 (d, J = 8.4 Hz, 2H, H-3',5'), 7.33 (d, J = 8.4 Hz, 2H, H-2',6'), 9.61 (s, 1H, 4'-OH), 12.10 (s, 1H, 5-OH).

Synthesis of 7-(3-chloropropoxy)-5-hydroxy-2-(4hydroxyphenyl)chroman-4-one (**2**)

According to same procedure as preparation of **1**, compound **2** was obtained as a pale yellowish solid, yield 58%, mp 96.0–98.0 °C. ¹H NMR (400 MHz, CDCl₃): δ 2.19–2.25 (m, 2H, H-2″), 2.79 (dd, J = 3.2, 17.2 Hz, 1H, H-3*cis*), 3.09 (dd, J = 10.2, 17.2 Hz, 1H, H-3*trans*), 3.71 (t, J = 6.4 Hz, 2H, H-3″), 4.13 (t, J = 6.0 Hz, 2H, H-1″), 5.36 (dd, J = 2.8, 12.8 Hz, 1H, H-2), 6.04 (d, J = 2.4 Hz, 1H, H-8), 6.07 (d, J = 2.4 Hz, 1H, H-6), 6.87–6.91 (m, 2H, H-3′, 5′), 7.32–7.35 (m, 2H, H-2′, 6′).

Synthesis of 7-methoxy-5-hydroxy-2-[4-(2-bromoethoxy) phenyl]chroman-4-one (7)

To a solution of 7-methoxy-5-hydroxy-2-(4-hydroxyphenyl) chroman-4-one (4.29 g, 15 mmol) in 150 mL acetone, potassium carbonate (2.07 g, 15 mmol) and 1,2-dibromoethane (56.4 g, 0.3 mol) was added successively. The resulting mixture was stirred at 80 °C for 60 h. Acetone was removed by rotavaper. 100 mL ethyl acetate was added to the residue, and washed by H₂O (20 mL × 3). The organic solution was concentrated. The residue was purified with a silica gel column and was eluted with Petroleum ether/EtOAc 5:1 to afford 7 (1.14 g, 19%): mp 125.5–126.7 °C. ¹H NMR (400 MHz, CDCl₃): δ 2.79 (dd, J = 2.4, 16.4 Hz, 1H, H-3*cis*), 3.09 (dd, J = 13.6, 16.4 Hz, 1H, H-3*trans*), 3.66 (t, J = 6.0 Hz, 2H, H-2″), 3.81(s, 3H, OCH₃), 4.32 (t, J = 6.0 Hz,

2H, H-1"), 5.37 (dd, J = 12.4, 14.4 Hz, 1H, H-2), 6.05 (d, J = 2.4 Hz, 1H, H-8), 6.08 (d, J = 2.4 Hz, 1H, H-6), 6.97 (d, J = 8.0 Hz, 2H, H-3', 5'), 7.39 (d, J = 8.0 Hz, 2H, H-2', 6').

Synthesis of 7-methoxy-5-hydroxy-2-[4-(2-choloropropoxy) phenyl]chroman-4-one (8)

According to same procedure as preparation of **7**, Compound **8** was obtained as a white solid, yield 24%: mp 88.0–90.0 °C. ¹H NMR (400 MHz, CDCl₃): δ 2.22–2.28 (m, 2H, H-2"), 2.79 (dd, J = 3.2, 17.2 Hz, 1H, H-3*cis*), 3.10 (dd, J = 13.2, 17.2 Hz, 1H, H-3*trans*), 3.78 (t, J = 6.4 Hz, 2H, H-3"), 3.81 (s, 3H, OCH₃), 4.14 (t, J = 6 Hz, 2H, H-1"), 5.37 (dd, J = 3.2, 13.2 Hz, 1H, H-2), 6.04 (d, J = 2.4 Hz, 1H, H-8), 6.07 (d, J = 2.4 Hz, 1H, H-6), 6.95–6.97 (m, 2H, H-3', 5'), 7.37–7.39 (m, 2H, H-2', 6').

General procedure for the preparation of **3a–3d**, **4a–4c**, **9a**, **9b**, **10a**, **10b**

To a solution of the halide compound (1, 2, 7 or 8, 1 mmol) in 5 mL DMF, a secondary amine (5 mmol) was added. The resulting solution was stirred at 80 °C for 70 min, then cooled to room temperature. The solution was diluted with 30 mL H₂O and extracted with ethyl acetate (10 mL \times 3). The combined organic phase was concentrated and the resulting residue was purified on a silica gel column eluted with Petroleum ether/EtOAc to afford the substituted product.

7-(2-Diethylaminoethoxy)-5-hydroxy-2-(4-hydroxyphenyl) chroman-4-one (**3a**)

White solid, yield 62%, $R_f = 0.35$ (MeOH/CH₂Cl₂ = 1:10, v/v), mp 121.0–123.0 °C. ¹H NMR (400 MHz, DMSO-d₆): δ 0.95 (t, J = 6.8 Hz, 6H, H-2^{'''}), 2.49–2.54 (tetra, J = 6.8Hz, 4H, H-1"'), 2.70-2.75 (m, 3H, H-3cis), 3.33-3.35 (m, 1H, H-3*trans*), 4.06 (t, J = 6.4 Hz, 2H, H-1"), 5.48 (dd, J =2.8, 12.8 Hz, 1H, H-2), 6.06 (d, J = 2.0 Hz, 1H, H-8), 6.08 (d, J = 2.0 Hz, 1H, H-6), 6.79 (d, J = 8.8 Hz, 2H, H-3', 5'), 7.32 (d, J = 8.8 Hz, 2H, H-2', 6'), 9.60 (s, 1H, 4'-OH), 12.10 (s, 1H, 5-OH). ¹³C NMR (100 MHz, DMSO-d₆): δ 12.1 (C-2"'), 42.3 (C-3), 47.1 (C-1"'), 51.2 (C-2"), 67.3 (C-1"), 78.8 (C-2), 94.4 (C-8), 95.2 (C-6), 102.8 (C-4a), 115.4 (C-3', 5'), 128.6 (C-2',6'), 128.9 (C-1'), 158.0 (C-4'), 163.1 (C-8b), 163.4 (C-5), 166.9 (C-7), 197.1 (C=O). IR $(\text{KBr, cm}^{-1}) V_{\text{max}}$: 3421.4, 3122.3, 2970.9, 1634.9, 1568.9, 1514.6, 1398.1, 1161.2, 1087.4, 838.8. HRMS (ESI) m/z [M + H]⁺ calcd. for C₂₁H₂₆O₅N, 372.1811; found 372.1808.

7-(2-Morpholinoethoxy)-5-hydroxy-2-(4-hydroxyphenyl) chroman-4-one (**3b**)

White solid, yield 90%, $R_f = 0.45$ (ethyl acetate/petroleum ether = 2:1, v/v), mp 192.5–193.5 °C. ¹H NMR (400 MHz, DMSO-d₆): δ 2.44 (t, J = 4.4 Hz, 4H, H-1^{'''}), 2.66 (t, J =6.4 Hz, 2H, H-2"), 2.74 (dd, J = 2.8, 17.2 Hz, 1H, H-3*cis*), 3.34 (dd, J = 12.8, 17.2 Hz, 1H, H-3*trans*), 3.56 (t, J = 4.4Hz, 4H, H-2^{'''}), 4.13 (t, J = 5.6 Hz, 2H, H-1^{''}), 5.48 (dd, J = 2.8, 12.8 Hz, 1H, H-2), 6.08 (d, J = 2.0 Hz, 1H, H-8), 6.10 (d, J = 2.0 Hz, 1H, H-6), 6.79 (d, J = 8.8 Hz, 2H, H-3', 5'), 7.32 (d, J = 8.8 Hz, 2H, H-2', 6'), 9.60 (s, 1H, 4'-OH), 12.10 (s, 1H, 5-OH). ¹³C NMR (100 MHz, DMSO- d_6): δ 42.3 (C-3), 53.8 (C-1"'), 56.9 (C-2"), 66.3 (H-2"'), 66.5 (C-1"), 78.9 (C-2), 94.5 (C-8), 95.4 (C-6), 102.9 (C-4a), 115.5 (C-3', 5'), 128.7 (C-2', 6'), 129.0 (C-1'), 158.1 (C-4'), 163.2 (C-8b), 163.5 (C-5), 166.9 (C-7), 197.3 (C=O). IR (KBr, cm^{-1}) V_{max} : 3440.8, 2959.2, 2924.3, 2862.14, 1643.9, 1522.3, 1440.8, 1306.8, 1172.5, 1115.1, 831.7, 590.3. HRMS (ESI) m/z [M + H]⁺ calcd. for C₂₁H₂₄O₆N, 386.1604; found 386.1603.

7-(2-(Piperidin-1-yl)ethoxy)-5-hydroxy-2-(4hydroxyphenyl)chroman-4-one (**3c**)

White solid, yield 71%, $R_f = 0.35$ (MeOH/CH₂Cl₂ = 1:10, v/v), mp 168.0–170.0 °C. ¹H NMR (400 MHz, DMSO-d₆): δ 1.35–1.39 (m, 2H, H-3'''), 1.41–1.49 (m, 4H, H-2''', 4'''), 2.39 (t, J = 2.0 Hz, 4H, H-1^{'''}, 5^{'''}), 2.61 (t, J = 6.0 Hz, 2H, H-2"), 2.73 (dd, J = 3.2, 17.2 Hz, 1H, H-3*cis*), 3.33 (dd, J = 12.8, 17.2 Hz, 1H, H-3*trans*), 4.11 (t, J = 5.6 Hz, 2H, H-1"), 5.47 (dd, J = 2.8, 12.8 Hz, 1H, H-2), 6.07 (d, J = 2.4Hz, 1H, H-8), 6.09 (d, J = 2.4 Hz, 1H, H-6), 6.79 (d, J =8.8 Hz, 2H, H-3', 5'), 7.32 (d, J = 8.8 Hz, 2H, H-2', 6'), 9.61 (s, 1H, 4'-OH), 12.10 (s, 1H, 5-OH). ¹³C NMR (100 MHz, DMSO-d₆): δ 23.9 (C-3^{'''}), 25.5 (C-2^{'''}, 4^{'''}), 42.0 (C-3^{'''}, 5^{'''}), 54.3 (C-1""), 57.0 (C-2"), 66.3 (C-1"), 78.6 (C-2), 94.2 (C-8), 95.1 (C-6), 102.5 (C-4a), 115.1 (C-3', 5'), 128.3 (C-2', 6'), 128.7 (C-1'), 157.7 (C-4'), 162.8 (C-8b), 163.2 (C-5), 166.6 (C-7), 196.8 (C=O). IR (KBr, cm⁻¹) V_{max} : 3433.0, 3118.5, 2935.9, 1638.8, 1576.7, 1522.3, 1440.8, 1196.1, 1168.9, 1087.4, 834.9. HRMS (ESI) m/z $[M + H]^+$ calcd. for C₂₂H₂₆O₅N, 384.1811; found 384.1810.

7-(2-Dimethylanimoethoxy)-5-hydroxy-2-(4-hydroxyphenyl) chroman-4-one (**3d**)

White solid, yield 30%, $R_f = 0.35$ (MeOH/CH₂Cl₂ = 1:10, v/v), mp 165.0–167.0 °C. ¹H NMR (400 MHz, DMSO- d_6): δ 2.16 (s, 6H, NCH₃), 2.73 (dd, J = 2.4, 17.2 Hz, 1H, H-3cis), 3.30 (dd, J = 12.8, 17.2 Hz, 1H, H-3trans), 3.32 (t, J = 5.6 Hz, 2H, H-2") 4.08 (t, J = 5.6 Hz, 2H), 5.47 (dd, J = 2.4, 12.8 Hz, 1H, H-2), 6.07 (d, J = 2.4 Hz, 1H, H-8), 6.09

(d, J = 2.4 Hz, 1H, H-6), 6.78 (d, J = 8..4 Hz, 2H, H-3',5'), 7.32 (d, J = 8.4 Hz, 2H, H-2',6'), 12.02 (br, 1H, 5-OH). ¹³C NMR (100 MHz, DMSO- d_6): δ 41.9 (C-3), 45.3 (N-CH₃), 57.2 (C-2''), 66.4 (C-1''), 78.5 (C-2), 94.1 (C-8), 94.9 (C-6), 102.5 (C-4a), 115.0 (C-3', 5'), 128.3 (C-2', 6'), 128.6 (C-1'), 157.6 (C-4'), 162.8 (C-8b), 163.1 (C-5), 166.6 (C-8), 196.7 (C=O). IR (KBr, cm⁻¹) V_{max} : 34.25.3, 3137.9, 2951.5, 1634.9, 1572.8, 1518.4, 1401.9, 1277.7, 1165.0, 1087.4, 838.8. HRMS (ESI) m/z [M + H]⁺ calcd. for C₁₉H₂₂O₅N, 344.1498; found 344.1496.

7-(3-Diethylaminopropoxy)-5-hydroxy-2-(4-hydroxyphenyl) chroman-4-one (**4a**)

Pale yellowish oil, 39.0%, $R_f = 0.35$ (MeOH/CH₂Cl₂ = 1:10, v/v), ¹H NMR (400 MHz, CDCl₃): δ 1.05 (t, J = 7.2Hz, 6H, H-2"'), 1.90-1.96 (m, 2H, H-2"), 2.56-2.65 (m, 6H, H-1", H-3"), 2.77 (dd, J = 2.8, 17.2 Hz, 1H, H-3*cis*), 3.08 (dd, J = 12.8, 17.2 Hz, 1H, H-3*trans*), 4.00 (t, J = 6.0 Hz, 2H, H-1"), 5.33 (dd, J = 2.8, 12.8 Hz, 1H, H-2), 6.02 (d, J = 2.4 Hz, 1H, H-8), 6.05 (d, J = 2.4 Hz, 1H, H-6), 6.83-6.86 (m, 2H, H-3', 5'), 7.29-7.32 (m, 2H, H-2', 6'), 12.00 (br, 1H, 5-OH). ¹³C NMR (100 MHz, CDCl₃): δ 10.7 (C-2"), 25.7 (C-2"), 43.0 (C-3), 46.4 (C-1"'), 48.8 (C-3"), 66.6 (C-1"), 79.0 (C-2), 94.4 (C-8), 95.5 (C-6), 103.0 (C-4a), 116.0 (C-3',5'), 127.8 (C-2',6'), 129.3 (C-1'), 157.3 (C-4'), 162.9 (8b), 164.0 (C-5), 167.2 (C-7), 196.2 (C=O). IR (KBr, cm⁻¹) V_{max}: 3163.6, 2969.6, 2918.6, 1649.1, 1564.0, 1513.0, 1455.1, 1390.4, 1298.5, 1267.9, 1159.0, 1090.9, 832.2, 736.9, 594.0. HRMS (ESI) $m/z [M + H]^+$ calcd. for C₂₂H₂₈O₅N, 386.1967; found 386.1962.

7-(3-Morpholinopropoxy)-5-hydroxy-2-(4-hydroxyphenyl) chroman-4-one (**4b**)

Pale yellowish oil, yield 76%, $R_{\rm f} = 0.36$ (ethyl acetate/petroleum ether = 2:1, v/v), ¹H NMR (400 MHz, CDCl₃): δ 1.97 (m, 2H, H-2"), 2.52-2.55 (m, 6H, H-1"', H-3"), 2.75 (dd, J = 2.8, 17.2 Hz, 1H, H-3*cis*), 3.07 (dd, J = 13.2, 17.2 Hz, 1H, H-3*trans*), 3.75 (t, J = 4.4 Hz, 4H, H-2^{'''}), 4.01 (t, J = 6.4 Hz, 2H, H-1"), 5.30 (dd, J = 2.8, 13.2 Hz, 1H, H-2), 6.00 (d, J = 2.0 Hz, 1H, H-8), 6.04 (d, J = 2.0 Hz, 1H, H-6),6.83 (d, J = 8.4 Hz, 2H, H-3', 5'), 7.27 (d, J = 8.4 Hz, 2H, H-2', 6'), 12.00 (br, 1H, 5-OH). ¹³C NMR (400 MHz, CDCl₃): δ 25.7 (C-2"), 43.0 (C-3), 53.5 (C-1""), 55.2 (C-3"), 66.4 (C-2"'), 66.6 (C-1"), 78.9 (C-2), 94.5 (C-8), 95.4 (C-6), 103.0 (C-4a), 115.7 (C-3', 5'), 127.9 (C-2', 6'), 129.7 (C-1'), 156.7 (C-4'), 162.9 (C-8b), 163.9 (C-5), 167.2 (C-7), 196.1 (C=O). IR (KBr, cm^{-1}) V_{max} : 3425.2, 3188.3, 2955.3, 2850.5, 2807.8, 1646.6, 1572.8, 1518.4, 1440.8, 1301.0, 1165.0, 1095.2, 834.9. HRMS (ESI) $m/z [M + H]^+$ calcd. for C₂₂H₂₆O₆N, 400.1760; found 400.1757.

7-(3-(Piperidin-1-yl)propoxy)-5-hydroxy-2-(4hydroxyphenyl)chroman-4-one (**4c**)

Pale vellowish oil, vield 81%, $R_f = 0.35$ (MeOH/CH₂Cl₂ = 1:10, v/v), ¹H NMR (400 MHz, CDCl₃): δ 1.47 (br, 2H, H-3""), 1.62-1.68 (m, 4H, H-2""), 1.96-2.03 (m, 2H, H-2"), 2.52–2.55 (m, 6H, H-1^{'''}, 3^{''}), 2.75 (dd, J = 2.8, 17.2 Hz, 1H, H-3*cis*), 3.07 (dd, J = 13.2, 17.2 Hz, 1H, H-3*trans*), 3.98 (t, J = 6.0 Hz, 2H, H-1"), 5.31 (dd, J = 2.8, 13.2 Hz, 1H, H-2), 5.98 (d, J = 2.0 Hz, 1H, H-8), 6.03 (d, J = 2.0 Hz, 1H, H-6), 6.82 (d, J = 7.2 Hz, 2H, H-3', 5'), 7.27 (d, J = 7.2Hz, 2H, H-2', 6'), 12.01 (br, 1H, 5-OH). ¹³C NMR (100 MHz, CDCl₃): δ 23.9 (C-3"'), 25.1 (C-2"'), 25.8 (C-2"), 43.0 (C-3), 54.3 (C-1"'), 55.6 (C-3"), 66.6 (C-1"), 79.0 (C-2), 94.4 (C-8), 95.5 (C-6), 103.0 (C-4a), 115.9 (C-3',5'), 127.8 (C-2',6'), 129.3 (C-1'), 157.3 (C4'), 162.9 (C-8b), 163.9 (C-5), 167.1 (C-7), 196.2 (C=O). IR (KBr, cm⁻¹) V_{max}: 3440.1, 3123.3, 2927.5, 2856.3, 2799.4, 1642.4, 1571.2, 1514.2, 1439.5, 1297.1, 1158.2, 1087.1, 837.9. HRMS (ESI) m/z [M + H]⁺ calcd. for C₂₃H₂₈O₅N, 398.1967; found 398.1964.

5-Hydroxy-7-methoxy-2-(4-(2-diethylaminoethoxy)phenyl) chroman-4-one (**9***a*)

Pale yellowish oil, 72%, $R_f = 0.45$ (MeOH/CH₂Cl₂ = 1:10, v/v), ¹H NMR (400 MHz, CDCl₃): δ 1.08 (t, J = 7.2 Hz, 6H, H-2^{'''}), 2.63-2.69 (m, 4H, H-1^{'''}), 2.78 (dd, J = 2.8, 17.2 Hz, 1H, H-3*cis*), 2.90 (t, J = 6.4 Hz, 2H, H-2"), 3.10 (dd, J = 12.8, 17.2 Hz, 1H, H-3*trans*), 3.80 (s, 3H, OCH₃), 4.08 (t, J = 6.4 Hz, 2H, H-1"), 5.36 (dd, J = 2.8, 12.8 Hz, 1H, H-2), 6.04 (d, J = 2.0 Hz, 1H, H-8), 6.06 (d, J = 2.0 Hz, 1H, H-6), 6.95 (d, J = 8.8 Hz, 2H, H-3',5'), 7.36 (d, J = 8.8 Hz, 2H, H-2',6'), 12.03 (s, 1H, 5-OH). ¹³C NMR (100 MHz, CDCl₃): δ 11.8 (C-2^{'''}), 43.1 (C-3), 47.8 (C-1^{'''}), 51.7 (C-2^{''}), 55.6 (OCH₃), 66.7 (C-1'), 79.0 (C-2), 94.1 (C-8), 95.0 (C-6), 103.1 (C-4a), 114.8 (C-3', 5'), 127.6 (C-2', 6'), 130.3 (C-1'), 159.3 (C-4'), 162.9 (C-8b), 164.1 (C-5), 167.9 (C-7), 196.0 (C=O). IR (KBr, cm⁻¹) V_{max}: 2970.9, 1642.7, 1572.8, 1518.4, 1444.7, 1374.8, 1293.2, 1250.5, 1153.4, 1091.3, 1025.2, 827.2, 737.9. HRMS (ESI) $m/z [M + H]^+$ calcd. for C₂₂H₂₈O₅N, 386.1967; found 386.1961.

5-Hydroxy-7-methoxy-2-(4-(2-morpholinoethoxy)phenyl) chroman-4-one (**9b**)

White solid, yield 82%, $R_f = 0.40$ (ethyl acetate/petroleum ether = 2:1, v/v), mp 101.6–102.7 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.60 (br, 4H, H-1‴), 2.78 (dd, J = 3.2, 17.2 Hz, 1H, H-3*cis*), 2.83 (t, J = 5.6 Hz, 2H, H-2″), 3.08 (dd, J = 13.2, 17.2 Hz, 1H, H-3*trans*), 3.75 (t, J = 4.4 Hz, 4H, H-2‴), 3.80 (s, 3H, OCH₃), 4.15 (t, J = 5.6 Hz, 2H, H-1″), 5.36 (dd, J = 3.2, 13.2 Hz, 1H, H-2), 6.04 (d, J = 2.0 Hz, 1H, H-8),

6.07 (d, J = 2.0 Hz, 1H, H-6), 6.94–6.98 (m, 2H, H-3', 5'), 7.36–7.39 (m, 2H, H-2', 6'), 12.03 (s, 1H, 5-OH). ¹³C NMR (100 MHz, CDCl₃): δ 43.1 (C-3), 54.0 (C-1'''), 55.6 (OCH₃), 57.5 (C-2''), 65.8 (C-2'''), 66.8 (C-1''), 78.9 (C-2), 94.1 (C-8), 95.0 (C-6), 103.0 (C-4a), 114.8 (C-3',5'), 127.7 (C-2',6'), 130.5 (C-1'), 159.1 (C-4'), 162.8 (8b), 164.0 (C-5), 167.8 (C-7), 195.9 (C=O). IR (KBr, cm⁻¹) V_{max} : 3436.9, 3126.2, 2951.5, 2869.9, 2800.0, 1627.2, 1572.8, 1514.6, 1301.0, 1207.8, 1153.4, 1114.6, 1087.4. HRMS (ESI) m/z [M + H]⁺ calcd. for C₂₂H₂₆O₆N, 400.1760; found 400.1758.

5-Hydroxy-7-methoxy-2-(4-(3-diethylaminopropoxy)phenyl) chroman-4-one (**10a**)

Pale yellowish oil, yield 45%, $R_f = 0.45$ (MeOH/CH₂Cl₂ = 1:10, v/v), ¹H NMR (400 MHz, CDCl₃): δ 1.05 (t, J = 7.2Hz, 6H, H-2"'), 1.98 (m, 2H, H-2"), 2.56-2.58 (m, 4H, H-1"'), 2.63 (t, J = 7.4 Hz, 2H, H-3"), 2.78 (dd, J = 3.2, 17.2 Hz, 1H, H-3*cis*), 3.09 (dd, *J* = 13.6, 17.2 Hz, 1H, H-3*trans*), 3.80 (s, 3H, OCH₃), 4.03 (t, J = 6.0 Hz, 2H, H-1"), 5.36 (dd, J = 2.8, 12.8 Hz, 1H, H-2), 6.04 (d, J = 2.4 Hz, 1H, H-8), 6.07 (d, J =2.4 Hz, 1H, H-6), 6.94 (d, J = 8.8 Hz, 2H, H-3',5'), 7.36 (d, J = 8.8 Hz, 2H, H-2',6'), 12.04 (s, 1H, 5-OH). ¹³C NMR (100) MHz, CDCl₃): δ 11.6 (C-2"'), 26.8 (C-2"), 43.1 (C-3), 46.9 (C-1""), 49.2 (C-3"), 55.6 (OCH₃), 66.3 (C-1'), 79.0 (C-2), 94.1 (C-8), 95.0 (C-6), 103.0 (C-4a), 114.7 (C-3',5'), 127.6 (C-2',6'), 130.1 (C-1'), 159.4 (C-4'), 162.8 (C-8b), 164.0 (C-5), 167.8 (C-7), 196.1 (C=O). IR (KBr, cm⁻¹) V_{max} : 3444.7, 3130.5, 2964.8, 2923.4, 1642.7, 1570.2, 1515.0, 1397.6, 1297.5, 1204.3, 1156.0, 1090.4, 834.9. HRMS (ESI) m/z [M $+H^{+}$ calcd. for C₂₁H₃₀O₅N, 400.2124; found 400.2121.

5-Hydroxy-7-methoxy-2-(4-(3-morpholinopropoxy)phenyl) chroman-4-one (**10b**)

White solid, yield 85%, $R_{\rm f} = 0.40$ (ethyl acetate/petroleum ether = 2:1, v/v), mp 126.9–127.9 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.00 (m, 2H, H-2"), 2.52 (m, 6H, H-1"", 3"), 2.78 (dd, J = 2.4, 17.2 Hz, 1H, H-3cis), 3.09 (dd, J = 13.2, 17.2)Hz, 1H, H-3trans), 3.74 (t, 4H, H-2"), 3.80 (s, 3H, OCH₃), 4.05 (t, J = 6.0 Hz, 2H, H-1"), 5.36 (dd, J = 2.4, 12.4 Hz, 1H, H-2), 6.04 (d, J = 2.0 Hz, 1H, H-8), 6.06 (d, J = 2.0 Hz, 1H, H-6), 6.94 (d, J = 8.4 Hz, 2H, H-3', 5'), 7.36 (d, J = 8.4 Hz, 2H, H-2', 6'), 12.03 (s, 1H, 5-OH). ¹³C NMR (400 MHz, CDCl₃): δ 26.3 (C-2"), 43.1 (C-3), 53.7 (C-1""), 55.4 (C-3"), 55.6 (OCH₃), 66.2 (C-1"), 66.9 (C-2"), 78.9 (C-2), 94.2 (C-8), 95.0 (C-6), 103.1 (C-4a), 114.7 (C-3', 5'), 127.6 (C-2', 6'), 130.3 (C-1'), 159.4 (C-4'), 162.8 (C-8b), 164.1 (C-5), 167.9 (C-7), 195.9 (C=O). IR (KBr, cm^{-1}) V_{max} : 3433.0, 3126.2, 2955.3, 2846.6, 1631.1, 1580.6, 1522.3, 1382.5, 1273.8, 1207.8, 1157.3, 1087.4, 819.4. HRMS (ESI) $m/z [M + H]^+$ calcd. for $C_{23}H_{28}O_6N$, 414.1917; found 414.1913.

7-Diethylaminocarbonylmethoxy-5-hydroxy-2-(4hydroxyphenyl)chroman-4-one (5)

To a solution of NG (272 mg, 1 mmol) in 20 mL acetone, was added N,N-diethyl-2-chloro-acetoamide (150 mg, 1 mmol), potassium carbonate (69 mg, 0.5 mmol) and potassium iodide (16 mg, 0.1 mmol). The resulting mixture was stirred at 45 °C for 2 days. 40 mL H₂O was added to the mixture. The precipitates collected by filtration was washed with ethyl acetated to afford 5 (240 mg, 62%) as white solid: $R_{\rm f} = 0.55$ (ethyl acetate/petroleum ether = 2:1, v/v), mp 229.5–230.7 °C, ¹H NMR (400 MHz, DMSO-d₆): δ 1.02 (t, J = 7.2 Hz, 3H, CH_{3a}), 1.13 (t, J = 7.2 Hz, 3H, CH_{3b}), 2.71 (dd, J = 2.8, 17.2 Hz, 1H, H-3*cis*), 3.28 (m, 4H, NCH₂), 3.31 (dd, J = 12.8, 17.2 Hz, 1H, H-3*trans*), 4.86 (s, 2H, H-1"), 5.48 (dd, J = 2.8, 12.8 Hz, 1H, H-2), 6.04 (d, J = 2.0 Hz, 1H, H-8), 6.07 (d, J = 2.0 Hz, 1H, H-6), 6.79 (d, J = 8.0 Hz, 2H, H-3', 5', 7.32 (d, J = 8.0 Hz, 2H, H-2', 6'), 9.57 (s, 1H, 4'-OH), 12.07 (s, 1H, 5-OH). ¹³C NMR (100 MHz, DMSO-d₆): δ 13.2 (CH_{3a}), 14.3 (CH_{3b}), 42.3 (NCH₂), 66.2 (C-1"), 78.9 (C-2), 94.6 (C-8), 95.5 (C-6), 102.9 (C-4a), 115.4 (C-3', 5'), 128.7 (C-2', 6'), 129.0 (C-1'), 157.9 (C-4'), 163.0 (C-8b), 163.1 (C-5), 165.6 (C-7), 166.6 (2''-C=O), 197.2 (4-C = O). IR (KBr, cm⁻¹) V_{max} : 3231.0, 2974.8, 2932.0, 1634.9, 1576.7, 1518.4, 1448.5, 1161.2, 1141.7, 1087.4, 842.7. HRMS (ESI) m/z [M + H]⁺ calcd. for C₂₁H₂₄O₆N, 386.1604; found 386.1607.

Antiproliferative assay

Cell growth inhibition was analyzed using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. Briefly, cells cultured overnight on 96-well plates were washed twice with PBS, administered media containing different concentrations of NG derivatives and incubated for 48 h. Twenty microliter of MTT stock solution (5 mg/mL) were added to each well, followed by incubation for an additional 4 h. Blue formazans were eluted from cells by the addition of 100 µL of DMSO with gentle shaking for 10 min at room temperature. Absorbencies were measured at 570 nm using an enzyme-linked immunosorbent assay reader. Data were collected from three separate experiments and the percentage of NG derivatives induced cell growth inhibition was determined by comparison to DMSO-treated control cells. Growth inhibition rate was calculated in the following way: GI (%) = (A_0) $-A_1$ / A_0 *100, where A_0 is the absorbance of the control, A_1 is the absorbance of tested samples. The GI₅₀ values were calculated using SPSS software.

Results and discussion

Tertiary amino group is a common moiety in drugs. Introduction of a tertiary amino group could enhance the hydrophilicity of a chemical entity through the corresponding salt form and in the meanwhile may improve its bioactivity. Here, we attempted to introduce a tertiary amino moiety to NG. As we know, the 7-hydroxy group of NG can be selectively alkylated. As shown in Scheme 1, nucleophilic substitution with 1,2-dibromoethane or 1-bromo-3chloropropane gave alkylated halo-intermediate 1 and 2 respectively, which were further substituted with a secondary amine to afford tertiary amino group bearing NG derivatives 3a-3d, 4a-4c. 7-methylated NG 6 was obtained using methyl iodide as electrophile. Further alkylation with 1,2-dibromoethane or 1-bromo-3-chloropropane give the 4' substituted halo-intermidiates 7 and 8, respectively. In a similar manner, tertiary amino group bearing NG derivatives 9a, 9b, 10a, and 10b were also obtained using a secondary amine. Meanwhile, compound 5, a NG derivative bearing an amide side chain at 7-hydroxy group was also prepared (Scheme 1).

The antiproliferative activities of the synthesized NG derivatives were evaluated by MTT method on four human cancer cell lines, namely, MCF-7 human breast cancer cells, HCT116 human colon cancer cells, Hela human cervical cancer cell and A549 human Lung epithelial carcinoma cells. The growth inhibitory rate at $100 \,\mu M \, (GI_{100 \mu M})$ and GI₅₀ value for each compound on each cell line were collected. As shown in Table 1, NG showed very weak antiproleferative activity to these four cancer cell lines even at 100 µM concentration (entry 13, Table 1). As we expected, the tertiary amino side chain bearing NG dereivatives generally exihibited much stronger growth inhibitory effect on these cell lines than NG did while the amide side chain bearing compound 5 did not show any enhanced activity (entry 1-11 and 12, Table 1). Among these compounds, 10a exhibited the most potent growth inhibitory activity to the tested cancer cell lines with GI₅₀ value of 15.0, 6.0, 8.7 and 11.4 µM to MCF7, HCT116, Hela and A549, respectively (entry 10, Table 1). Further SAR analysis indicated that: (1) the distance between N and O on the side chain has an notable influence on the antiproliferative activity, namely, the compounds with three methlenes between N and O (compounds 4a, 10a, entry 5 and 10, Table 1) showed stronger activity than the compounds with two methlenes between N and O (compound 3a, 9a, entry 1 and 8, Table 1); (2) a morphlino group is not a good tertiary amino moiety for the side chain, since the morpholino group bearing compounds 3b, 4b, 9b, and 10b (entry 2, 6, 9, 11, Table 1) showed much higher GI₅₀s than corresponding diethyl amino group bearing compounds 3a, 4a, 9a, and 10a did (entry 1, 5,8, 10, Table 1).



Scheme 1 Synthesis of naringenin derivatives

Table 1Antiproliferativeactivity of NG derivatives onhuman cancer cell lines, $GI_{100\mu M}$ (%), GI_{50} (μM)

Entry	Compound	MCF-7		HCT116		Hela		A549	
		$GI_{100\;\mu M}$	GI ₅₀						
1	3a	44	>100	93	22.3	94	55.2	92	52.0
2	3b	0	>100	55	107.0	0	>100	16	>100
3	3c	48	>100	91	35.1	93	63.9	93	48.6
4	3d	35	>100	93	43.7	88	66.3	86	87.3
5	4a	99	26.5	92	11.0	92	25.1	92	20.6
6	4b	19	>100	82	75.7	0	>100	28	>100
7	4c	88	44.4	92	12.9	93	28.7	92	25.1
8	9a	99	19.7	89	10.5	92	10.0	94	17.7
9	9b	11	>100	85	68.8	8	>100	22	>100
10	10a	99	15.0	90	6.0	94	8.7	91	11.4
11	10b	30	>100	29	>100	11	>100	16	>100
12	5	0	>100	19	>100	6	>100	1	>100
13	Naringenin	4	>100	20	>100	0	>100	0	>100

 $GI_{100\mu M}$ (%) = Growth inhibition rate at 100 μ M; GI_{50} (μ M) = Median growth inhibition concentration (μ M)

Conclusion

In summary, by attaching a tertiary amino side chain on 7hydroxy group or 4'-hydroxy group of naringenin, several novel NG derivatives were prepared. Some of these naringenin derivatives exhibited remarkably enhanced antiproliferative activity to four tested human cancer cell lines. The mechanism behind the enhanced sensitivity of these NG derivatives towards cancer cells will be further explored.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no competing interests.

References

- Arul D, Subramanian P (2013) Naringenin (citrus flavonone) induces growth inhibition, cell cycle arrest and apoptosis in human hepatocellular carcinoma cells. Pathol Oncol Res 19:763–770
- Bak Y, Kim H, Kang J-W, Lee DH, Kim MS, Park YS, Kim J-H, Jung K-Y, Lim Y, Hong J, Yoon D-Y (2011) A synthetic naringenin derivative, 5-Hydroxy-7,4'-diacetyloxyflavanone-N-phenyl Hydrazone (N101-43), induces apoptosis through up-regulation of Fas/FasL expression and inhibition of PI3K/Akt signaling pathways in non-small-cell lung cancer cells. J Agric Food Chem 59:10286–10297
- Benavente-Garcia O, Castillo J, Marin FR, Ortuno A, Del Rio JA (1997) Uses and properties of citrus flavonoids. J Agric Food Chem 45:4505–4515
- Chen J-W, Zhu Z-Q, Hu T-X, Zhu D-Y (2002) Structure-activity relationship of natural flavonoids in hydroxyl radical-scavenging effects. Acta Pharmacol Sin 23:667–672
- Felgines C, Texier O, Morand C, Manach C, Scalbert A, Regerat F, Remesy C (2000) Bioavailability of the flavanone naringenin and its glycosides in rats. Am J Physiol Gastrointest Liver Physiol 279:G1148–G1154
- Fuhr U, Klittich K, Staib AH (1993) Inhibitory effect of grapefruit juice and its bitter principal, naringenin, on CYP1A2 dependent metabolism of caffeine in man. Br J Clin Pharmacol 35:431–436
- Heo HJ, Kim DO, Shin SC, Kim MJ, Kim BJ, SHIN DH (2004) Effect of antioxidant flavanone, naringenin, from *citrus junos* on neuroprotection. J Agric Food Chem 52:1520–1525
- Kanno S, Tomizawa A, Hiura T, Osanai Y, Shouji A, Ujibe M, Ohtake T, Kimura K, Ishikawa M (2005) Inhibitory effects of naringenin on tumor growth in human cancer cell lines and sarcoma S-180implanted mice. Biol Pharm Bull 28:527–530

- Kawaii S, Tomono Y, Katase E, Ogawa K, Yano M (1999) HL-60 differentiating activity and flavonoid content of the readily extractable fraction prepared from citrus juices. J Agric Food Chem 47:128–135
- Kim H, Lee E, Kim J, Kim J, Lim H, Lee C-H, Ahn J-H, Chong Y, Lim Y (2007) Binding study of naringenin derivatives and cyclin dependent kinase 2. Bull Korean Chem Soc 28:1413–1415
- Lee E-R, Kang Y-J, Choi H-Y, Kang G-H, Kim J-H, Kim B-W, Han YS, Nan S-Y, Paik H-D, Park Y-S, Cho S-G (2007) Induction of apoptotic cell death by synthetic naringenin derivatives in human lung epithelial carcinoma A549 cells. Biol Pharm Bull 30:2394–2398
- Lee SH, Park YB, Bae KH, Bok SH, Kwon YK, Lee ES, Choi MS (1999) Cholesterol-lowering activity of naringenin via inhibition of 3-hydroxy-3-methylglutaryl coenzyme A reductase and acyl coenzyme A: cholesterol acyltransferase in rats. Ann Nutr Metab 43:173–180
- Liu Z, Wei W, Gan C, Huang Y, Liu S, Zhou M, Cui J (2013) Semisynthesis and cytotoxicity of *E*-naringenin oximes from naringin. Chin J Org Chem 33:2551–2558
- Moon YJ, Wang X, Morris ME (2006) Dietary flavonoids: Effects on xenobiotic and carcinogen metabolism. Toxicol in Vitro 20:187–210
- Park JH, Jin CY, Lee BK, Kim GY, Choi YH, Jeong YK (2008) Naringenin induces apoptosis through downregulation of Akt and caspase-3 activation in human leukemia THP-1 cells. Food Chem Toxicol 46:3684–3690
- Sabarinathan D, Mahalakshmi P, Vanisree AJ (2010) Naringenin promote apoptosis in cerebrally implanted C6 glioma cells. Mol Cell Biochem 345:215–222
- Sabarinathan D, Vanisree AJ (2013) Plausible role of naringenin against cerebrally implanted C6 glioma cells in rats. Mol Cell Biochem 375:171–178
- Semalty A, Semalty M, Singh D, Rawat MSM (2010) Preparation and characterization of pospholipid complexes of naringenin for effective drug delivery. J Incl Phenom Macrocycl Chem 67:253–260
- Yoon H, Kim TW, Shin SY, Park MJ, Yong Y, Kim DW, Islam T, Lee YH, Jung K-Y, Lim Y (2013) Design, synthesis and inhibitory activities of naringenin derivatives on human colon cancer cells. Bioorg Med Chem Lett 23:232–238