

Antifertility Agents. 12. Structure-Activity Relationship of 3,4-Diphenylchromenes and -chromans¹

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Synthesis and antiimplantation activity of variously substituted 2,2-dialkyl-3,4-diphenylchromenes and 3,4-*cis*- and *trans*-chromans derived from them are described. Pregnancy-inhibiting activity in rats was exhibited by a number of these compounds, which was particularly marked in the case of 3,4-*trans*-3-phenyl-4-*p*-(β -pyrrolidinoethoxy)phenyl-7-methoxychroman (32), the corresponding 2,2-dimethyl analog 34, and 3-phenyl-4-*p*-(β -pyrrolidinoethoxy)phenyl-7-methoxychromene (26). The structure-activity relationship of these compounds is discussed.

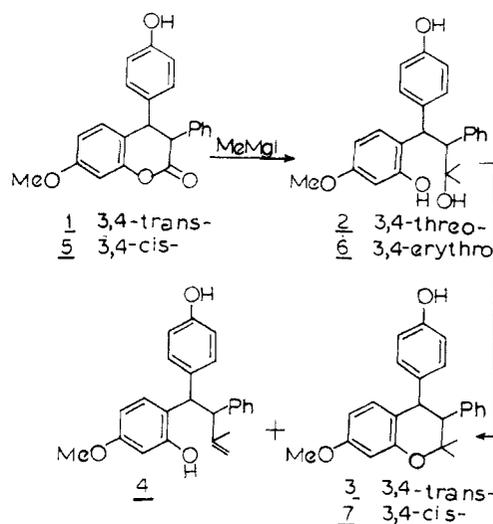
In a continuing search for nonsteroidal pregnancy-inhibiting agents,² the synthesis and antiimplantation activity of 2,3-diphenylbenzofurans, having an ω -*tert*-aminoethoxy chain at an appropriate position, were reported earlier.^{2a} This led to a detailed study of 2,2-dialkyl-3,4-diphenylchromenes and 2,2-dialkyl-3,4-*cis*- and -*trans*-3,4-diphenylchromans as possible antifertility agents. Their syntheses and biological evaluation are reported in this communication. During the course of this work Carney et al.^{3a} reported the preparation of *trans*-3-phenyl-4-*p*-(β -pyrrolidinoethoxy)phenyl-7-methoxychroman by Friedel-Crafts alkylation of phenol with 3-phenyl-7-methoxy-4-chromanol. The preparation of substituted 3,4-diphenylchromans and -chromenes and their estrogenic and antifertility activity were also reported by Carney et al.^{3b}

cis- and *trans*-3,4-diphenylchromans were synthesized by acid-catalyzed cyclization of *erythro*- and *threo*-2,3-diphenyl-4-(*o*-hydroxy)phenyl-1-propanols, respectively, following the method described earlier.^{2d} The synthesis of 2,2-dialkyl-3,4-diphenylchromans was first attempted through the reaction of alkylmagnesium halides on 3,4-diphenyldihydrocoumarins⁴ (Scheme I), prepared in turn from the corresponding coumarins by treatment with Raney nickel alloy in alkali. *trans*-3-Phenyl-4-(*p*-hydroxyphenyl)-7-methoxy-3,4-dihydrocoumarin (1) on treatment with methylmagnesium iodide gave the *threo*-carbinol 2 in good yield. Treatment of 2 with MeOH-HCl or polyphosphoric acid gave the required *trans*-chroman 3 in only 15 and 25% yields, respectively, and the corresponding butene 4 was formed as the major product. This butene 4 could not be converted to the chroman under different conditions tried.

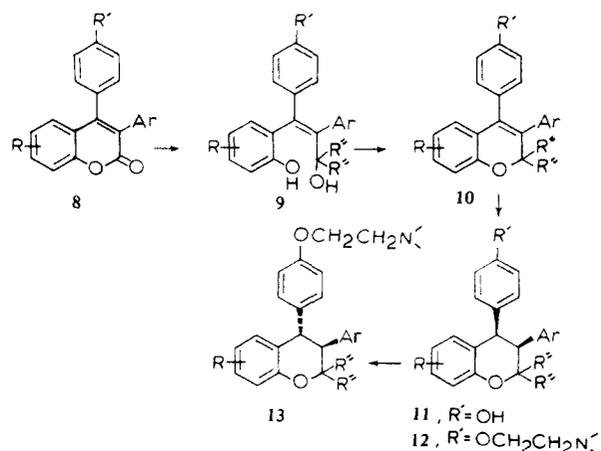
The *erythro*-butan-2-ol 6, obtained in a similar manner from *cis*-dihydrocoumarin 5, was found to be still more recalcitrant to cyclization, giving only about 4% yield of the required *cis*-chroman 7. In the rotamer population of the *erythro*-carbinol 6, the conformer shown in Figure 1 would be the most predominant as this would have the least gauche interactions of all the possible rotamers. The spatial geometry of this rotamer would be unfavorable for cyclization which is the likely reason for the poor yield obtained in the cyclization of the *erythro*-carbinol 6 to the *cis*-chroman 7.

In view of the difficulty experienced in preparing chromans 3 and 7 by cyclization of carbinols 2 and 6, the alternative route shown in Scheme II, starting from 2,2-dialkyl-3,4-diphenylchromenes 10, was next investigated. The chromenes 10 were very conveniently synthesized by the reaction of 3,4-diphenylcoumarins with excess Grignard reagents⁵ when the corresponding diols 9 were obtained, which cyclized very readily to the required chromenes 10 by treatment with mineral acids or heating; this cyclization was particularly facile when electron-releasing groups were present in the phenyl rings in conjugation with the $\Delta^{3,4}$ in the starting coumarin, when even during work-up of the

Scheme I



Scheme II

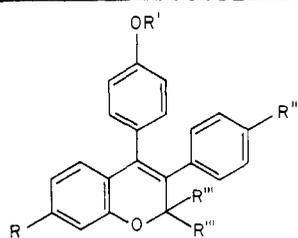


Grignard reaction product the chromenes were formed. The use of limiting quantities of Grignard reagent gave a mixture of products including the corresponding 2,3-diphenylbenzofurans.⁶

The chromenes 10 on catalytic hydrogenation gave the *cis*-chromans 11 in excellent yield and were converted to the corresponding β -*tert*-aminoethoxy ethers 12 by standard methods. 12 when treated with BuLi in Me₂SO isomerized almost quantitatively to the thermodynamically more stable *trans*-chromans 13. This method thus provides a convenient route for the synthesis of both *cis*- and *trans*-chromans from a common intermediate.

In the NMR spectrum of the chromene 16 both the CH₃ groups resonate at 84 Hz (CDCl₃), while in chromans the CH₃ signals appear as two peaks at 74 and 98 Hz in the *cis*-chroman 33 and at 74 and 82 Hz in the *trans*-chroman 34. The CH₃ signals thus appearing at 98 and 82 Hz were diagnostic for *cis* and *trans* diastereomers and were helpful

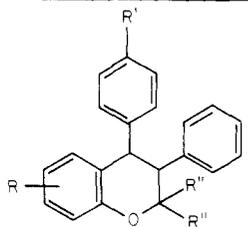
Table I



| Compd no. | R | R' | R'' | R''' | Mp, °C | Yield, % | Mol formula | Analyses | Antifertility ED ₁₀₀ (rat), mg/kg |
|-----------------|--|---|-----|------|--------|----------|--|----------|--|
| 14 | OMe | H | H | Me | 172 | 78 | C ₂₄ H ₂₀ O ₃ | C, H | Inactive |
| 15 | OMe | CH ₂ CH ₂ N(Et) ₂ | H | Me | 215 | 87 | C ₃₀ H ₃₅ NO ₃ ·HCl | C, H, N | 10 |
| 16 | OMe | CH ₂ CH ₂ -c-NC ₄ H ₉ | H | Me | 216 | 85 | C ₃₀ H ₃₃ NO ₃ ·HCl | C, H, N | 5 |
| 17 | H | H | H | Me | 152 | 73 | C ₂₃ H ₂₀ O ₂ | C, H | Inactive |
| 18 | H | CH ₂ CH ₂ -c-NC ₄ H ₉ | H | Me | 224 | 69 | C ₂₉ H ₃₁ NO ₂ ·HCl | C, H, N | 10 |
| 19 | OH | Me | H | Me | 145 | 75 | C ₂₄ H ₂₂ O ₃ | C, H | Inactive |
| 20 | OCH ₂ CH ₂ -c-NC ₄ H ₉ | Me | H | Me | 118 | 76 | C ₃₀ H ₃₃ NO ₃ ·HCl | C, H, N | 10 |
| 21 | OMe | H | Cl | Me | 205 | 70 | C ₂₄ H ₂₁ ClO ₃ | C, H | Inactive |
| 22 | OMe | CH ₂ CH ₂ -c-NC ₄ H ₉ | Cl | Me | 258 | 82 | C ₃₀ H ₃₂ ClNO ₃ ·HCl | C, H, N | 5 |
| 23 | OMe | Me | OMe | Me | 133 | 78 | C ₂₆ H ₂₆ O ₄ | C, H | Inactive |
| 24 | OMe | H | H | Et | 96 | 45 | C ₂₆ H ₂₆ O ₃ | C, H | Inactive |
| 25 | OMe | CH ₂ CH ₂ -c-NC ₄ H ₉ | H | Et | 202 | 85 | C ₃₂ H ₃₇ NO ₃ ·HCl | C, H, N | 10 |
| 26 ^a | OMe | CH ₂ CH ₂ -c-NC ₄ H ₉ | H | H | | | | | 0.15 |

^a This compound was supplied by E. Merk, Darmstadt, for comparison of biological activity.

Table II



| Compd no. | R | R' | R'' | Isomer | Antifertility ED ₁₀₀ (rat), mg/kg |
|-----------|-------|--|-----|--------|--|
| 28 | 6-Me | H | H | Cis | Inactive |
| 30 | 6-Me | H | H | Trans | Inactive |
| 31 | 7-OMe | OCH ₂ CH ₂ -c-NC ₄ H ₉ | H | Cis | 1 |
| 32 | 7-OMe | OCH ₂ CH ₂ -c-NC ₄ H ₉ | H | Trans | 0.25 |
| 7 | 7-OMe | OH | Me | Cis | Inactive |
| 33 | 7-OMe | OCH ₂ CH ₂ -c-NC ₄ H ₉ | Me | Cis | Inactive |
| 3 | 7-OMe | OH | Me | Trans | Inactive |
| 34 | 7-OMe | OCH ₂ CH ₂ -c-NC ₄ H ₉ | Me | Trans | 0.25 |
| 35 | 7-OMe | OH | Et | Cis | Inactive |
| 36 | 7-OMe | OCH ₂ CH ₂ -c-NC ₄ H ₉ | Et | Cis | Inactive |
| 37 | 7-OMe | OCH ₂ CH ₂ -c-NC ₄ H ₉ | Et | Trans | 5.0 |

Table III

| Compd no. | Dose, mg/kg | Uterotrophic act. ^a | Vaginal opening | Vaginal smear |
|-----------------|-----------------------|--------------------------------|-----------------|---------------|
| Control | | 13.6 ± 0.8 (6) | - | |
| Estrogen | 1 μg/rat ^b | 74.5 ± 3.0 (6) | + | +++ |
| 14 | 10 | 49.2 ± 4.2 (5) | + | + to ++ |
| 15 | 10 | 42.0 ± 2.0 (6) | + | - to + |
| 24 | 10 | 39.2 ± 2.4 (8) | + | - |
| 25 | 10 | 35.4 ± 2.8 (7) | + | - |
| 26 ^c | 10 | 37.4 ± 1.8 (7) | + | - |
| 31 | 10 | 50.6 ± 2.0 (7) | + | ++ |
| 31 | 5 | 43.7 ± 2.3 (7) | + | - to ++ |
| 32 | 10 | 48.3 ± 3.9 (7) | + | - to + |
| 33 | 10 | 20.5 ± 1.3 (6) | - | |
| 34 | 10 | 48.3 ± 3.1 (7) | + | ++ to +++ |
| 35 | 10 | 14.6 ± 0.4 (7) | - | |
| 36 | 10 | 16.5 ± 1.1 (6) | - | |
| 37 | 10 | 46.6 ± 2.0 (7) | + | + to +++ |

^a Uterine weight in mg, mean ± SE; number of animals in parentheses. ^b Immature rats of 25-30-g weight.

^c Compound 26 is reported to be positive in the Allen-Doisy test according to E. Merck, Darmstadt.

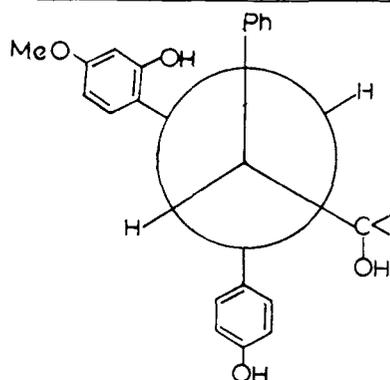


Figure 1.

in monitoring the stereochemical purity of the samples.

The compounds thus prepared are listed in Tables I and II.

Biological Activity. The compounds were tested for

their antifertility activity in pregnant female albino rats of proven fertility. The day on which the vaginal smears showed the presence of spermatozoa was considered day 1 of pregnancy. In the primary screening the compounds were fed for 5 days from days 1-5 of pregnancy using five animals in each group. The compounds were macerated with gum acacia and administered orally to animals. The results were scored as positive only if implantations were totally absent in both uterine horns, examined on day 10 of pregnancy; control animals had an average of seven implants. The initial dose for each compound was 20 mg/kg/day unless otherwise mentioned. Compounds inactive at this dose level were dropped. Results of antifertility screening are given in Tables I and II.

Estrogenic activity of the compounds was tested in immature female rats weighing 25-30 g by increase in uterine weight⁷ and vaginal cornification⁸ and compared with estrone as standard, and the results are described in Table III. The compounds were administered orally twice

daily for 3 days and the animals killed on the fourth day. The uteri were separated from uterotubal and cervical junctions and weighed fresh after pressing out the uterine fluid between folds of a blotting paper. Vaginal smears were examined before autopsy and scored as -, diestrus smear, leucocytes only; +, mixture of leucocytes and epithelial cells; ++, proestrus smear, nucleated or nucleated and cornified cells; and +++, estrus smear cornified cells only.

Failure to induce vaginal opening⁹ by any particular compound was considered as complete absence of estrogenic activity.

Most of the compounds showed weak estrogenic activity as compared to estrone. The antifertility potential of the compounds does not seem to be due to estrogenicity alone since compound 34^{2b} (under detailed investigation) showed estrogenic, antiestrogenic, and antiprogesterational property.

In conformity with earlier results only compounds with a basic side chain showed antiimplantation activity and pyrrolidine residues in the side chain yielded more active compounds than the diethylamino moiety.¹⁰ Introduction of a chloro radical in the para position of the 3-phenyl group as in 22 did not materially increase the activity in chromenes while removal of the 7-OMe radical as in 18 decreased the activity. It is of note that deletion of the substituents at the 2 position of chromenes (compound 26) markedly increased activity.

In the chroman series the trans basic ether 32 showed activity approaching that of the most active chromene. The corresponding cis isomer 31 shows but one-fourth the activity of the trans compound. Substitution of the methyl group at the 2 position (compound 34) has no influence on the activity of the trans isomer. The dimethyl cis isomer 33, on the other hand, is inactive. Substitution of the 2 position of the trans compound by the larger ethyl group, compound 37, decreases potency by a factor of 20. The corresponding cis isomer 36 is again inactive. Examination of Dreiding models of the isomeric chromans suggests that the trans isomers retain considerable planarity even when substituted at position 2; the same substitution induces considerable distortion in the cis isomers.

Experimental Section

All melting points were determined in a sulfuric acid bath and are uncorrected. The structure of the products was checked routinely by ir, uv, and NMR spectroscopy run on Perkin-Elmer Infracord 137, Unicam, and Varian A-60D spectrometers, respectively.

The following experiments represent typical experimental procedures employed in the preparation of the above-mentioned compounds.

2,2-Dimethyl-3-phenyl-4-(*p*-hydroxyphenyl)-7-methoxychromene (14). A solution of 3-phenyl-4-*p*-hydroxyphenyl-7-methoxycoumarin⁴ (2.2 g, 0.006 mol) in tetrahydrofuran (40 ml) was gradually added to methylmagnesium iodide [from 8.5 g (0.05 mol) of CH₃I and 1.2 g (0.049 g-atom) of Mg] in dry ether (30 ml). The mixture was heated under reflux for 4 hr and poured over crushed ice containing ammonium chloride (10 g) and hydrochloric acid (5 ml). The reaction mixture was extracted with ethyl acetate, the extract was washed with water to neutrality and dried (Na₂SO₄), the solvent distilled off, and the syrup crystallized from benzene-petroleum ether: yield 1.8 g (78%); mp 172°C.

2,2-Dimethyl-3-phenyl-4-*p*-(β-pyrrolidinoethoxy)phenyl-7-methoxychromene (16). A mixture of 14 (0.5 g, 0.0016 mol), pyrrolidinoethyl chloride hydrochloride (0.5 g, 0.0029 mol), anhydrous potassium carbonate (2.0 g), and dry acetone (50 ml) was heated under reflux for 15 hr. The solvent was removed, the residue extracted with benzene, and the extract washed with water to neutrality and dried (Na₂SO₄). The benzene extract was passed through a short column of basic alumina; the solvent distilled off and the residue was converted to its hydrochloride and crystallized

from ethanol-ether: yield 0.60 g (76%); mp 216°.

erythro-2,3-Diphenyl-3-(2-tosyloxy-5-methyl)phenyl-1-O-tosylpropanol (27). A mixture of *erythro*-2,3-diphenyl-3-(2-hydroxy-5-methyl)phenylpropan-1-ol¹¹ (2 g, 0.006 mol) and *p*-toluenesulfonyl chloride (3.4 g, 0.018 mol) in dry pyridine (15 ml) was heated on a steam bath for 3 hr, poured over ice, and extracted with benzene. The organic layer was washed with 3 *N* HCl and then with water to neutrality, dried (Na₂SO₄), and concentrated and the residue was crystallized from methanol: yield 2 g (53%); mp 140°. Anal. (C₃₆H₃₄O₆S₂) C, H.

cis-3,4-Diphenyl-6-methylchroman (28). A mixture of the ditosylate 27 (1.8 g, 0.0028 mol) in alcoholic KOH (KOH, 10 g in 200 ml of ethanol) was refluxed for 3 hr, the solvent distilled off, and residue extracted with benzene. The benzene extract was washed with water to neutrality, dried (Na₂SO₄), and concentrated and the residue was crystallized from benzene-methanol: yield 0.9 g (93%); mp 117°. Anal. (C₂₂H₂₀O) C, H.

threo-2,3-Diphenyl-3-(2-tosyloxy-5-methyl)phenyl-1-O-tosylpropanol (29). This was similarly prepared from the corresponding *threo*-2,3-diphenyl-3-(2-hydroxy-5-methyl)phenylpropan-1-ol¹¹ in 56% yield and crystallized from benzene-hexane: mp 195°. Anal. (C₃₆H₃₄O₆S₂) C, H.

trans-3,4-Diphenyl-6-methylchroman (30). Cyclization of 16 by treatment with ethanolic KOH as described for 28 gave 30 in 91% yield which was crystallized from benzene-hexane: mp 117°. Anal. (C₂₂H₂₀O) C, H.

erythro-2-Methyl-3-phenyl-4-(*p*-hydroxyphenyl)-4-(2-hydroxy-4-methoxy)phenylbutan-2-ol (6). A 3 *M* solution of MeMgCl (6.5 ml) in THF was added to a solution of *cis*-3-phenyl-4-*p*-acetoxyphenyl-7-methoxy-3,4-dihydrocoumarin (1.5 g, 0.004 mol) in anisole (25 ml) and the reaction mixture stirred at 100° for 24 hr. The reaction mixture was then cooled, treated with ammonium chloride solution, and extracted with ether, the extract was washed with water and dried (Na₂SO₄), the solvent distilled off, and the residue was crystallized from THF-benzene, yield 1.22 g (80%), and recrystallized from THF-benzene: mp 192°. Anal. (C₂₄H₂₆O₄) C, H.

cis-2,2-Dimethyl-3-phenyl-4-*p*-hydroxyphenyl-7-methoxychroman (7). (a) A solution of the butanol 6 (1 g) in PPA (15 g) was heated at 75–80° for 0.5 hr. The reaction mixture was treated with water and the solid which separated was crystallized from benzene: yield 0.04 g (4%); mp 180°. Anal. (C₂₄H₂₄O₃) C, H.

(b) Hydrogenation of 14 (23.1 g) in 300 ml of THF in the presence of Pd/C catalyst [4.5 g of dry, 5% Pd/C and 12.8 g of 5% Pd/C (w/w 50/50)] at 60° and 100 psi of pressure gave 19.5 g (84%) of 7.

cis-2,2-Dimethyl-3-phenyl-4-*p*-(β-pyrrolidinoethoxy)phenyl-7-methoxychroman (33). β-Pyrrolidinoethyl chloride hydrochloride (9.8 g, 0.57 mol) was added to a warm solution of 7 (20.74 g, 0.058 mol) in NaOH (5.8 g, 0.145 mol), water (16.5 ml), and 2-propanol (104 ml) and the mixture was heated at 50° for 4 hr under vigorous stirring. The reaction solution was cooled and diluted with cold water (200 ml) and the solid which separated was collected by filtration, washed successively with 30% 2-propanol (10 ml) and cold water (40 ml), and dried: yield 23.7 g (90%); mp 118–121°.

The hydrochloride of 33 crystallized from ethanol-ether: mp 196°. Anal. (C₃₀H₃₆ClNO₃) C, H, N.

trans-2,2-Dimethyl-3-phenyl-4-(*p*-hydroxyphenyl)-7-methoxychroman (3). (a) A solution of *trans*-3-phenyl-4-*p*-hydroxyphenyl-7-methoxy-3,4-dihydrocoumarin⁴ (3.46 g, 0.01 mol) in dry THF was added under stirring to CH₃MgI prepared from CH₃I (7.10 g, 0.05 mol) and Mg (1.33 g, 0.05 g-atom) in dry ether. The reaction mixture was refluxed for 6 hr and then worked up as described above for 6. The oily product thus obtained was found to be a complex mixture (five spots) on TLC. Chromatography of this mixture on silica gel using increasing proportions of chloroform in benzene as eluent gave 3 in 15% yield (0.55 g) which was crystallized from benzene: mp 264°. Anal. (C₂₄H₂₄O₃) C, H.

Further development of the column gave *threo*-2-methyl-3-phenyl-4-(*p*-hydroxyphenyl)-4-(2-hydroxy-4-methoxy)phenylbutan-2-ol (2) in 20% yield (0.75 g) which was crystallized from benzene-hexane: mp 180°. Anal. (C₂₄H₂₆O₄) C, H.

(b) A solution of the butanol 33 (1 g) in PPA (15 g) was heated

at 75–80° for 0.5 hr. The reaction mixture was decomposed with water and the separated solid was crystallized from benzene: yield 0.22 g (25%).

(c) The butanol 2 when heated under reflux with EtOH–HCl (4%) gave 3 in 15% yield.

trans-2,2-Dimethyl-3-phenyl-4-p-(β-pyrrolidinoethoxy-phenyl)-7-methoxychroman (34). (a) Alkylation of 3 with β-pyrrolidinoethyl chloride hydrochloride as mentioned above for 7 gave 34 in 92% yield which was converted to its HCl salt and crystallized from ethanol–ether: mp 164°. Anal. (C₃₀H₃₆ClNO₃) C, H, N.

(b) BuLi (35 ml of 19.85%) in THF was added to a suspension of 33 (23.6 g, 0.05 mol) in 250 ml of anhydrous Me₂SO at room temperature under nitrogen atmosphere under good stirring. The resulting red solution was stirred for further 3 hr and then decomposed with 50 ml of cold water. The reaction mixture was then poured over 1.5 l. of an ice–water mixture and extracted with ether. The organic layer was washed well with water and dried (Na₂SO₄), the solvent distilled off, and the residue converted to its HCl salt and crystallized from ethanol–ether: yield 24.8 g (97.4%); mp 164°.

cis-2,2-Diethyl-3-phenyl-4-(p-hydroxyphenyl)-7-methoxychroman (35). High-pressure hydrogenation of 24 following the procedure used for 7 yielded 35 in 68% yield. The solid was recrystallized from benzene–hexane: mp 186°. Anal. (C₂₆H₂₈O₃) C, H.

cis-2,2-Diethyl-3-phenyl-4-p-(β-pyrrolidinoethoxy)-phenyl-7-methoxychroman (36). Alkylation of 35 with β-pyrrolidinoethyl chloride hydrochloride as mentioned above for 33 gave a 93% yield of 36 as a solid compound which was recrystallized from benzene–hexane: mp 133–133.5°. Anal. (C₃₂H₃₉NO₃) C, H, N.

trans-2,2-Diethyl-3-phenyl-4-p-(β-pyrrolidinoethoxy)-phenyl-7-methoxychroman (37). Isomerization of 36 using BuLi

in Me₂SO as mentioned for 34 yielded 81.3% of 37 which was converted to its HCl salt and crystallized from 2-propanol: mp 227°. Anal. (C₃₂H₄₀ClNO₃) C, H, N.

Acknowledgment. We are thankful to Dr. Jim Bolger for his help in the reduction of chromene 14 and its subsequent isomerization which greatly facilitated the preparation of 34.

References and Notes

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Relationship of Molecular Structure to in Vivo Scintigraphic Distribution Patterns of Carbon-11 Labeled Compounds. 3. [¹¹C]Hydantoin

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The preparation and γ-scintigraphic evaluation of the in vivo distribution patterns in dogs of a series of structurally related hydantoin labeled with the positron emitting, 20.4 min half-life radionuclide, carbon-11 are described. Carbon-11 labeled HCN was collected in water or aqueous Me₂SO containing carrier KCN following cyclotron bombardment of 99% N₂–1% H₂ gas mixture with 22 MeV protons for 1 hr at 25–35 μA. Five ¹¹C-labeled 5,5-dialkylhydantoin, three [¹¹C]diarylhydantoin, five [¹¹C]-5-alkyl-5-phenylhydantoin, and five [¹¹C]spirohydantoin were prepared by heating, generally under pressure, a mixture of ¹¹C-labeled KCN, which was produced by isotopic exchange with carrier KCN, the corresponding aldehyde or ketone, and excess (NH₄)₂CO₃ in aqueous ethanol or Me₂SO solvent. The ¹¹C-labeled hydantoin were dissolved in 1–1.5% aqueous NaOH for intravenous administration to dogs. Total synthesis time was 70–106 min and 1–59 mCi of final product was available for conducting serial in vivo imaging for up to 2 hr or more with the γ-scintillation camera. Carbon-11 activity from all compounds showed initial blood-pool distribution with variable concentration of activity in the brain, lungs, liver, and kidney. All of the ¹¹C-labeled diethylhydantoin, and most having one phenyl substituent, and one having a hexamethylene moiety showed initial accumulation of ¹¹C activity in brain. Carbon-11 labeled 5,5-diphenylhydantoin (dilantin) showed the greatest qualitative accumulation of activity in the brain. Those ¹¹C-labeled hydantoin having a carboxyl substituent showed prominent renal concentration and urinary excretion of activity. Most ¹¹C-labeled hydantoin showed a progressive homogenous whole body distribution of activity in all cellular tissues of the body. The relatively uniform distribution of activity in cellular tissues and slow excretion from the body support the thesis that the pharmacologic action of the hydantoin is related to physical effects on biomembranes rather than to specific chemical interactions with cell constituents.

Data presently available from experiments using ¹⁴C-labeled compounds are insufficient to predict adequately the relationship between molecular structure of an organic compound and its in vivo distribution pattern as studied by scintigraphic techniques. This is so for

several reasons. The inherent low specific activity of ¹⁴C-labeled compounds makes ¹⁴C an unsuitable label for evaluation of in vivo distribution patterns of molecules whose in vivo distribution demonstrates a strong “carrier” effect in the range of the quantity of ¹⁴C-labeled material