

- Matsumoto, T. (1988) *Phytochemistry* **27**, 2931.
- 4 Akihisa, T., Thakur, S., Rosenstein, F. U. and Matsumoto, T. (1986) *Lipids* **21**, 39
 - 5 Akihisa, T., Ghosh, P., Thakur, S., Rosenstein, F. U. and Matsumoto, T. (1986) *J. Am. Oil Chem. Soc.* **63**, 653.
 - 6 Akihisa, T., Shimizu, N., Ghosh, P., Thakur, S., Rosenstein, F. U., Tamura, T. and Matsumoto, T. (1987) *Phytochemistry* **26**, 1693
 - 7 Doyle, P. J., Patterson, G. W., Dutky, S. R. and Thompson, M. J. (1972) *Phytochemistry* **11**, 1951
 - 8 Knapp, F. F., Phillips, D. O., Goad, L. J. and Goodwin, T. W. (1972) *Phytochemistry* **11**, 3497.
 - 9 Akihisa, T., Shimizu, N., Tamura, T. and Matsumoto, T. (1986) *Lipids* **21**, 491
 - 10 Farnes, M., Cocallemen, S. and Soulier, J. (1988) *Lipids* **23**, 349.

Phytochemistry, Vol 28, No 4, pp 1273–1275, 1989
Printed in Great Britain

0031-9422/89 \$3.00 + 0.00
Pergamon Press plc

A PREGNANE TRIGLYCOSIDE ESTER FROM *DREGEA SINENSIS* VAR *CORRUGATA*

JIN QIDUAN, ZHOU QIANLAN and MU QUANZHANG

Kunming Institute of Botany, Academia Sinica, Kunming, China

(Received in revised form 22 July 1988)

Key Word Index—*Dregea sinensis* var *corrugata*, Asclepiadaceae, dregeoside, steroid, pregnane glycoside ester

Abstract—A new pregnane glycoside ester, dregeoside, was isolated from the dried rhizome of *Dregea sinensis* var *corrugata*. On the basis of chemical reactions and spectroscopic evidence, the structure was established as 12-O-benzoyl-drevogenin-3-O- β -D-oleandropyranosyl(1 \rightarrow 4)-O- β -D-cymaropyranosyl(1 \rightarrow 4)-O- β -D-cymaropyranoside

INTRODUCTION

In a previous paper [1], we reported the isolation and structure elucidation of a drevogenin from the rhizomes of *Dregea sinensis*. As a continuation of the studies on this plant, we present the spectral and chemical evidence for the structure of a new triglycoside (1)

RESULTS AND DISCUSSION

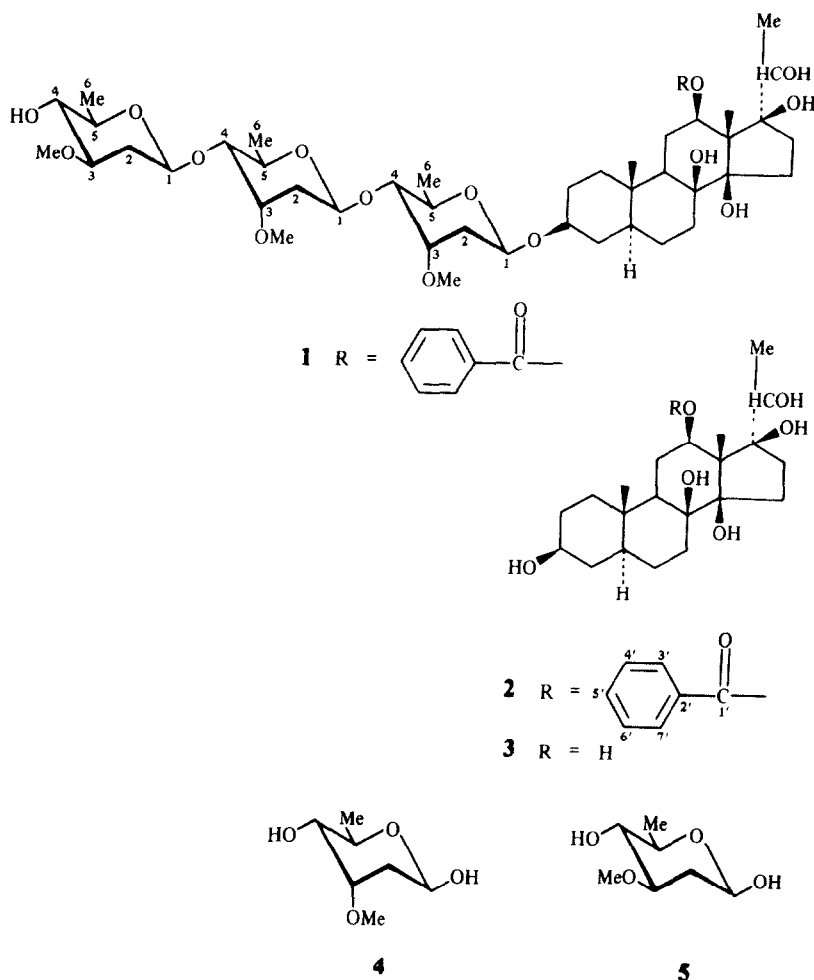
Dregeoside (1) was isolated by gel column chromatography and reverse-phase chromatography. The mass spectrum of compound 1 indicated that molecular formula was $C_{49}H_{76}O_{16}$ (FAB-MS $[M]^+$ at m/z 920). The UV spectrum showed absorption at 230 nm. Its IR spectrum showed the presence of methoxy group (2925, 1440 cm^{-1}) and $-C-O-C-O-$ (1195, 1160, 1080, 1050 cm^{-1}). Confirmation of the triglycoside structure of compound 1 and the position of its monobenzoate ester group on the genin moiety was provided by the 1H NMR (400 MHz) spectrum of 1 which indicated the presence of three methoxy group at 3.38 (3H, s, OMe), 3.45 (6H, s, 2 \times OMe) and three methyl group at 1.23 (6H, d, $J=6$ Hz, sec. 2 \times Me), 1.34 (3H, d, $J=6$ Hz, sec. Me). A methyl group and a methoxy group could be assigned to C-3 and C-6, respectively, of a deoxy sugar

As cymaropyranose possesses only two hydroxy groups at C-1 and C-4, the sugar sequence in 1 was linear. We assigned the ^{13}C NMR signals of the sugar chain in 1 as shown in Table 1 in comparison with the data on the ^{13}C NMR chemical shifts of methyl β -D-cymaroside and

α,β -D-oleandroside [2] [3]. From the ^{13}C NMR chemical shifts of the anomeric carbons the D-cymarose and D-oleandrose moieties in 1 are suggested to have a β -configuration at C-1. The ^{13}C NMR data of 1 is presented in Table 1. Mass spectral fragment ion peaks at m/z 145 ($C_7H_{13}O_3$), 144 ($C_7H_{12}O_3$), and 499 ($C_7H_{13}O_3-C_7H_{12}O_3-C_7H_{12}O_4$) suggested that there were three deoxy sugars in the molecule.

Mild acid hydrolysis of the acetate of 1 with dilute sulphuric acid yielded 4-O-acetyl-oleandrose, cymarose and the monoacetate of 2 which was identical with an authentic sample as determined by TLC and GLC. Oleandrose was indicated to be the terminal sugar.

Mild acid hydrolysis of 1 using the earlier reported method of Mannich and Siewert [4] yielded drevogenin (2) and cymarose (4), oleandrose 5 ($[\alpha]_D$ mp, TLC) [5]. Alkaline hydrolysis of 2 yielded deacyldrevogenin (3), which was identical with authentic dihydrosarcostin (mp, mmp, and IR) [6]. The monobenzoate nature of the ester function in 1 was supported by its IR, UV (λ_{max}^{EtOH} 282 nm, $\log \epsilon$ 3.05) and 1H and ^{13}C NMR spectral data. The difference of $C_{21}H_{36}O_9$ between the formula of glycoside 1 and its aglycone 2 indicated that 1 was a triglycoside. The same conclusion could be drawn from the mass spectrum of 1 which recorded fragment ions for a trisaccharide unit (m/z 450) and the genin moiety (m/z 470) besides the prominent fragment ions of drevogenin monobenzoate giving ions for benzoic acid at m/z 122 and the other expected ions of the drevogenin moiety including the fragment ions due to the sequential losses of its four molecules of water at m/z 452, 434, 416, and 398. In



the light of the above evidence the structure of dregeoside was established as 12-*O*-benzoyl-drevogenin-3-*O*- β -D-oleandropyranosyl(1 \rightarrow 4)-*O*- β -D-cymaropyranosyl (1 \rightarrow 4)-*O*- β -D-cymaropyranoside

EXPERIMENTAL

Mps: uncorr, ^1H and ^{13}C NMR 400 MHz. TMS as int standard TLC and CC used CHCl_3 -MeOH (49/1), reverse phase-CC MeOH- H_2O (4/1)

Plant extraction Shade-dried powdered rhizome (5.2 kg) of *Dregea sinensis* var. *corrugata* were extracted with EtOAc on a hot bath. The aq. concentrate was fractionated with different organic solvents to afford a petrol extract (20 g) and a CHCl_3 extract (11.5 g), i.e. crude glycoside, which was subjected to CC on silica gel with CHCl_3 -MeOH (49/1) and further on reversed-phase gel with MeOH- H_2O (4/1) to afford **1** (120.5 mg, yield 0.0011%).

Dregeoside 1, Mp 125–128°, $[\alpha]_D^{25} + 43.2^\circ$ (MeOH, c 0.15) (Found C, 63.91, H, 8.26, $\text{C}_{49}\text{H}_{76}\text{O}_{16}$ requires C, 63.81, H, 8.15%). It showed positive Liebermann-Burchard and Keller-Kiliani reactions. UV λ_{max} nm (log ϵ) 231 (4.23), 274 (3.12), 282 (3.05). IR ν_{max} cm^{-1} 3540 (ass. OH group), 1710 (C=O of benzoate ester), 1368 (Me def), 710 (C-H def arom). ^1H NMR (400 MHz, CDCl_3) 7.49 (2H, t , $J = 7.9$ Hz, 4', 6'-CH), 7.62 (1H, tt , $J = 7.9$, 1.2 Hz, 5'-CH), and 8.05 (2H, dd , $J = 7.9$, 1.2 Hz, 3', 7'-CH), 4.47 (1H, dd , $J = 10$, 2 Hz, 1-CH of β -D-cymaropyranose),

4.74 (1H, dd , $J = 10$, 2 Hz, 1-CH of β -D-cymaropyranose), and 4.82 (1H, dd , $J = 10$, 2 Hz, 1-CH, of β -D-oleandropyranose), 4.70 (1H, dd , $J = 9$, 5 Hz, 12-H), 4.80 (1H, q , $J = 6.5$ Hz, 20-H), 3.39 (3H, s , OMe), 3.44 (3H, s , $2 \times$ OMe), 1.34 (3H, d , $J = 6$ Hz, sec Me), 1.25 (6H, d , $J = 6$ Hz, sec $2 \times$ Me), 1.24 (3H, s , 18-Me), 0.96 (3H, s , 19-Me). FABMS m/z 920 $[\text{M}]^+$, 488 $[\text{M} - \text{trisaccharide}]^+$ (3), 470 $[\text{488} - \text{H}_2\text{O}]^+$ (6), 452 $[\text{488} - 2\text{H}_2\text{O}]^+$ (2), 443 $[\text{488} - 45]^+$ (4), 383 $[\text{488} - 105]^+$ (8), 366 $[\text{488} - \text{BzOH}]^+$ (100), 348 $[\text{488} - \text{BzOH} - \text{H}_2\text{O}]^+$ (5), 321 $[\text{488} - \text{BzOH} - \text{MeCHOH}]^+$ (4), sugar fragments 450 (dicymarose and oleandrose) $^+$ (3), 354 $[\text{450} - 3\text{MeOH}]^+$ (24), 290 (rearranged disaccharide fragment) $^+$ (100), 162 [cymarose or oleandrose] $^+$ (21), 130 $[\text{162} - \text{MeOH}]^+$ (22), 86 (21).

Mild acid hydrolysis of 1. A soln of **1** (20 mg) in MeOH (10 ml) was mixed with 0.05 M H_2SO_4 (1.5 ml) and warmed for 30 min at 50°. MeOH was then removed under red pres. The aq. concentrate was repeatedly extracted with CHCl_3 and the organic layer washed in turn with H_2O , 1 M Na_2CO_3 and H_2O , dried over Na_2SO_4 and evapd to afford the genin **2** (12 mg) $[\alpha]_D^{25} + 63^\circ$ (MeOH, c 0.2). ^1H NMR (400 Hz) of **2** 8.05–7.49 (5H, m , aromatic), 1.91 (3H, s , 21-Me), 0.96 (3H, s , 19-Me), 1.24 (3H, s , 18-Me), 4.80 (1H, q , $J = 6.5$ Hz, 20-H). The aq. hydrolysate was neutralized with freshly pptd BaCO_3 , filtered and concd under red pres to yield the syrupy soln. This gave a positive Keller-Kiliani reaction, the sugars were identical with authentic samples by comparison on FLC and GLC of the TMS-sugar

Table 1 ^{13}C NMR chemical shifts of dregeoside (CDCl_3 , TMS, ppm)

Aglycone moiety			Sugar moiety		Methyl glycoside
C-1	38 12 <i>t</i>		Cymarose		V
2	33 13 <i>t</i>	C-1	96 65	C-1	99 4
3	70 91 <i>d</i>	2	37 30	2	35 1
4	38 92 <i>t</i>	3	77 40	3	78 5
5	45 41 <i>d</i>	4	83 70	4	74 0
6	34 01 <i>t</i>	5	68 75	5	71 0
7	34 23 <i>t</i>	6	18 58	6	18 9
8	76 81 <i>s</i>	C-3-OMe	58 47	C-1-OMe	57 8
9	46 62 <i>d</i>		Cymarose	C-1-OMe	56 0
10	38 13 <i>s</i>	C-1	100 30		VII
11	28 81 <i>t</i>	2	38 01	C-1	101 0
12	74 25 <i>d</i>	3	77 40	2	36 6
13	56 05 <i>s</i>	4	83 14	3	81 3
14	83 80 <i>s</i>	5	68 75	4	76 2
15	34 05 <i>t</i>	6	18 58	5	72 6
16	32 75 <i>t</i>	C-3-OMe	58 47	6	18 4
17	88 15 <i>s</i>		Oleandrose	C-1-OMe	56 9
18	11 82 <i>q</i>	C-1	102 04	C-3-OMe	56 0
19	16 15 <i>q</i>	2	37 30		IX
20	69 52 <i>d</i>	3	81 20	C-1	98 7
21	16 70 <i>q</i>	4	76 77	2	35 1
C-1'	165 85 <i>s</i>	5	73 50	3	79 0
2'	130 32 <i>s</i>	6	18 09	4	76 6
3'	130 32 <i>d</i>	C-3-OMe	56 80	5	68 4
4'	129 21 <i>d</i>			6	18 4
5'	133 42 <i>d</i>			C-1-OMe	56 9
6'	129 21 <i>d</i>			C-3-OMe	54 3
7'	130 32 <i>d</i>				

cymarose (R_f 0 46) and oleandrose (R_f 0 28); relative retention time (min): 0 44 (cymarose), 0 37 (oleandrose)

Alkaline hydrolysis of compound 2. Compound 2 (8 mg) was dissolved in 5% methanolic KOH (1 5 ml) and refluxed for 5 hr. After adding H_2O (1 0 ml) MeOH was removed under red pres. The aq. concentrate was extracted with CHCl_3 -MeOH (9 1), dried over Na_2SO_4 , filtered and evapd to dryness yielding 3 (4 0 mg) which crystallized from MeOH, mp 245–248°, $[\alpha]_D^{25} + 51^\circ$ (MeOH, c 0 15). It was identified as deacyldrevogenin by comparison with an authentic sample of dihydrosarcostin ($[\alpha]_D$, TLC, mmp) [6]

Acknowledgement—We wish to thank the instrument group of our Institute for UV, IR, ^1H and ^{13}C NMR, MS and elemental analysis

REFERENCES

- 1 Jin Qiduan and Mu Quanzhang (1987) *Acta Bot Yunnan.* **9**, 227
- 2 Fumiko Abe and Tatsuo, Yamauchi (1978) *Chem. Pharm Bull.* **26**, 3023
- 3 Keiji Wada, Koji Hayashi, Hiroshi Mitsuhashi and Hideo Bando (1979) *Chem. Pharm Bull.* **27**, 2254
- 4 Rangaswami, S. and Reichstein, T. (1949) *Helv. Chem. Acta.* **32**, 939
- 5 Ajay, S., Bhatnagar, W., Stocklin and Reichstein, T. (1968) *Helv. Chem. Acta* **51**, 133
- 6 Jaeggi, K. A., Weiss, E. K. und Reichstein, T. (1963) *Helv. Chem. Acta* **46**, 695