

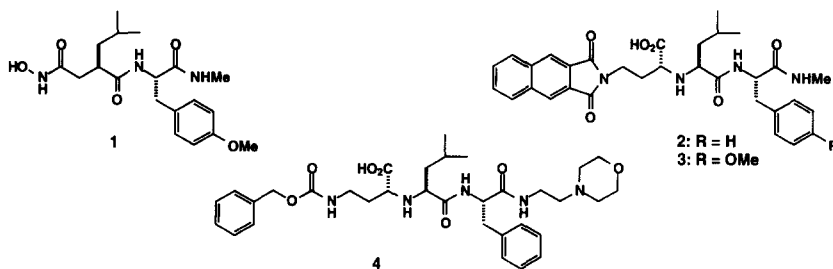


## INHIBITORS OF MMP-1: AN EXAMINATION OF P<sub>1</sub>' C $\alpha$ GEM-DISUBSTITUTION IN THE N-CARBOXYALKYLAMINE AND GLUTARAMIDE CARBOXYLATE SERIES.

Ralph P. Robinson,\* Brian J. Cronin, Kathleen M. Donahue, Brian P. Jones, Lori L. Lopresti-Morrow, Peter G. Mitchell, James P. Rizzi, Lisa M. Reeves and Sue A. Yocum  
Pfizer Central Research, Eastern Point Road, Groton, CT 06340

**Abstract.** Modification of the N-carboxyalkylamine **3** by independent replacement of the P<sub>1</sub>' NH group for CH<sub>2</sub> and introduction of P<sub>1</sub>' gem-cyclohexyl substitution affords compounds **5** and **6a** which retain appreciable activity against MMP-1 (IC<sub>50</sub>s = 0.023  $\mu$ M and 0.09  $\mu$ M, respectively). The glutaramide **7a** which incorporates both these structural changes also retains potent activity (IC<sub>50</sub> = 0.038  $\mu$ M). Copyright © 1996 Elsevier Science Ltd

**Introduction.** Orally active inhibitors of matrix metalloproteinases (MMP's), particularly interstitial collagenase (MMP-1), are sought as drugs for the treatment of osteoarthritis.<sup>1</sup> Functionality which can ligate the catalytic zinc atom in the active site of the enzyme is essential for inhibition of MMP's. The most widely employed zinc ligand has been the hydroxamate group which, when incorporated onto the appropriate dipeptide equivalent, affords compounds (e.g., the succinamide hydroxamate **1**)<sup>2</sup> having very high intrinsic potency. However, other classes of MMP inhibitors based on alternative zinc ligands remain as attractive targets for drug discovery due to the high rates of clearance which have generally characterized the hydroxamates *in vivo*.



In this paper, we report on our effort to discover potent and novel carboxylate-based inhibitors of MMP-1. Simple substitution of hydroxamate for carboxylate in compounds such as **1** is known to markedly reduce activity against MMP-1. However, potent carboxylate inhibitors of MMP's are known; in particular, a series of N-carboxyalkyl dipeptides, exemplified by **2** and **3**, are potent inhibitors of MMP-1.<sup>3</sup> We found **2** to lack appreciable activity *in vivo* due to poor absorption and pharmacokinetics.<sup>4</sup> Nonetheless, we chose **2** as a starting point for an investigation of carboxylate inhibitors of MMP-1 reasoning that certain structural modifications might improve *in vivo* activity without loss of potency. In particular, replacement of the P<sub>1</sub>' NH for CH<sub>2</sub> appeared likely to improve absorption by decreasing the number of hydrogen bond donors.<sup>5,6</sup> Additionally, since the 2,3-naphthalimide group of **2** is susceptible to hydrolysis giving the corresponding carboxynaphthylamide and since

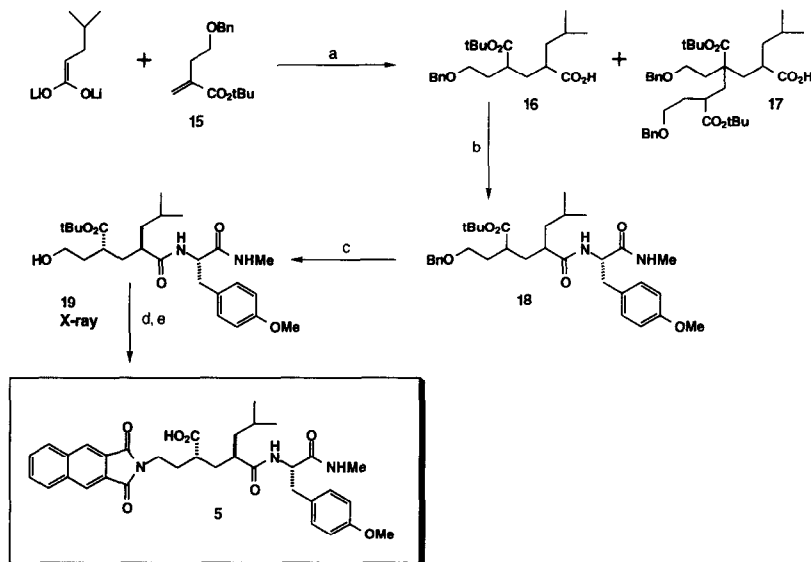
the mechanism of this hydrolysis is thought to involve autocatalysis by the P<sub>1</sub>' NH,<sup>2</sup> replacement of this NH group by CH<sub>2</sub> offered the potential for increased stability. The other structural modification we chose to explore was P<sub>1</sub>' C<sub>α</sub> gem-disubstitution which would hinder enzymatic hydrolysis of the P<sub>1</sub>'-P<sub>2</sub>' amide bond should this play a role in metabolism.<sup>6</sup> In the preceding paper, we examined the effect of P<sub>1</sub>' C<sub>α</sub> gem-disubstitution on the inhibition of MMP-1 by a series of succinamide hydroxamates.<sup>7</sup> While this substitution uniformly reduced potency against collagenase, P<sub>1</sub>' C<sub>α</sub> gem-cyclohexyl substitution was significantly less deleterious to activity than other P<sub>1</sub>' C<sub>α</sub> gem-disubstitutions, including gem-cyclopentyl. Thus, we were particularly interested in exploring P<sub>1</sub>' gem-cyclohexyl substitution, both in the N-carboxyalkylamine series and the related glutaramides (P<sub>1</sub>' NH replaced by CH<sub>2</sub>). Newly available X-ray crystallographic information (in particular, the crystal structure of the N-carboxyalkylamine **4** bound to 19 kDa collagenase),<sup>8</sup> in conjunction with molecular modeling studies, gave support to P<sub>1</sub>' C<sub>α</sub> gem-cyclohexyl substitution being an acceptable structural modification in both carboxylate series.

**Chemistry.** The glutaramide **5** in which the P<sub>1</sub>' NH of **3** is replaced by CH<sub>2</sub> was prepared as shown in Scheme 1. The C-C bond-forming step, involving conjugate addition of the lithiodianion of 4-methylvaleric acid to *tert*-butyl 1-(2-benzyloxyethyl)acrylate (**15**)<sup>9</sup>, gave **16** (a mixture of racemic diastereomers) as well as the unwanted bis-Michael adduct **17**. Separation of the components of this rather complex mixture was not attempted. However, after coupling with H-L-Tyr(Me)NHMe, chromatographic separation of **18** (four diastereomers) from the products arising from **17** could be achieved and, after hydrogenolysis of the O-benzyl group, partial chromatographic separation of the four diastereomeric alcohols was possible. Fortunately, the crystalline alcohol **19** eluted ahead of the other three diastereomers and workable quantities of this intermediate could be readily obtained in a pure state. An X-ray crystallographic analysis of **19** established the 2*R*, 4*S* stereochemistry corresponding to that of **2** and **3**. Final conversion of **19** to **5** proceeded uneventfully as shown.

The diastereomeric P<sub>1</sub>' C<sub>α</sub> gem-cyclohexyl glutaramides **7a,b** and the gem-cyclopentyl glutaramides **8a,b** were prepared by routes closely analogous to that in Scheme 1. In the case of gem-cyclopentyl, the conjugate addition of the lithiodianion of cyclopentanecarboxylic acid to **15** was quite efficient (78%). In contrast, the reaction of the lithiodianion of cyclohexanecarboxylic acid with **15** gave unacceptable yields of the desired conjugate addition product. However, by using the dilithio dienolate of cyclohexanecarboxylic acid, a 78% yield was obtained of **20** (a mixture of 2 diastereomers), the product of α-attack by the dienolate (Scheme 2). Following coupling to H-Tyr(Me)NHMe, hydrogenation of the cyclohexene double bond was achieved concurrently with hydrogenolysis of the O-benzyl ether. In both the gem-cyclohexyl and gem-cyclopentyl instances, separation of the C-4 diastereomers was achieved just before final cleavage of the *tert*-butyl ester. The stereochemistry at C-4 in compounds **7a,b** and **8a,b** was not assigned spectroscopically but could be inferred from the activity against collagenase (*vide infra*).

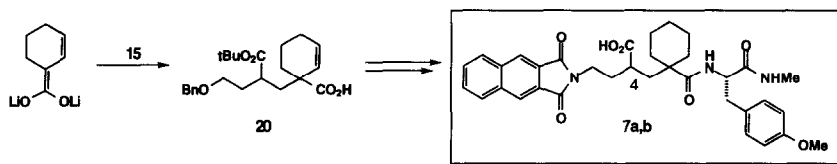
The substituted and unsubstituted phthalimide derivatives **9-11** were prepared similarly to **6a,b** by substituting 2,3-naphthalimide with the appropriate imide in the Mitsunobu step. Compound **12**, bearing only a methyl group at P<sub>1</sub>, was prepared analogously starting with the conjugate addition of cyclohexanecarboxylic acid dienolate to *tert*-butyl methacrylate. Compounds **13** and **14** were obtained by coupling of **20** to benzylamine and N-(2-aminoethyl)morpholine respectively, followed by hydrogenation (with accompanying hydrogenolysis of the benzyl ether), Mitsunobu reaction with 2,3-naphthalimide and *tert*-butyl ester cleavage.

Scheme 1



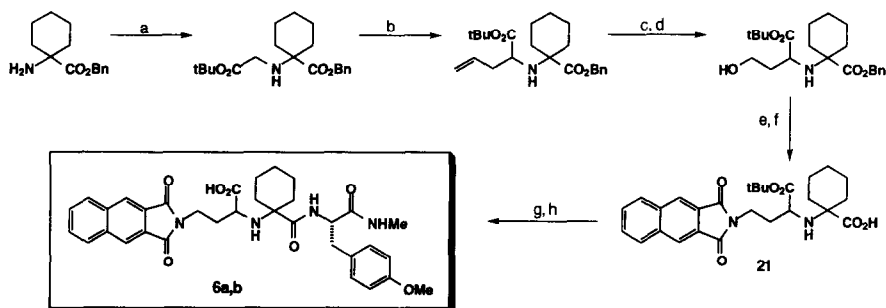
**Reagents and conditions:** a) THF/-78° (form dianion with 2.2 equiv. LDA); b) H-Tyr(Me)NHMe·HCl/DEC/HOBT/Et<sub>3</sub>N/CH<sub>2</sub>Cl<sub>2</sub>; c) H<sub>2</sub>/Pd(OH)<sub>2</sub>(C)/AcOH; d) n'imide/PPh<sub>3</sub>/DEAD/THF; e) TFA

Scheme 2



The diastereomeric P<sub>1</sub>' C<sub>α</sub> gem-cyclohexyl N-carboxyalkylamine analogues **6a,b** were prepared in a straightforward manner as shown in Scheme 3. Separation of the diastereomers was carried out by chromatography immediately prior to cleavage of the *tert*-butyl ester.

Scheme 3



**Reagents and conditions:** a) BrCH<sub>2</sub>CO<sub>2</sub>tBu/NaI/iPr<sub>2</sub>NEt/DMF/50°; b) LDA/THF/-78° then allyl bromide/-78° to 0°; c) NaO<sub>4</sub>/cat.OsO<sub>4</sub>/dioxane/H<sub>2</sub>O; d) NaBH<sub>4</sub>/MeOH; e) n'imide/PPh<sub>3</sub>/DEAD/THF; f) H<sub>2</sub>/10%Pd(C)/AcOH; g) H-Tyr(Me)NHMe·HCl/DEC/HOBT/Et<sub>3</sub>N/CH<sub>2</sub>Cl<sub>2</sub>; h) TFA

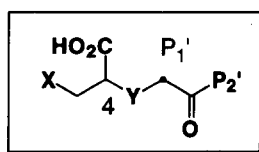


Table 1

Cmpd	X	Y	P <sub>1</sub> '	P <sub>2</sub> '	IC <sub>50</sub> (μM) <sup>12</sup>
					MMP-1 <sup>a</sup>
2 <sup>b</sup>	N'imide-CH <sub>2</sub> <sup>c</sup>	NH		PheNHMe	0.034 (n = 2)
3	N'imide-CH <sub>2</sub>	NH		Tyr(Me)NHMe	0.023 (0.0031; 3)
5 <sup>d</sup>	N'imide-CH <sub>2</sub>	CH <sub>2</sub>		Tyr(Me)NHMe	0.023 (0.014; 3)
6a <sup>e</sup>	N'imide-CH <sub>2</sub>	NH		Tyr(Me)NHMe	0.09 (0.039; 4)
6b <sup>e</sup>	N'imide-CH <sub>2</sub>	NH		Tyr(Me)NHMe	0.97 (0.46; 4)
7a <sup>e</sup>	N'imide-CH <sub>2</sub>	CH <sub>2</sub>		Tyr(Me)NHMe	0.038 (0.0065; 4)
7b <sup>e</sup>	N'imide-CH <sub>2</sub>	CH <sub>2</sub>		Tyr(Me)NHMe	0.14 (0.018; 4)
8a <sup>e</sup>	N'imide-CH <sub>2</sub>	CH <sub>2</sub>		Tyr(Me)NHMe	0.26 (0.12; 4)
8b <sup>e</sup>	N'imide-CH <sub>2</sub>	CH <sub>2</sub>		Tyr(Me)NHMe	0.94 (0.35; 4)
9 <sup>f</sup>		CH <sub>2</sub>		Tyr(Me)NHMe	3.2 (1.3; 3)
10 <sup>f</sup>		CH <sub>2</sub>		Tyr(Me)NHMe	0.57 (0.14; 3)
11 <sup>f</sup>		CH <sub>2</sub>		Tyr(Me)NHMe	0.14 (0.72; 3)
12 <sup>f</sup>	H	CH <sub>2</sub>		Tyr(Me)NHMe	>30 (n = 3)
13 <sup>g</sup>	N'imide-CH <sub>2</sub>	CH <sub>2</sub>		NHCH <sub>2</sub> Ph	3.0 (1.0; 3)
14 <sup>g</sup>	N'imide-CH <sub>2</sub>	CH <sub>2</sub>		NHCH <sub>2</sub> CH <sub>2</sub> N	12 (5.3; 3)

a) Avg. IC<sub>50</sub>s: μM (s.d.; n); b) R stereochemistry at C-4; c) N'imide = 2,3-naphthalimide; d) S stereochemistry at C-4; e) Single diastereomer, unassigned stereochemistry at C-4; f) 1:1 mixture of diastereomers; g) Racemic.

**Results and Discussion.** Replacement of the P<sub>1</sub>' NH group of **3** with CH<sub>2</sub> to give the glutaramide **5** did not result in a change in potency.<sup>10</sup> Both compounds exhibited IC<sub>50</sub>s = 0.023 μM (Table 1). The protonated P<sub>1</sub>' NH groups of **3** likely makes hydrogen bonds to Ala(182) and Glu(219) of MMP-1 based on the X-ray crystal structure of **4** bound in the active site of the 19 kDa enzyme.<sup>8</sup> Evidently, the free energy gained by removing the hydrophobic CH<sub>2</sub> of **5** from water can compensate for the loss of these hydrogen bonds.<sup>11</sup> Replacement of the P<sub>1</sub>' isobutyl of **3** for gem-cyclohexyl to give **6a** or **6b** was associated with decreased potency. However, assuming the more active isomer **6a** to have the same stereochemistry at C-4 as **3**, the loss of activity was only about 4-fold, a loss markedly less than that (80-fold) resulting from the same structural perturbation in the succinamide hydroxamate series.<sup>7</sup> Incorporation of a gem-cyclohexyl ring at P<sub>1</sub>' in the glutaramide series was even less detrimental to activity, the glutaramide **7a** (IC<sub>50</sub> = 0.038 μM) being only about 1.5 times less potent than the P<sub>1</sub>' isobutyl glutaramide **5**. We speculate that the increased activity of the glutaramides **7a** and **7b** relative to the N-carboxyalkylamines **6a** and **6b**, respectively, may reflect an ability of the bound glutaramides, being less constrained by hydrogen bonding, to more freely adopt a conformation favorable for projection of the cyclohexyl ring into the S<sub>1</sub>' pocket of the enzyme. Paralleling the structure-activity relationships seen in the P<sub>1</sub>' C<sub>α</sub> disubstituted succinamide hydroxamate series, the P<sub>1</sub>' gem-cyclopentyl glutaramides **8a,b** were less potent than the corresponding gem-cyclohexyl analogues (**7a,b**), although the difference in activity (about 7-fold) was not as large as that between the corresponding hydroxamates (about 20-fold).<sup>7</sup>

Compounds **9-14** were prepared to explore structural changes at P<sub>1</sub> and P<sub>2</sub>' in the gem-cyclohexyl glutaramide series. The goal here was to retain potency and, at the same time, maximize the potential for achieving oral bioavailability by reducing the molecular weight and the number of hydrogen bond donors. Activity against MMP-1 was very sensitive to the structure of the P<sub>1</sub> imide group as demonstrated by compounds **9-11**; compound **12**, bearing only a methyl group at P<sub>1</sub>, was essentially inactive. While the structure-activity relationships at P<sub>1</sub> reflect those seen with the corresponding analogues of **2**,<sup>3</sup> this does not appear to be true for analogues with structural modification at P<sub>2</sub>'. Thus, in the gem-cyclohexyl glutaramide series, replacement of the P<sub>2</sub>' Tyr(Me)NHMe residue with simple secondary amide groups (e.g., **13**, P<sub>2</sub>' = NHCH<sub>2</sub>Ph) gave poorly active compounds while in the N-carboxyalkylamine series P<sub>2</sub>' secondary amide analogues displayed good activity against MMP-1 (IC<sub>50</sub>'s <0.1 μM).<sup>3</sup>

**Conclusion.** Significant activity against MMP-1 (IC<sub>50</sub> < 0.1 μM) is retained following modification of the N-carboxyalkylamine **3** by independent replacement of the P<sub>1</sub>' NH group for CH<sub>2</sub> (giving **5**, equipotent to **3**) and introduction of P<sub>1</sub>' gem-cyclohexyl substitution (giving **6a**). Furthermore, the compound incorporating both these structural changes (glutaramide **7a**) also retains potent activity and is only slightly less active (1.5 -fold) than **3** or **5**. The latter finding significantly contrasts the 80-fold loss of potency against MMP-1 seen by replacement of P<sub>1</sub>' isobutyl for gem-cyclohexyl in the succinamide hydroxamate series.<sup>7</sup> This result underscores the care that must be taken in assuming parallel structure-activity relationships between inhibitors of different structural classes. To our knowledge compounds **6a** and **7a** are the first potent inhibitors of MMP-1 (IC<sub>50</sub> < 0.1 μM) having gem-disubstitution at P<sub>1</sub>'. The inclusion of this structural feature should be considered in designing inhibitors of MMP-1 and other matrix metalloproteinases.

**Acknowledgment.** We thank Drs. Lawrence Reiter and Todd Blumenkopf for their review of the manuscript and the Pfizer Scientific Advisory Committee for support of B. P. J. as an undergraduate research fellow.

### Footnotes and References

- 1) For recent reviews on matrix metalloproteinases and their inhibitors, see: (a) Morphy, J. R.; Millican, T. A.; Porter, J. R *Current Med. Chem.* **1995**, *2*, 743. (b) Beckett, R. P.; Davidson, A. H.; Drummond, A. H.; Huxley, P.; Whittaker, M. *Drug Discovery Today*, **1996**, *1*, 16.
- 2) Brown, F. K.; Brown, P. J.; Bickett, D. M.; Chambers, C. L.; Davies, H. G.; Deaton, D. N.; Drewry, D.; Foley, M.; McElroy, A. B.; Gregson, M.; McGeehan, G. M.; Myers, P. L.; Norton, D.; Salovich, J. M.; Schoenen, F. J.; Ward, P. *J. Med. Chem.*, **1994**, *37*, 674.
- 3) Dickens, J. P.; Donald, D. K.; Kneen, G.; McCay, W. R. U.S. Patent 4,599,361.
- 4) Lopez-Anaya, A. unpublished results.
- 5) Similar substitution of P<sub>1</sub>' NH with CH<sub>2</sub> in a series of N-carboxyalkyl dipeptide inhibitors of MMP-3 has recently been reported to improve oral activity : Chapman, K. T.; Durette, P. L.; Caldwell, C. G.; Sperow, K. M.; Niedzwiecki, L. M.; Harrison, R. K.; Saphos, C.; Christen, A. J.; Olszewski, J. M.; Moore, V. L.; MacCoss, M.; Hagmann, W. K. *Biorg. Med. Chem. Lett.* **1996**, *6*, 803.
- 6) Introduction of P<sub>1</sub>' C $\alpha$  gem-disubstitution and substitution of P<sub>1</sub>' NH with CH<sub>2</sub> were key structural modifications of an N-carboxyalkylamine leading to the discovery of candoxatril, an orally active prodrug of the NEP inhibitor candoxatrilat: (a) James, K.; Alabaster, C. T.; Barclay, P. L.; Barnish, I. T.; Blackburn, K. J.; Brown, D.; Campbell, S. F.; Cussans, N. J.; Danilewicz, J. C.; Palmer, M. J.; Terrett, N. K.; Samuels, G. M. R.; Wythes, M. J. *Perspect. Med. Chem.* **1993**, *45*. (b) Danilewicz, J. C.; Barclay, P. L.; Barnish, I. T.; Brown, D.; Campbell, S. F.; James, K.; Samuels, G. M. R.; Terrett, N. K.; Wythes, M. J. *Biochem. Biophys. Res. Comm.* **1989**, *164*, 58.
- 7) Robinson, R. P.; Ragan, J. A.; Cronin, B. J.; Donahue, K. M.; Lopresti-Morrow, L. L.; Mitchell, P. G.; Reeves, L. M.; Yocum, S. A. preceding paper in this issue.
- 8) Lovejoy, B.; Cleasby, A.; Hassell, A. M.; Longley, K.; Luther, M. A.; Weigl, D.; McGeehan, G.; McElroy, A. B.; Drewry, D.; Lambert, M. H.; Jordan, S. R. *Science* **1994**, *263*, 375.
- 9) Prepared by a 3 step sequence starting from *tert*-butyl ethyl malonate: 1) alkylation with 1-benzyloxy-2-bromoethane (NaH/DMF/25°); 2) ethyl ester hydrolysis (KOH/H<sub>2</sub>O/dioxane/ 25°); 3) Mannich reaction of the mono-acid with paraformaldehyde and piperidine (pyridine/70°); see Stetter, H.; Kuhlmann, H. *Synthesis* **1979**, 30.
- 10) Replacement of P<sub>1</sub>' NH for CH<sub>2</sub> was generally associated with a modest loss of potency in a series of glutaramide-based inhibitors of MMP-3 (ref. 5).
- 11) (a) Holland, D. R.; Barclay, P. L.; Danilewicz, J. C.; Matthews, B. W.; James, K. *Biochemistry* **1994**, *33*, 51. (b) Morgan, B. P.; Scholtz, J. M.; Ballinger, M. D.; Zipkin, I. D.; Bartlett, P. A. *J. Am. Chem. Soc.* **1991**, *113*, 297.
- 12) Enzyme assay: Recombinant full-length MMP-1 was activated with trypsin and assayed using a quenched fluorescent peptide substrate: Bickett, D. M.; Green, M. D.; Berman, J.; Dezube, M.; Howe, A. S.; Brown, P. J.; Roth, J. T.; McGeehan, G. M. *Anal. Biochem.* **1993**, *212*, 58.

(Received in USA 20 May 1996; accepted 24 June 1996)