PREPARATION OF BOTH ENANTIOMERS OF A CHIRAL LACTONE THROUGH COMBINED

MICROBIOLOGICAL REDUCTION AND OXIDATION.

An application of an enantioselective biological Baeyer-Villiger reaction

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<u>Abstract</u>: The Baeyer-Villiger-like oxidation of (**R**,**S**)-2,2,5,5-tetramethyl-4-hydroxy -cyclohexanone by several fungal strains was highly enantioselective, affording a rearranged (**S**)-hydroxy-γ-lactone. The recovery of the nearly optically pure (**R**)-hydroxyketone allowed its conversion to the enantiomeric (**R**)-hydroxylactone through a classical Baeyer-Villiger oxidation.

The microbiological reduction of 2,2,5,5-tetramethyl-1,4-cyclohexanedione <u>1</u> to the optically pure (S)-ketol <u>2</u>, a valuable synthon for the synthesis of optically active <u>cis</u>-chrysanthemic acid, has been precedently described^{1,2}. When submitted to <u>m</u>-chloroperbenzoic acid oxidation, in the conditions of a usual Baeyer-Villiger reaction, the (S)-ketol <u>2</u> is converted quantitatively to a presumed seven-membered lactone <u>3</u> (Scheme 1) which rearranges spontaneously to the hydroxy- γ -lactone <u>4</u>, $[\alpha]_{D}^{21}$ -110° (c 1.6, MeOH), retaining the initial chirality²; <u>4</u> can be easily dehydrated by POCl₃ treatment in HMPA-pyridine³ to a 2:1 mixture of unsaturated (S)-lactones <u>5</u>, $[\alpha]_{D}^{21}$ -107.5° (c 1, MeOH) and <u>6</u>, mp 48-49°C (from EtOAc-cyclohexane), $[\alpha]_{D}^{21}$ -86.6° (c 0.7, MeOH)⁴⁻⁶.



Scheme 1: i, reduction by C.lunata; ii, MCPBA/CH₂Cl₂; iii, POCl₃, C₆H₅N/HMPA,100°C.

As the hydroxylactone $\underline{4}$ and the unsaturated lactones $\underline{5}$ and $\underline{6}$ may constitute other valuable chiral synthons, or at least interesting models for the preparation of asymmetric lactones, it was worthwhile to have a method for the preparation of both enantiomers. As none of the fungal or yeast strains tested were able to reduce the dione $\underline{1}$ to the (R)-ketol², the dione was submitted first to <u>m</u>-chloroperbenzoic acid (1 equiv.) oxidation to give 75% of the keto-lactone $\underline{7}$ ⁵, mp 129-130°C (from cyclohexane), which, in turn, was submitted to fungal reduction (Scheme 1): the optically pure (S)-hydroxylactone 4 was again obtained.

On the other hand, it was observed that, during the fungal reduction of the dione to the ketol, if glucose (the reducing equivalent source) was omitted, the already formed (S)-ketol $\underline{2}$ was slowly converted to the same hydroxylactone $\underline{4}$, indicating the occurrence of an oxygenase-mediated reaction analogous to the Baeyer-Villiger oxidation. Such a reaction has been occasionally described for bacterial strains adapted to growth on cyclic alkanes, ketones or alcohols^{7,8}, or fungi acting on steroidal ketones⁹; the camphor ketolactonase complex¹⁰, and more recently cyclohexanone oxygenase¹¹, both flavoprotein enzymes, have been extensively purified and the mechanism of oxygen insertion closely examined¹². Only scattered informations about the stereospecificity of these enzymes, and their use for the synthesis of chiral lactones, have been published: for example, the specificity of camphor ketolactonases is complete and exclusive towards the D(+) or the L(-) camphor enantiomers, but much less exacting with respect to the substituants in the 5-position¹³. As a rule, these monooxygenases seem to participate in induced degradative pathways which are used by microorganisms to metabolize cyclic alkane, alkanone or alkanol molecules.



Scheme 2: i, <u>C.lunata</u>; ii, MCPBA/CH₂Cl₂; iii, POCl₃, C₆H₅N/HMPA, 100°C.

It was found, after optimizing the incubation conditions, specially by increasing the aeration of the culture medium, that the oxidation of the (S)-ketol $\underline{2}$ was fairly rapid, allowing a quantitative accumulation of the lactone $\underline{4}$, without apparent further degradation. Moreover, when a 2 g/liter solution of the racemic ketol $\underline{rac-2}$ (obtained by careful reduction of the dione $\underline{1}$ with limited amounts of NaBH₄ ¹⁴) was submitted to the oxidation by a Curvularia lunata suspension ¹⁵, the (S)-ketol disappeared rapidly (fig.1), leaving in a few



Figure 1: Time course of the conversion of racemic 2,2,5,5-tetramethyl -4-hydroxy-cyclohexanone to the (S)-hydroxylactone 4. Aliquots from the incubation with C.lunata were extracted, derivatized with isopropyl isocyanate and chromatographed at 170°C on a chiral column 4.

days the unchanged nearly optically pure (**R**)-ketol. Thus it appears that the Baeyer-Villiger-like oxidation of <u>rac-2</u> by <u>C.lunata</u> is able to discriminate between two stereoisomers differing only in a stereochemical feature located in a relatively remote position from the reaction center. After usual processing of the extracted mixture, a simple chromatographic separation on silicagel H_{60} (cyclohexane-EtOAc, 6:4) afforded the (**S**)-lactone <u>4</u> ($[\alpha]_D^{21}$ -89°, c 1.5, MeOH, 63% yield) and the unchanged (**R**)-ketol ($[\alpha]_D^{21}$ +86.9°, c 1.25, MeOH , 97% ee, 83% yield), which was then treated by <u>m</u>-chloroperbenzoic acid in the usual conditions to give the (**R**)-hydroxylactone <u>ent-4</u>, $[\alpha]_D^{21}$ +105.9° (c 0.96, MeOH); this lactone was in turn converted to the (**R**)-lactones <u>ent-5</u>, $[\alpha]_D^{21}$ +107.1° (c 0.57, MeOH) and <u>ent-6</u>, $[\alpha]_D^{21}$ +84° (c 0.6, MeOH), by the same dehydrating treatment³ (Scheme 2). The reactions described constitute a useful synthesis of some alkyl substituted chiral lactones and are currently extended to other less substituted prochiral 1,4- and 1,3-cyclic diketones, the reduction/oxidation of which should allow the generation of optically pure lactones containing more than one asymmetric carbon atom.

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References and Notes

1- D.Buisson, R.Azerad, G.Revial and J.d'Angelo, <u>Tetrahedron Lett.</u>, **25**, 1984, 6005. 2- J.d'Angelo, G.Revial, R.Azerad and D.Buisson, <u>J.Org.Chem.</u>, **51**, 1986, 40. 3- S.Torii, T.Inokuchi and R.Oi, <u>J.Org.Chem.</u>, **48**, 1983, 1944. The method described in the same reference and using a rhodium salt catalyst for a quantitative conversion of methylene-compounds like <u>5</u> to dimethylallyl-compounds like <u>6</u> was inefficient; however, slightly modified dehydrating conditions (pyridine added before POCl₃) afforded lactones <u>5</u> and <u>6</u> in a 91:9 ratio. On the contrary, a 14:86 ratio was obtained by refluxing the lactone <u>4</u> with <u>p</u>-toluenesulfonic acid in benzene.

4- Optical purity of products were best assessed by GL-chromatographic separation of isopropyl urethane derivatives (see ref.16) of enantiomers on a Chrompack (25 m x 0.25 mm) XE-60-polysiloxane S-valine-S- α -phenylethylamide column, operated at 170 °C with helium (1.5 bar) as carrier gas (compound, absolute configuration, retention time): lactone <u>4</u>, S, 52.5 min₁; R, 51.8 min.; ketol <u>2</u>(see ref.2), S, 17.0 min.; R, 17.4 min.

min; **k**, 51.8 min.; ketol 2(see ref.2), **s**, 1/.0 min.; **k**, 1/.4 min. 5- H-NMR data (250 MHz, CDCl₃): 4, δ 4.32 (dd, 1H,-CH-O, J₁= 10, J₂= 2 Hz), 2.40, 2.28 (2d, 2H,-CH₂-CO-, J= 17 Hz), 1.74 (dd, 1H,-CH-O, J₁= 15 Hz, J₂= 10 Hz), 1.68 (s, 1H,-OH), 1.62 (dd, 1H,-CH-, J₁= 15 Hz, J₂= 2 Hz), 1.30, 1.28, 1.12, 0.99 (4s, 12H,-CH₃). 5, δ 4.86, 4.83 (2m, 2H,=CH₂), 4.22 (dd, 1H,-CH-O, J₁= 16.5, J₂= 10 Hz), 2.41, 2.30 (2d, 2H,-CH₂-CO-, J= 16.5 Hz), 2.23 (m, 2H,-CH₂), 1.79 (s, 3H,CH₃-C=), 1.16, 1.03 (2s, 6H,-CH₃). 6, δ 5.18 (dq, 1H,-CH=, J₁= 9.5, J₂= 1.3 Hz), 4.80 (d, 1H,-CH-O, J= 9.5 Hz), 2.41, 2.32 (2d, 2H,-CH₂-CO-, J= 16.7 Hz), 1.79, 1.71 (2d, 6H, CH₃-C=, J= 1.3 Hz), 1.11, 1.0 (2s, 6H,-CH₃). 7, δ ²2.98, 2.75 (2s, 4H, -CH₂-O), 1.49, 1.20 (2s, 12H,-CH₃).

6- Analýtical samples of 5 and 6 were obtained by preparative GLC (10% XE60 on Gerapack 30 column, 3.3 m x 0.6 cm, carrier gas He, temperature 150°C).

7- W.H.Bradshaw, H.E.Conrad, E.J.Corey, I.C.Gunsalus and D.Lednicer, J.Am.Chem.Soc., 81, 1959, 5507; H.E.Conrad, R.DuBus and I.C.Gunsalus, <u>Biochem.Biophys.Res.Commun.</u>, 6, 1961, 293.
8- D.B.Norris and P.W.Trudgill, <u>Biochem.J.</u>, 121, 1971, 363; M.Griffin and P.W.Trudgill, <u>Biochem.J.</u>, 129, 1972, 595; Y.Hasegawa, K.Hamano, H.Obata and T.Tokuyama, <u>Agric.Biol.Chem.</u>, 46, 1982, 1139; M.K.Trower, R.M.Buckland, R.Higgins and M.Griffin, <u>Appl.environ.Microbiol.</u>, 49, 1985, 1282.

9- J.Fried, R.W.Thoma and A.Klingsberg, <u>J.Am.Chem.Soc.</u>, **75**, 1953, 5764; D.H.Peterson, S.H.Eppstein, P.D.Meister, H.C.Murray, H.M.Leigh, A.Weintraub and L.M.Reineke,

J.Am.Chem.Soc., **75**, 1953, 5768; R.L.Prairie and P.Talalay, <u>Biochemistry</u>, **2**, 1963, 203; E.Itagaki, <u>J.Biochem.</u>, **99**, 1986, 815, 825.

10- C.A.Yu and I.C.Gunsalus, <u>J.Biol.Chem.</u>, 244, 1969, 6149; D.G.Taylor and P.W.Trudgill, <u>J.Bacteriol.</u>, 165, 1986, 489.

11- N.A.Donoghue, D.B.Norris and P.W.Trudgill, Europ.J.Biochem., 63, 1976, 175.

12- C.C.Ryerson, D.P.Ballou and C.Walsh, Biochemistry, 21, 1982, 2644.

13- H.E.Conrad, R.DuBus, M.J.Namvedt and I.C.Gunsalus, J.Biol.Chem., 240, 1965, 495 and ref. cited therein.

14- The optimal conditions for the monoreduction of 1 were found with 1.5 equiv. of NaBH, in $EtOH-H_0O$ solution (yield 80-90%, after silicagel chromatographic separation from the diol and residual dione). For a quantitative reduction of the monoenolate, see J.d Angelo and G.Revial, <u>Tetrahedron Lett.</u>, 24, 1983, 2103.

15- <u>Curvularia lunata NRRL</u> 2380 was grown at 27°C in a liquid medium (see ref.2). After 72 hours, the grown mycelium was recovered by filtration and washed with a 0.3 M pH 7.0 phosphate buffer. The mycelium was resuspended in half the culture volume of the same buffer, added with the ketol and agitated in an orbital shaker at 27°C and 330 rpm. 16- W.A.König, W.Franck and I.Benecke, J.Chromatogr., 239, 1982, 227.