STUDIES OF THE KNOEVENAGEL REACTION OF 2,5-ANHYDRO-3,4-O-ISOPROPYLIDENE-D-ARABINOSE

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ABSTRACT

The Knoevenagel reactions of 2,5-anhydro-3,4-O-isopropylidene-Darabinose with methyl acetoacetate or 2,4-pentanedione were studied in connection with the further cyclisation of the reaction products to the spiro-glycosides (4R,5R,8R)-2,2-dimethyloxolano[4,3-d][1,3]dioxolane-5-spiro-2'-(4-methoxycarbonyl-5-methyl-2,3-dihydrofuran), (4R,5S,8R)-2,2-dimethyloxolano[4,3-d]-[1,3]dioxolane-5-spiro-2'-(4-methoxycarbonyl-5-methyl-2,3-dihydrofuran), and (4R,5S,8R)-2,2-dimethyloxolano[4,3-d][1,3]dioxolane-5-spiro-2'-(4-acetyl-5-methyl-2,3-dihydrofuran). A different C-glycoside, (4R,5S)-5-(2,3-O-isopropylidene- α -D-erythrofuranosyl)-4-methoxycarbonyl-3-methylcyclohex-2-en-1-one, results from the prolonged Knoevenagel reaction of 1 with 2 equiv. of methyl acetoacetate. Configurations and conformations have been assigned on the basis of ¹H-n.m.r. data.

INTRODUCTION

The Knoevenagel reaction of aldehydo sugars with active methylene compounds has been studied in connection with the syntheses of modified carbohydrates¹. We have reported² several examples of this reaction on 2,3-O-isopropylidene-D-glyceraldehyde, including the first examples of spiro compounds resulting from the easy cyclisation of the normal reaction products (the α,β - and β,γ -unsaturated derivatives and the corresponding enols).

The reaction has been extended³ to other aldehydo sugars, and we now report results with 2,5-anhydro-3,4-O-isopropylidene-D-arabinose⁴ (1).

RESULTS AND DISCUSSION

The reaction of 1 with an equimolecular amount of methyl acetoacetate in the presence of piperidine as catalyst gave 63% of a mixture of the isomers 2-4. ¹H-N.m.r. spectroscopy indicated this mixture to contain 2 (~25%), 3 (~60%), and 4 (~15%). The greater abundance of 3 may be explained by greater steric hindrance



at the α face in 2, which is lessened in 3 by formation of the exocyclic double-bond.

Similar reaction of 1 with 2,4-pentanedione gave the corresponding mixture 5–7, but with a higher proportion of the enolic form 7 (~90%), probably because of minor steric hindrance of the exocyclic double-bond, as noted above, and the higher enolic character of β -diketones.

The outcome of the treatment of the mixture 2-4 with trifluoroacetic acid in carbon tetrachloride depended on the temperature and reaction time. Thus, for a reaction time of ~45 min at room temperature with 2% acid, the two isomers 8 and 9 were formed and isolated by column chromatography. Longer reaction times, higher temperatures, or higher concentrations of acid gave only the thermodynamic product 9. Treatment of the kinetic product 8 with 0.01% acid effected almost quantitative isomerisation into 9 (g.l.c. analysis).

The configurations at the spiro-carbon (anomeric centre) in 8 and 9 were assigned tentatively on the basis of the following considerations. The quasi E_0 conformation reported⁵ for several 2,3-O-isopropylideneglycofuranosyl C-glycosides having endo substituents at C-1 and/or C-4 suggests the same conformation for 8 and 9. An E_0 conformation with the spiro-carbon slightly up can resemble the more stable conformer, as shown in 13 and 14. The two hydrogens of the dihydrofuran methylene group of 14 have non-symmetrical environments, as reflected by the ¹Hn.m.r. data. The more polar isomer 13 (by t.l.c.) would be expected to have O-1' endo, as in some α -D-ribofuranosides⁶.

Greater differences in chemical shift between the signals for the isopropylidene methyl groups would be expected for 13 because of the deshielding effect of the *endo* O-1' on the *endo*-methyl group, which accords with the ¹H-n.m.r. data. The higher thermodynamic stability of 9 (14) reflects the anomeric effect associated with glycosides having O-1,2 *trans*-diaxial.

Likewise, treatment of the mixture 5-7 with trifluoroacetic acid in carbon tetrachloride yielded a single compound, which was characterised as (4R, 5S, 8R)-



2,2-dimethyloxolano[4,3-d][1,3]dioxolane-5-spiro-2'-(4-acetyl-5-methyl-2,3-dihyd-rofuran) (11); isomer 10 was not detected even under very mild conditions of reaction. The ¹H-n.m.r. spectrum of 11 was very similar to that of 9; consequently, the same spiro configuration was assigned.

Basic hydrolysis of 9 gave the expected spiro-acid 12 in excellent yield, and its stability was consistent with an enolic derivative of a β -keto acid. The similarity of the ¹H-n.m.r. spectra of 9, 11, and 12 allowed the assignment of configuration to 12, which was confirmed by the conversion of 11 into 12 by the haloform reaction.

When 1 was treated under Knoevenagel reaction conditions with 2 equiv. of methyl acetoacetate for a prolonged period, the main product was crystalline and the structure 15 was assigned by analogy with similar products⁷ of the Knoevenagel reaction.



Fig. 1. 200-MHz, 1-D ¹H-N.m.r. spectrum of **15**: $H_a = H-2$, $H_b = H-3'$, $H_c = H-2'$, $H_d = H-4'$ endo, $H_c = OMe$, $H_f = H-4$, $H_g = H-4'$ exo, $H_h = H-1'$, $H_1 = H-5$, $H_j = H-6$ cis to CO_2Me , $H_k = H-6$ trans to CO_2Me , $H_1 = Me-C=C$, H_m endo-Me of CMe_2 , H_n exo-Mc of CMe_2 .



Fig. 2. 200-MHz, 2-D ¹H-n.m.r. data (WP-200SY) for 15; J-resolved 2-D experiment.

The ¹H-1-D (Fig. 1) and ¹³C-n.m.r. spectra of 15 confirmed the assigned structure as well as the configuration and conformation of the sugar moiety. The coupling constants $J_{1',2'}$ 3.5, $J_{2',3'}$ 6.25, $J_{3',4'endo}$ 0, and $J_{3',4'exo}$ 3.5 Hz are characteristic of a 2,3-O-isopropylidene- α -glycofuranosyl C-nucleoside derivative with a quasi E_0 conformation (C-2 slightly up) that lessens the interaction of the bulkier endo-aglycon group. The alternative ⁰E conformation does not accord with the $J_{3',4'endo}$ value of 0 Hz, whereas the $J_{1',2'}$ value of 3.5 Hz is characteristic of α -⁰E structures.

Configurational and conformational analysis of the aglycon was based on 200-MHz, J-resolved, 2-D (Fig. 2) and homonuclear, scalar-shift-correlated experiments (COSY, Fig. 3, and NOESY, Fig. 4). Fig. 2 shows the J-resolved cross-sections for the 13 multiplets present [methyl group (e) at 3.652 p.p.m. shows no coupling]; the multiplets **f**,**g**,**h**, and **i**,**j** were strongly overlapping in the normal 1-D spectrum (Fig. 1). The signal assignments were made by considering the shift correlations shown in Fig. 2, where several important correlations are labelled.

Several important points can be made about 15. Numerous long-range couplings were detected which were not resolvable in the 1-D spectrum, for example, **a**-l, **f**-l, **f**-i, **f**-k, **h**-k, **h**-f, **i**-h, **i**-k, and **i**-j. Due to these couplings, the complexity of signals **a**-l is increased (Fig. 2). In the corresponding stacked plot of Fig. 3, a few weak correlations (**a**-f, **b**-d, **c**-d) can be seen that do not appear in the contour



Fig. 3. Scalar shift-correlated experiment for 15 (COSY-90, N-type selection). Contour plot.





Fig. 4. Symmetry correlation of NOESY experiment for 15, to remove F_1 -noise and quad imperfections.

plot. These weak couplings can also be observed in Fig. 2. The methyl groups **m** and **n** are coupled and show distinguishable n.O.e. effects (Fig. 4), but their assignments are still tentative.

The configurations of the chiral centers C-4 and C-5 as well as the conformation in the cyclohex-2-en-1-one moiety in 15 result in a half-twist conformation, similar to that accepted for cyclohex-2-en-1-one, having C-1,2,3,4,6 in plane and C-5 out of plane. Taking into account the long-range coupling observed between **f** and **k** (suggesting W-geometry) and the absence of a $J_{5,6}$ trans-diaxial coupling constant, only two spatial representations (16 and 17) seemed to be logical, each of which has a trans-diaxial relationship at C-4,5. Structure 17 was rejected on the basis of the n.O.e. effects observed (Fig. 4) between **m** and **e**, and between **n** and **l**, which are compatible only with the absolute configuration 16 for the cyclohex-2en-1-one moiety.

EXPERIMENTAL

General methods. — Melting points were determined with a Gallenkamp instrument and are uncorrected. Optical rotations were measured with a Perkin-Elmer 241 polarimeter, using a 1-dm standard cell. I.r. spectra were recorded with a Beckman Aculab 4 spectrophotometer. U.v. spectra were recorded with a Beckman DB-GT spectrophotometer. ¹H-N.m.r. spectra were recorded for solutions in CCl₄ or CDCl₃ (internal Me₄Si) with a Perkin-Elmer Hitachi R-24B (60 MHz) or Bruker WP-200SY (200 MHz) spectrometer. The ¹³C-n.m.r. spectrum was recorded with a Bruker WP-200SY spectrometer. Chemical shifts are given on the δ scale, and coupling constants in Hz. The mass spectrum was recorded with a Kratos MS-25 spectrometer. Column chromatography was performed on Silica Gel 60 (0.063–0.200 mm) (Merck). T.I.c. was performed on Silica Gel GF₂₅₄ (Merck) with detection by u.v. absorption or by charring with sulphuric acid. All evaporations were performed in a rotary evaporator under diminished pressure at 40°. G.l.c. was carried out on a Hewlett-Packard 5710A chromatograph equipped with a flame-ionisation detector and a stainless-steel column (50.0 cm \times 3.0 mm i.d.) packed with 10% of UCW-982 on Chromosorb WAW-DMCS B 79 (80-100 mesh). The helium flow-rate was 30 mL/min, the injection-port temperature 250°, and the zone-detector temperature 300°.

Piperidine-catalysed reaction of 2,5-anhydro-3,4-O-isopropylidene-Darabinose (1) with methyl acetoacetate. — When 1 (11.6 g, 67 mmol), methyl acetoacetate (7.83 g, 67.5 mmol), and piperidine (0.6 mL) were mixed, the temperature rose to 50°. The mixture was kept in the dark for 24 h, and then distilled to yield 1 (2.86 g), and a second fraction (11.36 g, 63%), b.p. 120–130°/0.2 Torr, which appeared to be a mixture of (a) the α,β -unsaturated ketoester 2 [¹H-n.m.r. data (CCl₄): δ 6.82, 6.68 (2 d, J 6 Hz, H-3 of Z- and E-isomers), 2.29, and 2.24 (2 s, MeCO of Z- and E-isomers)]; (b) the β,γ -unsaturated ketoester 3 [¹H-n.m.r. data: δ 2.15 and 2.10 (2 s, MeCO of Z- and E-isomers)]; and (c) the enol 4 [¹Hn.m.r. data: δ 12.5 (bs, OH), and 1.92 (s, Me-C=)].

The n.m.r. data indicated the mixture to contain **2** (25%), **3** (60%), and **4** (15%). It had $\nu_{\text{max}}^{\text{film}}$ 3300–3200, 2990, 2960, 1740, 1720, 1630, 1430, 1370, 1240, 1200, 1140, 1080, 1050, 980, 850, and 720 cm⁻¹.

Anal. Calc. for C₁₃H₁₈O₆: C, 57.77; H, 6.71. Found: C, 57.96; H, 6.89.

Piperidine-catalysed reaction of 1 *with pentane-2,4-dione.* — This reaction was performed as described above, starting from 1 (2.6 g, 15 mmol), pentane-2,4-dione (1.54 g, 13.5 mmol), and piperidine (0.25 mL), to give 1 (0.6 g, 23%) and a mixture of isomers 5–7 (1.6 g, 48.2% from unrecovered 1), b.p. 130–140°/0.1 Torr. The n.m.r. data indicated 7 to be the principal product (>90%), which had ν_{max}^{film} 3400–3200, 2990, 2920, 1720, 1680, 1620, 1450, 1420, 1380, 1270, 1220, 1150, 1100, 1050, 990, 850, and 720 cm⁻¹. ¹H-N.m.r. data (CCl₄) for 7: δ 16.2 (bs, OH), 5.30 (s, H-3), 2.05 (s, Me-C=), 1.98 (s, MeCO), 1.38, and 1.25 (2 s, Me₂C).

Anal. Calc. for C₁₃H₁₈O₅: C, 61.40; H, 7.14. Found: C, 61.65; H, 7.14.

(4R,5S,8R)-2,2-Dimethyloxolano[4,3-d][1,3]dioxolane-5-spiro-2'-(4-methoxycarbonyl-5-methyl-2,3-dihydrofuran) (9) and its 5(R) epimer (8). — A solution of the mixture 2-4 (2 g, 7.4 mmol) in CCl₄ (10 mL) and trifluoroacetic acid (0.2 mL) was kept at room temperature. After 45 min, t.l.c. (ethyl acetate-hexane, 1:3) revealed two products (R_F 0.4 and 0.6). The mixture was then washed with saturated, aqueous Na₂CO₃ (10 mL) and cold water (10 mL), dried (Na₂SO₄), filtered, and concentrated. Column chromatography (ethyl acetate-hexane, 1:4) of the residue on silica gel (150 g) gave 9 (0.588 g, 30%) and 8 (0.664 g, 34%).

Compound **8** had m.p. 52–54° (from ethyl acetate–hexane), $[\alpha]_{\rm D}^{20}$ +20° (*c* 1, chloroform), $R_{\rm F}$ 0.4 (ethyl acetate–hexane, 1:3); $\lambda_{\rm max}^{\rm EtOH}$ 245 nm (ε 26,000); $\nu_{\rm max}^{\rm film}$ 2990–2925, 2850, 1700, 1650, 1460, 1445, 1385, 1350, 1295, 1260, 1245, 1220, 1200, 1160, 1120, 1100, 1080, 1020, 980, 940, 900, 860, 800, 780, 760, and 680 cm⁻¹. ¹H-N.m.r. data (200 MHz, CDCl₃): δ 4.88 (ddd, 1 H, J 7, 5, and 3 Hz, H-8), 4.52 (d, 1 H, J 7 Hz, H-4), 4.20 (dd, 1 H, J 10.5 and 5 Hz, H-7), 4.10 (dd, 1 H, J 10.5 and 3 Hz, H-7'), 3.70 (s, 3 H, MeO), 3.01, 2.93 (2 dq, 2 H, J 16 and 2 Hz, CH₂-C=), 2.27 (t, 3 H, J 2 Hz, Me-C=), 1.60, and 1.37 (2 s, 6 H, Me₂C).

Anal. Calc. for C₁₃H₁₈O₆: C, 57.77; H, 6.71. Found: C, 58.19; H, 6.56.

Compound **9** had m.p. 72–74° (from hexane), $[\alpha]_D^{20} = -150°$ (*c* 0.1, chloroform), $R_F 0.6$; $\lambda_{max}^{MeOH} 245$ nm ($\varepsilon 16,000$); $\nu_{max}^{KBT} 2950$, 2900, 2850, 1700, 1650, 1460, 1440, 1380, 1330, 1310, 1280, 1270, 1230, 1210, 1190, 1160, 1135, 1100, 1090, 1065, 1045, 1000, 965, 930, 890, 865, 840, 820, 790, 765, 750, and 675 cm⁻¹. ¹H-N.m.r. data: $\delta 4.84$ (ddd, 1 H, J 6, 3.5, and 1 Hz, H-8), 4.50 (d, 1 H, J 6 Hz, H-4), 3.96 (dm, 1 H, J 10 Hz, H-7endo), 3.90 (dd, 1 H, J 10 and 3.5 Hz, H-7exo), 3.63 (s, 3 H, MeO), 3.20 (dq, 1 H, J 16.5 and 2 Hz, H-3'endo), 2.78 (dq, 1 H, J 16.5 and 2 Hz, H-3'exo), 2.12 (t, 3 H, J 2 Hz, Me-C=), 1.40, and 1.27 (2 s, 6 H, Me₂C).

Anal. Found: C, 57.90; H, 6.78.

Treatment of the mixture 2-4 (3 g) with boiling CCl_4 (15 mL) and trifluoroacetic acid (0.3 mL) for 30 min afforded 9 (2 g, 67%; after recrystallisation from hexane).

A solution of **8** (2.16 mg) in CCl₄ (0.2 mL) was added to a solution (2 μ L) of 1% trifluoroacetic acid in CCl₄, and the reaction was monitored by g.l.c. (oven temperature, 200°). After 42 min, practically complete epimerisation (99.8%) of **8** (T 2.78 min) into **9** (T 1.90 min) had occurred.

(4R,5S,8R)-2,2-Dimethyloxolano[4,3-d][1,3]dioxolane-5-spiro-2'-(4-carboxy-5-methyl-2,3-dihydrofuran) (12). — Aqueous 10% potassium hydroxide (4 mL) was added to a solution of 9 (0.5 g, 1.85 mmol) in ethanol (2 mL). After stirring for 50 h, t.l.c. (ethyl acetate-hexane, 1:2) showed the absence of 9. The reaction mixture was then extracted with ether (2 × 3 mL), the aqueous layer was acidified (Congo Red) with 10% hydrochloric acid, and the resulting solid was collected, washed with cold water, and recrystallised from ethanol or ether, to give 12 (400 mg, 85%), m.p. 173–175°, $[\alpha]_D^{20} - 177.5°$ (c 0.1, chloroform); λ_{max}^{EtOH} 243 nm (ϵ 26,000); ν_{max}^{EB} 3400–3200, 2990, 2920, 2850, 2650, 2590, 1660, 1620, 1435, 1375, 1365, 1320, 1300, 1260, 1235, 1215, 1190, 1175, 1150, 1125, 1080, 1050, 1030, 990, 960, 945, 875, 850, 835, 805, 780, 765, 740, and 660 cm⁻¹. ¹H-N.m.r. data (60 MHz, CDCl₃): δ 11.20 (bs, 1 H, COOH), 4.80 (m, 1 H, H-8), 4,45 (d, 1 H, J 6 Hz, H-4), 3.90 (m, 2 H, H-7,7'), 3.25 (dq, 1 H, J 16 and 2 Hz, H-3'*endo*), 2.80 (dq, 1 H, J 16 and 2 Hz, H-3'*exo*), 2.14 (t, 3 H, J 2 Hz, Me-C=), 1.41, and 1.30 (2 s, 6 H, Me₂C).

Anal. Calc. for C₁₃H₁₆O₆: C, 56.24; H, 6.29. Found: C, 56.26; H, 6.30.

(4R, 5S, 8R)-2,2-Dimethyloxolano[4,3-d][1,3]dioxolane-5-spiro-2'-(4-acetyl-5-methyl-2,3-dihydrofuran) (11). — The mixture of 5–7 (2.7 g), CCl₄ (25 mL), and trifluoroacetic acid (0.5 mL) was boiled under reflux (45 min) and then washed with saturated, aqueous Na₂CO₃, dried (Na₂SO₄), and concentrated. The residue was purified by column chromatography (ethyl acetate-hexane, 1:4), to give syrupy 11 (1.05 g, 42%), $[\alpha]_D^{20}$ –157° (c 1, chloroform), R_F 0.42 (ethyl acetate-hexane, 1:2); λ_{max}^{EtOH} 265 nm (ϵ 26,000); ν_{max}^{fiim} 2990–2940, 2865, 2845, 1745, 1740, 1670, 1630, 1600, 1450, 1420, 1380, 1360, 1260, 1210, 1180, 1150, 1130, 1100, 1050, 1030, 1000, 960, 920, 880, 810, 785, 750, and 690 cm⁻¹. ¹H-N.m.r. data (60 MHz, CCl₄): δ 4.70 (m, 1 H, H-8), 4.40 (d, 1 H, J 5 Hz, H-4), 3.76 (m, 2 H, H-7,7'), 3.10 (dq, 1 H, J 15 and 2 Hz, H-3'endo), 2.70 (dq, 1 H, J 15 and 2 Hz, H-3'exo), 2.00 (s, 6 H, MeCO and Me-C=), 1.32, and 1.20 (2 s, 6 H, Me₂C).

Anal. Calc. for C₁₃H₁₈O₅: C, 61.41; H, 7.13. Found: C, 61.32; H, 7.01.

The 2,4-dinitrophenylhydrazone of 11 had m.p. 182° (from ethanol).

Anal. Calc. for $C_{19}H_{22}N_4O_8$: C, 52.54; H, 5.10; N, 12.89. Found: C, 52.54; H, 4.80; N, 12.88.

To a solution of 11 (100 mg) in 1,4-dioxane (5 mL) were added aqueous 10% sodium hydroxide (1 mL) and then a solution of iodine (1 g) and potassium iodide (2 g) in water (8 mL), dropwise, until the red colour persisted after 10 min at 60°. Aqueous 10% sodium hydroxide was then added to eliminate the excess of iodine. After removal of iodoform, the filtrate was neutralised with 10% hydrochloric acid and extracted with ether (2 \times 10 mL). The combined extracts were washed with aqueous Na₂SO₃, dried, and concentrated, to give 12 (30 mg, 30%) (see above).

(4R,5S)-5-(2,3-O-Isopropylidene-α-D-erythrofuranosyl)-4-methoxycarbonyl-3-methylcyclohex-2-en-1-one (15). — A mixture of 1 (1 g, 5.78 mmol), methyl acetoacetate (1.35 g, 11.6 mmol), and piperidine (0.2 mL) was kept at room temperature for at least 4 weeks. Crystallisation occurred slowly. The product was purified by column chromatography (ethyl acetate-hexane, 1:3) on silica gel (100 g), to give **15** as white needles (1.6 g, 89.3%), m.p. 178-180° (from ethanol), $[\alpha]_D^{20}$ +184° (c 0.1, chloroform), R_F 0.2 (ethyl acetate-hexane, 1:1); λ_{max}^{EtOH} 235 nm (ε 17,800); ν_{max}^{KBT} 2980, 2955, 2920, 2845, 1725, 1710, 1440, 1380, 1265, 1210, 1170, 1100, 1080, 1060, 1045, 1015, 995, 945, 900, and 855 cm⁻¹. N.m.r. data: ¹H (200 MHz, CDCl₃), δ 5.905 (bs, 1 H, H-2), 4.688 (dd, 1 H, J 6.25 and 3.5 Hz, H-3'), 4.534 (dd, 1 H, J 6.25 and 3.5 Hz, H-2'), 3.942 (d, 1 H, J 10.5 Hz, H-4'endo), 3.652 (s, 3 H, MeO), 3.447 (bd, 1 H, J 4.0 Hz, H-4), 3.373 (dd, 1 H, J 10.5 and 3.5 Hz, H-4'exo), 3.340 (m, 1 H, H-1'), 2.727 (m, 1 H, H-5), 2.717 (m, 1 H, J 11.5 Hz, H-6 cis to COOMe), 2.433 (m, 1 H, J 11.5 Hz, H-6 trans to COOMe), 1.976 (d, 3 H, J 1 Hz, Me-C=). 1.361 (s, 3 H, endo-Me of CMe₂), and 1.228 (s, 3 H, exo-Me of CMe₂); ¹³C (CDCl₃), δ 197.7 (C-1), 170.8 (C=O of ester), 156.7 (C-3), 128.4 (C-2), 112.2 (O-C-O), 82.8, 80.9, 80.0, 73.0 (C-2',3',1',4'), 52.1 (OMe), 47.4 (C-4), 36.5 (C-5), 34.9 (C-6), 26.0, 24.7 (Me of Me₂C), and 23.5 (Me on C-3). Mass spectrum: m/z 310 (M⁺), 295 (M⁺ – Me), 279 (M⁺ – OMe), and 252 (M⁺ – CO₂Me + H).

Anal. Calc. for C₁₆H₂₂O₆: C, 61.98; H, 7.15. Found: C, 61.80; H, 7.35.

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REFERENCES

- F. J. LOPEZ APARICIO, M. YRUELA ANTIÑOLO, AND F. GARCIA GONZALEZ, An. R. Soc. Esp. Fis. Quim., Ser. B, 54 (1953) 705-714; H. ZINNER, E. WITTEMBURG, AND G. REMBARZ, Chem. Ber., 92 (1959) 1614-1617; N. K. KOCHETKOV AND B. I. DMITRIEV, Chem. Ind. (London), (1962) 2147-2148; Izv. Akad. Nauk SSSR, Ser. Khim., (1962) 1262-1263; F. MICHEEL AND W. MOELLER, Justus Liebigs Ann. Chem., 670 (1963) 63-68; F. ALONSO CERMEÑO, A. M. GONZALEZ NOGAL, AND F. J. LOPEZ APARICIO, An. R. Soc. Esp. Fis. Quim., Ser. B, 68 (1972) 285-292; F. J. LOPEZ APARICIO, M. GOMEZ GUILLEN, AND I. IZQUIERDO CUBERO, *ibid.*, 72 (1976) 938-945; 73 (1977) 1168-1176.
- 2 F. J. LOPEZ APARICIO, F. J. LOPEZ HERRERA, AND J. SANCHEZ BALLESTEROS, Carbohydr. Res., 69 (1979) 55-70; F. J. LOPEZ APARICIO AND F. J. LOPEZ HERRERA, An. R. Soc. Esp. Fis. Quim., Ser. B, 72 (1976) 931-937; F. J. LOPEZ APARICIO, I. IZQUIERDO CUBERO, AND M. D. PORTAL OLEA, Carbohydr. Res., 115 (1983) 250-253.
- 3 F. J. LOPEZ HERRERA, Tetrahedron Lett., (1980) 4963-4966.
- 4 A. B. FOSTER AND W. G. OVEREND, J. Chem. Soc., (1951) 680–684; F. J. LOPEZ HERRERA, C. GOMEZ PEREZ, AND M. VALPUESTA FERNANDEZ, An. Quim., Ser. C, in press.
- 5 H. OHRUI AND S. EMOTO, J. Org. Chem., 42 (1977) 1951–1957; F. J. LOPEZ HERRERA AND C. URAGA BAELO, submitted to Carbohydr. Res.
- 6 H. OHRUI, G. H. JONES, J. G. MOFFATT, M. L. MADDOX, A. T. CHRISTENSEN, AND S. K. BYRAM, J. Am. Chem. Soc., 97 (1975) 4602–4613.
- 7 G. JONES, Org. React, 15 (1967) 249-254.