Chemical Synthesis and Biological Evaluation of Second-Generation Palmerolide A Analogues

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In this communication, second-generation analogues of palmerolide A, a recently reported cytotoxic agent against melanoma cancer cells, were rationally designed, synthesized, and biologically evaluated. Structural variations of the enamide side chain and the C1–C8 segment of palmerolide A revealed a narrow structure–activity relationship window of the newly synthesized compounds. In addition, mechanistic and pharmacological studies were performed to assess the therapeutic potential of palmerolide A.

Introduction

Palmerolide A (1) is a cytotoxic marine macrolide recently isolated from the circumpolar tunicate Synoicum adareanum around Anvers Island on the Antarctic Peninsula.^[1] Reported by the Baker team in April 2006, it was found to exhibit potent and selective growth inhibitory properties against melanoma cancer cells UACC-62 (LC₅₀ = 18 nM), and its mechanism of action correlates well with other enamide containing V-ATPase inhibitors.^[1] Synthetic efforts in the palmerolide A (1) arena have culminated its total syntheses,^[2] and approaches toward several key synthetic intermediates.^[3] Furthermore, structure-activity relationship (SAR) studies^[4] led to the identification of benzamide derivative 2 and deoxygenated analogue 3 with an eightfold increase in potency and comparable potency, respectively, relative to parent natural product 1. Continuing our chemical-biology investigations in this area, here we report the synthesis and biological evaluation of palmerolide E (10),^[5] and second-generation analogues including side-chain analogues 19-24 and C1-C8 analogues 49a-d, against a selected panel of cancer and normal cell lines. Mechanistic and pharmacological studies of 1 and benzamide 2 were also carried out to assess the therapeutic potential of the palmerolide family of antiproliferative agents.

Results and Discussion

The synthetic strategy towards the designed analogues is in accordance to our previously described technology in the total synthesis of palmerolide A (1),^[2b,2c] its stereochemical congeners and analogues,^[4] as outlined in Figure 1. In view of the possible role of the enamide moiety involved in the reversible inhibition of the V-ATPase,^[6] palmerolide E (10) and α , β -unsaturated methyl ester 11 could serve as negative controls in the biological studies against the enamide-containing compounds. As shown in Scheme 1, the preparation of 10 and α , β -unsaturated methyl ester 11 began with removal of the two MOM ether moieties from aldehyde 8^[2b,2c] (HCl, 52% yield) to afford dihydroxy aldehyde 9, which underwent smooth ring-closing metathesis^[7] in the presence of Grubbs II catalyst to furnish 10 in 62% yield. Treatment of 10 with Ph₃P=CHCO₂Me smoothly delivered enoate 11 in 55% yield as a single geometric isomer.

On the basis of our previous studies,^[4] the biologically inconsequential C7 hydroxy substituent of the palmerolide A core structure prompted us to employ structurally simplified 7-deoxyvinyl iodide $12^{[4]}$ to pursue further SAR investigations concerning the palmerolide enamide side chain. At the same time, we previously concluded that a lipophilic enamide side chain was necessary for retaining the potent biological activity, that is, benzamide **2**. Therefore, under our carefully optimized conditions, application of the Buchwald copper-catalyzed coupling (CuI, K₂CO₃ and *N*,*N'*-dimethylethylenediamine)^[8] engaging macrocyclic iodide **12** and primary amides **13–18** afforded palmerolide A analogues **19–24** in moderate to average yields (26– 51%) as summarized in Table 1.

Next, we returned our attention to the SAR exploration in the C1–C8 domain of palmerolide A. These second-generation analogues were designed to probe (i) the importance of the C2–C3 olefin in the α , β -unsaturated macrolactone (i.e., **49a**), (ii) the possibility of replacing the metabolically labile aliphatic side chain with a conformationally rigid aromatic moiety (i.e., **49b**), (iii) the effect of extending the C₁–

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Figure 1. Structure of palmerolide A (1), benzamide (2), and 7-deoxypalmerolide A (3), and retrosynthetic analysis of palmerolide structure 1a leading to building blocks 5, 6, 7a, 7b, 7c, and 7d. TBS = *tert*-butyldimethylsilyl.



Scheme 1. Synthesis of palmerolide E (10) and α , β -unsaturated methyl ester analogue 11. Reagents and conditions: (a) HCl (3.0 N aq., 20.0 equiv.), EtOH, 23 \rightarrow 80 °C, 50 min, 52%; (b) Grubbs II cat. (0.05 equiv.), CH₂Cl₂, 23 °C, 1 h, 62%; (c) Ph₃P=CHCO₂Me (2.5 equiv.), CH₂Cl₂, 23 °C, 2 h, 55%. MOM = methoxymethyl.

C8 aliphatic backbone (i.e., **49c**), and (iv) the effect of the sterically demanding C5 *gem*-dimethyl substituents (i.e., **49d**). In accordance to our generalized retrosynthetic blueprint for the preparation of the palmerolide family of compounds (Figure 1), the synthesis of the required building blocks and their unifications leading to analogues **49a**–**d** are outlined in Schemes 2–4. In our previously reported syntheses of palmerolide structures,^[2b,2c] the C10 hydroxy group was protected as its MOM ether throughout the synthetic sequence and removed prior to the macrocyclic ring closure

Table 1. Preparation of palmerolide A analogues $19\mathchar`-24$ through copper-mediated coupling reactions. $^{[a]}$



[a] Reagents and conditions: **13–18** (2.0 equiv.), CuI (1.5 equiv.), K_2CO_3 (5.0 equiv.), N,N'-dimethylethylenediamine (3.0 equiv.), DMF, 23 °C, 1 h. DMF = N,N'-dimethylformamide.

under the olefin metathesis conditions. The MOM ether could be removed at a later stage under mild acid conditions (HCl/MeOH or TMSCl/MeOH); however, this transformation was routinely complicated with the formation of the C10-C11 cyclic carbonate as a byproduct.^[2b,2c] Therefore, we thought to circumvent this late-stage counterproductive event by exchanging the MOM ether to a TBS ether early in the synthetic sequence (Scheme 2). In this instance, starting from MOM ether 25,^[2b,2c] a two-step protecting group interconversion (CBr₄; TBSCl) smoothly delivered TBS ether 27 in 58% yield over the two steps. Further elaboration of the alkyne terminus in 27 to key building block vinyl stannane 6 was carried out by following our previously reported palladium-catalyzed hydrostannation procedure,^[9] through intermediate bromide 28, in 69% overall yield as a single geometric isomer. Stille^[10] coupling between vinyl stannane 6 and vinyl iodide 5^[2b,2c] afforded tetraene 29 in 79% yield, which serves as a common intermediate for all ensuing synthetic investigations concerning the variation of the C1-C8 segment of palmerolide A.



Scheme 2. Synthesis of tetraene **29**. Reagents and conditions: (a) CBr₄ (0.1 equiv.), *i*PrOH, 23 \rightarrow 80 °C, 2 h, 62%; (b) TBSCI (1.3 equiv.), imidazole (2.5 equiv.), 4-DMAP (cat.), DMF, 23 °C, 5 h, 94%; (c) NBS (1.1 equiv.), AgNO₃ (0.1 equiv.), acetone, 23 °C, 1 h, 98%; (d) Pd(dba)₂ (0.05 equiv.), PPh₃ (0.22 equiv.), *n*Bu₃SnH (2.2 equiv.), THF, 23 °C, 1 h, 70%; (e) Pd(PPh₃)₄ (0.1 equiv.), CuCl (5.0 equiv.), LiCl (6.0 equiv.), DMSO, 23 °C, 8 h, 79%. 4-DMAP = *N*,*N'*-dimethylaminopyridine, NBS = *N*-bromosuccinimide; dba = dibenzylideneacetone.

Variation of the C1-C8 segment of palmerolide A also called for the preparation of carboxylic acids 7a-d, and their syntheses are shown in Scheme 3. Acid fragment 7a was prepared through conjugate reduction of enoate 30 (NaBH₄, CuCl), followed by saponification of resulting ethyl ester 31 (KOH, 75% yield over the two steps). Aromatic acid 7b was synthesized from phenolic benzaldehyde 32 through intermediate triflate 33 (Tf₂O, 46% yield), followed by palladium-mediated cross-coupling with allyltributyltin (34, 99% yield), homologation to enoate 35 (35% vield), and saponification (LiOH, 95% vield). Two-carbon homologated acid 7c was synthesized through sequential oxidation of alcohol 36 (PCC; then NaClO₂) in 64% yield over the two steps. Finally, preparation of gem-dimethyl acid 7d began with conjugate addition of 3-butenylmagnesium bromide to α,β -unsaturated diester 38 followed by thermal decarboxylation to give ethyl ester 40 in 92% yield over the two steps. Two-carbon homologation of ethyl ester 40 involved a reduction/oxidation/Wittig olefination sequence (DIBAL-H, DMP, Ph₃P=CHCO₂Me), through the intermediacy of alcohol 41, to furnish α , β -unsaturated methyl ester 42 (70% overall yield). Finally, saponification of methyl ester 42 (LiOH) afforded desired acid 7d in 95% vield.

With building blocks **29** and **7a–d** in hand, the next task became their unification and further elaboration to targeted analogues **49a–d**, as summarized in Scheme 4. Thus, Yama-guchi esterification^[11] (2,4,6-trichlorobenzoyl chloride, Et₃N, 4-DMAP) between alcohol **29** and acid fragments **7a–d** afforded esters **43a** (69%), **43b** (70%), **43c** (72%), and **43d** (60%), respectively. Selective removal of the primary TBS ether in **43a–d** under the mild action of TFA, followed by oxidation (DMP) of resulting alcohols **44a–d** gave corre-



Scheme 3. Synthesis of carboxylic acids 7a-d. Reagents and conditions: (a) NaBH₄ (10.0 equiv.), CuCl (0.75 equiv.), MeOH/THF (4:7), 0 °C, 6 h, 91%; (b) KOH (5.0 equiv.), dioxane/ H_2O , (4:1), 23 °C, 12 h, 82%; (c) Tf₂O (1.2 equiv.), pyridine (2.0 equiv.), CH₂Cl₂, $0 \rightarrow 23$ °C, 3 h, 46%; (d) Pd(PPh₃)₄ (0.1 equiv.), LiCl (3.0 equiv.), allyltributyltin (1.1 equiv.), 1,4-dioxane, 100 °C, 16 h, 99%; (e) trimethyl phosphonoacetate (1.2 equiv.), LiBr (4.0 equiv.), Et₃N (2.0 equiv.), THF, 23 °C, 16 h, 35%; (f) LiOH (3.0 equiv.), THF/MeOH/H₂O (2:1:1), 23 °C, 4 h, 95%; (g) PCC (1.6 equiv.), CH2Cl2, 23 °C, 16 h, 88 %; (h) NaClO2 (3.0 equiv.), 2-methyl-2-butene (10 equiv.), NaH₂PO₄ (5.0 equiv.), tBuOH/H₂O (1:1), 0 °C, 2 h, 73%; (i) 3-butenylmagnesium bromide (2.0 equiv.), CuCl $(0.05 \text{ equiv.}), \text{ THF}, -30 \rightarrow 23 \text{ °C}, 1.5 \text{ h}, 97\%; (j) \text{ LiCl} (2.0 \text{ equiv.}),$ H₂O (1.0 equiv.), DMSO, reflux, 24 h, 95%; (k) DIBAL-H (1.4 M in CH2Cl2, 2.5 equiv.), CH2Cl2, -78 °C, 1.5 h, 90%; (l) DMP, (1.1 equiv.), NaHCO₃ (5.0 equiv.), CH₂Cl₂ 23 °C, 0.5 h; (m) trimethyl phosphonoacetate, (1.1 equiv.), LiBr (4.0 equiv.), Et₃N (2.0 equiv.), 23 °C, 12 h, 78 % over two steps; (n) LiOH (3.0 equiv.), THF/MeOH/H₂O (2:1:1), 23 °C, 4 h, 95%. PCC = pyridinium chlorochromate, Tf = trifluoromethanesulfonyl.

sponding aldehydes **45a** (77%), **45b** (68%), **45c** (72%), and **45d** (50%). Further elaboration of aldehydes **45a–d** to macrocyclic ring-closing metathesis precursors **47a–d** involved Takai olefination^[12] (CrCl₂, CHI₃; **46a–d**) and TBS ether removal (HCl aq.; **47a**: 76%, **47b**: 40%, **47c**: 42%, and **47d**: 44% yield over the two steps). With allylic alcohols **47a–d** in hand, their ring-closing-metatheses^[7] under the action of Grubbs II catalyst (CH₂Cl₂, 0.005 M,

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25 °C) proceeded smoothly to afford macrocycles **48a** (61%), **48b** (60%), **48c** (60%), and **48d** (60%), respectively. Finally, installation of the enamide moiety onto vinyl iodides **48a–d** through the established copper-catalyzed crosscoupling protocol^[8] provided C1–C8 palmerolide A analogues **49a** (56%), **49b** (28%), **49c** (26%), and **49d** (31%), respectively.



Scheme 4. Synthesis of palmerolide A analogues 49a-d employing acid fragments 7a-d, respectively. Reagents and conditions: (a) 2,4,6-trichlorobenzoyl chloride (1.1 equiv.), Et₃N (2.0 equiv.), 7a or 7b or 7c or 7d (1.2 equiv.), 4-DMAP (1.0 equiv.), toluene, 23 °C, 12 h, 43a: 69%; 43b: 70%; 43c: 72%; 43d: 60%; (b) TFA (10% aq., 1.0 equiv.), THF, 0 °C, 12 h, 44a: 91%; 44b: 90%; 44c: 90%; 44d: 65%; (c) Dess-Martin periodinane (1.1 equiv.), NaHCO₃ (5.0 equiv.), CH₂Cl₂, 23 °C, 20 min, 45a: 85%; 45b: 75%; 45c: 80%; 45d: 75%; (d) CrCl₂ (10.0 equiv.), CHI₃ (3.0 equiv.), THF/dioxane (1:6), 23 °C, 2 h, 46a: 87% (>95:5 *E*/*Z*); 46b: 55% (>95:5 *E*/*Z*); 46c: 60% (>95:5 E/Z); 46d: 60% (>95:5 E/Z); (e) HCl (3.0 N aq., 2.0 equiv.), EtOH, 23 °C, 12 h, 47a: 87%; 47b: 73%; 47c: 70%; 47d: 74%; (f) Grubbs II cat. (0.05 equiv.), CH₂Cl₂, 23 °C, 1 h, 48a: 61%, **48b**: 60%; **48c**: 60%; **48d**: 60%; (g) 3-methyl-2-butenamide (2.0 equiv.), CuI (1.5 equiv.), K₂CO₃ (6.0 equiv.), N,N'-dimethylethylenediamine (3.0 equiv.), DMF, 23 °C, 1 h, 49a: 56%; 49b: 28%; **49c**: 26%; **49d**: 31%. TFA = trifluoroacetic acid.

The synthesized compounds were tested against a panel of cancer cells, including breast (MCF-7), melanoma (UACC-62), CNS (SF268), lung (NCI-H460), ovarian (1A9), Taxol®-resistant ovarian (PTX22),^[13] epothilone-resistant ovarian (A8),^[14] and human melanocyte (HM2) cells using doxorubicin, Taxol[®], and natural palmerolide A (1) as standards; the results are summarized in Table 2. As expected and on the basis of the hypothesized reversible V-ATPase inhibition involving the enamide side chain,^[6] both palmerolide E (10) and methyl ester 11 showed complete lost of activity up to 10 µM against all cells tested. Sidechain analogues 19-24 revealed some interesting findings. Benzamide 19 exhibited the most potent activity, an observation similar to that previously reported for its C7 oxygenated relative (compound 2). In stark contrast, biphenyl 20 and 2,4-dimethoxy 23 showed significantly lower activities, whereas 4-fluoro 21, 4-trifluoromethyl 22, and 3,5-dimethoxy 24 displayed modest activities. This seems to suggest a very narrow SAR window concerning the steric environment of the aromatic side chain, even for 4-fluoro analogue 21. It is interesting to note that 2,4-dimethoxy and 3,5-dimethoxy analogues 23 and 24 exhibited drastically different cytotoxic activities, possibly attributed to the electron-donating groups at the *ortho* and *para* positions of analogue 23, which shift the equilibrium towards the protonated enamide rather than the enamide/V-ATPase complex.^[6] The biological activities of C1-C8 analogues 49a-d also revealed little SAR flexibility concerning this domain of the palmerolide A core structure. In this instance, whereas C2-C3 saturated analogue 49a displayed an approximately 4-10-fold reduction in activity across all cell lines examined, aromatic analogue 49b, two-carbon homologated analogue 49c, and gem-dimethyl analogue 49d all exhibited a dramatic reduction in their biological activities. Once again, this indicates that a subtle increase in the steric environment around the C1-C8 domain of palmerolide A cannot be tolerated at the active site of the enzyme. The therapeutic indexes of the palmerolide family of compounds were also determined by comparing their cytotoxicities against the melanoma cancer cell line (UACC-62) and normal human melanocyte (HM-2). Synthetic palmerolide A (1) and analogues 2, 3, 19, 21, and 24 all exhibited excellent selectivity towards UACC-62, with benzamide analogue 2 being the most potent and most selective compound prepared to date.

Finally, mechanistic and pharmacological investigations of palmerolide A (1) and benzamide analogue 2 were carried out to further assess the therapeutic potential of the palmerolide family of antitumor agents.^[15] In the Apo-BrdU TUNEL assay (to detect the 3'-hydroxy ends of fragmented DNA in apoptotic cells), palmerolide A (1) displayed cell killing effect resembling a late-apoptotic pathway. Furthermore, direct evidence in support of the V-AT-Pase inhibitory properties of palmerolide A (1) was obtained through LysoSensor studies (monitor pH level changes of the lysosomes and acidic organelles), in comparison with concanamycin, a known V-ATPase inhibitor. Palmerolide A (1) and benzamide analogue 2 were also found to inhibit CYP3A4 metabolic activity (IC₅₀ = 1.182

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Table 2.	Cytotoxicity	of natural an	nd synthetic	palmerolides	against	selected	cancer	and norma	l cell lines.
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Entry	Cell line/		$GI_{50}^{[a]}[\mu M]$							
2	Compound	UACC-62 ^[b]	MCF-7 ^[b]	SF268 ^[b]	NCI-H460 ^[b]	IA9[c]	PTX22 ^[c]	A8 ^[c]	HM-2 ^[d]	SI ^[e]
1	1	0.061	0.068	0.040	0.014	0.070	0.067	0.059	6.177	101.3
		± 0.011	± 0.006	±0.012	± 0.004	± 0.009	± 0.004	± 0.015	± 1.272	
2	2	0.009	0.008	0.007	0.007	0.008	0.029	0.007	2. 309	256.6
		± 0.000	± 0.001	± 0.001	± 0.001	± 0.001	± 0.011	± 0.001	±1.269	
3	3	0.076	0.153	0.069	0.069	0.087	0.194	0.091	14.006	184. 3
		± 0.010	± 0.087	± 0.004	± 0.004	± 0.006	± 0.078	± 0.004	± 4.589	
4	10	>10	>10	>10	>10	>10	>10	>10	>10	
5	11	>10	9.061	>10	>10	>10	>10	>10	>10	
			±0.174							
6	19	0.055	0.061	0.045	0.027	0.072	0.075	0.065	8.256	150.1
		± 0.011	± 0.004	± 0.008	± 0.017	± 0.004	± 0.003	± 0.003	± 2.222	
7	20	5.823	6.320	8.181	0.701	7.309	7.035	6.217	56.641	9.7
		±0.625	± 0.978	±0.134	± 0.028	±1.172	± 0.858	± 0.472	± 0.989	
8	21	0.227	0.339	0.282	0.069	0.646	0.631	0.486	27.186	119.8
		± 0.048	± 0.093	± 0.074	± 0.005	± 0.098	± 0.060	±0.122	±4.734	
9	22	0.882	0.864	0.936	0.582	3.004	2.232	0.785	23.701	26.9
		± 0.041	± 0.086	± 0.041	± 0.072	± 0.764	±0.491	± 0.029	±1.541	
10	23	9.026	5.452	>10	5.093	>10	9.206	8.317	42.201	4.7
		±0.423	± 0.659		± 0.018		± 0.240	±0.299	± 8.480	
11	24	0.753	0.855	0.789	0.517	0.869	0.863	0.710	>100	>132.8
		± 0.113	± 0.006	± 0.111	± 0.048	± 0.077	± 0.161	± 0.013		
12	49a	0.750	0.849	0.687	0.465	0.820	0.723	0.807	8.020	10.7
		±0.123	± 0.083	±0.121	± 0.086	± 0.066	± 0.056	± 0.075	±1.243	
13	49b	4.958	2.357	2.686	0.845	7.084	5.489	4.269	7.787	3.8
									± 0.889	
14	49c	6.857	5.708	6.793	5.41	8.102	6.66	5.626	8.619	1.2
		± 0.071							± 0.361	
15	49d	>10	>10	>10	>10	_	_	_	>10	_
16	Taxol	0.015	0.006	0.024	0.006	0.006	0.083	0.035	0.061	4.1
		± 0.007	± 0.000	± 0.008	± 0.001	± 0.000	± 0.007	± 0.016	± 0.010	
17	Doxorubicin	0.207	0.057	0.160	0.008	0.035	0.159	0.063	0.172	0.8
		±0.091	± 0.007	± 0.053	± 0.001	± 0.010	±0.051	± 0.013	± 0.083	

[a] Antiproliferative effects of tested compounds against human tumor cell lines and drug-resistant cell lines in a 48 h growth inhibition assay using the sulforhodamine B staining methods. Human cancer cell lines: melanoma (UACC-62); breast (MCF-7), CNS (SF268), lung (NCI-H460), ovarian (IA9) and its drug resistant mutant PTX22 (taxol-resistant) and A8 (epithilone resistant), normal melanocyte (HM-2). Growth inhibition of 50% (GI₅₀) is calculated as the drug concentration, which caused a 50% reduction in the net protein increase in control cells during the drug incubation. GI₅₀ values for each compound represent mean of 2–5 independent experiments \pm SE. [b] These cell lines were provided by the National Cancer Institute (NCI), Division of Cancer Treatment and Diagnosis (DCTD). [c] These cell lines were provided by Professor Paraskevi Giannakakou, Weill Medical College of Cornell University. [d] These cells were purchased from Cell Research Corporation (Singapore). [e] Selectivity index, SI (GI₅₀ HM-2/GI₅₀ UACC-62).

and 0.956 μ M, respectively) and verapamil-stimulated Pgp-ATPase activity (IC₅₀ = 1.220 and 1.384 μ M, respectively) and inhibited hERG channel at 3.5 μ M (with E4031 as the positive control).

Conclusions

In conclusion, the design and synthesis of second-generation palmerolide A analogues with structural variations in the enamide side chain and the C1–C8 aliphatic backbone were accomplished. The synthesized compounds were evaluated against a panel of cancer and normal cell lines, revealing a narrow SAR window concerning both of these structural domains. The beneficial effect of the benzamide group was once again confirmed through C7 deoxygenated analogue **19**. The most active members of the palmerolide analogues also exhibited promising therapeutic indexes when compared between cancerous (UACC-62) and normal (HM-2) cell lines. In conjunction with the preliminary mechanistic and pharmacological findings of palmerolide A (1) and benzamide 2, in vivo biological evaluations of selected candidate(s) will be carried out, and the results of these studies will be reported in due course.

Supporting Information (see footnote on the first page of this article): Experimental section; details of the cytotoxicity assay, APO-BrdU TUNEL assay, pH-lysosensor assay, verapamil-stimulated Pgp-ATPase assay, Vivid[®]CYP3A4 assay, and PredictorTM hERG fluorescence polarization assay; ¹H and ¹³C NMR spectra of compounds.

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- [15] For methods and results of the mechanistic and pharmacological studies of palmerolide A (1) and benzamide analogue 2, see the Supporting Information.

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