Preliminary communication

Synthesis of a branched D-glucoheptaose: the repeating unit of extracellular α -D-glucan 1355-S of Leuconostoc mesenteroides NRRL B-1355*

TOMOYA OGAWA** and TOSHIAKI KABURAGI

The Institute of Physical and Chemical Research, Wako-shi, Saitama, 351 (Japan) (Received September 16th, 1982; accepted for publication, September 28th, 1982)

In 1980, Misaki *et al.*² proposed for α -D-glucan 1355-S of *Leuconostoc* mesenteroides B-1355 the ramified structure 1, which involves an alternating arrangement of $(1 \rightarrow 6)$ - and $(1 \rightarrow 3)$ - α -D-glucopyranosyl linkages.



1 ($G = \alpha$ -D-glucopyranosyl residue)

As part of a project on the synthesis of branched D-glucans of biological interest, we describe here a stereoselective synthesis of D-glucoheptaose 2, the repeating unit of 1. The key intermediates for the convergent synthesis of 2 were designed to be the partially protected D-glucotriose 3 as the D-glucosyl acceptor, and the specifically protected, Dglucobiosyl chloride 4 as the α -D-glucosyl donor.

Synthesis of 4 was achieved as follows, starting from allyl α -D-glucopyranoside³; benzylidenation thereof with⁴ benzaldehyde and ZnCl₂ afforded an 80% yield of the 4,6-benzylidene acetal 5, $[\alpha]_D$ *** +112.3°. Monobenzylation of 5 by the stannylidene method⁵ gave a 74% yield of the 2-benzyl ether 6, m.p. 75–76°, $[\alpha]_D$ +145.1°, along with a 9.2% yield of the 3-benzyl ether, m.p. 143.5–144.5°, $[\alpha]_D$ +95.1°. Reductive cleavage of the benzylidene group of 6 with⁶ LiAlH₄–AlCl₃ gave a 71% yield of the dibenzyl ether 7, m.p. 57–58°, $[\alpha]_D$ +126.1°. Selective monoacetylation of 7 with AcCl–pyridine–CH₂Cl₂ afforded an 80% yield of acetate 8, $[\alpha]_D$ +107.1°. D-Glucosylation of 8 with 9 in the pre-

^{*}Glucan Synthesis, Part III. For Part II, see ref. 1.

^{**}To whom enquiries should be addressed.

^{***}Values of $[\alpha]_D$ were measured for CDCl₃ solutions at 25°, unless noted otherwise. Compounds having $[\alpha]_D$ recorded gave satisfactory data for elemental analyses.



sence of ⁷ AgOSO₂CF₃—Me₂NCONMe₂ afforded a 78% yield of a mixture of 10 {[α]_D +71.5°; δ_{C} (CDCl₃): 97.44 (¹J_{CH} 169.9 Hz, C-1b) and 95.06 (¹J_{CH} 167.0 Hz, C-1a)} and its β anomer {[α]_D +42.2°; δ_{C} (CDCl₃): 102.51 (¹J_{CH} 164.6 Hz, C-1b) and 95.10 (¹J_{CH} 168.9 Hz, C-1a)}, in the ratio of 5:2. Deallylation of 10 in the presence of PdCl₂ in MeOH gave hemiacetal 11, which was transformed into chloride 4 in 88% yield by treatment⁸ with SOCl₂-HCONMe₂.



For the synthesis of the D-glycosyl acceptor 3, a highly stereoselective introduction¹ of two α -D-glucopyranosyl groups, at O-3 and O-6, of 12 is required. In this respect, use of the D-glucopyranosyl chloride⁹ 13 carrying a 6-O-acetyl group was expected to be efficient¹⁰. Thus, D-glucosylation of 12 with 13 in the presence of AgOSO₂CF₃ and molecular sieves 4A gave 14 {[α]_D +82.2°; δ_C (CDCl₃): 97.34 (¹J_{CH} 170.9 Hz, C-1b, C-1c) and 94.33 (¹J_{CH} 166.0 Hz, C-1a)}; and deacetylation of 14 with MeOH–MeONa afforded a 41% yield of 3, [α]_D +87.7°; R_F 0.39 in 2:1 toluene--EtOAc.

Finally, the highly stereoselective glycosylation of 3 with 4 (containing a 6-O-acetyl group) was performed at -20° in the presence of AgOSO₂CF₃ and molecular sieves 4A in CH₂Cl₂, to give the completely protected D-glucoheptaose 15, which was deacetylated to give a 36% yield of 16, $[\alpha]_{D}$ +93.0°, R_{F} 0.43 in 3:1 toluene-EtOAc. Catalytic hydrogenolysis of 16 in the presence of 10% Pd-C in MeOH-THF afforded the desired D-glucoheptaose 2, $[\alpha]_{D}$ +196.0° (c 0.25, H₂O); R_{F} 0.21 in 3:3:2 iPrOH-EtOH-H₂O; δ_{H} (D₂O): 5.334 (bs, 3 H, H-1c,1f,1g), 4.976 (bs, 3 H. H-1b,1d,1e), 5.266 (d, J 4 Hz, H-1a α), and 4.683 (d, J 8.4 Hz, H-1a β); δ_{C} (D₂O): 99.50 (¹J_{CH} 171.9 Hz, C-1c,1f,1g), 98.25 (¹J_{CH} 170.9 Hz, C-1b,1d,1e), 96.43 (C-1a β), and 92.57 (C-1a α). The ¹³C-n.m.r. data



for synthetic 2 were in good agreement with those of the natural D-glucan reported by Seymour and Knapp¹¹, thus supporting the proposed structure 1.

In conclusion, the target molecule, namely, D-glucoheptaose (2), was synthesized in a stereocontrolled way by employing the D-glucobiosyl chloride 4 and the D-glucotrioside 3 as the key intermediates.

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