### Accepted Manuscript

Title: Photorelease of La<sup>3+</sup> with turn-on fluorescent detection

Author: Yongchao Hao Xu Zhang Yi Chen

PII: DOI: Reference:	S1010-6030(14)00203-2 http://dx.doi.org/doi:10.1016/j.jphotochem.20 JPC 9671	14.05.007
To appear in:	Journal of Photochemistry and Photobiology	A: Chemistry
Received date: Revised date: Accepted date:	31-3-2014 5-5-2014 13-5-2014	

霐

Photochemistry

Photobiology

Please cite this article as: Y. Hao, X. Zhang, Y. Chen, Photorelease of La<sup>3+</sup> with turnon fluorescent detection, *Journal of Photochemistry and Photobiology A: Chemistry* (2014), http://dx.doi.org/10.1016/j.jphotochem.2014.05.007

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

# Photorelease of La<sup>3+</sup> with turn-on fluorescent detection

Yongchao Hao, Xu Zhang and Yi Chen\*

Key Laboratory of Photochemical Conversion and Optoelectronic Materials, Technical Institute of Physics and Chemistry, The Chinese Academy of Sciences, Beijing, 100190, China. Tel: +86 10 82543595; Fax: +86 10 62564049. E-mail: <u>vichen@mail.ipc.ac.cn</u>

Keywords: photorelease, La<sup>3+</sup>, fluorescent monitoring, photochemistry.

Abstract: Photorelease of  $La^{3+}$  with turn on fluorescent detection is described. Complex (1) is synthesized by treatment 2,2'-(((ethane-1,2-diylbis(oxy)bis(2,1-phenylene))bis(methanylylidene)) bis(azanylylidene))diphenol with  $LaCl_3 \cdot 7H_2O$  in 1:1 molar ratio. Upon irradiation with UV light, 1 undergoes  $La^{3+}$  release and converts to 1,2-bis(2-(benzoxazol-2-yl)phenoxy)ethane (2). It is found that 2 exhibits strong fluorescence emission and the turn-on fluorescence is detected during the photo-conversion of 1 to 2, which is beneficial for monitoring the photo-release of  $La^{3+}$ . Moreover, an excellent (R = 0.9963) linear correlation between the ratio of optical density (OD<sub>1</sub>/OD<sub>2</sub>) with irradiation time is obtained when 1 is photo-converted to 2, which is beneficial for controlling or regulating the quantity of  $La^{3+}$ -released.

#### 1. Introduction

The light-activated release provides a powerful tool for the rapid introduction of a variety of species to biological systems with spatial and temporal control, allowing the time-resolved study of the ensuing events [1-5]. Metal photo-release has currently attracted considerable attention

because of its potential since it will be a valuable tool for the delivery of biologically active metals, which are an essential cofactor in numerous enzymes [6-9], or of toxic metal ions to study the mechanisms of metal-ion trafficking and applications, such as chemotherapy, by means of inducing cell death by catalyzing DNA and RNA cleavage or by binding active sites of proteins competition with essential metals [10-13].

Lanthanum ( $La^{3+}$ ) plays an important role in biological systems [14,15]. Lanthanum is a highvalence rare-earth metal used as a substitute for calcium as a blocker [16,17] of several nonselective ion channels in different systems based on the blocking activity of lanthanum including connexin hemichannels, murin frontal cortex networks, tobacco BY-2 cells, or the outward K<sup>+</sup> channel [18-21]. Although lanthanides have been known for their diversity in biological effects [22-24], and the application of lanthanides in medicine has high potential [25-28], the mechanism of the biological effects of lanthanides is not clearly known [29-31], specially at molecular and genetic level. Lanthanides  $(Ln^{3+})$  including lanthanum  $(La^{3+})$  are exogenous ions, the concentration of  $Ln^{3+}$  has great influenced on their biological effects [32,33]. Therefore, it is required that the development of a simple and efficient way to delivery exogenous lanthanides  $(Ln^{3+})$  to biological systems. Photo-trigged release is a promising tool, and can fulfill to molecules/ions release with spatial, temporal and quantitative control. In this paper, we report a prototype for  $La^{3+}$  photo-release. Complex 1 is prepared by employing 2,2'-(((ethane-1,2-diylbis(oxy)bis(2,1-phenylene))bis(methanylylidene))bis(azanylylidene))diphenol as ligand (Scheme 1), it is found that upon irradiation the solution of 1 in DMSO, the bound La<sup>3+</sup> can be released and the ligand is converted to 1,2-bis(2-(benzoxazol-2-yl)phenoxy)ethane 2. It is also found that accompanying the process of photoconversion, turn on fluorescence is observed, which is beneficial to monitor the photorelease of  $La^{3+}$ .



**Scheme 1.** Outline of photorelease of La<sup>3+</sup> form complex **1.** 

#### 2. Experimental

#### 2.1. General

<sup>1</sup>H NMR spectra were recorded at 400 MHz with TMS as an internal reference and DMSO-d<sub>6</sub> as solvent. HRMS spectra were recorded with Q-Exactive MS spectrometer (Thermo Fisher). UV absorption spectra and fluorescence spectra were measured on an absorption spectrophotometer (Hitachi U-3010) and a fluorescence spectrophotometer (F-2500), respectively. IR spectra (KBr) were recorded on spectrophotometers Excalibur 3100 (Varian). All chemicals for synthesis were purchased from commercial suppliers, and solvents were purified according to standard procedures. Reactions were monitored by TLC silica gel plate (60F-254). Column chromatography was performed on silica gel (70-230 mesh). A UV lamp (36 watt) was used as light sources for photo-release (irradiation energy: 4.3 mW/cm<sup>2</sup>).

2.2. Materials.

#### 2.2.1. Synthesis of complex 1.

The complex **1** was synthesized by reaction of lanthanum (III) salt and the ligand in amount equal to metal: ligand molar ratio of 1:1. The synthetic route for **1** was presented in Scheme 2, and the detailed procedures were as follows. (a) To the solution of salicylaldehyde (6.1 g, 50

mmol) and 1,2-dibromoethane (4.7 g, 25 mmol) in acetonitrile (150 ml) was added potassium carbonate (6.9 g, 50 mmol) and potassium iodide (0.4 g, 2.5 mmol). The solution mixture was refluxed. When no starting material (salicylaldehyde) was detectable by TLC plate, the solution was concentrated and the crude product was added to water (100 ml) and the solution mixture was extracted with dichloromethane (50 ml  $\times$  3). The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, the resulting solution was concentrated and the crude product was purified by flash column chromatography (petroleum ether / ethyl acetate, 2:1) to afford intermed product dialdehyde in 80% yield. (b) A solution of dialdehyde (0.54 g, 2 mmol) in absolute ethanol (20 ml) was heated to dissolved completely, to the solution was added a solution of 2-aminophenol (436 mg, 4 mmol) in absolute ethanol (10 ml), the solution mixture was reflexed till the dialdehyde was disappeared. The crude product was purified by recrystallization from ethanol to afford ligand in 80% yiled. <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>, ppm): 7.47 (d, J = 7.7 Hz, 2H), 7.29 (t, J = 7.5 Hz, 2H), 7.10 (d, J = 7.9 Hz, 2H), 6.97 (t, J = 7.5 Hz, 2H), 6.95-6.82 (m, 4H), 6.80 (s, 3.25 Hz)2H), 6.78-6.68 (m, 2H), 6.62-6.51 (m, 4H), 4.51 (d, J = 3.2 Hz, 4H). HRMS (EI) Calcd for  $C_{28}H_{24}N_2O_4$ , (M<sup>+</sup>): 452.1736. Found: 452.1741. (c) To the boiling solution of ligand (452 mg, 1.0 mmol) in ethanol (100 ml) was added dropwise a solution of lanthanum (III) chloride heptahydrate (371 mg, 1.0 mmol) in ethanol (10 ml). The mixture solution was refluxed for 1 hr and then cooled down to ambient temperature. The yellow crystalline was filtered off, washed with ethanol (10 ml  $\times$  3), and pure complex 1 was obtained after vacuum-dried without further purification. Yield: 96%. M.p. > 300 °C. IR (v.  $cm^{-1}$ ): 3570.14, 3556.87, 3410.64 (br), 1645.21, 1601.90, 1573.20, 1286.23, 1182.07. <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>, ppm): 8.74 (s, 2H), 7.90-

7.78 (m, 2H), 7.54 (t, J = 7.6 Hz, 2H), 7.29 (t, J = 7.8 Hz, 4H), 7.21 (dd, J = 7.5, 1.8 Hz, 2H),

6.95 (t, *J* = 7.5 Hz, 2H), 6.85-6.78 (m, 4H), 4.61 (s, 4H). HRMS (EI) calcd for C<sub>28</sub>H<sub>22</sub>ClLaN<sub>2</sub>O<sub>4</sub> 3H<sub>2</sub>O (M<sup>+</sup>+Na): 701.0547. Found: 701.4920.



Scheme 2. Synthetic route for complex 1.

#### 2.2.2 Synthesis of photo-product 2

The synthetic route for **2** was presented in Scheme 3, and the detailed procedure was as follows. A mixture of **1** (226 mg, 0.5 mmol) and lead acetate, Pb(OAc)<sub>4</sub> (244 mg, 0.55 mmol), dissolved in 20 ml of CHCl<sub>3</sub> was refluxed till no starting material was detected by TLC plate. The solution was filtered and the filtrate was extracted with CH<sub>2</sub>Cl<sub>2</sub>-H<sub>2</sub>O (v/v = 1:1, 10 ml × 3). The conbined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>. The resulting solution was concentrated and the crude product was purified by flash column chromatography (petroleum ether/CH<sub>2</sub>Cl<sub>2</sub>, 1:1) to afford **2**. Yield: 15%. <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>): 8.55 (dt, *J* = 8.0, 1.6 Hz, 2H), 7.72-7.70 (m, 2H), 7.50-7.31 (m, 10H), 7.11 (t, *J*<sub>1</sub> = 7.6 Hz, *J*<sub>2</sub> = 7.2 Hz, 2H), 4.62 (s, 4H). TOF-MS (EI) calcd for C<sub>28</sub>H<sub>20</sub>N<sub>2</sub>O<sub>2</sub>S<sub>2</sub> (M<sup>+</sup>): 480.0966. Found: 480.0991.



Scheme 3. Synthesis of 2.

#### 3. Results and discussion

### 3.1. Photorelease of $La^{3+}$

Photorelease of La<sup>3+</sup> from complex **1** was performed in DMSO solution. Upon irradiation with 254 nm light, the absorption of complex **1** at  $\lambda_{max} = 352$  nm ( $\varepsilon = 1.5 \times 10^4$  L mol<sup>-1</sup> cm<sup>-1</sup>, in DMSO) decreased with the increase of irradiation time, accompanying this process, a new band at  $\lambda_{max} = 320$  nm appeared (Fig. 1). The absorption of 320 nm attributed to the photo-product **2**, whose structure is suggested in Scheme 1. Further study found that the photorelease of La<sup>3+</sup> from complex **1** could also be performed in other organic solvents such as ethanol (EtOH) and N,N-dimethylformamide (DMF), the obtained results were similar to that in DMSO. It was worth noting that the photorelease of La<sup>3+</sup> could not performed in aqueous solution since **1** was insoluble in water. The investigation of **1** in mixture solution of H<sub>2</sub>O-DMSO found that the photorelease of La<sup>3+</sup> could be conducted in H<sub>2</sub>O-DMSO (v/v = 5/95) mixture solution with H<sub>2</sub>O not more than 5%.



Fig. 1. Absorption changes of 1 (50 $\mu$ M, in DMSO) with UV light irradiation (irradiation wavelength:  $\lambda = 254$  nm, irradiation periods: 0, 2, 4, 6, 8, 10, 12, 14, 16, 18, 20 min).

The kinetics of photorelease of  $La^{3+}$  was investigated by determination the optical density (OD) changes of **1** with different irradiation time according to Fig. 1. The ratio of  $OD_{352 nm}$  (maximum absorption of complex **1**) to  $OD_{320 nm}$  (maximum absorption of photo-product **2**) as function of irradiation time was presented in Fig. 2. The linear correlation between the  $OD_{320nm}$  /  $OD_{352nm}$  with irradiation time is excellent (R = 0.9963) in the process of  $La^{3+}$ -photorelease till it was completed. Photorelease with ratiometric detection suggested that the amount of  $La^{3+}$  release could be manipulated via controlling irradiation time, which is beneficial for the regulation of  $La^{3+}$  release in terms of quantity, location and time.



Fig. 2. The plot of optical density changes of 1 (50 $\mu$ M, in DMSO) with different irradiation time (irradiation wavelength:  $\lambda = 254$  nm, irradiation periods: 0, 2, 4, 6, 8, 10, 12, 14, 16, 18, 20 min). OD<sub>320 nm</sub>: optical density at 320 nm, OD<sub>352nm</sub>: optical density at 352 nm.

### 3.2. Fluorescence monitoring of photorelease of $La^{3+}$

In term of our previously study [34,35], phenolic Schiff bases are photo-converted to benzoxazole derivatives accompanying metal-ion photo-release, and a strong fluorescence is detected at the same time. Complex **1** showed no fluorescence in DMSO solution using 352 nm or 320 nm as excitation wavelength. Upon irradiating the solution of **1** in DMSO, the turn on fluorescence ( $\lambda_{em} = 364$  nm), which attributed to the emission of photo-product **2**, was observed. As presented in Fig. 3, the fluorescence was increased with increase of irradiation time till the photorelease of La<sup>3+</sup> was completed, which provides a convenient method to monitor the process of photorelease of La<sup>3+</sup>.



Fig. 3. Fluorescence changes of 1 (50 $\mu$ M, in DMSO) with UV light irradiation (irradiation wavelength:  $\lambda = 254$  nm, irradiation periods: 0, 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22 min, excitation wavelength:  $\lambda_{ex} = 290$  nm).

To check the chemical structure of photo-product **2** and elucidate the fluorescence properties of photo-product **2**, pure **2** was prepared by treatment of the ligand with Pb(OAc)<sub>4</sub> in CHCl<sub>3</sub> (Scheme 3) (detailed procedure see experimental section). The TLC (thin layer chromatography) test showed that both photo-product **2** and pure **2** has the same value of  $R_f$  ( $R_f = 0.26$ , elute: ethyl acetate/petroleum = 1 : 2, v/v) at the TLC plate. Moreover, it was found that both absorption and emission profiles of **2** were similar to those of pure **2**, and absorption ( $\lambda_{max} = 320$  nm) and emission ( $\lambda_{em} = 364$  nm) maxima were at the same position (Fig.s 1 and 3 vs. Fig. 4). Obtained results confirmed that photo-product **2** has the same chemical structure as that of pure **2**. Further study found that **2** exhibited strong fluorescence in DMSO, and a large fluorescence quantum yield ( $\phi_f = 0.76$ ) was obtained using quinine sulphate as a standard.



Fig. 4. Absorption and fluorescence of 2 in DMSO solution (50  $\mu$ M, irradiation wavelength:  $\lambda_{ex} = 290$  nm).

#### 3.3. Possible mechanism for the photoreaction

A possible mechanism for the photoreaction consists of a two-step sequence involving the formation of benzoxazoline followed by oxidation and dehydration (Scheme 4) [36]. Preliminary investigation found that benzoxazoline formation is a prerequisite to convert a Schiff base into benzoxazoline by photon induction. Control experiments showed that oxygen in the air played a minor role in the conversion reaction. A similar conversion was obtained (detection by absorption spectroscopy) when the irradiation of 1 was carried out in an air saturated solution or in degassed solution, although the mechanism for the photoreaction in the absence of oxygen is not known. It is worth noting that complex 1 is stable at ambient temperature and no significant change (detection by absorption spectral) was detected when a solution of 1 was kept in the dark, which indicates that the conversion of 1 into 2 did not occur in the absence of light.



Scheme 4. A possible pathway for the photo-conversion of 1 into 2.

#### 3.4. Control experiments

The binding of **2** with  $La^{3+}$  was investigated. No significant absorption changes were detected upon addition of  $LaCl_3 7H_2O$  to a solution of **2** in DMSO, which indicated that no (or only weak) binding between **2** and  $La^{3+}$ . To confirm this suggestion, <sup>1</sup>H NMR studies of **2** with and without addition of  $La^{3+}$  were performed. It was found that <sup>1</sup>H NMR spectrum of **2** with addition of  $La^{3+}$  agreed quite well with that of **2** without addition of  $La^{3+}$ , except for the signal at  $\delta = 3.3$  ppm, which is attributed to the proton of H<sub>2</sub>O resulting from the addition of  $LaCl_3 7H_2O$ . As presented in Fig. 5, the signals of **2** with addition of  $La^{3+}$  (above), assigned to the aromatic protons in the range of 7.93 – 7.05 ppm, are exactly as same as those of **2** without addition of  $La^{3+}$  (below), no significant chemical shifts were detected. These results confirmed that there was no (or only a very weak) binding between **2** and  $La^{3+}$ .



**Fig. 5**. Partial <sup>1</sup>H NMR spectra of **2** with (a) and without (b) addition of LaCl<sub>3</sub>  $7H_2O$  (solvent: DMSO-d<sub>6</sub>).

Further investigation found that the fluorescence of **2** was not affected by  $La^{3+}$ . As presented in Fig. 6, the emission spectral of **2** (both profile and strength) with addition of  $La^{3+}$  was exactly as same as that of **2** without addition of  $La^{3+}$ , no significant fluorescence changes were detected, which indicated that  $La^{3+}$  did not quench the fluorescence of **2**. In addition, control experiments showed that **2** is photostable, and no distinct changes in absorption or fluorescence was detected after the solution of **2** was irradiated for 60 min with 320 nm light. No significant binding between **2** and  $La^{3+}$  and no distinct fluorescence quench/decrease are beneficial for application of photorelease of  $La^{3+}$  with fluorescent monitoring.



Fig. 6. Fluorescence of 2 (50 $\mu$ M, in DMSO) with (red) and without (black) addition of LaCl<sub>3</sub> 7H<sub>2</sub>O (100 $\mu$ M). Irradiation wavelength:  $\lambda_{ex} = 290$  nm.

### 4. Conclusions

A platform for La<sup>3+</sup>-photorelease with fluorescence monitoring has been developed. Although the system presented herein has some shortcomings including unable to be performed in aqueous solution, irradiation, excitation and detection in short wavelength region, the system has some advantages such as controlled photo-release and turn on fluorescence monitor. Controlled photorelease system is great beneficial for the drug delivery in terms of quantity, location and time as improved control maximizes therapeutic effect while minimizing side effects. From a practical viewpoint, it is desirable for the system to be used in buffer solution and with visible light irradiation, excitation and detection. Modifying molecular structure to meet the requirements, and exploring the applications in biological processing will be the subject of future studies.

#### Acknowledgments

This work was supported by the National Natural Science Foundation of China (No 21302197) and National Basic Research Program of China (2010CB934103).

#### References

[1] M. Rothlingshofer, K. Gorska, N. Winssinger, Nucleic acid-templated energy transfer leading to a photorelease reaction and its application to a system displaying a nonlinear response, Journal of the American Chemical Society 133 (2011) 18110-18113.

[2] C.A. Terry, M.-J. Fernandez, L. Gude, A. Lorente, K.B. Grant, Physiologically relevant concentrations of NaCl and KCl increase DNA photocleavage by an N-substituted 9-aminomethylanthracene dye, Biochemistry 50 (2011), 10375-10389.

[3] A. Biton, A. Ezra, J. Kasparkova, V. Brabec, E. Yavin, DNA photocleavage by DNA and DNA–LNA amino acid–dye conjugates, Bioconjugate Chemistry 21 (2010) 616-621.

[4] W.-Y. Yang, S. Roy, B. Phrathep, Z. Rengert, R. Kenworthy, D.A.R. Zorio, I.V. Alabugin, Engineering pH-gated transitions for selective and efficient double-strand DNA photocleavage in hypoxic tumors, Journal of Medicinal Chemistry 54 (2011) 8501-8516.

[5] L. Poon, W. Zandberg, D. Hsiao, Z. Erno, D. Sen, B.D. Gates, N.R. Branda, Photothermal release of single-stranded DNA from the surface of gold nanoparticles through controlled denaturating and Au–S bond breaking, ACS Nano 4 (2010) 6395-6403.

[6] K.L. Ciesienski, K.L. Hass, M.G. Dickens, Y.T. Tesema, K.J. Franz, A photolabile ligand for light-activated release of caged copper, Journal of the American Chemical Society 130 (2008) 12246-12247.

[7] G.C.R. Ellis-Davies, Caged compounds: photorelease technology for control of cellular chemistry and physiology, Nature Methods 4 (2007) 619-628.

[8] M.J. Clarke, F. Zhu, D.R. Frasca, Non-platinum chemotherapeutic metallopharmaceuticals, Chemical Reviews 99 (1999) 2511-2534.

[9] K. Ciesienski, K.J. Franz, Keys for unlocking photolabile metal-containing cages, Angewandte Chemie International Edition 50 (2011) 814-824.

[10] D. Mishra, A. Mehta, S.J.S. Flora, Reversal of arsenic-induced hepatic apoptosis with combined administration of DMSA and its analogues in guinea pigs: role of glutathione and linked enzymes, Chemical Research in Toxicology 21 (2008) 400-407.

[11] R.L. Bouffant, O. Mulner-Lorillon, J. Morales, P. Cornier, R. Belle, Chromium(III) triggers the DNA-damaged checkpoint of the cell cycle and induces a functional increase of 4E-BP, Chemical Research in Toxicology 21 (2008) 542-549.

[12] M. Vinken, L. Ceelen, T. Vanhaecke, V. Rogiers, Inhibition of gap junctional intercellular communication by toxic metals, Chemical Research in Toxicology 23 (2010) 1862-1867.

[13] L. Doan, B. Handa, N.A. Roberts, K. Klumpp, Metal ion catalysis of RNA cleavage by the influenza virus endonuclease, Biochemistry 38 (1999) 5612-5619.

[14] C. Canavese, C. Mereu, M. Nordio, E. Sabbioni, S. Aime, Blast from the past: the aluminum's ghost on the lanthanum salts, Current Medicinal Chemistry 12 (2005) 1631–1636.

[15] L. Feng, H. Xiao, X. He, Z. Li, F. Li, N. Liu, Y. Zhao, Y. Huang, Z. Zhang, Z. Chai, Neurotoxicological consequence of long-term exposure to lanthanum, Toxicology Letters 165 (2006) 112–120.

[16] J. Hu, X. Jia, Q. Li, X. Yang, K. Wang, Binding of La<sup>3+</sup> to calmodulin and iIts effects on the interaction between calmodulin and calmodulin binding peptide, polistes mastoparan, Biochemistry 43 (2004) 2688-2698.

[17] Y. Ye, H.-W. Lee, W. Yang, S. Shealy, J.J. Yang, Probing site-specific calmodulin calcium and lanthanide affinity by grafting, Journal of the American Chemical Society 127 (2005) 3743-3750.

[18] R.J. Thompson, N. Zhou, B.A. MacVicar, Ischemia opens neuronal gap junction hemichannels, Science 312 (2006) 924–927.

[19] A. Gramowski, K. Jugelt, O.H. Schroder, D.G. Weiss, S. Mitzner, Acute functional neurotoxicity of lanthanum(III) in primary cortical networks, Toxicology Sciences 120 (2011) 173–183.

[20] C. Lachaud, D. Da Silva, V. Cotellea, P. Thuleau, T.C. Xiong, A. Jauneau, C. Briere, A. Graziana, Y. Bellec, J.-D. Faure, R. Ranjeva, C. Mazars, Nuclear calcium controls the apoptoticlike cell death induced by D-erythro-sphinganine in tobacco cells, Cell Calcium 47 (2010) 92–100.

[21] L.H. Wang, N. Jiang, B. Zhao, X.D. Li, T.H. Lu, X.L. Ding, X.H. Huang, Structural basis for the decrease in the outward potassium channel current induced by lanthanum, JBIC Journal of Biological Inorganic Chemistry 15 (2010) 989–993.

[22] C.H. Evans, Biochemistry of Lanthanides, Plenum Press, New York, 1990.

[23] J.S.W. Tsang, A.A. Neverov, R.S. Brown, La<sup>3+</sup>-catalyzed methanolysis of hydroxypropyl-*p*nitrophenyl phosphate as a model for the RNA transesterification reaction, Journal of the American Chemical Society 125 (2003) 1559-1566.

[24] J. Sabin, G. Prieto, P.V. Messina, J.M. Ruso, R. Hidalgo-Alvarez, F. Sarmiento, On the effect of  $Ca^{2+}$  and  $La^{3+}$  on the colloidal stability of liposomes, Langmuir 21 (2005) 10968-10975.

[25] K. Wang, R.C. Li, Y. Cheng, B. Zhu, Lanthanides—the future drugs? Coordination Chemical Reviews 190-192 (1999) 297-308.

[26] G. Zhao, F. Li, H. Li, H. Lin, Synthesis, characterization and biological activity of complexes of lanthanum(III) with 2-(1'-phenyl- 2'-carboxyl-3'-aza-*n*-butyl)-1,10-phenanthroline and 2-(1'-*p*-phenol-2'-carboxyl-3'-aza-*n*-butyl)-1,10-phenanthroline, Bioorganic Medicinal Chemistry 15 (2007) 533-540.

[27] I. Kostova, G. Momekov, T. Tzanova, M. Karaivanova, Synthesis, characterization, and cytotoxic activity of new lanthanum (III) complexes of biscoumarins, bioinorganic chemistry and applications 2006 (2006) 25651.

[28] G. Karthikeyan, K. Mohanraj, K.P. Elango, K. Girishkumar, Synthesis and spectral characterization of lanthanide complexes with sulfamethoxazole and their antibacterial activity, Russian Journal Coordination Chemistry 32 (2006) 380-385.

[29] S. Yu, J. Hu, X. Yang, K. Wang, Z.M. Qian, La<sup>3+</sup>-induced extracellular signal-regulated kinase (ERK) signaling via a metal-sensing mechanism linking proliferation and apoptosis in NIH 3T3 cells Biochemistry 45 (2006) 11217-11225.

[30] S.L. Shorte, J.G. Schofield, The effect of extracellular polyvalent cations on bovine anterior pituitary cells Evidence for a  $Ca^{2+}$ -sensing receptor coupled to release of intracellular calcium stores, Cell Calcium 19 (1996) 43-57.

[31] K. Matsumura, M. Omiyama, Hydrolysis of phosphatidylinositol by rare earth metal ion as a phospholipase C mimic, Journal of Inorganic Biochemistry 55 (1994) 153-156.

[32] Q. Zeng, J.G. Zhu, H.L. Cheng, Phytotoxicity of lanthanum in rice in haplic acrisols and cambisols, Ecotoxicology and Environmental Safety 64 (2006) 226-233.

[33] P. Shi, G.C. Chen, Z.W. Huang, Effects of  $La^{3+}$  on the active oxygen-scavenging enzyme activities in cucumber seedling leaves, Russian Journal of Plant Physiology 52 (2005) 294-297.

[34] X. Zhang, Y. Chen, Phototriggered metal-ion release from phenolic schiff bases: A system for metal-ion photodelivery, ChemPhysChem 10 (2009) 1993-1995.

[35] X. Zhang, Y. Chen, Photo-controlled Zn<sup>2+</sup> release system with dual binding-sites and turnon fluorescence, Physical Chemistry Chemical Physics 12 (2010) 1177-1181.

[36] E. Tauer, K.H. Grellmann, Photochemical and thermal reactions of aromatic Schiff bases, The Journal of Organic Chemistry, 46 (1981) 4252-4258.

Received when the second

Graphical Abstract



### Highlights

- (1) A platform for  $La^{3+}$ -photorelease with turn on fluorescence monitoring has been developed.
- (2) La-complex is synthesized by employing a Schiff base derivative as ligand.
- (3) La-complex undergoes photorelease of La<sup>3+</sup> and photo-converts to fluorescent benzoxazole derivative.

20 Page 20 of 20